



Diagnostic and Management Issues in Patients with Late-Onset Ornithine Transcarbamylase Deficiency

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Abstract: Ornithine transcarbamylase deficiency (OTCD) is the most common inherited disorder of the urea cycle and, in general, is transmitted as an X-linked recessive trait. Defects in the OTC gene cause an impairment in ureagenesis, resulting in hyperammonemia, which is a direct cause of brain damage and death. Patients with late-onset OTCD can develop symptoms from infancy to later childhood, adolescence or adulthood. Clinical manifestations of adults with OTCD vary in acuity. Clinical symptoms can be aggravated by metabolic stressors or the presence of a catabolic state, or due to increased demands upon the urea. A prompt diagnosis and relevant biochemical and genetic investigations allow the rapid introduction of the right treatment and prevent long-term complications and mortality. This narrative review outlines challenges in diagnosing and managing patients with late-onset OTCD.

Keywords: late-onset OTC deficiency; urea cycle disorders; clinical manifestations; adults

1. Introduction

The urea cycle is the physiological primary pathway for removing nitrogen, a toxic byproduct of amino acid metabolism. Consisting of five enzymes, one cofactor producer and two mitochondrial transport molecules in the mammalian liver, the urea cycle converts ammonia to urea for urinary excretion [1,2]. Urea cycle disorders (UCD) are monogenic disorders caused by decreased function in any of the eight components of this cycle. Common complications of UCD include the accumulation of ammonia, which is neurotoxic [3–7], and hepatic dysfunction. The most prevalent UCD is an ornithine transcarbamylase deficiency (OTCD) [8–11]. The anabolic OTC enzyme is responsible for the transfer of a carbamoyl group from carbamoyl phosphate to the amino group of L-ornithine, producing citrulline in an early step of the urea cycle [12,13].

OTCD was first described in 1962 by Russell regarding two girls at the age of 1 year and 8 months and 6 years [3]. The estimated prevalence is between 1:14,000 and 80,000 [14]. The X-linked inheritance of OTCD is unique with the remaining UCDs inherited in an autosomal recessive manner. Due to its X-linked recessive inheritance, OTCD tends to present earlier, and more severely, in males [11]. However, patient phenotype can vary due to random X inactivation in females and hypomorphic variants in OTC that cause a partial enzyme deficiency and later onset of symptoms in males [7,15]. While the other UCDs are



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). included on newborn screening panels, the diagnosis of OTCD is challenging. The early presentation of the classical OTCD makes newborn screening unhelpful in these patients. In addition, the late-onset patients may be very difficult to diagnose on a metabolite analysis alone. There are no common mutations as such and therefore full gene sequencing needs to be completed and include a promoter analysis.

Thus, we use age at the presentation and severity of the phenotype to classify patients into two categories, neonatal and late-onset. Current evidence estimates 30% of cases are neonatal onset and 70% are late-onset [16]. The neonatal presentation is severe [16], leading to coma and death [17]. It commonly presents during the first few days of life [18] and affects mainly hemizygous males with a complete enzyme deficiency [7,19]. This category is rare in females (7%) [20]. Neonates present with severe rapid-onset neurological manifestations, leading to death within the first few hours if untreated and within a few months despite medical management. Of the survivors, many develop severe neurological consequences [18].

Those with late-onset OTCD can develop symptoms anytime from infancy to later childhood, adolescence or adulthood [21]. Hemizygous males with a partial enzyme deficiency and around 20% of heterozygous females fall into this category. More than 80% of heterozygous females remain asymptomatic throughout life [4,8,9,22]. Clinical manifestations in this form of OTC are nonspecific and are mainly due to hyperammonemia. They include neurological (encephalopathy and migraine headaches), psychiatric (bipolar-disorder-like symptoms) or gastrointestinal symptoms (hepatic dysfunction and cyclic vomiting). All symptoms can present as a combination [23–26]. Symptoms can be triggered by stressors such as prolonged fasting, certain medications and pregnancy, and may lead to death if not treated immediately [27].

In this narrative review, we outline challenges in diagnosing and managing patients with late-onset OTCD. Prompt treatment of OTCD can be lifesaving but can only follow a successful diagnosis. An overview of the upcoming therapeutic developments for this rare condition is a novel aspect of the review.

2. Literature Review

A systematic literature search was performed in Pubmed, Embase and CINAHL ending on 1 April 2023. Original studies, reviews, case reports, case series, conference abstracts and proceedings including patients with known late-onset OTCD were eligible for inclusion. Data from abstracts that were not written in English were excluded.

The data extraction was performed from each full text by two authors (M.I. and K.S.).

The search was performed using combinations of several keywords: "Ornithine Transcarbamylase deficiency", "pregnancy", "transition", "therapy", "gene therapy", "dietetic modifications", "clinical outcome", "mortality", "acute decompensation", "laboratory investigations" and "genetic counselling" with Boolean operators "AND" and "OR" for a thorough search. We performed a descriptive narrative synthesis.

3. Diagnosis

3.1. Clinical Manifestations as Diagnostic Clues

Recognizing the symptoms of late-onset OTCD is challenging because it is a rare disease with a highly heterogeneous, episodic presentation [19,24,28]. Therefore, the prompt diagnosis and the rapid introduction of dietetic modifications, followed by appropriate treatment with ammonia scavengers, are essential to reduce the mortality and improve the quality of life [17].

Clinical manifestations of adults with OTCD vary in acuity. Gastrointestinal symptoms include vomiting, abdominal pain and hepatic dysfunction. Patients may present with chronic migraine headaches or neurological signs of hyperammonemia such as lethargy, disorientation, agitation, confusion, reduced consciousness and prolonged generalized seizures. An initial presentation with psychiatric symptoms can occur and OTCD should be considered in adolescents with new-onset psychosis or bipolar-like symptoms [21,26,27,29–32].

3.2. Triggering Factors

Clinical symptoms can be aggravated by metabolic stressors (an increased protein consumption including the use of sport supplements, infection, trauma, fever, severe illness and acute stress) or presence of a catabolic state (intrapartum and immediate postpartum, menstruation, gastric bypass surgery, starvation and fasting), or due to increased demands upon the urea cycle (systemic corticosteroids, rapid weight loss, gastrointestinal bleeding trauma and chemotherapy) [19,21,26,27,31,33–36]. Apart from steroids (dexamethasone, hydrocortisone and methylprednisolone) and chemotherapy, anticonvulsants (valproate and topiramate), metoclopramide and azathioprine may increase demands on the urea cycle, thus triggering an initial OTCD presentation [37].

In addition, underlying morbidities such as cardiac dysfunction and nonalcoholic fatty liver disease also unmask the OTCD in adults [24]. As hyperammonemia may become apparent in these patients during acute metabolic decompensation [21], it is important to consider and initiate the relevant investigations.

3.3. Laboratory Work Up

As the clinical presentation may be non-specific, the diagnosis should be confirmed with the biochemical and genetic testing [4]. A patient presenting with the symptoms listed above should have a blood ammonia level measured, using a sample collected through a free-flowing venipuncture and placed immediately on ice. Ammonia levels should also be drawn from any patient presenting with encephalopathy and respiratory alkalosis as hyperammonemia irritates the central respiratory drive and increases the respiratory rate. Late-onset OTCD is not uncommon to appear in adults with hyperammonemia, especially with no background of underlying liver cirrhosis [21,38].

Once hyperammonemia is established, diagnostic biochemical investigations including plasma amino acids and urine organic acids should be performed. Plasma amino acids will aid the diagnosis—showing high plasma glutamine and low plasma citrulline—and urine organic acids should show orotic aciduria [13,24,25,33,39]. The urine extraction of orotic acid results from excess carbamoylphosphate, which diverts into the pyrimidine pathway [16]. Orotic acid quantification in urine following allopurinol or protein loading, and/or a pedigree analysis, can be used as an alternative for female carriers [27]. However, the sensitivity and specificity of the allopurinol test is 91% and 70%, respectively. As protein loading can trigger symptoms, it is better to avoid it [29]. Orotic acid concentration can, however, be raised in other conditions such as lysinuric protein intolerance, Rett syndrome, certain forms of liver disease, certain forms of cancer and secondary to the use of certain medications and alcohol [40].

Laboratory investigations may only be diagnostic during an acute, symptomatic phase, therefore increasing the importance of a genetic diagnosis [12,27]. Approximately 80–90% of cases are detected through a mutation analysis [25,27,41,42]. In some laboratories, the likelihood of not confirming the mutation in a biochemically confirmed OTC family is as high as 30% (personal observation). Functional assays including enzyme activity in the liver and intestinal mucosa are helpful [27], although they are more invasive and require a specialized biochemical laboratory. In most males with the absence of mutation, an analysis of liver enzymes is diagnostic. However, functional studies may be unreliable due to random X inactivation leading to a mosaic distribution of residual OTC activity [43].

With advances in gene sequencing technology, novel mutations are detected over time [15]. More than 500 disease causing mutations have been reported in the OTC gene so far [44]. Of all, single base substitutions account for 70–84%, 12% are small-fragment deletions or insertions and the rest are large fragment deletions [14,44]. Several mutations have been reported in the promoter and enhancer region [41]. New high-throughput assays are in development to aid in classifying variants by functional status [45]. In general, the complete dysfunction of enzymes is due to the distortion of the reading frame of the coding sequence via nonsense, insertion and deletion. The substitution of an amino acid in either the active site or the hydrophobic core leads to the partial dysfunction of the enzyme [14,46].

Approximately 60% of hemizygous males with the severe neonatal form have mutation around the active site of the enzyme. The rest show mutation at the periphery, present later with less severe manifestations [8]. However, this classification is not definitive as presentation can vary among two individuals with the same OTC variant [46,47].

4. Management

4.1. Common Complications in Adults

The management of patients with OTCD focuses on the prevention of both acute and chronic complications [36]. Acute hyperammonemia leads to encephalopathy and neuronal death. Glutamatergic receptors in the brain become overstimulated by excess glutamate and glutamine, which affects neuronal function. During chronic hyperammonemia, increased GABAergic signaling results in reduced neurocognitive functions [36].

Consequently, OTCD patients frequently show long-term neuropsychological complication such as a learning disability, intellectual disability, attention deficit disorder and executive function deficit [29,48,49] and chronic psychiatric problems [19]. Fine motor, executive and cognitive flexibility and inhibition ability functions were shown to be affected in asymptomatic and symptomatic patients but no measurable differences were noted in attention, language or verbal memory [48,49]. Late-onset OTCD patients may present with lethargy, "fogginess of their brain" or psychosis. As a result of persistently raised plasma ammonia, the cognitive function may be affected, which becomes more apparent during acute illness when plasma ammonia raises significantly higher than usual. In such situations, their capacity to make decisions is limited. Any clinical decisions regarding the patient's health, e.g., procedures and scans under sedation, are made by the medical team in the patient's best interest.

4.2. Multidisciplinary Team

The involvement of the multidisciplinary team (MDT) is very important for the successful management [49,50]. This MDT comprises an intensive care team, a neurologist, a surgical team, a nutritionist, a metabolic specialist, pharmacy staff, social workers, a learning disability nurse, a hepatologist and genetic counselors [2]. The multispecialty and multidisciplinary approach facilitates an early diagnosis and comprehensive management, and results in positive clinical outcomes.

4.3. Dietetic Support

In a significant number of cases, patients are protein averse and often follow a vegetarian diet or only consume small quantities of high-biological-value proteins such as meat and fish (a natural protein intake not higher than 0.8 g/kg of body weight). A detailed diet history is important to understand current nutritional intake and to inform dietetic advice individually tailored to the needs of the patient.

If patients are not protein averse, then they are advised to limit the intake of protein. Recommendations on protein intake are informed by current protein intake, blood tests including ammonia and plasma amino acid concentrations and clinical symptoms. If protein intake is too low, a total protein or an essential amino acid supplement may be recommended if dietary intake cannot be adjusted to meet the patient's needs. A reduction in protein intake will be advised if ammonia is raised above the recommended range or if high ammonia is indicated with clinical symptoms in the absence of an ammonia result.

Due to their restricted diet, patients can be at risk of deficiencies in iron, zinc, copper, calcium, cobalamin and essential fatty acids. Key micronutrients should therefore be monitored, and replaced if below the normal range. It is of particular importance to monitor zinc, which acts as a cofactor for the OTC enzyme [50].

Tube feeding (gastrostomy) is more common in individuals with early-onset OTCD, to ensure the adequate intake of energy and protein. Indications for a nasogastric tube in acute OTCD include difficult swallowing, the refusal of food, gastrointestinal discomfort, a poor palatability of drugs and supplements if required. A gastrostomy is recommended

when tube feeding is to be continued for more than a short time and/or overnight feeds are required [50].

4.4. Family Screening—Genetic Counsellors

The genetic counselor role is pivotal in educating the family in the inheritance of this rare disease and to help them to identify other members at risk with genetic testing [2]. Genetic testing of newborn offspring with OTC carriers is useful to genetic counselling and to inform early intervention [51]. The counselling needs to include experts in OTCD because it is impossible to predict the severity of disease in female carriers. The clinical picture in heterozygous females varies among individuals both in terms of the onset of the disease and its severity, which results from the genotype and the degree of inactivation of the mutated X chromosome in hepatocytes. Up to 85% of heterozygous females do not develop symptoms of hyperammonemia during their lifetime [37]. In rare cases, de novo mutations have been reported in up to 26% of male probands [52], meaning their mothers are not carriers for the mutation.

4.5. Transition from Pediatric to Adult Hospital

With the advances in diagnostic technology and therapeutic options, affected children reach adulthood. Therefore, a well-driven, planned transition of adolescents and young adults from child-centered care to an adult-oriented care system is essential. Successful transition will lead to continuity in care and provide metabolic stability [53]. In the process of transition and the transfer of care, it is essential to identify whether the young adult recognizes potential triggers of a hyperammonemic episode and would know how to seek medical care for concerning symptoms.

However, different challenges are faced in the management and social requirements vary from childhood to adulthood [54,55]. A lack of guidelines on the transition care leads to anxiety among young adults and their families [56,57].

The required facilities and adequate specialist training of adult care physicians in Inherited Metabolic Diseases (IMD) management are difficult to access in most countries and non-existent in some [55]. As a result, young adults remain under pediatricians' care long-term or, in some cases, are lost to follow up [57,58].

The common challenges around the transition to adult services include autonomy and becoming independent, which can lead to a poor adherence to the medicine and diet [56]. An additional challenge in transition is the possible use of alcohol and illicit substances. These can increase the risk of decompensation events. Changes in dose and frequency of medication and the volume of diet are influenced with age, growth spurts, the magnitude of the disease and associated co-morbidities. Limitations in the availability of amino acid supplements and a dietetic regimen can form obstacles to engagement during the transfer of care, as can changes of medications in the adult hospital [24,53].

A study by Ladha et al. [53] has shown that the total average transition readiness assessment questionnaire (TRAQ) score for subjects with UCDs, including OTCD, in this cohort was low at 2.96/5.0, which was significantly lower than the published TRAQ score from young people without any ongoing debilitating conditions [53]. It was concluded that the individuals with a UCD and low IQ may benefit from personalized education in keeping with domains of the TRAQ [53].

OTCD patients may have a wide range of cognitive impairment and behavioral problems. Further arrangements and refinements should be considered for patients who have a poor neurocognitive outcome in order to have a successful transition [55,57]. A tailored approach is necessary for timing, planning and implementing the transition for each patient.

For individuals with late-onset OTCD that have been mostly asymptomatic in their youth and adolescence, and who, in many cases, are not required to follow even an emergency regimen during illness, the underlying diagnosis may have a lower impact on their overall knowledge about transition.

4.6. Preimplantation Genetic Diagnosis

A preimplantation genetic diagnosis (PGD) is more beneficial over a prenatal diagnosis either with chorionic villous sampling or genetic amniocentesis with molecular testing for unfortunate families. PGD allows for the preselection of an OTC-free embryo for implantation and prevents the unnecessary therapeutic termination of the pregnancy, mortality of offspring and intense trauma to the family [23,34,59–61].

4.7. Pregnancy

Pregnancy is a risk for hemizygous females because the catabolic state, both intraand post-partum, is a critical trigger for hyperammonemia [36,62,63]. Anorexia, nausea, vomiting and dehydration in the first trimester increase the risk of catabolism and it should be managed aggressively both medically and dietetically, in the form of extra calories, fluids and medications to manage nausea and vomiting. Management can be intravenous if indicated [23,63]. Protein levels rise during the immediate post-partum period due to uterine involution and patients must be managed carefully to prevent hyperammonemia. This management would usually include a protein restriction, to improve ammonia levels, and additional calories, to reduce the complicating risk of catabolism.

Uncommon clinical manifestations such as psychosis during post-partum can also appear in OTCD [29]. However, it should not be misdiagnosed as post-partum depression. The diagnosis of OTCD in mothers has been previously made after their sons developed symptoms of an acute illness and hyperammonemia [23,34,62].

A lack of proper planning and limited time for multidisciplinary work are main risks of the unexpected OTCD diagnosis during pregnancy [21,34]. It may result in detrimental consequences for the mother and their baby.

4.8. Mortality

Mortality is a key indicator for the further evaluation of the diagnosis of the late-onset phenotype [21,28]. A meta-analysis by Burgard et al. (2016) has shown that all UCDs, apart from females with OTCD, have a high risk of early-onset manifestations and neonatal death [20]. Despite significant diagnostic and therapeutic improvements, the mortality rate has not changed in several decades. In older patients, the risk of mortality is very high if high ammonia is not identified and early treatment is not initiated [64]. The initial peak value of >1000 μ mol/L shows a higher mortality compared to <500 μ mol/L [21,65,66]. The family history is often very relevant, as the diagnosis of late-onset OTCD can be made in an adult person after a grandchild presents with a severe illness. As an example, the late presentation of OTCD in a 62-year-old man did not raise a suspicion of a rare disease until their grandson was confirmed to be affected with the same mutation. The delayed diagnosis was made post-mortem with an undetectable enzyme activity in their liver [64].

Reported causes of mortality in late-onset OTCD include hyperammonemic encephalopathy, cerebral edema and cerebellar herniation, an elevated intracranial pressure (ICP) and status epilepticus [20,22,23,32].

5. Current and Upcoming Therapies

For acute management, three points are to be considered—the removal of ammonia, reversing the catabolic state and avoiding exogenous protein and the initiation of nitrogen scavengers [4,12,27,30,35,43,67,68].

The removal of ammonia with hemofiltration is the recommended choice before transferring to a specialized center when the ammonia level is >200 μ g/dL [26,28,67]. The provision of caloric support with 10–20% of IV glucose, and SMOF or intralipids, if required to achieve the calorie requirement, is necessary to prevent the catabolism [67,68]. An exogenous protein supply should be avoided temporarily but not for more than 48 h. Restarting protein intake is then recommended to prevent the endogenous protein catabolism [67].

If the dietetic management is not sufficient, then sodium phenylacetate and sodium benzoate are used as nitrogen scavengers with L-citrulline (oral preparation) or L-arginine hydrochloride (IV and oral preparation) supplementation [4]. Glycerol phenylbutyrate has been shown to improve the overall metabolic control [69]. Ammonia is eliminated via an alternative pathway by sodium phenylbutyrate and sodium benzoate. Arginine activates the urea cycle [8,26] and also controls the proteolysis and in turn reduces the urea production [68].

Long-term management mainly focuses on a nutrient-controlled diet, especially with protein, the supplementation of L-arginine and L-carnitine, ammonia scavengers whenever indicated [68] and the avoidance of triggers [17]. L-carnitine crosses the blood–brain barrier and helps reduce the level of ammonia through reaction cascades [28].

Orthotopic liver transplantation (OLT) is an option for patients who are affected by severe, recurrent attacks or failure for the medications to reduce ammonia. Patients who underwent liver transplantation showed a good outcome and improved quality of life [70]. The long-term neurocognitive outcome needs to be closely monitored after liver transplantation to provide the best supportive care [71].

5.1. Hepatocyte Transplantation

This mode of therapy is a substitute for OLT but, whilst it shows less surgical complications, the effect is transient, and frequent repeat hepatocyte transfusions are required as well as similar levels of immunosuppression [72]. Mesenchymal stem cells that differentiate into hepatocytes theoretically have more capacity to proliferate in a diseased liver and are highly immunotolerant compared to mature implanted hepatocytes [52,73]. However, there is no evidence from clinical trials that this method works in humans [74].

5.2. Upcoming Therapies

OTCD is an appealing candidate for novel therapies including gene addition, mRNA therapy and genome editing [75]. The first gene therapy clinical trial targeting OTCD was initiated in the late 1990s using adenoviral vectors. This attempt ended with the death of one of the participants due to a severe immune response [76].

Efforts continued with safer viral vectors including adeno-associated virus (AAV) after the 2000s. Early pre-clinical proof of concept studies with hepatotropic AAV vectors showed metabolic correction and prolonged survival [77]. More recent studies showed that AAVmediated gene addition can also prevent chronic liver damage and fibrosis [78]. A phase I/II safety and dose-finding study (CAPtivate, ClinicalTrials.gov Identifier NCT02991144) using AAV8 recruited adult patients and has been completed without any serious treatment-related adverse events. A long-term follow-up study (ClinicalTrials.gov Identifier NCT03636438) is ongoing. In total, 7/11 treated patients were determined as responders and remained metabolically stable from 2 to 4.5 years. Four complete responders have required neither ammonia scavengers nor dietary restrictions after treatment [79]. A phase III study is currently recruiting 50 participants who are 12 years and older (ClinicalTrials.gov Identifier NCT05345171).

As AAVs mostly do not integrate into the host genome and mainly remain episomal, AAV DNA is lost during the cell division. This is still an important hurdle to achieve sustained transduction in the growing liver [80]. To overcome this, engineered AAV capsids with a higher transduction efficiency have been developed. AAVLK03 was found to have 10 times higher transduction rates compared to AAV8 [81]. Therefore, it has been a strong candidate to treat OTCD and, after proving safety in preclinical studies, clinical translation is ongoing [82]. A phase I/II open-label, multicenter clinical trial (HORACE, ClinicalTrials.gov Identifier NCT05092685), aiming to treat pediatric patients, is at the pre-recruiting stage.

mRNA technology is another promising gene therapy approach for OTCD. However, as it provides a rapid and transient expression of the protein, repeated administration is necessary for long-term efficacy [83]. The multi-dose systemic administration of mRNA provided metabolic correction as well as an improved survival in a murine model of OTCD [84]. A phase Ia randomized, double-blinded, placebo-controlled study (ClinicalTrials.gov Identifier NCT04416126) using a single dose of OTC mRNA has just been completed. A phase

Ib study (ClinicalTrials.gov Identifier NCT04442347) is currently active and has recruited 12 adults. A phase II study (ClinicalTrials.gov Identifier NCT055260660 using multi-dose OTC mRNA is now recruiting both adolescents and adults.

Both in vivo and ex vivo genome editing strategies are emerging for OTCD. AAVdelivered CRISPR-Cas9-mediated gene editing was found to be effective both in a OTCdeficient murine model and primary human hepatocytes [85–87]. Ex vivo CRISPR-corrected human hepatocytes provided metabolic correction in a murine model [88]. This preclinical success is encouraging for clinical translation.

6. Conclusions

The early diagnosis of late-onset OTCD is important as even asymptomatic patients are at risk for a life-threatening hyperammonemic crisis and can benefit from a tailored UCD management plan. Comorbidities and dietetic preferences may play a role in masking or unveiling a late-onset OTCD. The condition requires unique clinical care, including the prevention and treatment of acute metabolic decompensation; a patient-centered approach; continued education regarding symptom recognition and adherence to the therapy and diet; highly specialized management through pregnancy; and genetic counselling regarding the risks of the condition. There is a clinical need for diagnostic and disease monitoring biomarkers. Upcoming therapies will bring meaningful clinical benefits to the patients and improve their quality of life. An increase in residual enzymatic OTC activity above the therapeutic target of 5% might reduce mortality and the incidence of a severe OTCD phenotype.

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References

- 1. Çelik, K.; Terek, D.; Olukman, Ö.; Kağnıcı, M.; Keskin Gözmen, Ş.; Serdaroğlu, E.; Çalkavur, Ş.; Arslanoğlu, S. Urea Cycle Disorders in Neonates: Six Case Reports. *J. Pediatr. Res.* **2017**, *4*, 85–89. [CrossRef]
- Baker, J.; Hitchins, L.; Vucko, E.; Havens, K.; Becker, K.; Arduini, K. Variable disease manifestations and metabolic management within a single family affected by ornithine transcarbamylase deficiency. *Mol. Genet. Metab. Rep.* 2022, 33 (Suppl. 1), 100906. [CrossRef] [PubMed]
- Ratnakumari, L.; Qureshi, I.A.; Butterworth, R.F. Evidence for cholinergic neuronal loss in brain in congenital ornithine transcarbamylase deficiency. *Neurosci. Lett.* 1994, 178, 63–65. [CrossRef] [PubMed]
- Ben-ari, Z.; Dalal, A.; Morry, A.; Pitlik, S.; Zinger, P.; Cohen, J.; Fattal, I.; Galili-Mosberg, R.; Tessler, D.; Baruch, R.G.; et al. Case Report Adult-onset ornithine transcarbamylase (OTC) deficiency unmasked by the Atkins' diet. *J. Hepatol.* 2010, 52, 292–295. [CrossRef]
- 5. Ye, X.; Robinson, M.B.; Batshaw, M.L.; Furth, E.E.; Smith, I.; Wilson, J.M. Prolonged Metabolic Correction in Adult Ornithine Transcarbamylase-deficient Mice with Adenoviral Vectors. J. Biol. Chem. **1996**, 271, 3639–3646. [CrossRef] [PubMed]
- Butterworth, R.F. Effects of hyperammonaemia on brain function. J. Inher. Metab. Dis. 1998, 21 (Suppl. 1), 6–20. [CrossRef] [PubMed]
- Laróvere, L.E.; Silvera Ruiz, S.M.; Arranz, J.A.; Dodelson de Kremer, R. Mutation Spectrum and Genotype–Phenotype Correlation in a Cohort of Argentine Patients with Ornithine Transcarbamylase Deficiency: A Single-Center Experience. J. Inborn Errors Metab. Screen. 2018, 6, 1–5. [CrossRef]
- 8. Gyato, K.; Wray, J.; Huang, Z.J.; Yudkoff, M.; Batshaw, M.L. Metabolic and neuropsychological phenotype in women heterozygous for ornithine transcarbamylase deficiency. *Ann. Neurol.* **2004**, *55*, 80–86. [CrossRef]

- 9. Tuchman, M.; Jaleel, N.; Morizono, H.; Sheehy, L.; Lynch, M.G. Mutations and polymorphisms in the human ornithine transcarbamylase gene. *Hum. Mutat.* **2002**, *19*, 93–107. [CrossRef]
- Feigenbaum, A. Challenges of managing ornithine transcarbamylase deficiency in female heterozygotes. *Mol. Genet. Metab. Rep.* 2022, 33, 100941. [CrossRef]
- 11. Seker Yilmaz, B.; Baruteau, J.; Arslan, N.; Aydin, H.I.; Barth, M.; Bozaci, A.E.; Brassier, A.; Canda, E.; Cano, A.; Chronopoulou, E.; et al. Three-Country Snapshot of Ornithine Transcarbamylase Deficiency. *Life* **2022**, *12*, 1721. [CrossRef] [PubMed]
- 12. Jin, X.; Zeng, X.; Zhao, D.; Jiang, N. Liver transplantation in rare late-onset ornithine transcarbamylase deficiency with central nervous system injury: A case report and review of the literature. *Brain Behav.* **2022**, *12*, e2765. [CrossRef] [PubMed]
- 13. Couchet, M.; Breuillard, C.; Corne, C.; Rendu, J.; Morio, B.; Schlattner, U.; Moinard, C. Ornithine Transcarbamylase—From Structure to Metabolism: An Update. *Front. Physiol.* **2021**, *12*, 748249. [CrossRef] [PubMed]
- 14. Nguyen, H.H.; Khanh Nguyen, N.; Dung Vu, C.; Thu Huong Nguyen, T.; Nguyen, N.L. Late-Onset Ornithine Transcarbamylase Deficiency and Variable Phenotypes in Vietnamese Females with OTC Mutations. *Front. Pediatr.* **2020**, *8*, 321. [CrossRef] [PubMed]
- 15. Caldovic, L.; Abdikarim, I.; Narain, S.; Tuchman, M.; Morizono, H. Genotype-Phenotype Correlations in Ornithine Transcarbamylase Deficiency: A Mutation Update. *J. Genet. Genom.* **2015**, *42*, 181–194. [CrossRef] [PubMed]
- Brassier, A.; Gobin, S.; Arnoux, J.B.; Valayannopoulos, V.; Habarou, F.; Kossorotoff, M.; Servais, A.; Barbier, V.; Dubois, S.; Touati, G.; et al. Long-term outcomes in Ornithine Transcarbamylase deficiency: A series of 90 patients. *Orphanet J. Rare Dis.* 2015, 10, 58. [CrossRef]
- 17. Liu, J.; Dong, L.; Wang, Y.; Zhang, M. Two novel mutations of ornithine transcarbamylase gene identified from three Chinese neonates with ornithine transcarbamylase deficiency. *Int. J. Clin. Exp. Med.* **2015**, *8*, 2656–2661.
- Wilnai, Y.; Blumenfeld, Y.J.; Cusmano, K.; Hintz, S.R.; Alcorn, D.; Benitz, W.E.; Berquist, W.E.; Bernstein, J.A.; Castillo, R.O.; Concepcion, W.; et al. Prenatal treatment of ornithine transcarbamylase deficiency. *Mol. Genet. Metab.* 2018, 123, 297–300. [CrossRef]
- Andrews, A.; Roberts, S.; Botto, L.D. Benefits of tailored disease management in improving tremor, white matter hyperintensities, and liver enzymes in a child with heterozygous X-linked ornithine transcarbamylase deficiency. *Mol. Genet. Metab. Rep.* 2022, 33 (Suppl. 1), 100891. [CrossRef]
- Burgard, P.; Kölker, S.; Haege, G.; Lindner, M.; Hoffmann, G.F. Neonatal mortality and outcome at the end of the first year of life in early onset urea cycle disorders—Review and meta-analysis of observational studies published over more than 35 years. *J. Inherit. Metab. Dis.* 2016, 39, 219–229. [CrossRef]
- Yamamoto, S.; Yamashita, S.; Kakiuchi, T.; Kurogi, K.; Nishi, T.M.; Tago, M.; Yamashita, S.I. Late-Onset Ornithine Transcarbamylase Deficiency Complicated with Extremely High Serum Ammonia Level: Prompt Induction of Hemodialysis as the Key to Successful Treatment. Am. Case Rep. 2022, 23, e937658. [CrossRef] [PubMed]
- 22. Wraith, J.E. Ornithine Carbamoyltransferase Deficiency Disease. Arch. Dis. Child. 2001, 84, 84–88. [CrossRef] [PubMed]
- Torkzaban, M.; Haddad, A.; Baxter, J.K.; Berghella, V.; Gahl, W.A.; Al-Kouatly, H.B. Maternal ornithine transcarbamylase deficiency, a genetic condition associated with high maternal and neonatal mortality every clinician should know: A systematic review. Am. J. Med. Genet. A 2019, 179, 2091–2100. [CrossRef]
- 24. Abbott, J.; Senzatimore, M.; Atwal, P. A complex case of delayed diagnosis of ornithine transcarbamylase deficiency in an adult patient with multiple comorbidities. *Mol. Genet. Metab. Rep.* **2022**, *33* (Suppl. 1), 100916. [CrossRef]
- Hertzog, A.; Selvanathan, A.; Halligan, R.; Fazio, T.; de Jong, G.; Bratkovic, D.; Bhattacharya, K.; Tolun, A.A.; Bennetts, B.; Fisk, K. A serendipitous journey to a promoter variant: The c.-106C>A variant and its role in late-onset ornithine transcarbamylase deficiency. *JIMD Rep.* 2022, 63, 271–275. [CrossRef] [PubMed]
- Hidaka, M.; Higashi, E.; Uwatoko, T.; Uwatoko, K.; Urashima, M.; Takashima, H.; Watanabe, Y.; Kitazono, T.; Sugimori, H. Late-onset ornithine transcarbamylase deficiency: A rare cause of recurrent abnormal behavior in adults. *Acute Med. Surg.* 2020, 7, 2–5. [CrossRef]
- 27. Gascon-Bayarri, J.; Campdelacreu, J.; Estela, J.; Reñé, R. Severe Hyperammonemia in Late-Onset Ornithine Transcarbamylase Deficiency Triggered by Steroid Administration. *Case Rep. Neurol. Med.* **2015**, 2015, 453752. [CrossRef]
- Alameri, M.; Shakra, M.; Alsaadi, T. Fatal coma in a young adult due to late-onset urea cycle deficiency presenting with a prolonged seizure: A case report. J. Med. Case Rep. 2015, 9, 4–9. [CrossRef]
- 29. Stepien, K.M.; Geberhiwot, T.; Hendriksz, C.J.; Treacy, E.P. Challenges in diagnosing and managing adult patients with urea cycle disorders. *J. Inherit. Metab. Dis.* **2019**, *42*, 1136–1146. [CrossRef]
- Marquetand, J.; Freisinger, P.; Lindig, T.; Euler, S.; Gasser, M.; Overkamp, D. Ammonia and coma—A case report of late onset hemizygous ornithine carbamyltransferase deficiency in 68-year-old female. *BMC Neurol.* 2020, 20, 118. [CrossRef]
- 31. Roberts, D.L.; Galbreath, D.A.; Patel, B.M.; Ingall, T.J.; Khatib, A.; Johnson, D.J. Hyperammonemic Coma in an Adult due to Ornithine Transcarbamylase Deficiency. *Case Rep. Crit. Care* **2013**, 2013, 493216. [CrossRef] [PubMed]
- Pizzi, M.A.; Alejos, D.; Hasan, T.F.; Atwal, P.S.; Krishnaiengar, S.R.; Freeman, W.D. Adult Presentation of Ornithine Transcarbamylase Deficiency: 2 Illustrative Cases of Phenotypic Variability and Literature Review. *Neurohospitalist* 2019, *9*, 30–36. [CrossRef]
- 33. Lu, D.; Han, F.; Qiu, W.; Zhang, H.; Ye, J.; Liang, L.; Wang, Y.; Ji, W.; Zhan, X.; Gu, X.; et al. Clinical and molecular characteristics of 69 Chinese patients with ornithine transcarbamylase deficiency. *Orphanet J. Rare Dis.* **2020**, *15*, 340. [CrossRef] [PubMed]
- Mendez-Figueroa, H.; Lamance, K.; Sutton, V.R.; Aagaard-Tillery, K.; Van Den Veyver, I. Management of ornithine transcarbamylase deficiency in pregnancy. Am. J. Perinat. 2010, 27, 775–783. [CrossRef] [PubMed]

- 35. Anstey, J.R.; Haydon, T.P.; Ghanpur, R.B.; de Jong, G. Initial presentation of a urea cycle disorder in adulthood: An underrecognised cause of severe neurological dysfunction. *Med. J. Aust.* 2015, 203, 445–447. [CrossRef]
- 36. Strong, A.; Gold, J.; Gold, N.B.; Yudkoff, M. Hepatic Manifestations of Urea Cycle Disorders. *Clin. Liver Dis.* **2021**, *18*, 198–203. [CrossRef]
- Lichter-Konecki, U.; Caldovic, L.; Morizono, H.; Simpson, K. Ornithine transcarbamylase deficiency. In *GeneReviews*[®]; Adam, M.P., Ardinger, H.H., Pagon, R.A., Wallace, S.E., LJH, B., Stephens, K., Amemiya, A., Eds.; University of Washington: Seattle, WA, USA, 2016; pp. 1993–2018. Available online: http://www.ncbi.nlm.nih.gov/books/NBK154378/ (accessed on 30 May 2023).
- 38. Durer, S.; Durer, C.; Hoilat, G.J. Adult-onset ornithine transcarbamylase deficiency as a rare cause of fatal hyperammonaemia. *Lancet* **2021**, *398*, e11. [CrossRef]
- 39. Sysák, R.; Brennerová, K.; Krlín, R.; Štencl, P.; Rusňák, I.; Vargová, M. Effect of Ornithine Transcarbamylase (OTC) Deficiency on Pregnancy and Puerperium. *Diagnostics* **2022**, *12*, 415. [CrossRef]
- 40. Visek, W.J.; Shoemaker, J.D. Orotic acid, arginine, and hepatotoxicity. J. Am. Coll. Nutr. 1986, 5, 153–166. [CrossRef]
- Jang, Y.J.; LaBella, A.L.; Feeney, T.P.; Braverman, N.; Tuchman, M.; Morizono, H.; Ah Mew, N.; Caldovic, L. Disease-causing mutations in the promoter and enhancer of the ornithine transcarbamylase gene. *Hum. Mutat.* 2018, 39, 527–536. [CrossRef] [PubMed]
- 42. Cheng, L.; Liu, Y.; Wang, W.; Merritt, J.L.; Yeh, M. Hepatocellular Adenoma in a Patient with Ornithine Transcarbamylase Deficiency. *Case Rep. Hepatol.* **2019**, 2019, 2313791. [CrossRef] [PubMed]
- Häberle, J.; Burlina, A.; Chakrapani, A.; Dixon, M.; Karall, D.; Lindner, M.; Mandel, H.; Martinelli, D.; Pintos-Morell, G.; Santer, R.; et al. Suggested guidelines for the diagnosis and management of urea cycle disorders: First revision. J. Inherit. Metab. Dis. 2019, 42, 1192–1230. [CrossRef]
- 44. The Human Gene Mutation Database. Available online: https://www.hgmd.cf.ac.uk/ac/search.php (accessed on 27 May 2023).
- Lo, R.S.; Cromie, G.A.; Tang, M.; Teng, K.; Owens, K.; Sirr, A.; Kutz, J.N.; Morizono, H.; Caldovic, L.; Ah Mew, N.; et al. The functional impact of 1570 individual amino acid substitutions in human OTC. *Am. J. Hum. Genet.* 2023, *110*, 863–879. [CrossRef] [PubMed]
- Ausems, M.G.; Bakker, E.; Berger, R.; Duran, M.; van Diggelen, O.P.; Keulemans, J.L.; de Valk, H.W.; Kneppers, A.L.; Dorland, L.; Eskes, P.F.; et al. Asymptomatic and late-onset ornithine transcarbamylase deficiency caused by a A208T mutation: Clinical, biochemical and DNA analyses in a four-generation family. *Am. J. Med. Genet.* 1997, *68*, 236–239. [CrossRef]
- Choi, J.H.; Lee, B.H.; Kim, J.H.; Kim, G.H.; Kim, Y.M.; Cho, J.; Cheon, C.K.; Ko, J.M.; Lee, J.H.; Yoo, H.W. Clinical outcomes and the mutation spectrum of the OTC gene in patients with ornithine transcarbamylase deficiency. *J. Hum. Genet.* 2015, 60, 501–507. [CrossRef]
- 48. Kazmierski, D.; Sharma, N.; O'Leary, K.; Ochieng, P. Valproate-induced fatal acute hyperammonaemia-related encephalopathy in late-onset ornithine transcarbamylase deficiency. *BMJ Case Rep.* **2021**, *14*, e241429. [CrossRef]
- Sprouse, C.; King, J.; Helman, G.; Pacheco-Colón, I.; Shattuck, K.; Breeden, A.; Seltzer, R.; VanMeter, J.W.; Gropman, A.L. Investigating neurological deficits in carriers and affected patients with ornithine transcarbamylase deficiency. *Mol. Genet. Metab.* 2014, 113, 136–141. [CrossRef] [PubMed]
- Santos, C.D.; Ratzlaff, R.A.; Meder, J.C.; Atwal, P.S.; Joyce, N.E. Ornithine Transcarbamylase Deficiency: If at First You Do Not Diagnose, Try and Try Again. Case Rep. Crit. Care 2017, 2017, 8724810. [CrossRef]
- 51. Fujisawa, D.; Mitsubuchi, H.; Matsumoto, S.; Iwai, M.; Nakamura, K.; Hoshide, R.; Harada, N.; Yoshino, M.; Endo, F. Early intervention for late-onset ornithine transcarbamylase deficiency. *Pediatr. Int.* **2015**, *57*, e1–e3. [CrossRef]
- 52. Rüegger, C.M.; Lindner, M.; Ballhausen, D.; Baumgartner, M.R.; Beblo, S.; Das, A.; Gautschi, M.; Glahn, E.M.; Grünert, S.C.; Hennermann, J.; et al. Cross-sectional observational study of 208 patients with non-classical urea cycle disorders. *J. Inherit. Metab. Dis.* **2014**, *37*, 21–30. [CrossRef]
- 53. Ladha, F.A.; Le Mons, C.; Craigen, W.J.; Magoulas, P.L.; Marom, R.; Lewis, A.M. Barriers to a successful healthcare transition for individuals with urea cycle disorders. *Mol. Genet. Metab.* **2023**, *139*, 107609. [CrossRef] [PubMed]
- 54. Gariani, K.; Nascimento, M.; Superti-Furga, A.; Tran, C. Clouds over IMD? Perspectives for inherited metabolic diseases in adults from a retrospective cohort study in two Swiss adult metabolic clinics. *Orphanet J. Rare Dis.* **2020**, *15*, 210. [CrossRef] [PubMed]
- 55. Stepien, K.M.; Kieć-Wilk, B.; Lampe, C.; Tangeraas, T.; Cefalo, G.; Belmatoug, N.; Francisco, R.; Del Toro, M.; Wagner, L.; Lauridsen, A.G.; et al. Challenges in Transition from Childhood to Adulthood Care in Rare Metabolic Diseases: Results From the First Multi-Center European Survey. *Front Med.* **2021**, *8*, 652358. [CrossRef] [PubMed]
- 56. Abeyagunawardena, S.; Abeyagunawardena, A.; Rajindrajith, S. Transition from paediatric to adult care: An emerging challenge. *Sri Lanka J. Child. Health* **2021**, *50*, 334–337. [CrossRef]
- 57. Chabrol, B.; Jacquin, P.; Francois, L.; Broué, P.; Dobbelaere, D.; Douillard, C.; Dubois, S.; Feillet, F.; Perrier, A.; Fouilhoux, A.; et al. Transition from pediatric to adult care in adolescents with hereditary metabolic diseases: Specific guidelines from the French network for rare inherited metabolic diseases (G2M). *Arch. De Pediatr.* 2018, 25, 344–349. [CrossRef]
- White, P.H.; Cooley, W.C. Transitions Clinical Report Authoring Group; American Academy of Pediatrics; American Academy of Family Physicians; American College of Physicians. Supporting the Health Care Transition From Adolescence to Adulthood in the Medical Home. *Pediatrics* 2019, 143, e20183610.

- Lee, H.S.; Jin, H.J.; Hye, W.C.; Chun, K.L.; Yoo, H.W.; Mi, K.K.; Inn, S.K. Preimplantation genetic diagnosis for ornithine transcarbamylase deficiency by simultaneous analysis of duplex-nested PCR and fluorescence in situ hybridization: A case report. *J. Korean Med. Sci.* 2007, 22, 572–576. [CrossRef]
- 60. Verlinsky, Y.; Rechitsky, S.; Verlinsky, O.; Strom, C.; Kuliev, A. Preimplantation diagnosis for ornithine transcarbamylase deficiency. *Reprod. BioMed. Online* **2000**, *1*, 45–47. [CrossRef]
- 61. Ray, P.F.; Gigarel, N.; Bonnefont, J.P.; Attié, T.; Hamamah, S.; Frydman, N.; Vekemans, M.; Frydman, R.; Munnich, A. First specific preimplantation genetic diagnosis for ornithine transcarbamylase deficiency. *Prenat. Diag.* **2000**, *20*, 1048–1054. [CrossRef]
- Pinho, G.; Ross, G.; Krishnamoorthy, K.; Kresge, C.; Shih, L.Y.; Apuzzio, J.J.; Williams, S.F. Ornithine transcarbamylase deficiency and pregnancy: A case series and review of recommendations. *Case Rep. Women's Health* 2022, 34, e00390. [CrossRef]
- 63. Açıkalın, A.; Dişel, N.R.; Direk, E.Ç.; Ilgınel, M.T.; Sebe, A.; Bıçakçı, Ş. A rare cause of postpartum coma: Isolated hyperammonemia due to urea cycle disorder. *Am. J. Emerg. Med.* **2016**, *34*, 1894. [CrossRef]
- 64. Rohininath, T.; Costello, D.J.; Lynch, T.; Monavari, A.; Tuchman, M.; Treacy, E.P. Fatal presentation of ornithine transcarbamylase deficiency in a 62-year-old man and family studies. *J. Inherit. Metab. Dis.* **2004**, *27*, 285–288. [CrossRef] [PubMed]
- Thurlow, V.R.; Asafu-Adjaye, M.; Agalou, S.; Rahman, Y. Fatal ammonia toxicity in an adult due to an undiagnosed urea cycle defect: Under-recognition of ornithine transcarbamylase deficiency. *Ann. Clin. Biochem.* 2010, 47 Pt 3, 279–281. [CrossRef] [PubMed]
- 66. Imoto, K.; Tanaka, M.; Goya, T.; Aoyagi, T.; Takahashi, M.; Kurokawa, M. Corticosteroid suppresses urea-cycle-related gene expressions in ornithine transcarbamylase deficiency. *BMC Gastroenterol.* **2022**, *22*, 144. [CrossRef]
- 67. Cavicchi, C.; Donati, M.; Parini, R.; Rigoldi, M.; Bernardi, M.; Orfei, F.; Gentiloni Silveri, N.; Colasante, A.; Funghini, S.; Catarzi, S.; et al. Sudden unexpected fatal encephalopathy in adults with OTC gene mutations-Clues for early diagnosis and timely treatment. *Orphanet J. Rare Dis.* **2014**, *9*, 105. [CrossRef]
- 68. Redant, S.; Empain, A.; Mugisha, A.; Kamgang, P.; Attou, R.; Honoré, P.M.; De Bels, D. Management of late onset urea cycle disorders—A remaining challenge for the intensivist? *Ann. Intensive Care* **2021**, *11*, 2. [CrossRef]
- 69. Laemmle, A.; Stricker, T.; Häberle, J. Switch from Sodium Phenylbutyrate to Glycerol Phenylbutyrate Improved Metabolic Stability in an Adolescent with Ornithine Transcarbamylase Deficiency. *JIMD Rep.* **2017**, *31*, 11–14. [CrossRef]
- 70. Morioka, D.; Kasahara, M.; Takada, Y.; Shirouzu, Y.; Taira, K.; Sakamoto, S.; Uryuhara, K.; Egawa, H.; Shimada, H.; Tanaka, K. Current role of liver transplantation for the treatment of urea cycle disorders: A review of the worldwide English literature and 13 cases at Kyoto University. *Liver Transpl.* **2005**, *11*, 1332–1342. [CrossRef]
- Posset, R.; Gropman, A.L.; Nagamani, S.C.S.; Burrage, L.C.; Bedoyan, J.K.; Wong, D.; Berry, G.T.; Baumgartner, M.R.; Yudkoff, M.; Zielonka, M.; et al. Urea Cycle Disorders Consortium and the European Registry and Network for Intoxication Type Metabolic Diseases Consortia Study Group. Impact of Diagnosis and Therapy on Cognitive Function in Urea Cycle Disorders. *Ann. Neurol.* 2019, *86*, 116–128.
- 72. Iansante, V.; Mitry, R.R.; Filippi, C.; Fitzpatrick, E.; Dhawan, A. Human hepatocyte transplantation for liver disease: Current status and future perspectives. *Pediatr. Res.* 2018, *83*, 232–240. [CrossRef]
- 73. Soria, L.R.; Ah Mew, N.; Brunetti-Pierri, N. Progress and challenges in development of new therapies for urea cycle disorders. *Hum. Mol. Genet.* 2019, *28*, 42–48. [CrossRef] [PubMed]
- 74. Meyburg, J.; Opladen, T.; Spiekerkötter, U.; Schlune, A.; Schenk, J.P.; Schmidt, J.; Weitz, J.; Okun, J.; Bürger, F.; Omran, T.B.; et al. Human heterologous liver cells transiently improve hyperammonemia and ureagenesis in individuals with severe urea cycle disorders. J. Inherit. Metab. Dis. 2018, 41, 81–90. [CrossRef] [PubMed]
- Yilmaz, B.S.; Gurung, S.; Perocheau, D.; Counsell, J.; Baruteau, J. Gene therapy for inherited metabolic diseases. J. Mother. Child. 2020, 24, 53–64. [PubMed]
- Raper, S.E.; Chirmule, N.; Lee, F.S.; Wivel, N.A.; Bagg, A.; Gao, G.P.; Wilson, J.M.; Batshaw, M.L. Fatal systemic inflammatory response syndrome in a ornithine transcarbamylase deficient patient following adenoviral gene transfer. *Mol. Genet. Metab.* 2003, 80, 148–158. [CrossRef]
- 77. Moscioni, D.; Morizono, H.; McCarter, R.J.; Stern, A.; Cabrera-Luque, J.; Hoang, A.; Sanmiguel, J.; Wu, D.; Bell, P.; Gao, G.P.; et al. Long-term correction of ammonia metabolism and prolonged survival in ornithine transcarbamylase-deficient mice following liver-directed treatment with adeno-associated viral vectors. *Mol. Ther.* 2006, 14, 25–33. [CrossRef]
- Wang, L.; Bell, P.; Morizono, H.; He, Z.; Pumbo, E.; Yu, H.; White, J.; Batshaw, M.L.; Wilson, J.M. AAV gene therapy corrects OTC deficiency and prevents liver fibrosis in aged OTC-knock out heterozygous mice. *Mol. Genet. Metab.* 2017, 120, 299–305. [CrossRef]
- 79. Harding, C.O.; Geberhiwot, T.; Couce, M.L.; Tan, W.-H.; Khan, A.; Hualde, L.C.; Diaz, G.A.; Konczal, L.; Thomas, J.; Guffon, N.; et al. Safety and Efficacy of DTX301 in Adults with Late-Onset Ornithine Transcarbamylase (OTC) Deficiency: A Phase 1/2 Trial. In *Molecular Therapy*; Cell Press: Cambridge, MA, USA, 2022.
- 80. Baruteau, J.; Waddington, S.N.; Alexander, I.E.; Gissen, P. Gene therapy for monogenic liver diseases: Clinical successes, current challenges and future prospects. *J. Inherit. Metab. Dis.* **2017**, *40*, 497–517. [CrossRef] [PubMed]
- 81. Lisowski, L.; Dane, A.P.; Chu, K.; Zhang, Y.; Cunningham, S.C.; Wilson, E.M.; Nygaard, S.; Grompe, M.; Alexander, I.E.; Kay, M.A. Selection and evaluation of clinically relevant AAV variants in a xenograft liver model. *Nature* **2014**, *506*, 382–386. [CrossRef]

- Baruteau, J.; Cunningham, S.C.; Yilmaz, B.S.; Perocheau, D.P.; Eaglestone, S.; Burke, D.; Thrasher, A.J.; Waddington, S.N.; Lisowski, L.; Alexander, I.E.; et al. Safety and efficacy of an engineered hepatotropic AAV gene therapy for ornithine transcarbamylase deficiency in cynomolgus monkeys. *Mol. Ther. Methods Clin. Dev.* 2021, 23, 135–146. [CrossRef]
- 83. Martini, P.G.V.; Guey, L.T. A New Era for Rare Genetic Diseases: Messenger RNA Therapy. *Hum. Gene Ther.* **2019**, *30*, 1180–1189. [CrossRef]
- Prieve, M.G.; Harvie, P.; Monahan, S.D.; Roy, D.; Li, A.G.; Blevins, T.L.; Paschal, A.E.; Waldheim, M.; Bell, E.C.; Galperin, A.; et al. Targeted mRNA Therapy for Ornithine Transcarbamylase Deficiency. *Mol. Ther.* 2018, 26, 801–813. [CrossRef] [PubMed]
- Wang, L.; Yang, Y.; Breton, C.; Bell, P.; Li, M.; Zhang, J.; Che, Y.; Saveliev, A.; He, Z.; White, J.; et al. A mutation-independent CRISPR-Cas9-mediated gene targeting approach to treat a murine model of ornithine transcarbamylase deficiency. *Sci. Adv.* 2020, *6*, 5701. [CrossRef] [PubMed]
- Yang, Y.; Wang, L.; Bell, P.; McMenamin, D.; He, Z.; White, J.; Yu, H.; Xu, C.; Morizono, H.; Musunuru, K.; et al. A dual AAV system enables the Cas9-mediated correction of a metabolic liver disease in newborn mice. *Nat. Biotechnol.* 2016, 34, 334–338. [CrossRef] [PubMed]
- Ginn, S.L.; Amaya, A.K.; Liao, S.H.Y.; Zhu, E.; Cunningham, S.C.; Lee, M.; Hallwirth, C.V.; Logan, G.J.; Tay, S.S.; Cesare, A.J.; et al. Efficient in vivo editing of OTC-deficient patient-derived primary human hepatocytes. *JHEP Rep.* 2019, 2, 100065. [CrossRef] [PubMed]
- Zabulica, M.; Srinivasan, R.C.; Akcakaya, P.; Allegri, G.; Bestas, B.; Firth, M.; Hammarstedt, C.; Jakobsson, T.; Jakobsson, T.; Ellis, E.; et al. Correction of a urea cycle defect after ex vivo gene editing of human hepatocytes. *Mol. Ther.* 2021, 29, 1903–1917. [CrossRef]

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