

Review

Micro- and Macrovascular Effects of Inflammation in Peripheral Artery Disease—Pathophysiology and Translational Therapeutic Approaches

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Abstract: Inflammation has a critical role in the development and progression of atherosclerosis. On the molecular level, inflammatory pathways negatively impact endothelial barrier properties and thus, tissue homeostasis. Conformational changes and destruction of the glycocalyx further promote pro-inflammatory pathways also contributing to pro-coagulability and a prothrombotic state. In addition, changes in the extracellular matrix composition lead to (peri-)vascular remodelling and alterations of the vessel wall, e.g., aneurysm formation. Moreover, progressive fibrosis leads to reduced tissue perfusion due to loss of functional capillaries. The present review aims at discussing the molecular and clinical effects of inflammatory processes on the micro- and macrovasculature with a focus on peripheral artery disease.

Keywords: atherosclerosis; inflammation; peripheral artery disease; glycocalyx; endothelial dysfunction

1. Introduction

Cardiovascular disease (CVD) is the leading cause of mortality worldwide and accounts for millions of deaths globally [1]. CVD is associated with a significant impairment of quality of life and the prevalence of its main manifestations, such as coronary artery disease (CAD), cerebrovascular disease and peripheral artery disease (PAD), has been increasing steadily over the last two decades [1,2].

Atherosclerosis is considered the major driver of CVD. Formerly, atherosclerosis was thought of as a process primarily related to dyslipidaemia and the deposition of triglycerides and cholesterol [3]. However, besides lipid accumulation, more recent insights into the pathogenesis of atherosclerosis increasingly emphasise the role of inflammation and endothelial dysfunction as major drivers of atherogenesis [3–8]. Moreover, the mentioned pathomechanisms depend on each other and amplify each other's responses. Indeed, a central element initiating prothrombotic processes and herein, atherogenesis, remains glycocalyx destruction due to inflammatory processes [9]. In PAD, inflammation is also

triggered by ischaemia-reperfusion (I/R) injury promoting increased production of reactive oxygen species (ROS) [10], which contribute to endothelial dysfunction and microvascular pathology [11].

Chronic autoimmune diseases, which are associated with significantly elevated levels of systemic inflammation, e.g., rheumatoid arthritis, systemic lupus erythematosus, anti-phospholipid syndrome and antineutrophil cytoplasmic antibody (ANCA)-associated vasculitis (AAV), are associated with markedly increased prevalence of CVD [12–14]. Conversely, there is accumulating evidence that some agents with anti-inflammatory characteristics reduce cardiovascular risk significantly [15,16]. Canakinumab, a monoclonal antibody targeting interleukin (IL)-1 β [15], and colchicine, which attenuates leukocyte responsiveness by inhibition of tubulin polymerisation [16,17], have been shown to improve outcomes in CAD in randomized controlled trials [15,16]. While not yet implemented in regular clinical practice, there is increasing awareness for anti-inflammatory therapy in secondary prevention of CVD in the current guidelines [18].

The single most effective prevention of CVD, smoking cessation, lowers levels of systemic inflammation as assessed utilising biomarkers of inflammation and oxidative damage [19,20]. Statins, which are the most widely established agents for lipid control, also have been shown to exert significant immuno-modulatory influence by inhibiting the nuclear factor kappa B (NF- κ B) pathway and decreasing the expression of toll-like receptors (TLR) [21]. Smoking cessation and statins are both recommended in all patients with CVD [22].

Formation of aneurysms is also considered to be linked to atherosclerosis. Recently, the role of leukocytes and, especially, neutrophils in the development of aneurysms has been revisited [23,24]. Activation of matrix metalloproteinases (MMP), degradation of the extracellular matrix (ECM), smooth muscle apoptosis and oxidative stress all contribute to aneurysm formation and are mediated by cytokines secreted by leukocytes [23,25]. Interestingly, atherosclerosis and aneurysm formation do not always occur at the same locations. While the abdominal aorta, an area of predilection for aneurysm formation, is also prone to atherosclerosis, the external iliac artery, a common location for significant atherosclerosis, is very seldomly involved in the formation of aneurysms. Which cellular and non-cellular processes discern these two locations is currently unclear, however, the different embryologic origin of these vessels may be responsible for varying susceptibility to atherosclerosis and aneurysm formation, respectively [24].

This review aims to describe inflammatory pathomechanisms implicated in atherosclerotic processes of the macro- and microvasculature, their determinants and implications for interactions with the endothelium, leukocytes and non-cellular components involved in vascular homeostasis. In addition, therapeutic applications of anti-inflammatory concepts for the management of PAD are discussed.

2. Pathophysiology

2.1. Inflammation and Endothelial Dysfunction

Endothelial and vessel homeostasis is to a wide extent ensured by an intact glycocalyx coverage [26]. The endothelial glycocalyx is located at the luminal side of the cells and consists of membrane-bound proteoglycans and, together with adsorbed proteins, forms the endothelial surface layer [27]. Its components exert significant influence on the interactions between the blood and the endothelium, including rolling and diapedesis of leukocytes [28], platelet adhesion and activation [29], interaction with pro-coagulatory proteins [27], endothelial permeability [30] and the regulation of vascular tone [31].

Dysfunction and degradation of the endothelial glycocalyx allows low-density lipoproteins (LDLs) to accumulate in the endothelial wall [32]. Following aggregation, LDL is oxidised (oxLDL) and subsequently phagocytosed by macrophages, which transform into foam cells and thereby initiate the progressive process of atherogenesis [32]. In turn, the integrity of the endothelial glycocalyx is disturbed by vascular inflammation, there-

fore creating a vicious cycle of endothelial dysfunction, inflammation and progression of atherosclerosis [33].

The components of the glycocalyx also play a major role in the modulation of thromboinflammatory pathways [9,34]. Importantly, the glycocalyx barrier does not only cover endothelial cells, but functions as a protective barrier exhibiting steric and charge hindrance on blood components such as macrophages, erythrocytes, microspheres, tumour cells and microbes [35,36]. Similarly, neutrophils have been demonstrated to express syndecan-1 and syndecan-4, hyaluronan, srglycin and cluster of differentiation (CD) 44 in their surface layer [37]. These molecules are essential components of both the endothelial and the neutrophil surface layers and are thought to regulate neutrophil rolling and recruitment [37]. Modifications to the neutrophil surface layer, including shedding of the glycocalyx and formation of microvilli, are thought to regulate leukocyte behaviour by exposing receptor proteins and promoting leukocyte activation [36,38]. However, the exact interactions of the endothelial and the neutrophil surface layers remain to be completely elucidated [37].

Macrophage activation after phagocytosis may lead to macrophage extracellular trap (MET) formation, but the process might be dependent on the recognized pathogen [39,40]. On the other hand, inflammation triggers leukocyte activation, promoting neutrophil– and monocyte–platelet aggregate formation [41,42]. The process is perpetuated by ETosis and enhanced oxidative stress [43–45].

Moreover, activated platelets lead to a thrombin burst; thrombin is the strongest platelet agonist, mediating platelet activation via protease-activated receptors (PARs) 1 and 4 at subnanomolar concentrations [46]. Platelet aggregation through these pathways has been shown to be preserved despite adequate dual P2Y₁₂ inhibition in patients with acute coronary syndromes [47]. Moreover, thrombin also activates platelets via glycoprotein Ib [48]. Further, inflammation mediates platelet activation through other alternative signalling pathways, including damage-associated signalling through TLRs [34]. Human platelets express all 10 TLR receptors [49], and related inflammatory signalling leads, amongst others, to P-selectin expression, ROS formation and enhanced platelet–neutrophil contacts [50]. Moreover, TLR-induced endothelial activation results in endothelial dysfunction [51]. The complex interplay of TLR receptor signalling pathways leads through signalling cascades via toll-interleukin-1 receptor resistance (TIR) domain-containing adaptor proteins to gene expression altering via different transcription factors, such as nuclear factor-κB (NF-κB), activator protein 1 (AP-1), nuclear factor erythroid-2-related factor 2 (NRF2), activating transcription factor 2 (ATF2) and interferon regulatory factors (IRFs) [34]. In humans, there are five TIR adaptors, namely the myeloid differentiation primary response protein 88 (MyD88), TIR domain-containing adaptor protein (TIRAP), TRIF, TRIF-related adaptor molecule (TRAM) and TIR domain sterile alpha and HEAT/Armadillo motif (SARM) [34,52–55].

All human TLRs signal via MyD88 to mediate inflammatory cytokine production [56,57]. However, NF-κB and the IRFs can be activated via MyD88-dependent, as well as MyD88-independent, pathways [53,58,59]. TLR-induced NF-κB activation modulates the NLRP3 inflammasome, which is a major mediator of IL-1 family cytokine production [60,61]. NLRP3 activation is directly involved in endothelial dysfunction, and enhanced expression was found in the serum of PAD patients [62,63].

TLR-4-mediated signalling in platelets, neutrophils and macrophages also contributes to neutrophil extracellular trap (NET) and MET formation, respectively [64–67].

Some risk factors commonly associated with atherosclerosis and thromboembolic events are also thought to impair the integrity of the glycocalyx [33]. Chronic diseases, such as diabetes mellitus (DM) and chronic kidney disease, are often linked to inflammatory processes and promote glycocalyx disturbance [33,68–74].

Several pathophysiologic properties link atherosclerosis and DM [75]. First, DM-associated dyslipidaemia leads to increased triglyceride-rich lipoproteins (TLP) in serum [75]. Under physiologic circumstances, insulin regulates hepatic lipoprotein and triglyceride production, however, in DM, these regulatory properties are diminished due to hepatic in-

sulin resistance [75]. It has been demonstrated that not only the prevalence of lipoproteins, but also their modifications, can be considered essential for atherogenesis [76]. In a murine model of DM, an injection of LDL from diabetic patients resulted in a fourfold increase in arterial wall LDL retention compared to injected LDL from clinically healthy, non-diabetic control subjects [76].

Advanced glycation end-products (AGEs) are formed in patients with prolonged hyperglycaemia by non-enzymatic post-translational modification of proteins, lipids and nucleic acids [77]. AGEs promote inflammation by facilitating the activation of the endothelium, increasing cytokine release from macrophages, and ultimately, enhancing ROS production [10]. The latter are also key in I/R injury in PAD and contribute to inflammatory processes and endothelial dysfunction [10]. Ischaemia leads to succinate accumulation due to impaired mitochondrial citric acid cycle (TCA) [78,79]. Succinate can be transported to the cytosol, where, due to its excess, it leads to prolyl hydroxylase activity impairment and, in turn, to the stabilization and activation of the transcription factor hypoxia-inducible factor 1 (HIF-1) α . This pathway results in the expression of IL-1 β [80]. In addition, succinate accumulation is a hallmark of macrophage polarisation, occurring in the pro-inflammatory M1 macrophages [81].

Reperfusion leads to rapid reoxidation of succinate by succinate dehydrogenase, driving extensive ROS generation [82]. During I/R injury, NO bioavailability is decreased, and ROS activate the nucleotide oligomerization domain, leucine-rich repeat, and pyrin domain-containing protein 3 (NLRP3) inflammasomes, promote mitochondrial fission and endothelial microvesicle release, and change connexin/pannexin signalling [11]. As a result of the oxidative stress, I/R impairs capillary perfusion [11]. Furthermore, reduced NO levels promote M1 polarisation [83].

CVD including PAD is further linked to a reduced endothelial progenitor cell (EPC) number [84]. The inflammatory processes induced by uncontrolled oxidative stress also modify EPC function and thus impair endothelial regenerative potential [85]. In response to ischaemia, EPC release has been demonstrated to be markedly decreased in patients with PAD compared to healthy control subjects [86]. After mobilization, EPCs were shown to home to ischaemic tissue, facilitated by vascular growth factor (VEGF) and stromal cell-derived factor 1 (SDF-1) [87]. The latter binds to C-X-C chemokine receptor type 4 (CXCR-4) on EPCs [88].

EPCs have been shown to express gene transcripts coding for TLR 1–6, including the TLR-4 co-receptor CD14, TLR 8–10 and the TLR adaptor molecule myeloid differentiation factor 88 (MyD88) [89]. Hence, during inflammation, EPCs might also be modulated by TLR signalling pathways, such as TLR-4 mediated caspase 3 signalling promoting EPC apoptosis [85,90]. In addition, ROS formation triggers extracellular trap formation by different cells of the immune system such as neutrophils, eosinophiles, macrophages and mast cells, hereby influencing coagulability and vascular perfusion [34,91].

Coronavirus disease 2019 (COVID-19), which increases the risk of thromboembolic events during and after the infection [92], is also thought to impair the regular functioning of the glycocalyx [93,94]. The degradation of the glycocalyx is mediated by a complex interaction of cellular and non-cellular factors but is mainly driven by infection of endothelial cells by severe acute respiratory distress syndrome coronavirus type 2 (SARS-CoV-2) [95]. Subsequent endothelial inflammation and damage leads to disintegration of the glycocalyx, collagen exposure and, thereupon, activation of leukocytes and platelets [96]. These processes are thought to lead to an environment of thromboinflammation, which may ultimately trigger atherogenic processes and promote organ dysfunction [9,94].

2.2. Microparticles

Microparticles (MP) are cell-membrane-derived vesicles which are shed by, among others, endothelial cells, leukocytes, monocytes and platelets [97] at an increased rate upon cell activation due to oxidative injury, shear stress and apoptosis [98]. MPs can carry a plethora of cell-specific proteins and molecules such as receptors, lipids and both

mitochondrial desoxyribonucleic acid (DNA) and messenger ribonucleic acid (mRNA) [97]. MPs are thought to contribute to cell–cell communication as their surface is representative of the originator cell [97,99,100]. Novel diagnostic and therapeutic applications are currently under investigation and first results seem promising [101]. MP composition has been demonstrated to be altered in inflammatory conditions, where endothelial cells stimulated with tumour necrosis factor (TNF)- α secrete MPs rich in pro-inflammatory cytokines and chemokines [102]. Intercellular signalling via MPs is therefore considered to exert a significant regulatory role in vascular homeostasis [103,104].

Under physiologic conditions, endothelial nitric oxide (NO) synthetase maintains vascular homeostasis by regulation of vascular tone and inhibition of platelet function through NO [105]. In addition, NO promotes anti-inflammatory M2 macrophage polarisation and limits the pro-inflammatory M1 phenotype [83]. In conditions associated with CVD, e.g., hypertension, tobacco abuse and dyslipidaemia, the endothelial production of NO is drastically reduced, leading to increased platelet activation and leukocyte diapedesis [105–107].

As described above, endothelial dysfunction is generally considered the earliest stage of atherogenesis [108]. While at physiological levels, ROS serve as signalling molecules with effects on cell differentiation, growth and apoptosis, in higher levels, their ability to oxidise various molecules results in cellular dysfunction and inflammation [109,110]. Under normal conditions, ROS are generated by mitochondria in the course of the electron transport chain, by xanthin oxidase and uncoupled endothelial NO synthetase (eNOS), and by nicotinamide adenine dinucleotide phosphate (NADPH) oxidase [110]. The latter is especially important as a source of ROS in host-defence responses and inflammation [109].

ROS trigger a range of cellular responses, which include the activation of the NLRP3 inflammasome and consecutive IL-1 β activation, the inhibition of eNOS via peroxisome proliferator-activated receptor (PPAR)- γ and adenosine-monophosphate-kinase (AMPK), and increased expression of adhesion molecules and several pro-inflammatory cytokines [110–112]. ROS have also been demonstrated to activate the TLR-4-mediated NF- κ B signalling pathway [113] and, therefore, stimulate further ROS formation [110,114]. In addition, MPs also bind to TLR-4, and can induce NLRP3 inflammasome activation and IL-1 β expression through phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt) signalling [115]. By binding to TLR-4 on platelets, MPs also contribute to platelet activation [116].

Furthermore, MPs can aggravate ROS production by expressing NADPH oxidase [117,118], potentially creating a vicious cycle of self-sustained pro-atherogenic stimuli. Importantly, MPs can not only induce the release of pro-inflammatory cytokines and ROS, but can in fact act as a vehicle of transfer between donor and recipient cells, conferring both pro- as well as anti-inflammatory effects [119].

In particular, MPs derived from endothelial cells (EMP) and platelets (PMP) disrupt endothelial function and impair endothelium-induced vasodilation [120,121]. Formation of EMPs has been shown to correlate with carotid artery atherosclerotic plaque size in patients recovering from stroke [122] and promote inflammation [123]. Via regulation of macrophage functions, adipose-tissue-derived MPs facilitate foam cell formation, herein being central in the progression of atherosclerosis [124].

Depending on the donor cell and its state of activation, MPs express and transfer specific microRNAs (miRNA), which are thought to contribute to intercellular signalling [125–129]. miRNAs are single-stranded non-coding RNAs of up to 25 nucleotides, which bind to miRNA-response elements in untranslated regions of target genes, therefore regulating gene expression [130,131]. miRNA can also be found in plasma bound to proteins, such as argonaute 2 and high-density lipoprotein (HDL) [132,133].

Signalling via miRNA has been demonstrated to exert both pro- as well as anti-atherogenic effects on target cells and to regulate vascular inflammation, the formation of a neointima following stent implantation and endothelial regeneration [125,134,135]. In the context of cigarette smoking and PAD, the downregulation of miRNA-27b is independently associated with tobacco abuse and severity of PAD [128]. Following endovascular

angioplasty and stent implantation for PAD, miRNA-195 has been found to predict adverse ischaemic events and the need for target vessel revascularisation [134].

In another study, miRNA-30c-5p was shown to inversely correlate with levels of LDL and plaque development, while miRNA-30c-5p expression in MPs was inhibited via the scavenger receptor CD36 by oxLDL and, in turn, modulated macrophage IL-1 β release, caspase 3 and apoptosis [127]. Furthermore, miRNA-21 and miRNA-126 have also been independently associated with monocyte–platelet aggregate formation in acute coronary syndrome patients *in vivo*, as well as after TLR 1/2 activation [136]. In patients with CAD, MP miRNA enrichment and function was demonstrated to be impaired, which may contribute to disease progression [137]. Conversely, in an animal model of atherosclerosis, the incorporation of MPs of healthy controls resulted in improved EPC function due to miRNA transfer (miRNA-10a, miRNA-21, miRNA-126, miRNA-146a and miRNA-223) [138].

2.3. Neutrophil Extracellular Traps

Neutrophil extracellular traps (NETs)—web-like structures consisting of cell-free DNA—are extruded from neutrophils upon activation during inflammatory processes and consist of chromatin, histones and neutrophil granule proteins [139,140]. Previously, NETosis, which describes the process of neutrophils releasing NETs, was primarily regarded as a mechanism of the innate immune system to engulf and neutralise a wide range of extracellular pathogens including bacteria [139], viruses [141] and fungi [141]. However, NETosis is suggested to play a crucial role in inflammatory diseases including vasculitis [142], atherosclerosis and thrombosis [143].

There is increasing evidence that NETs contribute to endothelial dysfunction [144,145], glycocalyx degradation [9] and atherosclerosis [143] by generation of ROS and concomitant release of neutrophil granule proteins associated with atherogenesis, including neutrophil elastase and myeloperoxidase [146,147]. Vice versa, both enzymes also play a crucial role in the induction of NETosis [148,149]. Moreover, ROS stimulate the formation of pro-inflammatory MPs [150].

In atherosclerosis, oxLDL is also a potent stimulus for NET formation. Awasthi et al. have shown that incubation of neutrophils with oxLDL leads to NETosis in a time- and concentration-dependent manner [151]. OxLDL is likely to induce NETosis via TLR-2 and TLR-6, as their blockade resulted in significantly reduced NETosis [151]. Furthermore, the recognition of NETs promotes the production of an IL-1 β precursor in macrophages and the subsequent release of mature IL-1 β upon phagocytosis of oxLDL [152]. This, in turn, causes IL-17 production from T-cells [152]; IL-17 is a potent chemokine perpetuating the pro-atherogenic inflammatory environment [152]. In addition, oxidative stress induced by NET-associated enzymes, including myeloperoxidase and NO synthetase, is considered to promote oxidation of HDL, therefore rendering this inherently anti-atherosclerotic protein dysfunctional [153].

From a clinical perspective, NETs also offer relevant insight into the mechanisms of atherothrombosis [154,155]. Activated neutrophils and NETs were detected in about 90% of thrombi from patients with acute myocardial infarction and NET load correlated with infarct size and resolution of ST-segment elevation [155].

2.4. The Role of Inflammation in Aneurysm Formation

The most common location of aortic aneurysms is the infrarenal segment of the abdominal aorta [156]. While often asymptomatic, abdominal aortic aneurysms (AAA) are associated with significant mortality. In the UK, ruptured AAAs account for 7.5 and 3.7 deaths per 100,000 for men and women, respectively, while in the Mediterranean, these numbers are closer to 1.0–2.8 per 100,000 per year [157].

The presence of leukocytes [158,159], enzymes degrading ECM in the aortic wall [160,161] and excessive levels of inflammatory parameters [25] have been reported hallmarks of aneurysm formation. The risk factors associated with aneurysm formation are similar

to those for atherosclerosis, namely, among others, male sex, dyslipidaemia and tobacco use [162,163].

While DM is a common risk factor for atherogenesis [22], it is associated with a reduction of morbidity due to AAA by almost a third [164]. DM enhances atherosclerosis progression and vascular calcification [165,166]. The latter accounts for a higher cardiovascular risk and higher mortality in diabetic patients and those with chronic kidney disease [167,168].

The observed survival benefit in diabetic patients with AAA is not yet fully elucidated and may be attributed towards DM itself or concomitant metformin therapy [169], as randomised placebo-controlled trials investigating metformin-repurposing for the prevention of AAA formation and enlargement are still ongoing [170–172]. Furthermore, increased vascular calcification is linked to aortic aneurysmal wall stabilization and slower AAA progression [173].

The estimated rate of comorbidity of atherosclerosis and aneurysm formation is about 27–53% [174,175]. Atherosclerosis and aneurysm formation are both increasingly regarded as inflammatory diseases, as leukocyte and platelet activation is a key factor for the pathogenesis of both disease entities [176–178]. AAA pathogenesis is characterised by infiltration of the aortic wall by neutrophils, macrophages and lymphocytes [179]. Subsequently, secreted enzymes, proteases and cytokines lead to ECM degradation, e.g., of collagen and elastin fibres, and an increased rate of apoptosis of smooth muscle cells promoting destruction and dilation of the vessel wall [180].

Macrophages are thought to play a decisive role in AAA formation [178]. Accumulation of macrophages during aneurysm formation can be observed in all three layers of the vessel wall but is particularly pronounced in the adventitia and the intraluminal thrombus (ILT) [181,182]. While the role of different subsets of macrophages in the stages of AAA development is not yet fully elucidated, it is hypothesised that bone-marrow-derived macrophages extravasate into the aortic wall and contribute to inflammatory processes and early stages of AAA formation [178].

The recruitment of monocytes into the aortic wall has been shown to be largely dependent on monocyte chemotactic protein 1 (MCP-1) and IL-6 produced by aortic adventitial fibroblasts [183]. Tieu et al. have shown that recruited monocytes locally mature into macrophages, which in turn stimulate the activation of adjacent fibroblasts and the release of further pro-inflammatory cytokines, forming a vicious circle of macrophage–fibroblast activation [183,184].

The pathways involved in AAA monocyte recruitment are also thought to play a decisive role in atherogenesis [185]. The infusion of angiotensin 2 in an apolipoprotein-E-deficient mouse model prone to atherosclerosis was not only shown to increase the severity of atherosclerotic lesions, but also promote AAA formation [186]. Upon stimulation by angiotensin 2, aortic adventitial fibroblasts release MCP-1 and IL-6, which cause monocyte recruitment and differentiation, and cytokine release [183,184].

The chemokine receptor 2 (CCR-2) signal, which is induced by MCP-1, plays a central role in various inflammatory diseases, including cancer and CVD [187]. Tieu et al. have demonstrated that the knock-out of CCR-2 resulted in significantly reduced adventitial fibroblast proliferation in a murine model of AAA formation [183]. Conversely, the transfer of CCR-2 positive monocytes resulted in restored proliferation and restored AAA formation [183]. The MCP-1/CCR-2 axis is thought to be crucial to the initiation of atherogenesis by promoting monocyte accumulation in atherosclerotic lesions [183,184]. In addition, levels of MCP-1/CCR-2 expression are associated with plaque vulnerability [188].

The activation of TLR-2 and TLR-4 and their downstream signalling pathways, including, among others, MyD88, NF- κ B, and mitogen-activated protein kinase, is also considered a relevant driver of both aneurysm formation and atherosclerosis [34,189,190]. As a consequence, inhibition of the TLR-4/MyD88/NF- κ B pathway by statins conveys anti-inflammatory and anti-atherosclerotic properties [21].

Neutrophils are considered to be both regulators and effector cells of inflammation [191]. In the context of AAA formation, activated neutrophils contribute to chronic inflammation, mainly by releasing ROS, NETs, histones and neutrophil granule proteins [192–194].

The formation of an ILT is frequently observed in progressive AAA and a risk factor for AAA rupture [195,196]. An ILT with concomitant platelet activation contributes to inflammation, vessel remodelling and ECM degradation [197]. Platelets activated in the context of ILT formation secrete pro-inflammatory cytokines and chemokines, which in turn stimulate leukocyte recruitment, activation and, ultimately, AAA progression [196–199].

Klopf et al. have reviewed various parameters including neutrophil-derived markers of inflammation, e.g., gelatinase-associated lipocalin [200,201], neutrophil elastase [202], myeloperoxidase [203,204], MMP [205] and NETs [206], as potential biomarkers for prognosis in AAA [25]. While the exact mechanisms which lead to aortic wall inflammation and leukocyte recruitment are not yet fully elucidated, these findings illustrate the involved processes and may help establish a better understanding of both factors determining prognosis and potential new therapeutic targets in AAA [25].

Importantly, inflammatory processes evoked by different infections, e.g., *Porphyromonas gingivalis*, *Epstein–Barr virus*, *cytomegalovirus* or *papillomavirus*, are also being discussed as potential promoters of local inflammation and risk factors for aneurysm formation [207,208]. In fact, the presence of periodontal disease, mainly with *Porphyromonas gingivalis* [209], and the occurrence of periodontal bacteria in the bloodstream or in the vascular lesion is associated with AAA formation [210–212]. In patients with AAA, *cytomegalovirus* was detected about five times as often as in healthy volunteers and was associated with increased levels of pro-inflammatory TNF- α and higher rates of arterial hypertension and CAD [208,213].

In addition to inflammatory conditions, aneurysms may also occur on the basis of pathogenic gene variants [214]. The variants best established generally concern structural proteins, e.g., procollagen type III α 1, transforming growth factor β and fibrillin 1, as seen in Marfan syndrome [214].

2.5. Vasculitis

Vasculitides are a group of rare diseases characterised by auto-immune inflammation of blood vessels of various sizes [215]. The introduction of targeted immuno-modulatory agents has improved prognosis and reduced mortality due to exacerbated vasculitis or infection drastically [216]. In patients with vasculitis, CVD is now the most common cause of death [217,218]. In addition, a chronic inflammatory state is independently associated with long-term mortality in patients with Raynaud's phenomenon [219].

An acceleration of atherogenesis in patients with AAV has been previously demonstrated [220], and surrogate markers of endothelial dysfunction, e.g., endothelium-dependent dilation of the brachial artery or pulse-wave velocity, are increased in the context of AAV [221]. One study evaluated atherosclerotic plaque burden by means of ultrasound and found that, compared to a healthy control cohort, AAV patients had a significantly higher plaque burden in the abdominal aorta and the carotid and the femoral arteries [220]. It may be hypothesised that a continuous sub-clinical inflammatory state contributes to the acceleration of atherogenesis in these patients [222]. The shedding of the endothelial glycocalyx, endothelial dysfunction [223] with enhanced expression of leukocyte adhesion factors, and leukocyte-diapedesis into the vessel wall promote a pro-inflammatory and pro-coagulatory state [224,225]. Furthermore, risk factors commonly associated with atherosclerosis are more prevalent in patients with AAV [223,226].

However, it must be noted that solid evidence of accelerated atherosclerosis in vasculitides has thus far only been established for Kawasaki's disease, Takayasu's arteritis and, most prominently, AAV [222].

Despite advances in immuno-modulatory therapy and the application of novel biologic disease-modifying drugs, glucocorticoids are still frequently used for induction therapy and are associated with significant toxicity. Traditional risk factors for atherosclerosis, i.e.,

hypertension, hyperglycaemia and dyslipidaemia, are exacerbated in patients with frequent glucocorticoid intake [227]. Risk factor management for the prevention of cardiovascular events in these high-risk patients has been shown to be insufficient in many patients [14,228]. Even with advanced biologics, e.g., Janus kinase inhibitors, undesired cardiovascular effects may occur [229,230]. Evidence is conflicting and therapeutic benefits may depend on the specific disease entity [231,232].

2.6. The Influence of Inflammation on Angiogenesis, Arteriogenesis, and Collateralisation

While inflammation is generally regarded as deleterious in PAD, specific inflammatory pathways involved in atherogenesis also participate in tissue regeneration, angi- and arteriogenesis [233,234]. Angiogenesis is the process of the formation of new capillaries for improved tissue perfusion, while arteriogenesis describes the transformation of arterio-arteriolar anastomoses to fully functional collateral arteries [235]. Therefore, while angiogenesis primarily involves endothelial cells, arteriogenesis necessitates the proliferation, migration and transformation of vascular smooth muscle cells [235], the latter being promoted by inflammatory conditions [233].

Angiogenesis is induced by various cytokines, e.g., vascular growth factor (VEGF), fibroblast growth factor (FGF) and angiopoietin, and is regulated via HIF-1 [87,236]. The molecular pathways which lead to arteriogenesis additionally include a response to increased shear stress and blood flow in arterio-arteriolar anastomoses and require the recruitment of macrophages [87].

The recruitment of macrophages is regulated via intercellular adhesion molecule 1 (ICAM-1) and CCR-2 signalling and promoted by granulocyte-colony stimulating factor (G-CSF) and granulocyte macrophage-colony stimulating factor (GM-CSF) [87,237,238]. While the M1 macrophage population is largely responsible for tissue damage associated with inflammation, alternatively activated M2 macrophages modulate cell proliferation and transition and are involved in tissue regeneration by secretion of growth factors (VEGF, FGF), MMPs and NO [87,239]. However, pro-inflammatory M1 macrophages are especially considered crucial sources of VEGF-A in arteriogenesis [234]. In this context, inflammatory M1 macrophages upregulate the transcription of the pro-angiogenic VEGF-A isoform via autocrine IL-1 β -mediated activation of NF- κ B and signal transducer and activator of transcription 3 (STAT3) [234]. Conversely, in an IL-1 β knock-out mouse model, VEGF-transcription depending on HIF-1 alone was markedly decreased in comparison to a wild-type IL-1 β cohort, where VEGF transcription is promoted by both HIF-1 and IL-1 β -dependent pathways [234,240,241] (Table 1).

Table 1. Triggers of inflammation in peripheral artery disease.

	Triggers	Involved Pathways	Resultant Effects
Endothelial dysfunction	glycocalyx degradation [26] ROS [11] ETs [144,145]	eNOS ↓ [105] TLR/MyD88/MAPK/NF- κ B ↑ [34] thrombin/PAR-1 and 4 ↑ [46]	endothelial permeability ↑ [30] leukocyte rolling and diapedesis ↑ [28,107] platelet adhesion and activation ↑ [29] binding of anticoagulant mediators ↓ [27] endothelium-induced vasodilation ↓ [120,121] macrophage M1 polarisation ↑ [83]
(Oxidised) LDL accumulation	endothelial dysfunction and glycocalyx degradation [32] hyperlipidaemia [3]	scavenger receptor A [242,243] TLR-2 and -6/MyD88/MAPK/NF- κ B ↑ [151] NLRP3/IL-1 β ↑ [61,152] PKC/IRAK/MAPK ↑ [151]	NET formation ↑ [151] ROS ↑ [151] endothelial dysfunction ↑ [61] SMC proliferation ↑ [61] leukocyte recruitment ↑ [61] MCP-1 ↑ [61]

Table 1. Cont.

	Triggers	Involved Pathways	Resultant Effects
Oxidative stress and ROS	I/R injury [10] microparticles [120,121] NETs [146,147] NF-κB [244] AGE [10]	NF-κB ↑ [244] PPAR γ /AMPK/eNOS ↓ [245–247] NLRP3/IL-1 β ↑ [11] adiponectin ↓ [247–250]	endothelial dysfunction ↑ [11,110] ET and MP formation ↑ [91,150] SMC proliferation ↑ [251] ICAM-1, VCAM-1 ↑ [252]
Ischaemia and reperfusion	Impaired perfusion and resolution	TCA dysfunction with succinate accumulation [79] HIF-1 α /IL-1 β ↑ [80] NO bioavailability ↓ [78] ROS ↑ [82] EPC function ↓ [85]	ROS ↑ [11] macrophage M1 polarisation ↑ [83] microvascular perfusion ↓ [11] regenerative potential ↓ [85]
TLR activation	oxLDL [151] DAMPs (oxLDL, HSP, fibronectin, fibrinogen, hyaluronic acid derivate) [34,189] PAMPs (bacterial, viral and fungal) [34,189]	caspase 3 ↑ [85,90] TLR/MyD88/MAPK/NF-κB ↑ [34] NLRP3/IL-1 β ↑ [34]	EPC apoptosis ↑ [85,90] endothelial dysfunction ↑ [61] SMC proliferation ↑ [61] leukocyte recruitment ↑ [61] MCP-1 ↑ [61] aneurysm formation ↑ [253]

Abbreviations: AGE, advanced glycation end-products; AMPK, adenosine-monophosphate-kinase; DAMPs, danger-associated molecular patterns; eNOS, endothelial nitric oxide synthetase; EPC, endothelial progenitor cell; ET, extracellular trap; HIF-1 α , hypoxia-inducible factor 1 α ; HSP, heat-shock protein; I/R, ischaemia-reperfusion injury; ICAM-1, intercellular adhesion molecule 1; IL-1 β , interleukin 1 β ; IRAK, interleukin 1 receptor-associated kinase; MAPK, mitogen-activated protein kinase; MCP-1, monocyte chemoattractant protein 1; MP, microparticle; MyD88, myeloid differentiation factor 88; NETs, neutrophil extracellular traps; NF-κB, nuclear factor kappa B; NLRP3, nucleotide oligomerization domain, leucine-rich repeat, and pyrin domain-containing protein 3; oxLDL, oxidised low-density lipoprotein; PAMPs, pathogen-associated molecular patterns; PAR, protease-activated receptor; PKC, protein kinase C; PPAR γ , peroxisome proliferator-activated receptor γ ; ROS, reactive-oxygen species; SMC, smooth muscle cell; TCA, citric acid cycle; TLR, toll-like receptor; VCAM-1, vascular adhesion molecule 1; ↑, increase in expression or function; ↓ decrease in expression or function. Please note that TLR-signalling involved in immunothrombosis is complex and has many branching paths; therefore, we only depicted the predominantly used pathway [34].

3. Current and Novel Therapeutic Targets and Strategies

Current guidelines in PAD emphasise metabolic risk management, including reduction of LDL levels, antiplatelet therapy, management of hypertension, glycaemic control, smoking cessation and physical activity [254–258]. While some of these interventions also exert a positive influence on systemic and local levels of inflammation [20,259,260], therapeutic strategies which directly intercept pro-inflammatory signalling pathways may be promising and are only beginning to be established [18]. In the following, we highlight some of the anti-inflammatory drugs and concepts. An overview of current and novel therapeutic targets and strategies is provided in Table 2.

3.1. Statins

The management of dyslipidaemia and, specifically, reduction of elevated levels of LDL has been a cornerstone of preventive cardiovascular medicine for many years. Statins, which inhibit hepatic 3-hydroxy-3-methylglutaryl-coenzyme-A reductase, and therefore impair cholesterol synthesis [261], are first line agents in treatment of CVD [18,254].

However, there is accumulating evidence that the positive effects of statins on atherosclerosis go beyond LDL reduction [262]. In fact, statin therapy has been demonstrated to result in increased endothelial biosynthesis of NO with a positive effect on vascular tone and platelet aggregation, and even plaque stabilisation or regression [262]. It has been suggested that statins also interfere with various endothelial adhesion molecules and therefore reduce leukocyte transmigration [263]. In patients with AAA, the anti-inflammatory properties of simvastatin treatment were shown by reduced TNF- α as well as cyclosporine A levels and a decreased amount of phosphorylated extracellular-signal regulated kinases

(ERK) 1/2 [264,265]. Furthermore, a significant difference in the concentration of MMPs and their inhibitors was observed in aneurysmal wall tissue and ILT [266]. In addition, simvastatin reduced monocyte tissue factor expression in response to LPS treatment in healthy volunteers [267].

The anti-inflammatory effects of statins also include the reduction of c-reactive protein (CRP) concentration regardless of LDL levels [268]. Utilising fluorodeoxyglucose-positron emission tomography and computed tomography imaging, Tawakol et al. demonstrated a dose-dependent anti-inflammatory effect of atorvastatin in patients with suspected or proven atherosclerosis [269]. In a recent meta-analysis of three randomised controlled trials in patients receiving statins, inflammation, as assessed by CRP, was a stronger predictor than LDL for cardiovascular events and death [270].

Beyond CVD, the anti-inflammatory effects of statin therapy have also been demonstrated in other diseases. In chronic kidney disease, statins have resulted in lower levels of CRP [271], and in asthma, statins reduced both symptoms and biomarkers of inflammation [272].

3.2. Colchicine

Colchicine has been used for centuries for the treatment of inflammatory diseases, including gout and familial Mediterranean fever [17]. The pharmacodynamics of colchicine are complex, and they exert multiple effects on cellular signal transduction [17]. Colchicine has been demonstrated to reduce neutrophil chemotaxis by inhibition of the polymerisation of tubulin [273], reduce the expression of TNF- α [17,274], and attenuate the exocytosis of neutrophil granules [17,275]. Though less well elucidated, inhibitory effects of colchicine on the NLRP3 inflammasome have been observed and may inhibit the proliferation of smooth muscle cells as seen in the context of atherosclerosis [276].

The discovery of colchicine's pleiotropic effects on inflammation and atherosclerosis [277,278] have led to the initiation of the phase III randomised placebo-controlled Colchicine Cardiovascular Outcomes Trial (COLCOT) [279]. Therein, 4745 patients who have suffered from myocardial infarction within the previous 30 months were randomised to receive either 0.5 mg of colchicine or placebo. The risk for the primary endpoints of cardiovascular death and serious cardiovascular events was reduced significantly in the colchicine group, with a hazard ratio (HR) of 0.77 [279].

Based on these findings, the 2021 guidelines on CVD prevention by the European Society of Cardiology have now included a class IIb, level A recommendation to consider low-dose colchicine in secondary prevention of CVD [18].

In the future, colchicine may also be applied in the context of acute myocardial infarction. A recent study by Wang et al. showed that an infusion of colchicine-loaded nanoparticles subsequent to myocardial infarction reduced inflammation and myocardial infarct size by 45% on average [280]. These findings highlight the potential of colchicine as a promising anti-inflammatory agent in CVD, both in the acute and chronic settings [281].

3.3. Eicosapentaenoic Acid Ethyl Ester

Eicosapentaenoic acid ethyl ester and its purified prescription form icosapent ethyl (IPE) is an omega-3 fatty acid and has demonstrated several anti-inflammatory and anti-atherosclerotic properties [282]. In a large, randomised placebo-controlled trial in patients with established CVD or several risk factors for the development of CVD and elevated triglyceride levels, the addition of IPE to standard statin treatment resulted in a highly significant risk reduction (HR 0.75) for ischaemic events and cardiovascular death when compared to placebo [283]. Furthermore, Budoff et al. documented a significant reduction in plaque size in patients with established CAD who received IPE when compared to a control group [284].

The exact mechanisms leading to these results remain to be established, especially as allocation to the IPE cohort in the Reduction of Cardiovascular Events with EPA—Intervention Trial (REDUCE-IT) did not result in a relevant reduction of inflammatory parameters [283].

In other trials, however, a high-sensitivity CRP and lipoprotein-associated phospholipase A2 lowering effect has been documented [282].

It is hypothesised that protective effects with regard to CVD may be attributed to the production of the bioactive IPE metabolites thromboxane A3 and prostacyclin, which exert antithrombotic influence on platelets and promote endothelial vasodilation [285]. In addition, IPE integration in cellular membranes also seems to have a biophysical anti-atherogenic effect [285].

The reduction in TLP, which is observed under large doses of IPE, is also considered to have protective effects in CVD [286]. As these predominantly transport saturated fatty acids, which are thought to promote activation of the NLRP3 inflammasome, the reduction of TLP levels may also attenuate atherogenesis [285,287].

3.4. Canakinumab and Anakinra

The NLRP3 inflammasome is a pro-inflammatory signalling complex with pleotropic effects on cytokine release and cleavage of pro-interleukins [61]. Its activation is mediated by pathogen- and damage-associated molecular patterns including, among others, TLR-2 and TLR-4, and results in activation of the IL-1 pathway [61]. In the context of atherosclerosis, the NLRP3 inflammasome is activated by the recognition of oxLDL and cholesterol crystals via various receptors in macrophages. The subsequent release and formation of, among others, IL-1 β results in activation of endothelial cells, promotes the expression of adhesion molecules and the proliferation of smooth muscle cells and increases the production of MCP-1 [61].

The monoclonal antibody canakinumab, which targets IL-1 β , and anakinra, an IL-1 β receptor antagonist, are applied in various immune-mediated disorders [288] and were also considered potential therapeutics in atherosclerosis [289–292]. The pivotal Antiinflammatory Therapy with Canakinumab for Atherosclerotic Disease (CANTOS) trial has demonstrated a dose-dependent effect of canakinumab on systemic levels of inflammation in patients with previous myocardial infarction [15]. In this trial, 150 mg of canakinumab every three months reduced the risk of adverse cardiovascular events (HR: 0.85) compared to placebo, independently of lipid level lowering [15].

3.5. Glucocorticoids

Glucocorticoids are analogues of endogenous cortisone and constitute a cornerstone of the treatment of various chronic inflammatory conditions [293]. Glucocorticoids exert their pleiotropic effects by binding to intracellular steroid-receptor proteins and regulate gene expression and cellular signalling [294].

However, long-term glucocorticoid excess is also associated with significant undesirable effects including hyperglycaemia [295], arterial hypertension [296], obesity [297], dyslipidaemia [298] and dysregulation of the coagulation cascade [299], all of which are considered well-established risk factors for atherosclerosis and adverse cardiovascular events [294]. In patients diagnosed with Cushing's disease, which is characterised by endogenous overproduction of cortisone, a thickened intimal-medial layer and a lower systolic carotid artery lumen diameter [300] have been observed [301]. Furthermore, the ankle-brachial pressure index is elevated in Cushing's disease [302]. In large register studies, glucocorticoid prescription has been associated with significant and dose-dependent increases in the risk for CVD [301,303–305].

3.6. Antidiabetic Drugs

Several antidiabetic drugs have been proposed to promote anti-inflammatory pathways or inhibit pathways associated with inflammation [306].

Sodium-glucose cotransporter 2 (SGLT-2) inhibitors and glucagon-like peptide 1 (GLP-1) receptor agonists were originally developed for the control of hyperglycaemia in patients with DM type 2. SGLT-2 inhibitors prevent glucose reabsorption in the proximal tubule and cause glucosuria, therefore lowering glucose levels in serum [307].

Large-scale clinical trials have demonstrated that SGLT-2 inhibitors significantly reduce the risk for hospitalisation and cardiovascular death in patients with heart failure [308–311]. Today, SGLT-2 inhibitors constitute an integral component of heart failure therapy and are recommended in the most recent heart failure guidelines [312]. While the detailed molecular mechanisms of the observed clinical benefits are largely unknown, recent studies have proposed potential anti-inflammatory properties of these substances in addition to pleiotropic, metabolic and cardiovascular effects of SGLT-2 inhibitors, which have been described in detail by Hou et al. [313,314].

In murine models, SGLT-2 inhibitors were demonstrated to reduce the expression of MCP-1 and IL-1 β [315–317]. A reduction of inflammasome activation and the subsequent release of IL-1 β was also found in patients with DM [318]. Furthermore, macrophage behaviour seems to be influenced by SGLT-2 inhibitors as increased autophagy and cholesterol efflux were observed in a murine model. Therein, the modulation of an AMPK-dependent pathway resulted in attenuated atherosclerosis [319]. Among a plethora of metabolic effects, GLP-1 receptor agonists were also found to exert a robust anti-inflammatory effect by lowering the levels of ROS generation and reducing NF- κ B activation, as well as expression of mRNA coding for, among others, TNF- α , IL-1 β , TLR-2 and TLR-4 [320,321]. In apolipoprotein-E- and LDL-receptor-deficient mice, the application of liraglutide or semaglutide resulted in decreased aortic intima thickening and inhibited plaque progression compared to a control group [322]. Semaglutide was also demonstrated to alter the expression of genes associated with inflammation and atherogenesis including IL-6, chemokine ligand 2, MMPs and proteins relevant for cholesterol metabolism [322]. In vitro, GLP-1 receptor agonists have been shown to modulate macrophage behaviour and reduce the secretion of pro-inflammatory cytokines (e.g., interferon γ , TNF- β , IL-1 β , IL-2 and IL-6), and promote the release of anti-inflammatory IL-10 [323].

Clinical trials in patients with DM type 2 showed a consistent reduction in CRP, TNF- α and malondialdehyde [324]. Cardiovascular outcomes were also improved in patients with DM type 2 receiving GLP-1 receptor agonist treatment in some, but not all, clinical trials [325]. In a retrospective cohort study of patients with PAD and DM under either SGLT-2 inhibitors, GLP-1 receptor agonists or sulfonylureas, GLP-1 receptor agonist prescription was associated with a significantly lower rate of lower limb amputation [326]. Metformin used to be considered the established first-line therapy for patients with DM type 2 for decades [327]. In recent years, the attention has been increasingly focused on the effects of metformin beyond control of hyperglycaemia [328]. Though the exact pharmacodynamic properties of metformin have yet to be fully elucidated, there is some mechanistic evidence that metformin may attenuate atherogenesis [329,330]. In vitro studies have demonstrated that metformin attenuates foam cell formation and phagocytosis of oxLDL [331]. On a molecular level, metformin administration resulted in reduced expression of the macrophage scavenger receptor A and CD36, both of which are involved in oxLDL uptake [242,243,331]. The expression of inflammatory markers, including IL-1 β , IL-18, cysteinyl aspartate specific proteinase-1, NLRP3 and ROS, was reduced in macrophages treated with metformin [331]. In a rabbit model of atherosclerosis, treatment with metformin resulted in significantly decreased burden of atherosclerotic lesions with lower macrophage content [332]. In addition to reducing plasma levels of MCP-1, CRP and TNF- α , metformin also reduced the expression of mRNA coding for vascular adhesion molecule 1 and intercellular adhesion molecule 1, therefore ameliorating adhesion of monocytes to endothelial cells [332]. Furthermore, the activation of AMPK, the inhibition of NF- κ B expression and NET formation are also considered potentially anti-atherogenic properties of metformin [329,333,334].

Though there is some evidence that metformin therapy has a positive effect on cardiovascular outcomes in patients with DM type 2 [329,330,335], the data are hitherto contradictory for non-diabetic patients [336–339]. The ongoing Glucose Lowering in Non-diabetic hyperglycaemia trial (GLINT) [340,341] may help to determine the role of metformin in prevention of CVD in these patients [339].

Dipeptidyl peptidase 4 (DPP4) inhibitors or gliptins are established second-line anti-diabetic agents which exert their effect by inhibition of proteolysis of endogenous GLP-1 and glucose-dependent insulinotropic polypeptide [342,343].

DPP4 is involved in the cleavage of chemokines and cytokines, therefore potentially exhibiting a role in cell–cell communication [344]. Furthermore, it is suggested that DPP4 induces endothelial dysfunction and promotes the expression of TLR-2 and TLR-4 and subsequent activation of inflammatory pathways [344–346]. The inhibition of DPP4 is therefore considered to attenuate inflammation and improve endothelial function, potentially by stimulation of NO synthesis and reduction of endothelin 1 expression [344,347–349]. Gliptins have also been demonstrated to reduce the expression of vascular adhesion molecules and MCP-1, TNF- α , IL-1 β and IL-6, as well as LDL- or lipopolysaccharide-induced foam cell formation, most likely due to attenuation of NF- κ B and c-Jun N-terminal kinase signalling and AMPK phosphorylation [344,347]. There is evidence that DPP4 inhibitors can increase the number of circulating EPCs [349–351] and repress the activation of the NLRP3 inflammasome [349,352]. On a systemic level, reduced hepatic production of TLP [353] and an accelerated postprandial lipid metabolism [354] have been observed under DPP4 therapy [344].

While pre-clinical data may look promising and gliptins have been demonstrated to reduce established risk factors for CVD in patients with DM type 2, including dyslipidaemia and hypertension [344], randomised controlled trials have thus far failed to demonstrate a beneficial effect beyond glycaemic control with regard to cardiovascular outcomes [343,355–358].

Table 2. Potential novel applications of established therapeutic agents.

	Standard Application	Proposed Mechanism	Clinical Effect	Selected Evidence
Statins	LDL reduction secondary prevention of CVD	NO synthesis ↑ leukocyte adhesion ↓	cardiovascular events and death ↓	Tawakol et al. [269] Ridker et al. [270]
Colchicine	gout familial Mediterranean fever	leukocyte chemotaxis ↓ TNF- α ↓ exocytosis of neutrophil granules ↓ NLRP3 activation ↓	cardiovascular events and death following MI ↓	Tardif et al. [279] Chen et al. [281]
Icosapent ethyl	no previous application	active metabolites (thromboxane A3, prostacyclin) ↑ biophysical effect on cell membranes TLP ↓	cardiovascular events and death in established CVD or risk for CVD and hypertriglyceridemia plaque progression ↓	Bhatt et al. [283] Budoff et al. [284]
Glucocorticoids	various inflammatory conditions	modulation of gene transcription	risk of CVD including CAD, PAD ↑	Pujades-Rodriguez et al. [305] Macleod et al. [301]
IL-1 β antagonists	cryopyrin-associated periodic syndromes gout familial Mediterranean fever macrophage activation syndrome recurrent pericarditis rheumatoid arthritis systemic juvenile idiopathic arthritis	endothelial activation ↓ adhesion molecule expression ↓ smooth muscle cell proliferation ↓ MCP-1 ↓	cardiovascular events and death in patients with elevated CRP and MI	Ridker et al. [15]

Table 2. Cont.

	Standard Application	Proposed Mechanism	Clinical Effect	Selected Evidence
SGLT-2 inhibitors	DM type 2	NLRP3/IL-1 β /MCP-1 pathway ↓ AMPK pathway ↑ cholesterol efflux and autophagy in macrophages ↑	hospitalization and cardiovascular death in heart failure	McMurray et al. [308] Solomon et al. [359] Packer et al. [360] Anker et al. [311]
GLP-1 receptor agonists	DM type 2	ROS generation ↓ NF-κB activation ↓ INF-γ, MMP, TNF-β, IL-1 β , IL-2, IL-6 from macrophages ↓ IL-10 ↑	CRP, TNF-α ↓ Trials inconclusive	Bethel et al. [325]
Metformin	DM type 2	oxLDL phagocytosis ↓ scavenger receptor A, CD36 ↓ NLRP3, ROS, MCP-1, CRP, TNF-α NET formation ↓ NF-κB activation ↓ AMPK pathway ↑	all-cause death in DM type 2 and atherothrombosis ↓	Roussel et al. [335] GLINT (ongoing) [340,341]
DDP4 inhibitors	DM type 2	NO synthesis ↑ endothelin 1 ↓ MCP-1, TNF-α, IL-1 β , IL-6 ↓ NF-κB activation ↓ AMPK and c-Jun N-terminal kinase pathway ↑ NLRP3 activation ↓ TLP ↓	dyslipidaemia and hypertension in patients with DM type 2 ↓ cardiovascular death, MI, stroke in patients with DM type 2 ~	Rosenstock et al. [355] Green et al. [356] Scirica et al. [357] White et al. [358]

Abbreviations: AMPK, adenosine-monophosphate-kinase; CAD, coronary artery disease; CD, cluster of differentiation; CRP, c-reactive protein; CVD, cardiovascular disease; IL, interleukin; INF, interferon; LDL, low-density lipoprotein; MCP-1, monocyte chemotactic protein 1; MI, myocardial infarction; NF-κB, nuclear factor kappa B; NLRP3, nucleotide oligomerization domain, leucine-rich repeat, and pyrin domain-containing protein 3; NO, nitric oxide; oxLDL, oxidised low-density lipoprotein; PAD, peripheral artery disease; ROS, reactive oxygen species; TLP, triglyceride-rich lipoprotein; TNF, tumour necrosis factor; ↑, increase in expression or function; ↓, decrease in expression or function; ~, no or inconclusive effect.

3.7. Antiplatelet Therapy

Current antiplatelet regimes interfere with thromboinflammatory pathways; however, platelet reactivity is, to a wide extent, also determined by alternative platelet activation pathways despite adequate guideline-driven platelet inhibition [255,361–364]. Platelet activation and the formation of platelet–leukocyte aggregates is a hallmark of inflammatory atherosclerotic processes [198].

Recently, there is increasing evidence that platelet-to-lymphocyte ratio (PLR)—a simple marker calculated from the blood count—is related to platelet activation and ischemic events in CVD [198,365–367]. Moreover, a high PLR is also related to target vessel restenosis after revascularization in PAD [368].

Platelet reactivity can be modulated by various conditions such as age [369], sex [370], HDL levels [371,372] and cytochrome P450 2C9/2C19 polymorphism [373,374], but also anaemia [375,376]. The latter is often associated with chronic inflammation and implicated in both thrombotic and bleeding events [375,377]. Moreover, iron deficiency is associated with major adverse cardiovascular and leg events in PAD, suggesting anaemia as a possible therapeutic target [378].

Another aspect relevant for pain management of PAD patients with critical ischaemia is the drug interaction of morphine or fentanyl, as a decrease in plasma levels and/or antiplatelet effects of P2Y12 inhibitors can occur [379–382]. In contrast, morphine did not exert a significant effect on aspirin-mediated platelet inhibition [383].

Platelet activation, furthermore, has a high impact on platelet metabolism and redox balance [384]; hence, attenuation of platelet reactivity may have beneficial effects on redox processes.

3.8. Attenuation of Ischaemia-Reperfusion Injury

The counter regulation of the decrease in NO bioavailability due to I/R injury is one possible therapeutic approach to minimize endothelial dysfunction. Herein, dietary supplementation of NO donors, enhancers of NO availability, NO synthase inducers and antioxidants have been studied [385,386].

Interestingly, pleiotropic effects of statins include the increase in endothelial NO synthase expression and function [387]. In patients with AAAs, simvastatin reduced lipid peroxidation level as demonstrated by lower 4-hydroxy-trans-2-nonenal concentration [265]. Moreover, simvastatin has been shown to induce heme oxygenase 1 (HO-1), an enzyme with anti-inflammatory, antioxidant, antithrombotic, pro-angiogenic and antiapoptotic properties [388–391]. Induction of HO-1 can also be achieved by heme arginate infusion, which improves reperfusion patterns during I/R injury [392–394].

Further concepts to ameliorate I/R injury include a plethora of therapies, such as blocking of intercellular adhesion molecule 1, administration of polymerised albumin, colchicine, tocilizumab, anakinra and revacept and pre-, per- and postconditioning [395–400]. Exercise-induced I/R injury in PAD can also be attenuated using cilostazol, which results in a reduced expression of P-selectin, intercellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 (VCAM-1) [401]. P-selectin and ICAM-1, as well as VCAM-1, are known to promote leukocyte recruitment to sites of inflammation and are regulated via NF- κ B [402–404]. In addition, a double-blind randomised controlled trial of cilostazol in patients with PAD has demonstrated marked effects on EPC function, which may improve collateral vessel formation [405]. The homing of EPCs can be enhanced by SDF-1, which binds to CXCR-4 and has shown to improve ischemic tissue perfusion and increase capillary density in mice [88]. However, in the STOP-PAD trial (SDF-1 plasmid Treatment for Patients with Peripheral Artery Disease; a randomized, double-blind, placebo-controlled clinical trial), the injection of JVS-100, a non-viral DNA plasmid-based therapy encoding SDF-1, did not improve hemodynamic measures or wound healing at 3 months [406]. Another important cornerstone in the therapy of risk factors in PAD patients are angiotensin-converting enzyme inhibitors, which ameliorate (micro-)vessel perfusion by increasing nitric oxide production [407]. In addition, ROS formation is, amongst others, reduced by SGLT-2 inhibitors and GLP-1 receptor agonists [320,408,409]. One of the underlying mechanisms might be the reduction of succinate levels by SGLT-2 inhibition; however, on the contrary, the GLP-1 receptor agonist liraglutide elevates succinate levels despite ameliorating mitochondrial function [410,411].

3.9. Vascular Regeneration

The interactions between molecular signalling pathways involved in both inflammation and vascular regeneration imply a potential for therapeutic modulation. Different approaches tested so far include the application of GM-CSF [412], basic FGF [413] and plasmid-based SDF-1 gene therapy [406]; however, clinical trials were either negative or terminated prematurely due to adverse events largely attributed to inflammatory reactions. Novel application systems, including peptide-loaded microgels and microspheres, are hypothesised to help overcome inflammation-associated effects [414,415]. However, so far, these have been explored in murine models only [414,415].

The homing and angiogenic function of EPCs has been demonstrated to be modulated by inflammatory processes and can be improved by the inhibition of macrophage

inflammatory protein-1 β (MIP-1 β) [416]. This is especially relevant in the context of DM, where mononuclear cells and EPCs exhibit increased MIP-1 β secretion and consecutively impaired expression of VEGF, SDF-1 α and other pro-angiogenic cytokines [416].

Moreover, the abundance of cytokines during ischaemia is suggested to create an environmental condition facilitating transdifferentiation of fibroblasts into endothelial cells [87]. This is also promoted by an increased accessibility of DNA in the context of injury and ischaemia mediated by NF- κ B activation, a process referred to as transflammation [417]. Herein, therapeutic interaction could be conferred by modification of TLR signalling pathways, as shown for TLR-3 agonism in combination with EC growth factors, that result in fibroblast transdifferentiation into endothelial cells [418].

The modulation of macrophage polarisation and behaviour is also hypothesised to be a critical determinant and potential target following endovascular procedures for PAD [419]. An intervention to boost macrophage M2 polarisation may help to support endothelial repair by the release of proangiogenic signalling molecules, i.e., basic FGF-2, VEGF-A and transforming growth factor (TGF)- β [419]. This could ultimately improve outcomes following endovascular interventions in PAD [419].

Further studies on possible therapeutic concepts for the modulation of transdifferentiation, transflammation and vascular regeneration are warranted [87].

3.10. Physical Exercise

Regular physical exercise is a cornerstone in the prevention of CVD [18]. Besides improving endothelial function by increasing circulating EPC numbers [420], low intensity aerobic training also increases capillary density in skeletal muscle [421]. Furthermore, significant positive effects on established cardiovascular risk factors, e.g., hyperglycaemia [422], hypertension [423] and dyslipidaemia [421], have been demonstrated and a reduction in systemic markers of inflammation can be observed with physical exercise [424]. These include, among others, TNF- α and CRP, as well as the expression of vascular adhesion molecules, all of which are considered to be of crucial importance in the pathogenesis of atherosclerosis [424]. In addition, protective effects of previous physical activity may also improve outcomes following cardiovascular events [425,426].

4. Discussion

PAD is increasingly regarded as an inflammatory process affecting not only the macro- but also the microvasculature [10,255,258]. Herein, modification of glycocalyx conformation, charges and density leads to endothelial dysfunction [44]. Thromboinflammatory processes involving leukocyte and platelet activation, as well as ETosis, are central pathomechanisms in plaque formation [43,44].

Altered flow conditions due to plaque formation promote further disease progression by modulation of endothelial cell metabolism [427]. Upregulated 6-phosphofructokinase/2,6-bisphosphatase 3 (PFKFB3), which is a key enzyme in endothelial glycolysis, gives an impulse for angiogenesis with immature vessel formation, thus enhancing plaque vulnerability [427–429]. Moreover, rupture of the atheroma followed by atherothrombosis may also be triggered by (N)ETosis, as neutrophil, macrophage and mast cell activation play a critical role in atherosclerotic lesions [43,430,431].

In addition, NETs were shown to contribute to fibrous vascular occlusion [154]. This may also contribute to systemic microvessel rarefaction, which was observed in PAD and other CVDs [432–435].

NET release promoting subsequent microvascular thrombosis is regarded as a hallmark of atherosclerotic processes. The interplay of platelet activation, platelet-leukocyte aggregate formation, ETosis and ROS formation perpetuates thromboinflammation, resulting in altered microvascular fluid filtration, microthrombosis and finally, tissue necrosis (compare Figure 1) [9,11].

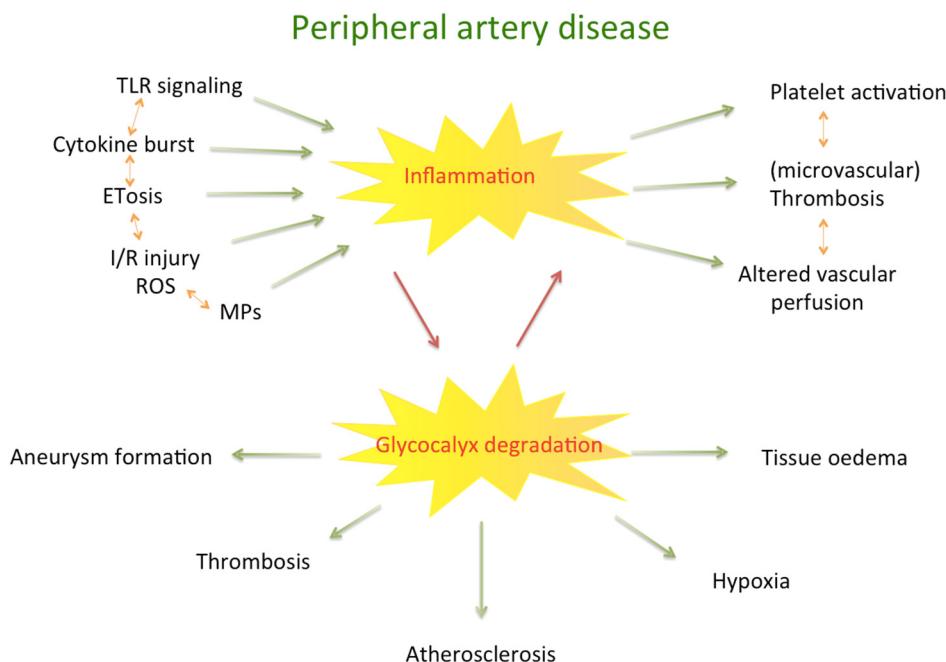


Figure 1. Pathophysiologic consequences of inflammation on the vasculature and adjacent tissue: Inflammatory processes promote endothelial dysfunction by glycocalyx degradation, leading to altered vascular homeostasis [9]. The pathological processes influence each other, perpetuating disease progression. ETosis, extracellular trap formation; I/R, ischaemia-reperfusion injury; MPs, microparticles; ROS, reactive oxygen species; TLR, toll-like receptor.

Capillary perfusion is also impaired by ROS formation during I/R injury, as it occurs during ischemic vascular diseases [11]. I/R injury also contributes to postischemic capillary no-reflow after successful arterial recanalization [11]. Attenuation of I/R injury to preserve microvascular haemodynamics [399] will be of importance for refinement of interventional PAD treatment.

Acute critical ischemia of the lower extremity often occurs in the setting of a total vessel occlusion, yet PAD patients encompass a wide spectrum of disease, including chronic (and often asymptomatic) disease courses, with progressive atherosclerosis, and development of collateral circuits [436]. In addition, it is also known that patients with lower extremity PAD are at higher risk of ischaemic events than those patients with isolated coronary artery disease [437]. Moreover, chronic inflammation, together with pro-thrombotic stimuli, re-endothelialisation, vascular smooth muscle cell migration and proliferation, as well as matrix remodelling, account for the limited patency of vascular stents and bypass grafts, herein presenting as a wider disease pathomechanism than the initial atherosclerotic lesion [438].

Inflammatory pathomechanisms might also differ during acute and chronic PAD processes. Herein, it should be emphasized that even different symptomatic PAD subpopulations, with regard to PAD severity and comorbidities, can experience a difference in therapeutic benefits [439]. The latter has been shown in the subanalysis of the COMPASS trial, where the combination therapy of aspirin and low-dose rivaroxaban conferred the highest estimated absolute risk reduction at 30 months in those patients with high-risk limb presentation or high-risk comorbidity at baseline [439]. Moreover, the results of the COMPASS trial also showed a greater reduction in major adverse limb events (MALE) in PAD patients with a high-risk limb presentation and a greater reduction in major adverse cardiac events (MACE) in PAD patients with high-risk comorbidity [439].

In the context of the SARS-CoV-2 pandemic and its long-term consequences, it should be noted that viral persistence promoting (subclinical) inflammation will have an impact on the vasculature and atherosclerosis [9,440–442]. The detection of (subclinical) inflammation

has recently been also proposed through measurements of cholinesterase levels, which are declined during inflammation and also linked to patient mortality [443,444]. In PAD patients, low levels of serum cholinesterase were associated with long-term adverse ischemic events after angioplasty and stenting of the superficial femoral artery [445]. However, more studies, in particular in comparison to established inflammatory markers, are needed.

In addition, it should be noted that inflammation does not only confer a physical limitation through atherosclerotic processes, but also impacts the patient's mood and personality functioning and can result in the development of vascular depression in patients with cardiovascular diseases [446].

Despite all pharmacotherapeutic progress, modulation of the fragile glycocalyx and, in consequence, preservation of endothelial cell function is demanding. Concepts to reduce endothelial dysfunction by interference with redox processes have hitherto only marginally been integrated into clinical practice [447]. However, different pharmacotherapies, such as statins, angiotensin converting enzyme inhibitors, GLP-1 receptor agonists or SGLT-2 inhibitors, may ameliorate inflammatory pathways [261,314,321,448]. The reduction of fibrosis and arterial stiffness may directly influence long-term pathogenesis [449].

While the therapeutic promotion of arteriogenesis and angiogenesis may be a promising concept to alleviate signs and symptoms of PAD, the complex interactions with inflammation complicate effective therapeutic applications so far. As it seems, some level of inflammation is required for effective neovascularisation; this is also observed in atherosclerotic plaques, where neovascularisation constitutes a hallmark of progression and plaque vulnerability [450].

In addition to pharmacotherapy, exercise training should remain a cornerstone in patients with stable PAD [18]. In particular, aerobic exercise training has been shown to upregulate microvessel perfusion [451]. Moreover, the number of circulating EPCs increases in patients with regular endurance training and is associated with improved endothelial function [420,452]. Therefore, future concepts should emphasize preventive strategies [18] including governmental-promoted exercise training and programs to raise awareness for cardiovascular risk factors.

One limitation of our review is the scarce evidence from randomised controlled clinical trials in patients with PAD; therefore, additional knowledge on inflammatory concepts is derived from animal studies and trials in patients with other vascular disease entities, where pathomechanisms might differ. However, where available, the literature regarding PAD is presented primarily. Moreover, in this review, we also wanted to highlight the common share of vascular diseases, including aneurysms, namely inflammatory processes leading to endothelial dysfunction. Further clinical trials are needed to give more insights into potentially unique local inflammatory pathomechanisms, which may be different throughout PAD stages and during acute and chronic ischaemic processes.

5. Conclusions

Inflammatory pathways have a critical role in the development, disease perpetuation and complications of atherosclerosis. Novel research results regarding disease pathophysiology imply the need for a paradigm shift in the therapeutic approach to atherosclerotic diseases. In the future, the attenuation of (subclinical) inflammatory processes will become equally important to other risk-factor management in the therapy of PAD. However, further studies regarding the long-lasting outcomes of PAD patients are warranted.

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References

- Roth, G.A.; Mensah, G.A.; Johnson, C.O.; Addolorato, G.; Ammirati, E.; Baddour, L.M.; Barengo, N.C.; Beaton, A.; Benjamin, E.J.; Benziger, C.P.; et al. Global Burden of Cardiovascular Diseases and Risk Factors, 1990–2019: Update From the GBD 2019 Study. *J. Am. Coll. Cardiol.* **2020**, *76*, 2982–3021. [CrossRef] [PubMed]
- Townsend, N.; Wilson, L.; Bhatnagar, P.; Wickramasinghe, K.; Rayner, M.; Nichols, M. Cardiovascular Disease in Europe: Epidemiological Update 2016. *Eur. Heart J.* **2016**, *37*, 3232–3245. [CrossRef] [PubMed]
- Woolf, N. The Pathology of Atherosclerosis with Particular Reference to the Effects of Hyperlipidaemia. *Eur. Heart J.* **1987**, *8* (Suppl. E), 3–14. [CrossRef] [PubMed]
- Hedin, U.; Matic, L.P. Recent Advances in Therapeutic Targeting of Inflammation in Atherosclerosis. *J. Vasc. Surg.* **2019**, *69*, 944–951. [CrossRef]
- Raggi, P.; Genest, J.; Giles, J.T.; Rayner, K.J.; Dwivedi, G.; Beanlands, R.S.; Gupta, M. Role of Inflammation in the Pathogenesis of Atherosclerosis and Therapeutic Interventions. *Atherosclerosis* **2018**, *276*, 98–108. [CrossRef]
- Geovanini, G.R.; Libby, P. Atherosclerosis and Inflammation: Overview and Updates. *Clin. Sci.* **2018**, *132*, 1243–1252. [CrossRef]
- Kong, P.; Cui, Z.Y.; Huang, X.F.; Zhang, D.D.; Guo, R.J.; Han, M. Inflammation and Atherosclerosis: Signaling Pathways and Therapeutic Intervention. *Signal Transduct. Target. Ther.* **2022**, *7*, 131. [CrossRef]
- Soehnlein, O.; Libby, P. Targeting Inflammation in Atherosclerosis—From Experimental Insights to the Clinic. *Nat. Rev. Drug Discov.* **2021**, *20*, 589–610. [CrossRef]
- Wadowski, P.P.; Panzer, B.; Józkowicz, A.; Kopp, C.W.; Gremmel, T.; Panzer, S.; Koppensteiner, R. Microvascular Thrombosis as a Critical Factor in Severe COVID-19. *Int. J. Mol. Sci.* **2023**, *24*, 2492. [CrossRef]
- Steven, S.; Daiber, A.; Dopheide, J.F.; Münzel, T.; Espinola-Klein, C. Peripheral Artery Disease, Redox Signaling, Oxidative Stress—Basic and Clinical Aspects. *Redox Biol.* **2017**, *12*, 787–797. [CrossRef]
- Yu, H.; Kalogeris, T.; Korthuis, R.J. Reactive Species-Induced Microvascular Dysfunction in Ischemia/Reperfusion. *Free Radic. Biol. Med.* **2019**, *135*, 182–197. [CrossRef] [PubMed]
- Mason, J.C.; Libby, P. Cardiovascular Disease in Patients with Chronic Inflammation: Mechanisms Underlying Premature Cardiovascular Events in Rheumatologic Conditions. *Eur. Heart J.* **2015**, *36*, 482–489. [CrossRef] [PubMed]
- Arida, A.; Protoplerou, A.; Kitas, G.; Sifakis, P. Systemic Inflammatory Response and Atherosclerosis: The Paradigm of Chronic Inflammatory Rheumatic Diseases. *Int. J. Mol. Sci.* **2018**, *19*, 1890. [CrossRef]
- Poledniczek, M.H. Coronary Artery Disease in Granulomatosis with Polyangiitis: A Review. *SN Compr. Clin. Med.* **2022**, *4*, 75. [CrossRef]
- Ridker, P.M.; Everett, B.M.; Thuren, T.; MacFadyen, J.G.; Chang, W.H.; Ballantyne, C.; Fonseca, F.; Nicolau, J.; Koenig, W.; Anker, S.D.; et al. Antiinflammatory Therapy with Canakinumab for Atherosclerotic Disease. *N. Engl. J. Med.* **2017**, *377*, 1119–1131. [CrossRef] [PubMed]
- Nidorf, S.M.; Fiolet, A.T.L.; Mosterd, A.; Eikelboom, J.W.; Schut, A.; Opstal, T.S.J.; The, S.H.K.; Xu, X.-F.; Ireland, M.A.; Lenderink, T.; et al. Colchicine in Patients with Chronic Coronary Disease. *N. Engl. J. Med.* **2020**, *383*, 1838–1847. [CrossRef]
- Deftereos, S.G.; Beerkens, F.J.; Shah, B.; Giannopoulos, G.; Vrachatis, D.A.; Giotaki, S.G.; Siasos, G.; Nicolas, J.; Arnott, C.; Patel, S.; et al. Colchicine in Cardiovascular Disease: In-Depth Review. *Circulation* **2022**, *145*, 61–78.
- Visseren, F.L.J.; MacH, F.; Smulders, Y.M.; Carballo, D.; Koskinas, K.C.; Bäck, M.; Benetos, A.; Biffi, A.; Boavida, J.M.; Capodanno, D.; et al. 2021 ESC Guidelines on Cardiovascular Disease Prevention in Clinical Practice. *Eur. Heart J.* **2021**, *42*, 3227–3337. [CrossRef]
- Darabseh, M.Z.; Maden-Wilkinson, T.M.; Welbourne, G.; Wüst, R.C.I.; Ahmed, N.; Aushah, H.; Selfe, J.; Morse, C.I.; Degens, H. Fourteen Days of Smoking Cessation Improves Muscle Fatigue Resistance and Reverses Markers of Systemic Inflammation. *Sci. Rep.* **2021**, *11*, 12286. [CrossRef]
- McElroy, J.P.; Carmella, S.G.; Heskin, A.K.; Tang, M.K.; Murphy, S.E.; Reisinger, S.A.; Jensen, J.A.; Hatsukami, D.K.; Hecht, S.S.; Shields, P.G. Effects of Cessation of Cigarette Smoking on Eicosanoid Biomarkers of Inflammation and Oxidative Damage. *PLoS ONE* **2019**, *14*, e0218386. [CrossRef]
- Koushki, K.; Shahbaz, S.K.; Mashayekhi, K.; Sadeghi, M.; Zayeri, Z.D.; Taba, M.Y.; Banach, M.; Al-Rasadi, K.; Johnston, T.P.; Sahebkar, A. Anti-Inflammatory Action of Statins in Cardiovascular Disease: The Role of Inflammasome and Toll-Like Receptor Pathways. *Clin. Rev. Allergy Immunol.* **2021**, *60*, 175–199. [CrossRef] [PubMed]
- Aboyans, V.; Ricco, J.B.; Bartelink, M.L.E.L.; Björck, M.; Brodmann, M.; Cohnert, T.; Collet, J.P.; Czerny, M.; De Carlo, M.; Debus, S.; et al. 2017 ESC Guidelines on the Diagnosis and Treatment of Peripheral Arterial Diseases, in Collaboration with the European Society for Vascular Surgery (ESVS). *Eur. Heart J.* **2018**, *39*, 763–816. [CrossRef] [PubMed]
- Yuan, Z.; Lu, Y.; Wei, J.; Wu, J.; Yang, J.; Cai, Z. Abdominal Aortic Aneurysm: Roles of Inflammatory Cells. *Front. Immunol.* **2020**, *11*, 609161. [CrossRef] [PubMed]
- Tilson, M.D. Decline of the Atherogenic Theory of the Etiology of the Abdominal Aortic Aneurysm and Rise of the Autoimmune Hypothesis. *J. Vasc. Surg.* **2016**, *64*, 1523–1525. [CrossRef] [PubMed]
- Klopff, J.; Brostjan, C.; Neumayer, C.; Eilenberg, W. Neutrophils as Regulators and Biomarkers of Cardiovascular Inflammation in the Context of Abdominal Aortic Aneurysms. *Biomedicines* **2021**, *9*, 1236. [CrossRef] [PubMed]

26. Reitsma, S.; Oude Egbrink, M.G.; Heijnen, V.V.T.; Megens, R.T.A.; Engels, W.; Vink, H.; Slaaf, D.W.; van Zandvoort, M.A.M.J. Endothelial Glycocalyx Thickness and Platelet-Vessel Wall Interactions during Atherogenesis. *Thromb. Haemost.* **2011**, *106*, 939–946. [[CrossRef](#)]
27. Reitsma, S.; Slaaf, D.W.; Vink, H.; Van Zandvoort, M.A.M.J.; Oude Egbrink, M.G.A. The Endothelial Glycocalyx: Composition, Functions, and Visualization. *Pflug. Arch.* **2007**, *454*, 345–359. [[CrossRef](#)]
28. Lipowsky, H.H. Protease Activity and the Role of the Endothelial Glycocalyx in Inflammation. *Drug Discov. Today Dis. Models* **2011**, *8*, 57. [[CrossRef](#)]
29. van der Poll, T.; Parker, R.I. Platelet Activation and Endothelial Cell Dysfunction. *Crit. Care Clin.* **2020**, *36*, 233–253. [[CrossRef](#)]
30. Dull, R.O.; Hahn, R.G. The Glycocalyx as a Permeability Barrier: Basic Science and Clinical Evidence. *Crit. Care* **2022**, *26*, 273. [[CrossRef](#)]
31. Fels, B.; Kusche-Vihrog, K. It Takes More than Two to Tango: Mechanosignaling of the Endothelial Surface. *Pflug. Arch.* **2020**, *472*, 419–433. [[CrossRef](#)] [[PubMed](#)]
32. Mitra, R.; O’Neil, G.L.; Harding, I.C.; Cheng, M.J.; Mensah, S.A.; Ebong, E.E. Glycocalyx in Atherosclerosis-Relevant Endothelium Function and as a Therapeutic Target. *Curr. Atheroscler. Rep.* **2017**, *19*, 63. [[CrossRef](#)] [[PubMed](#)]
33. Qu, J.; Cheng, Y.; Wu, W.; Yuan, L.; Liu, X. Glycocalyx Impairment in Vascular Disease: Focus on Inflammation. *Front. Cell Dev. Biol.* **2021**, *9*, 730621. [[CrossRef](#)]
34. Panzer, B.; Kopp, C.W.; Neumayer, C.; Koppensteiner, R.; Jozkowicz, A.; Poledniczek, M.; Gremmel, T.; Jilma, B.; Wadowski, P.P. Toll-like Receptors as Pro-Thrombotic Drivers in Viral Infections: A Narrative Review. *Cells* **2023**, *12*, 1865. [[CrossRef](#)] [[PubMed](#)]
35. Maschalidi, S.; Ravichandran, K.S. Phagocytosis: Sweet Repulsions via the Glycocalyx. *Curr. Biol.* **2021**, *31*, R20–R22. [[CrossRef](#)] [[PubMed](#)]
36. Imbert, P.R.C.; Saric, A.; Pedram, K.; Bertozzi, C.R.; Grinstein, S.; Freeman, S.A. An Acquired and Endogenous Glycocalyx Forms a Bidirectional “Don’t Eat” and “Don’t Eat Me” Barrier to Phagocytosis. *Curr. Biol.* **2021**, *31*, 77–89.e5. [[CrossRef](#)] [[PubMed](#)]
37. Marki, A.; Esko, J.D.; Pries, A.R.; Ley, K. Role of the Endothelial Surface Layer in Neutrophil Recruitment. *J. Leukoc. Biol.* **2015**, *98*, 503–515. [[CrossRef](#)]
38. Möckl, L. The Emerging Role of the Mammalian Glycocalyx in Functional Membrane Organization and Immune System Regulation. *Front. Cell Dev. Biol.* **2020**, *8*, 253. [[CrossRef](#)]
39. Doster, R.S.; Rogers, L.M.; Gaddy, J.A.; Aronoff, D.M. Macrophage Extracellular Traps: A Scoping Review. *J. Innate Immun.* **2018**, *10*, 3–13. [[CrossRef](#)]
40. Je, S.; Quan, H.; Yoon, Y.; Na, Y.; Kim, B.J.; Seok, S.H. Mycobacterium Massiliense Induces Macrophage Extracellular Traps with Facilitating Bacterial Growth. *PLoS ONE* **2016**, *11*, e0155685. [[CrossRef](#)]
41. Fu, G.; Deng, M.; Neal, M.D.; Billiar, T.R.; Scott, M.J. Platelet-Monocyte Aggregates: Understanding Mechanisms and Functions in Sepsis. *Shock* **2021**, *55*, 156–166. [[CrossRef](#)] [[PubMed](#)]
42. Kaiser, R.; Escaig, R.; Erber, J.; Nicolai, L. Neutrophil-Platelet Interactions as Novel Treatment Targets in Cardiovascular Disease. *Front. Cardiovasc. Med.* **2022**, *8*, 824112. [[CrossRef](#)] [[PubMed](#)]
43. Qi, H.; Yang, S.; Zhang, L. Neutrophil Extracellular Traps and Endothelial Dysfunction in Atherosclerosis and Thrombosis. *Front. Immunol.* **2017**, *8*, 928. [[CrossRef](#)]
44. Banerjee, S.; Mwangi, J.G.; Stanley, T.K.; Mitra, R.; Ebong, E.E. Regeneration and Assessment of the Endothelial Glycocalyx to Address Cardiovascular Disease. *Ind. Eng. Chem. Res.* **2021**, *60*, 17328–17347. [[CrossRef](#)]
45. Steven, S.; Frenis, K.; Oelze, M.; Kalinovic, S.; Kuntic, M.; Jimenez, M.T.B.; Vujacic-Mirska, K.; Helmstädtter, J.; Kröller-Schön, S.; Münz, T.; et al. Vascular Inflammation and Oxidative Stress: Major Triggers for Cardiovascular Disease. *Oxid. Med. Cell Longev.* **2019**, *2019*, 7092151. [[CrossRef](#)] [[PubMed](#)]
46. Ofosu, F.A.; Dewar, L.; Craven, S.J.; Song, Y.; Cedrone, A.; Freedman, J.; Fenton, J.W. Coordinate Activation of Human Platelet Protease-Activated Receptor-1 and -4 in Response to Subnanomolar Alpha-Thrombin. *J. Biol. Chem.* **2008**, *283*, 26886–26893. [[CrossRef](#)] [[PubMed](#)]
47. Wadowski, P.P.; Pultar, J.; Weikert, C.; Eichelberger, B.; Panzer, B.; Huber, K.; Lang, I.M.; Koppensteiner, R.; Panzer, S.; Gremmel, T. Protease-Activated Receptor-Mediated Platelet Aggregation in Acute Coronary Syndrome Patients on Potent P2Y12 Inhibitors. *Res. Pract. Thromb. Haemost.* **2019**, *3*, 383–390. [[CrossRef](#)]
48. Adam, F.; Guillen, M.C.; Jandrot-Perrus, M. Glycoprotein Ib-Mediated Platelet Activation. A Signalling Pathway Triggered by Thrombin. *Eur. J. Biochem.* **2003**, *270*, 2959–2970. [[CrossRef](#)] [[PubMed](#)]
49. Hally, K.; Fauteux-Daniel, S.; Hamzeh-Cognasse, H.; Larsen, P.; Cognasse, F. Revisiting Platelets and Toll-Like Receptors (TLRs): At the Interface of Vascular Immunity and Thrombosis. *Int. J. Mol. Sci.* **2020**, *21*, 6150. [[CrossRef](#)]
50. Niklaus, M.; Klingler, P.; Weber, K.; Koessler, A.; Kuhn, S.; Boeck, M.; Kobsar, A.; Koessler, J. Platelet Toll-Like-Receptor-2 and -4 Mediate Different Immune-Related Responses to Bacterial Ligands. *TH Open* **2022**, *6*, e156–e167. [[CrossRef](#)]
51. Salvador, B.; Arranz, A.; Francisco, S.; Córdoba, L.; Punzón, C.; Llamas, M.Á.; Fresno, M. Modulation of Endothelial Function by Toll like Receptors. *Pharmacol. Res.* **2016**, *108*, 46–56. [[CrossRef](#)] [[PubMed](#)]
52. O’Neill, L.A.J. DisSARMing Toll-like Receptor Signaling. *Nat. Immunol.* **2006**, *7*, 1023–1025. [[CrossRef](#)] [[PubMed](#)]
53. Akira, S.; Takeda, K. Toll-like Receptor Signalling. *Nat. Rev. Immunol.* **2004**, *4*, 499–511. [[CrossRef](#)] [[PubMed](#)]

54. Schilling, D.; Thomas, K.; Nixdorff, K.; Vogel, S.N.; Fenton, M.J. Toll-Like Receptor 4 and Toll-IL-1 Receptor Domain-Containing Adapter Protein (TIRAP)/Myeloid Differentiation Protein 88 Adapter-Like (Mal) Contribute to Maximal IL-6 Expression in Macrophages. *J. Immunol.* **2002**, *169*, 5874–5880. [CrossRef] [PubMed]
55. Carty, M.; Goodbody, R.; Schröder, M.; Stack, J.; Moynagh, P.N.; Bowie, A.G. The Human Adaptor SARM Negatively Regulates Adaptor Protein TRIF-Dependent Toll-like Receptor Signaling. *Nat. Immunol.* **2006**, *7*, 1074–1081. [CrossRef]
56. El-Zayat, S.R.; Sibai, H.; Mannaa, F.A. Toll-like Receptors Activation, Signaling, and Targeting: An Overview. *Bull. Natl. Res. Cent.* **2019**, *43*, 187. [CrossRef]
57. Liu, T.; Zhang, L.; Joo, D.; Sun, S.C. NF-KB Signaling in Inflammation. *Signal Transduct. Target. Ther.* **2017**, *2*, 17023. [CrossRef]
58. Rubio, D.; Xu, R.H.; Remakus, S.; Krouse, T.E.; Truckenmiller, M.E.; Thapa, R.J.; Balachandran, S.; Alcamí, A.; Norbury, C.C.; Sigal, L.J. Crosstalk between the Type 1 Interferon and Nuclear Factor Kappa B Pathways Confers Resistance to a Lethal Virus Infection. *Cell Host Microbe* **2013**, *13*, 701–710. [CrossRef]
59. Ernst, O.; Vaytaden, S.J.; Fraser, I.D.C. Measurement of NF-KB Activation in TLR-Activated Macrophages. *Methods Mol. Biol.* **2018**, *1714*, 67–78. [CrossRef]
60. Qiao, Y.; Wang, P.; Qi, J.; Zhang, L.; Gao, C. TLR-Induced NF-KB Activation Regulates NLRP3 Expression in Murine Macrophages. *FEBS Lett.* **2012**, *586*, 1022–1026. [CrossRef]
61. Grebe, A.; Hoss, F.; Latz, E. NLRP3 Inflammasome and the IL-1 Pathway in Atherosclerosis. *Circ. Res.* **2018**, *122*, 1722–1740. [CrossRef]
62. Bartoli-Leonard, F.; Zimmer, J.; Sonawane, A.R.; Perez, K.; Turner, M.E.; Kuraoka, S.; Pham, T.; Li, F.; Aikawa, M.; Singh, S.; et al. NLRP3 Inflammasome Activation in Peripheral Arterial Disease. *J. Am. Heart Assoc.* **2023**, *12*, e026945. [CrossRef]
63. Bai, B.; Yang, Y.; Wang, Q.; Li, M.; Tian, C.; Liu, Y.; Aung, L.H.H.; Li, P.; Yu, T.; Chu, X.M. NLRP3 Inflammasome in Endothelial Dysfunction. *Cell Death Dis.* **2020**, *11*, 776. [CrossRef] [PubMed]
64. Lee, Y.; Reilly, B.; Tan, C.; Wang, P.; Aziz, M. Extracellular CIRP Induces Macrophage Extracellular Trap Formation Via Gasdermin D Activation. *Front. Immunol.* **2021**, *12*, 780210. [CrossRef] [PubMed]
65. Hally, K.E.; Bird, G.K.; la Flamme, A.C.; Harding, S.A.; Larsen, P.D. Platelets Modulate Multiple Markers of Neutrophil Function in Response to in Vitro Toll-like Receptor Stimulation. *PLoS ONE* **2019**, *14*, e0223444. [CrossRef]
66. Clark, S.R.; Ma, A.C.; Tavener, S.A.; McDonald, B.; Goodarzi, Z.; Kelly, M.M.; Patel, K.D.; Chakrabarti, S.; McAvoy, E.; Sinclair, G.D.; et al. Platelet TLR4 Activates Neutrophil Extracellular Traps to Ensnare Bacteria in Septic Blood. *Nat. Med.* **2007**, *13*, 463–469. [CrossRef]
67. Zhang, D.; Chen, G.; Manwani, D.; Mortha, A.; Xu, C.; Faith, J.J.; Burk, R.D.; Kunisaki, Y.; Jang, J.E.; Scheiermann, C.; et al. Neutrophil Ageing Is Regulated by the Microbiome. *Nature* **2015**, *525*, 528–532. [CrossRef] [PubMed]
68. Katakami, N. Mechanism of Development of Atherosclerosis and Cardiovascular Disease in Diabetes Mellitus. *J. Atheroscler. Thromb.* **2018**, *25*, 27–39. [CrossRef]
69. Mulivor, A.W.; Lipowsky, H.H. Inflammation- and Ischemia-Induced Shedding of Venular Glycocalyx. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, *286*, H1672–H1680. [CrossRef]
70. Dogné, S.; Flamion, B.; Caron, N. Endothelial Glycocalyx as a Shield Against Diabetic Vascular Complications: Involvement of Hyaluronan and Hyaluronidases. *Arter. Thromb. Vasc. Biol.* **2018**, *38*, 1427. [CrossRef]
71. Lee, D.H.; Dane, M.J.C.; Van Den Berg, B.M.; Boels, M.G.S.; Van Teeffelen, J.W.; De Mutsert, R.; Den Heijer, M.; Rosendaal, F.R.; Van Der Vlag, J.; Van Zonneveld, A.J.; et al. Deeper Penetration of Erythrocytes into the Endothelial Glycocalyx Is Associated with Impaired Microvascular Perfusion. *PLoS ONE* **2014**, *9*, e96477. [CrossRef] [PubMed]
72. Rabelink, T.J.; De Zeeuw, D. The Glycocalyx—Linking Albuminuria with Renal and Cardiovascular Disease. *Nat. Rev. Nephrol.* **2015**, *11*, 667–676. [CrossRef] [PubMed]
73. Liew, H.; Roberts, M.A.; MacGinley, R.; McMahon, L.P. Endothelial Glycocalyx in Health and Kidney Disease: Rising Star or False Dawn? *Nephrology* **2017**, *22*, 940–946. [CrossRef] [PubMed]
74. Wadowski, P.P.; Kautzky-Willer, A.; Gremmel, T.; Koppensteiner, R.; Wolf, P.; Ertl, S.; Weikert, C.; Schörgenhofer, C.; Jilma, B. Sublingual Microvasculature in Diabetic Patients. *Microvasc. Res.* **2020**, *129*, 103971. [CrossRef] [PubMed]
75. Hirano, T. Pathophysiology of Diabetic Dyslipidemia. *J. Atheroscler. Thromb.* **2018**, *25*, 771–782. [CrossRef]
76. Hagensen, M.K.; Mortensen, M.B.; Kjolby, M.; Palmfeldt, J.; Bentzon, J.F.; Gregersen, S. Increased Retention of LDL from Type 1 Diabetic Patients in Atherosclerosis-Prone Areas of the Murine Arterial Wall. *Atherosclerosis* **2019**, *286*, 156–162. [CrossRef]
77. Singh, S.; Siva, B.V.; Ravichandiran, V. Advanced Glycation End Products: Key Player of the Pathogenesis of Atherosclerosis. *Glycoconj. J.* **2022**, *39*, 547–563. [CrossRef]
78. Palmieri, E.M.; Gonzalez-Cotto, M.; Baseler, W.A.; Davies, L.C.; Ghesquière, B.; Maio, N.; Rice, C.M.; Rouault, T.A.; Cassel, T.; Higashi, R.M.; et al. Nitric Oxide Orchestrates Metabolic Rewiring in M1 Macrophages by Targeting Aconitase 2 and Pyruvate Dehydrogenase. *Nat. Commun.* **2020**, *11*, 698. [CrossRef] [PubMed]
79. Vujic, A.; Koo, A.N.M.; Prag, H.A.; Krieg, T. Mitochondrial Redox and TCA Cycle Metabolite Signaling in the Heart. *Free Radic. Biol. Med.* **2021**, *166*, 287–296. [CrossRef]
80. Tannahill, G.M.; Curtis, A.M.; Adamik, J.; Palsson-McDermott, E.M.; McGettrick, A.F.; Goel, G.; Frezza, C.; Bernard, N.J.; Kelly, B.; Foley, N.H.; et al. Succinate Is an Inflammatory Signal That Induces IL-1 β through HIF-1 α . *Nature* **2013**, *496*, 238–242. [CrossRef]
81. O’Neill, L.A.J.; Pearce, E.J. Immunometabolism Governs Dendritic Cell and Macrophage Function. *J. Exp. Med.* **2016**, *213*, 15–23. [CrossRef]

82. Chouchani, E.T.; Pell, V.R.; Gaude, E.; Aksentijević, D.; Sundier, S.Y.; Robb, E.L.; Logan, A.; Nadtochiy, S.M.; Ord, E.N.J.; Smith, A.C.; et al. Ischaemic Accumulation of Succinate Controls Reperfusion Injury through Mitochondrial ROS. *Nature* **2014**, *515*, 431–435. [CrossRef] [PubMed]
83. Lee, W.J.; Tateya, S.; Cheng, A.M.; Rizzo-Deleon, N.; Wang, N.F.; Handa, P.; Wilson, C.L.; Clowes, A.W.; Sweet, I.R.; Bomsztyk, K.; et al. M2 Macrophage Polarization Mediates Anti-Inflammatory Effects of Endothelial Nitric Oxide Signaling. *Diabetes* **2015**, *64*, 2836–2846. [CrossRef] [PubMed]
84. Steiner, S.; Schaller, G.; Puttinger, H.; Födinger, M.; Kopp, C.W.; Seidinger, D.; Grisar, J.; Hörl, W.H.; Minar, E.; Vykytil, A.; et al. History of Cardiovascular Disease Is Associated with Endothelial Progenitor Cells in Peritoneal Dialysis Patients. *Am. J. Kidney Dis.* **2005**, *46*, 520–528. [CrossRef]
85. Ambasta, R.K.; Kohli, H.; Kumar, P. Multiple Therapeutic Effect of Endothelial Progenitor Cell Regulated by Drugs in Diabetes and Diabetes Related Disorder. *J. Transl. Med.* **2017**, *15*, 185. [CrossRef] [PubMed]
86. Sandri, M.; Beck, E.B.; Adams, V.; Gielen, S.; Lenk, K.; Höllriegel, R.; Mangner, N.; Linke, A.; Erbs, S.; Möbius-Winkler, S.; et al. Maximal Exercise, Limb Ischemia, and Endothelial Progenitor Cells. *Eur. J. Cardiovasc. Prev. Rehabil.* **2011**, *18*, 55–64. [CrossRef] [PubMed]
87. Cooke, J.P.; Meng, S. Vascular Regeneration in Peripheral Artery Disease. *Arter. Thromb. Vasc. Biol.* **2020**, *40*, 1627–1634. [CrossRef]
88. Yamaguchi, J.I.; Kusano, K.F.; Masuo, O.; Kawamoto, A.; Silver, M.; Murasawa, S.; Bosch-Marce, M.; Masuda, H.; Losordo, D.W.; Isner, J.M.; et al. Stromal Cell-Derived Factor-1 Effects on Ex Vivo Expanded Endothelial Progenitor Cell Recruitment for Ischemic Neovascularization. *Circulation* **2003**, *107*, 1322–1328. [CrossRef]
89. He, J.; Xiao, Z.; Chen, X.; Chen, M.; Fang, L.; Yang, M.; Lv, Q.; Li, Y.; Li, G.; Hu, J.; et al. The Expression of Functional Toll-like Receptor 4 Is Associated with Proliferation and Maintenance of Stem Cell Phenotype in Endothelial Progenitor Cells (EPCs). *J. Cell Biochem.* **2010**, *111*, 179–186. [CrossRef]
90. Matsumoto, Y.; Adams, V.; Walther, C.; Kleinecke, C.; Brugger, P.; Linke, A.; Walther, T.; Mohr, F.W.; Schuler, G. Reduced Number and Function of Endothelial Progenitor Cells in Patients with Aortic Valve Stenosis: A Novel Concept for Valvular Endothelial Cell Repair. *Eur. Heart J.* **2009**, *30*, 346–355. [CrossRef]
91. Stoiber, W.; Obermayer, A.; Steinbacher, P.; Krautgartner, W.D. The Role of Reactive Oxygen Species (ROS) in the Formation of Extracellular Traps (ETs) in Humans. *Biomolecules* **2015**, *5*, 702–723. [CrossRef]
92. Ali, M.A.M.; Spinler, S.A. COVID-19 and Thrombosis: From Bench to Bedside. *Trends Cardiovasc. Med.* **2021**, *31*, 143–160. [CrossRef]
93. Wadowski, P.P.; Jilma, B.; Kopp, C.W.; Ertl, S.; Gremmel, T.; Koppensteiner, R. Glycocalyx as Possible Limiting Factor in COVID-19. *Front. Immunol.* **2021**, *12*, 607306. [CrossRef] [PubMed]
94. Borrman, M.; Brandes, F.; Kirchner, B.; Klein, M.; Billaud, J.N.; Reithmair, M.; Rehm, M.; Schelling, G.; Pfaffl, M.W.; Meidert, A.S. Extensive Blood Transcriptome Analysis Reveals Cellular Signaling Networks Activated by Circulating Glycocalyx Components Reflecting Vascular Injury in COVID-19. *Front. Immunol.* **2023**, *14*, 1129766. [CrossRef] [PubMed]
95. Varga, Z.; Flammer, A.J.; Steiger, P.; Haberecker, M.; Andermatt, R.; Zinkernagel, A.S.; Mehra, M.R.; Schuepbach, R.A.; Ruschitzka, F.; Moch, H. Endothelial Cell Infection and Endotheliitis in COVID-19. *Lancet* **2020**, *395*, 1417–1418. [CrossRef] [PubMed]
96. Xu, S.W.; Ilyas, I.; Weng, J.P. Endothelial Dysfunction in COVID-19: An Overview of Evidence, Biomarkers, Mechanisms and Potential Therapies. *Acta Pharmacol. Sin.* **2023**, *44*, 695–709. [CrossRef]
97. Ratajczak, J.; Wysoczynski, M.; Hayek, F.; Janowska-Wieczorek, A.; Ratajczak, M.Z. Membrane-Derived Microvesicles: Important and Underappreciated Mediators of Cell-to-Cell Communication. *Leukemia* **2006**, *20*, 1487–1495. [CrossRef]
98. Chen, Y.T.; Yuan, H.X.; Ou, Z.J.; Ou, J.S. Microparticles (Exosomes) and Atherosclerosis. *Curr. Atheroscler. Rep.* **2020**, *22*, 23. [CrossRef]
99. Loyer, X.; Vion, A.C.; Tedgui, A.; Boulanger, C.M. Microvesicles as Cell-Cell Messengers in Cardiovascular Diseases. *Circ. Res.* **2014**, *114*, 345–353. [CrossRef]
100. Février, B.; Raposo, G. Exosomes: Endosomal-Derived Vesicles Shipping Extracellular Messages. *Curr. Opin. Cell Biol.* **2004**, *16*, 415–421. [CrossRef]
101. Kalluri, R.; LeBleu, V.S. The Biology, Function, and Biomedical Applications of Exosomes. *Science* **2020**, *367*, eaau6977. [CrossRef] [PubMed]
102. Hosseinkhani, B.; Kuypers, S.; van den Akker, N.M.S.; Molin, D.G.M.; Michiels, L. Extracellular Vesicles Work as a Functional Inflammatory Mediator Between Vascular Endothelial Cells and Immune Cells. *Front. Immunol.* **2018**, *9*, 1789. [CrossRef]
103. Wendt, S.; Goetzenich, A.; Goetsch, C.; Stoppe, C.; Bleilevens, C.; Kraemer, S.; Benstoem, C. Evaluation of the Cardioprotective Potential of Extracellular Vesicles—A Systematic Review and Meta-Analysis. *Sci. Rep.* **2018**, *8*, 15702. [CrossRef]
104. George, M.; Ganesh, M.R.; Sridhar, A.; Jena, A.; Rajaram, M.; Shanmugam, E.; Dhandapani, V.E. Evaluation of Endothelial and Platelet Derived Microparticles in Patients with Acute Coronary Syndrome. *J. Clin. Diagn. Res.* **2015**, *9*, OC09–OC13. [CrossRef]
105. Tousoulis, D.; Kampoli, A.-M.; Tentolouris Nikolaos Papageorgiou, C.; Stefanadis, C. The Role of Nitric Oxide on Endothelial Function. *Curr. Vasc. Pharmacol.* **2012**, *10*, 4–18. [CrossRef]
106. Huang, P.L.; Huang, Z.; Mashimo, H.; Bloch, K.D.; Moskowitz, M.A.; Bevan, J.A.; Fishman, M.C. Hypertension in Mice Lacking the Gene for Endothelial Nitric Oxide Synthase. *Nature* **1995**, *377*, 239–242. [CrossRef] [PubMed]
107. Gimbrone, M.A.; García-Cardeña, G. Endothelial Cell Dysfunction and the Pathobiology of Atherosclerosis. *Circ. Res.* **2016**, *118*, 620–636. [CrossRef] [PubMed]

108. Stary, H.C. Natural History and Histological Classification of Atherosclerotic Lesions: An Update. *Arter. Thromb. Vasc. Biol.* **2000**, *20*, 1177–1178. [[CrossRef](#)]
109. Mittal, M.; Siddiqui, M.R.; Tran, K.; Reddy, S.P.; Malik, A.B. Reactive Oxygen Species in Inflammation and Tissue Injury. *Antioxid. Redox Signal.* **2014**, *20*, 1126–1167. [[CrossRef](#)]
110. Incalza, M.A.; D’Oria, R.; Natalechchio, A.; Perrini, S.; Laviola, L.; Giorgino, F. Oxidative Stress and Reactive Oxygen Species in Endothelial Dysfunction Associated with Cardiovascular and Metabolic Diseases. *Vasc. Pharmacol.* **2018**, *100*, 1–19. [[CrossRef](#)]
111. Zhou, R.; Yazdi, A.S.; Menu, P.; Tschoopp, J. A Role for Mitochondria in NLRP3 Inflammasome Activation. *Nature* **2011**, *469*, 221–226. [[CrossRef](#)]
112. Bulua, A.C.; Simon, A.; Maddipati, R.; Pelletier, M.; Park, H.; Kim, K.Y.; Sack, M.N.; Kastner, D.L.; Siegel, R.M. Mitochondrial Reactive Oxygen Species Promote Production of Proinflammatory Cytokines and Are Elevated in TNFR1-Associated Periodic Syndrome (TRAPS). *J. Exp. Med.* **2011**, *208*, 519–533. [[CrossRef](#)]
113. Ryan, K.A.; Smith, M.F.; Sanders, M.K.; Ernst, P.B. Reactive Oxygen and Nitrogen Species Differentially Regulate Toll-Like Receptor 4-Mediated Activation of NF-KB and Interleukin-8 Expression. *Infect. Immun.* **2004**, *72*, 2123. [[CrossRef](#)] [[PubMed](#)]
114. Zhang, J.; Wang, X.; Vikash, V.; Ye, Q.; Wu, D.; Liu, Y.; Dong, W. ROS and ROS-Mediated Cellular Signaling. *Oxid. Med. Cell Longev.* **2016**, *2016*, 4350965. [[CrossRef](#)]
115. Qiu, Q.; Yang, Z.; Cao, F.; Yang, C.; Hardy, P.; Yan, X.; Yang, S.; Xiong, W. Activation of NLRP3 Inflammasome by Lymphocytic Microparticles via TLR4 Pathway Contributes to Airway Inflammation. *Exp. Cell Res.* **2020**, *386*, 111737. [[CrossRef](#)] [[PubMed](#)]
116. Jerez-Dolz, D.; Torramade-Moix, S.; Palomo, M.; Moreno-Castaño, A.; Lopez-Vilchez, I.; Hernandez, R.; Badimon, J.J.; Zafar, M.U.; Diaz-Ricart, M.; Escolar, G. Internalization of Microparticles by Platelets Is Partially Mediated by Toll-like Receptor 4 and Enhances Platelet Thrombogenicity. *Atherosclerosis* **2020**, *294*, 17–24. [[CrossRef](#)]
117. Zhang, W.; Liu, R.; Chen, Y.; Wang, M.; Du, J. Crosstalk between Oxidative Stress and Exosomes. *Oxid. Med. Cell Longev.* **2022**, *2022*, 3553617. [[CrossRef](#)] [[PubMed](#)]
118. Jansen, F.; Yang, X.; Franklin, B.S.; Hoelscher, M.; Schmitz, T.; Bedorf, J.; Nickenig, G.; Werner, N. High Glucose Condition Increases NADPH Oxidase Activity in Endothelial Microparticles That Promote Vascular Inflammation. *Cardiovasc. Res.* **2013**, *98*, 94–106. [[CrossRef](#)] [[PubMed](#)]
119. Mause, S.F.; Weber, C. Microparticles: Protagonists of a Novel Communication Network for Intercellular Information Exchange. *Circ. Res.* **2010**, *107*, 1047–1057. [[CrossRef](#)] [[PubMed](#)]
120. Ci, H.B.; Ou, Z.J.; Chang, F.J.; Liu, D.H.; He, G.W.; Xu, Z.; Yuan, H.Y.; Wang, Z.P.; Zhang, X.; Ou, J.S. Endothelial Microparticles Increase in Mitral Valve Disease and Impair Mitral Valve Endothelial Function. *Am. J. Physiol. Endocrinol. Metab.* **2013**, *304*, E695–E702. [[CrossRef](#)] [[PubMed](#)]
121. Densmore, J.C.; Signorino, P.R.; Ou, J.; Hatoum, O.A.; Rowe, J.J.; Shi, Y.; Kaul, S.; Jones, D.W.; Sabina, R.E.; Pritchard, K.A.; et al. Endothelium-Derived Microparticles Induce Endothelial Dysfunction and Acute Lung Injury. *Shock* **2006**, *26*, 464–471. [[CrossRef](#)] [[PubMed](#)]
122. Lukasik, M.; Rozalski, M.; Luzak, B.; Michalak, M.; Ambrosius, W.; Watala, C.; Kozubski, W. Enhanced Platelet-Derived Microparticle Formation Is Associated with Carotid Atherosclerosis in Convalescent Stroke Patients. *Platelets* **2013**, *24*, 63–70. [[CrossRef](#)]
123. Lin, Z.B.; Ci, H.B.; Li, Y.; Cheng, T.P.; Liu, D.H.; Wang, Y.S.; Xu, J.; Yuan, H.X.; Li, H.M.; Chen, J.; et al. Endothelial Microparticles Are Increased in Congenital Heart Diseases and Contribute to Endothelial Dysfunction. *J. Transl. Med.* **2017**, *15*, 4. [[CrossRef](#)] [[PubMed](#)]
124. Xie, Z.; Wang, X.; Liu, X.; Du, H.; Sun, C.; Shao, X.; Tian, J.; Gu, X.; Wang, H.; Tian, J.; et al. Adipose-Derived Exosomes Exert Proatherogenic Effects by Regulating Macrophage Foam Cell Formation and Polarization. *J. Am. Heart Assoc.* **2018**, *7*, e007442. [[CrossRef](#)]
125. Blaser, M.C.; Aikawa, E. Differential MiRNA Loading Underpins Dual Harmful and Protective Roles for Extracellular Vesicles in Atherogenesis. *Circ. Res.* **2019**, *124*, 467–469. [[CrossRef](#)] [[PubMed](#)]
126. Li, C.; Li, S.; Zhang, F.; Wu, M.; Liang, H.; Song, J.; Lee, C.; Chen, H. Endothelial Microparticles-Mediated Transfer of MicroRNA-19b Promotes Atherosclerosis via Activating Perivascular Adipose Tissue Inflammation in ApoE-/- Mice. *Biochem. Biophys. Res. Commun.* **2018**, *495*, 1922–1929. [[CrossRef](#)]
127. Ceolotto, G.; Giannella, A.; Albiero, M.; Kuppusamy, M.; Radu, C.; Simioni, P.; Garlaschelli, K.; Baragetti, A.; Catapano, A.L.; Iori, E.; et al. MiR-30c-5p Regulates Macrophage-Mediated Inflammation and pro-Atherosclerosis Pathways. *Cardiovasc. Res.* **2017**, *113*, 1627–1638. [[CrossRef](#)]
128. Pereira-Da-silva, T.; Napoleão, P.; Costa, M.C.; Gabriel, A.F.; Selas, M.; Silva, F.; Enguita, F.J.; Ferreira, R.C.; Carmo, M.M. Cigarette Smoking, MiR-27b Downregulation, and Peripheral Artery Disease: Insights into the Mechanisms of Smoking Toxicity. *J. Clin. Med.* **2021**, *10*, 890. [[CrossRef](#)]
129. Badacz, R.; Kleczyński, P.; Legutko, J.; Żmudka, K.; Gacoń, J.; Przewłocki, T.; Kabłak-Ziembińska, A. Expression of MiR-1-3p, MiR-16-5p and MiR-122-5p as Possible Risk Factors of Secondary Cardiovascular Events. *Biomedicines* **2021**, *9*, 1055. [[CrossRef](#)]
130. Wronska, A.; Kurkowska-Jastrzebska, I.; Santulli, G. Application of MicroRNAs in Diagnosis and Treatment of Cardiovascular Disease. *Acta Physiol.* **2015**, *213*, 60–83. [[CrossRef](#)]
131. Wang, M.; Zhang, W.; Zhang, L.; Wang, L.; Li, J.; Shu, C.; Li, X. Roles of MicroRNAs in Peripheral Artery In-Stent Restenosis after Endovascular Treatment. *Biomed. Res. Int.* **2021**, *2021*, 9935671. [[CrossRef](#)]

132. Arroyo, J.D.; Chevillet, J.R.; Kroh, E.M.; Ruf, I.K.; Pritchard, C.C.; Gibson, D.F.; Mitchell, P.S.; Bennett, C.F.; Pogosova-Agadjanyan, E.L.; Stirewalt, D.L.; et al. Argonaute2 Complexes Carry a Population of Circulating MicroRNAs Independent of Vesicles in Human Plasma. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 5003–5008. [[CrossRef](#)]
133. Vickers, K.C.; Palmisano, B.T.; Shoucri, B.M.; Shamburek, R.D.; Remaley, A.T. MicroRNAs Are Transported in Plasma and Delivered to Recipient Cells by High-Density Lipoproteins. *Nat. Cell Biol.* **2011**, *13*, 423–435. [[CrossRef](#)] [[PubMed](#)]
134. Stojkovic, S.; Jurisic, M.; Kopp, C.W.; Koppensteiner, R.; Huber, K.; Wojta, J.; Gremmel, T. Circulating MicroRNAs Identify Patients at Increased Risk of In-Stent Restenosis after Peripheral Angioplasty with Stent Implantation. *Atherosclerosis* **2018**, *269*, 197–203. [[CrossRef](#)] [[PubMed](#)]
135. Badacz, R.; Przewłocki, T.; Legutko, J.; Żmudka, K.; Kabłak-Ziembicka, A. MicroRNAs Associated with Carotid Plaque Development and Vulnerability: The Clinician’s Perspective. *Int. J. Mol. Sci.* **2022**, *23*, 15645. [[CrossRef](#)]
136. Stojkovic, S.; Wadowski, P.P.; Haider, P.; Weikert, C.; Pultar, J.; Lee, S.; Eichelberger, B.; Hengstenberg, C.; Wojta, J.; Panzer, S.; et al. Circulating MicroRNAs and Monocyte-Platelet Aggregate Formation in Acute Coronary Syndrome. *Thromb. Haemost.* **2021**, *121*, 913–922. [[CrossRef](#)] [[PubMed](#)]
137. Finn, N.A.; Eapen, D.; Manocha, P.; Al Kassem, H.; Lassegue, B.; Ghasemzadeh, N.; Quyyumi, A.; Searles, C.D. Coronary Heart Disease Alters Intercellular Communication by Modifying Microparticle-Mediated MicroRNA Transport. *FEBS Lett.* **2013**, *587*, 3456–3463. [[CrossRef](#)]
138. Alexandru, N.; Andrei, E.; Niculescu, L.; Dragan, E.; Ristoiu, V.; Georgescu, A. Microparticles of Healthy Origins Improve Endothelial Progenitor Cell Dysfunction via MicroRNA Transfer in an Atherosclerotic Hamster Model. *Acta Physiol.* **2017**, *221*, 230–249. [[CrossRef](#)]
139. Brinkmann, V.; Reichard, U.; Goosmann, C.; Fauler, B.; Uhlemann, Y.; Weiss, D.S.; Weinrauch, Y.; Zychlinsky, A. Neutrophil Extracellular Traps Kill Bacteria. *Science* **2004**, *303*, 1532–1535. [[CrossRef](#)]
140. Rada, B. Neutrophil Extracellular Traps. *Methods Mol. Biol.* **2019**, *1982*, 517–528. [[CrossRef](#)]
141. Schönrich, G.; Raftery, M.J. Neutrophil Extracellular Traps Go Viral. *Front. Immunol.* **2016**, *7*, 366. [[CrossRef](#)] [[PubMed](#)]
142. Arneth, B.; Arneth, R. Neutrophil Extracellular Traps (NETs) and Vasculitis. *Int. J. Med. Sci.* **2021**, *18*, 1532–1540. [[CrossRef](#)]
143. Nappi, F.; Bellomo, F.; Avtaar Singh, S.S. Worsening Thrombotic Complication of Atherosclerotic Plaques Due to Neutrophils Extracellular Traps: A Systematic Review. *Biomedicines* **2023**, *11*, 113. [[CrossRef](#)] [[PubMed](#)]
144. Gupta, A.K.; Joshi, M.B.; Philippova, M.; Erne, P.; Hasler, P.; Hahn, S.; Resink, T.J. Activated Endothelial Cells Induce Neutrophil Extracellular Traps and Are Susceptible to NETosis-Mediated Cell Death. *FEBS Lett.* **2010**, *584*, 3193–3197. [[CrossRef](#)]
145. Saffarzadeh, M.; Juenemann, C.; Queisser, M.A.; Lochnit, G.; Barreto, G.; Galuska, S.P.; Lohmeyer, J.; Preissner, K.T. Neutrophil Extracellular Traps Directly Induce Epithelial and Endothelial Cell Death: A Predominant Role of Histones. *PLoS ONE* **2012**, *7*, e32366. [[CrossRef](#)] [[PubMed](#)]
146. Nicholls, S.J.; Hazen, S.L. Myeloperoxidase, Modified Lipoproteins, and Atherogenesis. *J. Lipid Res.* **2009**, *50*, S346–S351. [[CrossRef](#)]
147. Alfaidi, M.; Wilson, H.; Daigneault, M.; Burnett, A.; Ridger, V.; Chamberlain, J.; Francis, S. Neutrophil Elastase Promotes Interleukin-1 β Secretion from Human Coronary Endothelium. *J. Biol. Chem.* **2015**, *290*, 24067–24078. [[CrossRef](#)]
148. Metzler, K.D.; Fuchs, T.A.; Nauseef, W.M.; Reumaux, D.; Roesler, J.; Schulze, I.; Wahn, V.; Papayannopoulos, V.; Zychlinsky, A. Myeloperoxidase Is Required for Neutrophil Extracellular Trap Formation: Implications for Innate Immunity. *Blood* **2011**, *117*, 953–959. [[CrossRef](#)]
149. Metzler, K.D.; Goosmann, C.; Lubojemska, A.; Zychlinsky, A.; Papayannopoulos, V. A Myeloperoxidase-Containing Complex Regulates Neutrophil Elastase Release and Actin Dynamics during NETosis. *Cell Rep.* **2014**, *8*, 883–896. [[CrossRef](#)]
150. Brodsky, S.V.; Zhang, F.; Nasjletti, A.; Goligorsky, M.S. Endothelium-Derived Microparticles Impair Endothelial Function in Vitro. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, *286*, H1910–H1915. [[CrossRef](#)]
151. Awasthi, D.; Nagarkoti, S.; Kumar, A.; Dubey, M.; Singh, A.K.; Pathak, P.; Chandra, T.; Barthwal, M.K.; Dikshit, M. Oxidized LDL Induced Extracellular Trap Formation in Human Neutrophils via TLR-PKC-IRAK-MAPK and NADPH-Oxidase Activation. *Free Radic. Biol. Med.* **2016**, *93*, 190–203. [[CrossRef](#)]
152. Warnatsch, A.; Ioannou, M.; Wang, Q.; Papayannopoulos, V. Inflammation. Neutrophil Extracellular Traps License Macrophages for Cytokine Production in Atherosclerosis. *Science* **2015**, *349*, 316–320. [[CrossRef](#)]
153. Smith, C.K.; Vivekanandan-Giri, A.; Tang, C.; Knight, J.S.; Mathew, A.; Padilla, R.L.; Gillespie, B.W.; Carmona-Rivera, C.; Liu, X.; Subramanian, V.; et al. Neutrophil Extracellular Trap-Derived Enzymes Oxidize High-Density Lipoprotein: An Additional Proatherogenic Mechanism in Systemic Lupus Erythematosus. *Arthritis Rheumatol.* **2014**, *66*, 2532–2544. [[CrossRef](#)]
154. Sharma, S.; Hofbauer, T.M.; Ondracek, A.S.; Chausheva, S.; Alimohammadi, A.; Artner, T.; Panzenboeck, A.; Rinderer, J.; Shafran, I.; Mangold, A.; et al. Neutrophil Extracellular Traps Promote Fibrous Vascular Occlusions in Chronic Thrombosis. *Blood* **2021**, *137*, 1104–1116. [[CrossRef](#)]
155. Mangold, A.; Alias, S.; Scherz, T.; Hofbauer, T.; Jakowitsch, J.; Panzenböck, A.; Simon, D.; Laimer, D.; Bangert, C.; Kammerlander, A.; et al. Coronary Neutrophil Extracellular Trap Burden and Deoxyribonuclease Activity in ST-Elevation Acute Coronary Syndrome Are Predictors of ST-Segment Resolution and Infarct Size. *Circ. Res.* **2015**, *116*, 1182–1192. [[CrossRef](#)] [[PubMed](#)]
156. Gillum, R.F. Epidemiology of Aortic Aneurysm in the United States. *J. Clin. Epidemiol.* **1995**, *48*, 1289–1298. [[CrossRef](#)] [[PubMed](#)]
157. Al-Balah, A.; Goodall, R.; Salciccioli, J.D.; Marshall, D.C.; Shalhoub, J. Mortality from Abdominal Aortic Aneurysm: Trends in European Union 15+ Countries from 1990 to 2017. *Br. J. Surg.* **2020**, *107*, 1459–1467. [[CrossRef](#)] [[PubMed](#)]

158. Treska, V.; Kocova, J.; Boudova, L.; Neprasova, P.; Topolcan, O.; Pecen, L.; Tonar, Z. Inflammation in the Wall of Abdominal Aortic Aneurysm and Its Role in the Symptomatology of Aneurysm. *Cytokines Cell Mol. Ther.* **2002**, *7*, 91–97. [CrossRef]
159. Beckman, E.N. Plasma Cell Infiltrates in Atherosclerotic Abdominal Aortic Aneurysms. *Am. J. Clin. Pathol.* **1986**, *85*, 21–24. [CrossRef] [PubMed]
160. Newmans, K.M.; Malon, A.M.; Shin, R.D.; Scholes, J.V.; Ramey, W.G.; Tilson, M.D. Matrix Metalloproteinases in Abdominal Aortic Aneurysm: Characterization, Purification, and Their Possible Sources. *Connect. Tissue Res.* **1994**, *30*, 265–276. [CrossRef]
161. Reilly, J.M.; Brophy, C.M.; Tilson, M.D. Characterization of an Elastase from Aneurysmal Aorta Which Degrades Intact Aortic Elastin. *Ann. Vasc. Surg.* **1992**, *6*, 499–502. [CrossRef]
162. Brown, P.M.; Zelt, D.T.; Sobolev, B.; Hallett, J.W.; Sternbach, Y. The Risk of Rupture in Untreated Aneurysms: The Impact of Size, Gender, and Expansion Rate. *J. Vasc. Surg.* **2003**, *37*, 280–284. [CrossRef]
163. Chaikof, E.L.; Dalman, R.L.; Eskandari, M.K.; Jackson, B.M.; Lee, W.A.; Mansour, M.A.; Mastracci, T.M.; Mell, M.; Murad, M.H.; Nguyen, L.L.; et al. The Society for Vascular Surgery Practice Guidelines on the Care of Patients with an Abdominal Aortic Aneurysm. *J. Vasc. Surg.* **2018**, *67*, 2–77.e2. [CrossRef]
164. Brady, A.R.; Thompson, S.G.; Fowkes, F.G.R.; Greenhalgh, R.M.; Powell, J.T. Abdominal Aortic Aneurysm Expansion: Risk Factors and Time Intervals for Surveillance. *Circulation* **2004**, *110*, 16–21. [CrossRef]
165. Kronmal, R.A.; McClelland, R.L.; Detrano, R.; Shea, S.; Lima, J.A.; Cushman, M.; Bild, D.E.; Burke, G.L. Risk Factors for the Progression of Coronary Artery Calcification in Asymptomatic Subjects. *Circulation* **2007**, *115*, 2722–2730. [CrossRef]
166. Liabeuf, S.; Olivier, B.; Vemeer, C.; Theuwissen, E.; Magdeleyns, E.; Aubert, C.E.; Brazier, M.; Mentaverri, R.; Hartemann, A.; Massy, Z.A. Vascular Calcification in Patients with Type 2 Diabetes: The Involvement of Matrix Gla Protein. *Cardiovasc. Diabetol.* **2014**, *13*, 85. [CrossRef]
167. Leow, K.; Szulc, P.; Schousboe, J.T.; Kiel, D.P.; Teixeira-Pinto, A.; Shaikh, H.; Sawang, M.; Sim, M.; Bondonno, N.; Hodgson, J.M.; et al. Prognostic Value of Abdominal Aortic Calcification: A Systematic Review and Meta-Analysis of Observational Studies. *J. Am. Heart Assoc.* **2021**, *10*, e017205. [CrossRef] [PubMed]
168. Rossi, A.; Targher, G.; Zoppini, G.; Ciccoira, M.; Bonapace, S.; Negri, C.; Stoico, V.; Faggiano, P.; Vassanelli, C.; Bonora, E. Aortic and Mitral Annular Calcifications Are Predictive of All-Cause and Cardiovascular Mortality in Patients with Type 2 Diabetes. *Diabetes Care* **2012**, *35*, 1781–1786. [CrossRef] [PubMed]
169. Niu, W.; Shao, J.; Yu, B.; Liu, G.; Wang, R.; Dong, H.; Che, H.; Li, L. Association Between Metformin and Abdominal Aortic Aneurysm: A Meta-Analysis. *Front. Cardiovasc. Med.* **2022**, *9*, 908747. [CrossRef] [PubMed]
170. Limiting AAA with Metformin (LIMIT) Trial—Full Text View—ClinicalTrials.gov. Available online: <https://clinicaltrials.gov/ct2/show/NCT04500756?cond=abdominal+aortic+aneurysm+metformin&draw=2&rank=3> (accessed on 5 March 2023).
171. Metformin Therapy in Non-Diabetic AAA Patients—Full Text View—ClinicalTrials.gov. Available online: <https://clinicaltrials.gov/ct2/show/NCT03507413?cond=abdominal+aortic+aneurysm+metformin&draw=2&rank=1> (accessed on 5 March 2023).
172. Metformin for Abdominal Aortic Aneurysm Growth Inhibition—Full Text View—ClinicalTrials.gov. Available online: <https://clinicaltrials.gov/ct2/show/NCT04224051?cond=abdominal+aortic+aneurysm+metformin&draw=2&rank=2> (accessed on 5 March 2023).
173. Klopff, J.; Fuchs, L.; Schernthaner, R.; Domenig, C.M.; Gollackner, B.; Brostjan, C.; Neumayer, C.; Eilenberg, W. The Prognostic Impact of Vascular Calcification on Abdominal Aortic Aneurysm Progression. *J. Vasc. Surg.* **2022**, *75*, 1926–1934. [CrossRef]
174. Kent, K.C.; Zwolak, R.M.; Egorova, N.N.; Riles, T.S.; Manganaro, A.; Moskowitz, A.J.; Gelijns, A.C.; Greco, G. Analysis of Risk Factors for Abdominal Aortic Aneurysm in a Cohort of More than 3 Million Individuals. *J. Vasc. Surg.* **2010**, *52*, 539–548. [CrossRef] [PubMed]
175. Ito, S.; Akutsu, K.; Tamori, Y.; Sakamoto, S.; Yoshimuta, T.; Hashimoto, H.; Takeshita, S. Differences in Atherosclerotic Profiles Between Patients with Thoracic and Abdominal Aortic Aneurysms. *Am. J. Cardiol.* **2008**, *101*, 696–699. [CrossRef]
176. Sun, W.; Zheng, J.; Gao, Y. Targeting Platelet Activation in Abdominal Aortic Aneurysm: Current Knowledge and Perspectives. *Biomolecules* **2022**, *12*, 206. [CrossRef] [PubMed]
177. Libby, P.; Ridker, P.M.; Maseri, A. Inflammation and Atherosclerosis. *Circulation* **2002**, *105*, 1135–1143. [CrossRef] [PubMed]
178. Raffort, J.; Lareyre, F.; Clément, M.; Hassen-Khodja, R.; Chinetti, G.; Mallat, Z. Monocytes and Macrophages in Abdominal Aortic Aneurysm. *Nat. Rev. Cardiol.* **2017**, *14*, 457–471. [CrossRef]
179. Houard, X.; Touat, Z.; Ollivier, V.; Louedec, L.; Philippe, M.; Sebbag, U.; Meilhac, O.; Rossignol, P.; Michel, J.B. Mediators of Neutrophil Recruitment in Human Abdominal Aortic Aneurysms. *Cardiovasc. Res.* **2009**, *82*, 532–541. [CrossRef]
180. Thompson, R.W.; Curci, J.A.; Ennis, T.L.; Mao, D.; Pagano, M.B.; Pham, C.T.N. Pathophysiology of Abdominal Aortic Aneurysms: Insights from the Elastase-Induced Model in Mice with Different Genetic Backgrounds. *Ann. N. Y. Acad. Sci.* **2006**, *1085*, 59–73. [CrossRef]
181. Rao, J.; Brown, B.N.; Weinbaum, J.S.; Ofstun, E.L.; Makaroun, M.S.; Humphrey, J.D.; Vorp, D.A. Distinct Macrophage Phenotype and Collagen Organization within the Intraluminal Thrombus of Abdominal Aortic Aneurysm. *J. Vasc. Surg.* **2015**, *62*, 585–593. [CrossRef]
182. Dutertre, C.A.; Clement, M.; Morvan, M.; Schäkel, K.; Castier, Y.; Alsac, J.M.; Michel, J.B.; Nicoletti, A. Deciphering the Stromal and Hematopoietic Cell Network of the Adventitia from Non-Aneurysmal and Aneurysmal Human Aorta. *PLoS ONE* **2014**, *9*, e89983. [CrossRef]

183. Tieu, B.C.; Ju, X.; Lee, C.; Sun, H.; Lejeune, W.; Recinos, A.; Brasier, A.R.; Tilton, R.G. Aortic Adventitial Fibroblasts Participate in Angiotensin-Induced Vascular Wall Inflammation and Remodeling. *J. Vasc. Res.* **2011**, *48*, 261–272. [CrossRef]
184. Tieu, B.C.; Lee, C.; Sun, H.; LeJeune, W.; Recinos, A.; Ju, X.; Spratt, H.; Guo, D.C.; Milewicz, D.; Tilton, R.G.; et al. An Adventitial IL-6/MCP1 Amplification Loop Accelerates Macrophage-Mediated Vascular Inflammation Leading to Aortic Dissection in Mice. *J. Clin. Investig.* **2009**, *119*, 3637–3651. [CrossRef]
185. Combadière, C.; Potteaux, S.; Rodero, M.; Simon, T.; Pezard, A.; Esposito, B.; Merval, R.; Proudfoot, A.; Tedgui, A.; Mallat, Z. Combined Inhibition of CCL2, CX3CR1, and CCR5 Abrogates Ly6C(Hi) and Ly6C(Lo) Monocytosis and Almost Abolishes Atherosclerosis in Hypercholesterolemic Mice. *Circulation* **2008**, *117*, 1649–1657. [CrossRef] [PubMed]
186. Daugherty, A.; Manning, M.W.; Cassis, L.A. Angiotensin II Promotes Atherosclerotic Lesions and Aneurysms in Apolipoprotein E-Deficient Mice. *J. Clin. Investig.* **2000**, *105*, 1605–1612. [CrossRef] [PubMed]
187. Deshmane, S.L.; Kremlev, S.; Amini, S.; Sawaya, B.E. Monocyte Chemoattractant Protein-1 (MCP-1): An Overview. *J. Interferon Cytokine Res.* **2009**, *29*, 313–326. [CrossRef]
188. Zhang, H.; Yang, K.; Chen, F.; Liu, Q.; Ni, J.; Cao, W.; Hua, Y.; He, F.; Liu, Z.; Li, L.; et al. Role of the CCL2-CCR2 Axis in Cardiovascular Disease: Pathogenesis and Clinical Implications. *Front. Immunol.* **2022**, *13*, 975367. [CrossRef]
189. Roshan, M.H.K.; Tambo, A.; Pace, N.P. The Role of TLR2, TLR4, and TLR9 in the Pathogenesis of Atherosclerosis. *Int. J. Inflamm.* **2016**, *2016*, 1532832. [CrossRef]
190. Yang, M.; Chen, Q.; Mei, L.; Wen, G.; An, W.; Zhou, X.; Niu, K.; Liu, C.; Ren, M.; Sun, K.; et al. Neutrophil Elastase Promotes Neointimal Hyperplasia by Targeting Toll-like Receptor 4 (TLR4)-NF-KB Signalling. *Br. J. Pharmacol.* **2021**, *178*, 4048–4068. [CrossRef] [PubMed]
191. Kolaczkowska, E.; Kubes, P. Neutrophil Recruitment and Function in Health and Inflammation. *Nat. Rev. Immunol.* **2013**, *13*, 159–175. [CrossRef]
192. Dalli, J.; Montero-Melendez, T.; Norling, L.V.; Yin, X.; Hinds, C.; Haskard, D.; Mayr, M.; Perretti, M. Heterogeneity in Neutrophil Microparticles Reveals Distinct Proteome and Functional Properties. *Mol. Cell Proteom.* **2013**, *12*, 2205–2219. [CrossRef]
193. Mortaz, E.; Alipoor, S.D.; Adcock, I.M.; Mumby, S.; Koenderman, L. Update on Neutrophil Function in Severe Inflammation. *Front. Immunol.* **2018**, *9*, 2171. [CrossRef]
194. Klopff, J.; Brostjan, C.; Eilenberg, W.; Neumayer, C. Neutrophil Extracellular Traps and Their Implications in Cardiovascular and Inflammatory Disease. *Int. J. Mol. Sci.* **2021**, *22*, 559. [CrossRef]
195. Arbănași, E.M.; Mureșan, A.V.; Coșarcă, C.M.; Arbănași, E.M.; Niculescu, R.; Voidăzan, S.T.; Ivănescu, A.D.; Hălmaciu, I.; Filep, R.C.; Mărginean, L.; et al. Computed Tomography Angiography Markers and Intraluminal Thrombus Morphology as Predictors of Abdominal Aortic Aneurysm Rupture. *Int. J. Environ. Res. Public Health* **2022**, *19*, 15961. [CrossRef] [PubMed]
196. Behr-Rasmussen, C.; Grøndal, N.; Bramsen, M.B.; Thomsen, M.D.; Lindholt, J.S. Mural Thrombus and the Progression of Abdominal Aortic Aneurysms: A Large Population-Based Prospective Cohort Study. *Eur. J. Vasc. Endovasc. Surg.* **2014**, *48*, 301–307. [CrossRef]
197. Kazi, M.; Thyberg, J.; Religa, P.; Roy, J.; Eriksson, P.; Hedin, U.; Swedenborg, J. Influence of Intraluminal Thrombus on Structural and Cellular Composition of Abdominal Aortic Aneurysm Wall. *J. Vasc. Surg.* **2003**, *38*, 1283–1292. [CrossRef] [PubMed]
198. Schrottmaier, W.C.; Mussbacher, M.; Salzmann, M.; Assinger, A. Platelet-Leukocyte Interplay during Vascular Disease. *Atherosclerosis* **2020**, *307*, 109–120. [CrossRef] [PubMed]
199. Rubenstein, D.A.; Yin, W. Platelet-Activation Mechanisms and Vascular Remodeling. *Compr. Physiol.* **2018**, *8*, 1117–1156. [CrossRef]
200. Houard, X.; Ollivier, V.; Louedec, L.; Michel, J.; Back, M. Differential Inflammatory Activity across Human Abdominal Aortic Aneurysms Reveals Neutrophil-Derived Leukotriene B4 as a Major Chemotactic Factor Released from the Intraluminal Thrombus. *FASEB J.* **2009**, *23*, 1376–1383. [CrossRef]
201. Karaolanis, G.; Moris, D.; Palla, V.V.; Karanikola, E.; Bakoyiannis, C.; Georgopoulos, S. Neutrophil Gelatinase Associated Lipocalin (NGAL) as a Biomarker. Does It Apply in Abdominal Aortic Aneurysms? A Review of Literature. *Indian J. Surg.* **2015**, *77* (Suppl. S3), 1313–1317. [CrossRef]
202. Petersen, E.; Wåberg, F.; Ängquist, K.A. Serum Concentrations of Elastin-Derived Peptides in Patients with Specific Manifestations of Atherosclerotic Disease. *Eur. J. Vasc. Endovasc. Surg.* **2002**, *24*, 440–444. [CrossRef]
203. Maguire, E.M.; Pearce, S.W.A.; Xiao, R.; Oo, A.Y.; Xiao, Q. Matrix Metalloproteinase in Abdominal Aortic Aneurysm and Aortic Dissection. *Pharmaceuticals* **2019**, *12*, 118. [CrossRef]
204. Hendy, K.; Gunnarson, R.; Golledge, J. Growth Rates of Small Abdominal Aortic Aneurysms Assessed by Computerised Tomography—A Systematic Literature Review. *Atherosclerosis* **2014**, *235*, 182–188. [CrossRef]
205. Selders, G.S.; Fetz, A.E.; Radic, M.Z.; Bowlin, G.L. An Overview of the Role of Neutrophils in Innate Immunity, Inflammation and Host-Biomaterial Integration. *Regen. Biomater.* **2017**, *4*, 55–68. [CrossRef]
206. Yan, H.; Zhou, H.F.; Akk, A.; Hu, Y.; Springer, L.E.; Ennis, T.L.; Pham, C.T.N. Neutrophil Proteases Promote Experimental Abdominal Aortic Aneurysm via Extracellular Trap Release and Plasmacytoid Dendritic Cell Activation. *Arter. Thromb. Vasc. Biol.* **2016**, *36*, 1660–1669. [CrossRef] [PubMed]
207. Delbosc, S.; Alsac, J.M.; Journe, C.; Louedec, L.; Castier, Y.; Bonnaure-Mallet, M.; Ruimy, R.; Rossignol, P.; Bouchard, P.; Michel, J.B.; et al. *Porphyromonas gingivalis* Participates in Pathogenesis of Human Abdominal Aortic Aneurysm by Neutrophil Activation. Proof of Concept in Rats. *PLoS ONE* **2011**, *6*, e18679. [CrossRef] [PubMed]

208. Jabłońska, A.; Zagrapan, B.; Paradowska, E.; Neumayer, C.; Eilenberg, W.; Brostjan, C.; Klinger, M.; Nanobachvili, J.; Huk, I. Abdominal Aortic Aneurysm and Virus Infection: A Potential Causative Role for *Cytomegalovirus* Infection? *J. Med. Virol.* **2021**, *93*, 5017–5024. [CrossRef] [PubMed]
209. Mysak, J.; Podzimek, S.; Sommerova, P.; Lyuya-Mi, Y.; Bartova, J.; Janatova, T.; Prochazkova, J.; Duskova, J. *Porphyromonas Gingivalis*: Major Periodontopathic Pathogen Overview. *J. Immunol. Res.* **2014**, *2014*, 476068. [CrossRef]
210. Salhi, L.; Rijkschroeff, P.; Van Hede, D.; Laine, M.L.; Teughels, W.; Sakalihasan, N.; Lambert, F. Blood Biomarkers and Serologic Immunological Profiles Related to Periodontitis in Abdominal Aortic Aneurysm Patients. *Front. Cell Infect. Microbiol.* **2022**, *11*, 766462. [CrossRef] [PubMed]
211. Salhi, L.; Sakalihasan, N.; Okroglic, A.G.; Labropoulos, N.; Seidel, L.; Albert, A.; Teughels, W.; Defraigne, J.O.; Lambert, F. Further Evidence on the Relationship between Abdominal Aortic Aneurysm and Periodontitis: A Cross-Sectional Study. *J. Periodontol.* **2020**, *91*, 1453–1464. [CrossRef]
212. Salhi, L.; Rompen, E.; Sakalihasan, N.; Laleman, I.; Teughels, W.; Michel, J.B.; Lambert, F. Can Periodontitis Influence the Progression of Abdominal Aortic Aneurysm? A Systematic Review. *Angiology* **2019**, *70*, 479–491. [CrossRef]
213. Gredmark-Russ, S.; Dzabic, M.; Rahbar, A.; Wanhanen, A.; Björck, M.; Larsson, E.; Michel, J.B.; Söderberg-Nauclér, C. Active *Cytomegalovirus* Infection in Aortic Smooth Muscle Cells from Patients with Abdominal Aortic Aneurysm. *J. Mol. Med.* **2009**, *87*, 347–356. [CrossRef]
214. Pinard, A.; Jones, G.T.; Milewicz, D.M. Genetics of Thoracic and Abdominal Aortic Diseases: Aneurysms, Dissections, and Ruptures. *Circ. Res.* **2019**, *124*, 588. [CrossRef]
215. La Rocca, G.; Del Frate, G.; Delvino, P.; Di Cianni, F.; Moretti, M.; Italiano, N.; Treppo, E.; Monti, S.; Talarico, R.; Ferro, F.; et al. Systemic Vasculitis: One Year in Review 2022. *Clin. Exp. Rheumatol.* **2022**, *40*, 673–687. [CrossRef]
216. Phillip, R.; Luqmani, R. Mortality in Systemic Vasculitis: A Systematic Review. *Clin. Exp. Rheumatol.* **2008**, *26* (Suppl. S51), S94–S104.
217. Wallace, Z.S.; Fu, X.; Harkness, T.; Stone, J.H.; Zhang, Y.; Choi, H. All-Cause and Cause-Specific Mortality in ANCA-Associated Vasculitis: Overall and According to ANCA Type. *Rheumatology* **2020**, *59*, 2308–2315. [CrossRef]
218. Hill, C.L.; Black, R.J.; Nossent, J.C.; Ruediger, C.; Nguyen, L.; Ninan, J.V.; Lester, S. Risk of Mortality in Patients with Giant Cell Arteritis: A Systematic Review and Meta-Analysis. *Semin. Arthritis Rheum.* **2017**, *46*, 513–519. [CrossRef]
219. Mueller, M.; Gschwandtner, M.E.; Gamper, J.; Giurgea, G.A.; Kiener, H.P.; Perkmann, T.; Koppensteiner, R.; Schlager, O. Chronic Inflammation Predicts Long-Term Mortality in Patients with Raynaud’s Phenomenon. *J. Intern. Med.* **2018**, *283*, 293–302. [CrossRef] [PubMed]
220. Chironi, G.; Pagnoux, C.; Simon, A.; Pasquinelli-Balice, M.; Del-Pino, M.; Gariepy, J.; Guillemin, L. Increased Prevalence of Subclinical Atherosclerosis in Patients with Small-Vessel Vasculitis. *Heart* **2007**, *93*, 96–99. [CrossRef] [PubMed]
221. Farrah, T.E.; Melville, V.; Czopek, A.; Fok, H.; Bruce, L.; Mills, N.L.; Bailey, M.A.; Webb, D.J.; Dear, J.W.; Dhaun, N. Arterial Stiffness, Endothelial Dysfunction and Impaired Fibrinolysis Are Pathogenic Mechanisms Contributing to Cardiovascular Risk in ANCA-Associated Vasculitis. *Kidney Int.* **2022**, *102*, 1115–1126. [CrossRef] [PubMed]
222. Clifford, A.H.; Cohen Tervaert, J.W. Cardiovascular Events and the Role of Accelerated Atherosclerosis in Systemic Vasculitis. *Atherosclerosis* **2021**, *325*, 8–15. [CrossRef]
223. Hilhorst, M.; Winckers, K.; Wilde, B.; Van Oerle, R.; Ten Cate, H.; Tervaert, J.W.C. Patients with Antineutrophil Cytoplasmic Antibodies Associated Vasculitis in Remission Are Hypercoagulable. *J. Rheumatol.* **2013**, *40*, 2042–2046. [CrossRef]
224. De Leeuw, K.; Sanders, J.S.; Stegeman, C.; Smit, A.; Kallenberg, C.G.; Bijl, M. Accelerated Atherosclerosis in Patients with Wegener’s Granulomatosis. *Ann. Rheum. Dis.* **2005**, *64*, 753–759. [CrossRef]
225. Shirai, T.; Hilhorst, M.; Harrison, D.G.; Goronzy, J.J.; Weyand, C.M. Macrophages in Vascular Inflammation—From Atherosclerosis to Vasculitis. *Autoimmunity* **2015**, *48*, 139–151. [CrossRef] [PubMed]
226. Wallace, Z.S.; Fu, X.; Liao, K.; Kallenberg, C.G.M.; Langford, C.A.; Merkel, P.A.; Monach, P.; Seo, P.; Specks, U.; Spiera, R.; et al. Disease Activity, Antineutrophil Cytoplasmic Antibody Type, and Lipid Levels in Antineutrophil Cytoplasmic Antibody-Associated Vasculitis. *Arthritis Rheumatol.* **2019**, *71*, 1879–1887. [CrossRef] [PubMed]
227. Proven, A.; Gabriel, S.E.; Orces, C.; Michael O’Fallon, W.; Hunder, G.G. Glucocorticoid Therapy in Giant Cell Arteritis: Duration and Adverse Outcomes. *Arthritis Rheum.* **2003**, *49*, 703–708. [CrossRef]
228. Bramlage, C.P.; Kröplin, J.; Wallbach, M.; Minguet, J.; Smith, K.H.; Lüders, S.; Schrader, J.; Patschan, S.; Gross, O.; Deutsch, C.; et al. Management of Cardiovascular Risk Factors in Patients with ANCA-Associated Vasculitis. *J. Eval. Clin. Pract.* **2017**, *23*, 747–754. [CrossRef]
229. Ytterberg, S.R.; Bhatt, D.L.; Mikuls, T.R.; Koch, G.G.; Fleischmann, R.; Rivas, J.L.; Germino, R.; Menon, S.; Sun, Y.; Wang, C.; et al. Cardiovascular and Cancer Risk with Tofacitinib in Rheumatoid Arthritis. *N. Engl. J. Med.* **2022**, *386*, 316–326. [CrossRef]
230. Smolen, J.S.; Landewé, R.B.M.; Bergstra, S.A.; Kerschbaumer, A.; Sepriano, A.; Aletaha, D.; Caporali, R.; Edwards, C.J.; Hyrich, K.L.; Pope, J.E.; et al. EULAR Recommendations for the Management of Rheumatoid Arthritis with Synthetic and Biological Disease-Modifying Antirheumatic Drugs: 2022 Update. *Ann. Rheum. Dis.* **2023**, *82*, 3–18. [CrossRef]
231. Ertus, C.; Scailteux, L.-M.; Lescoat, A.; Berthe, P.; Auffret, V.; Dupuy, A.; Oger, E.; Droitcourt, C. Major Adverse Cardiovascular Events in Patients Treated with Oral Janus Kinase Inhibitors for Atopic Dermatitis: A Systematic Review and Meta-Analysis. *Br. J. Dermatol.* **2023**, *6*, Ijad229. [CrossRef] [PubMed]

232. Hoisnard, L.; Pina Vegas, L.; Dray-Spira, R.; Weill, A.; Zureik, M.; Sbidian, E. Risk of Major Adverse Cardiovascular and Venous Thromboembolism Events in Patients with Rheumatoid Arthritis Exposed to JAK Inhibitors versus Adalimumab: A Nationwide Cohort Study. *Ann. Rheum. Dis.* **2023**, *82*, 182–188. [[CrossRef](#)]
233. Cooke, J.P. Inflammation and Its Role in Regeneration and Repair: A Caution for Novel Anti-Inflammatory Therapies. *Circ. Res.* **2019**, *124*, 1166. [[CrossRef](#)]
234. Mantsounga, C.S.; Lee, C.; Neverson, J.; Sharma, S.; Healy, A.; Berus, J.M.; Parry, C.; Ceneri, N.M.; López-Giráldez, F.; Chun, H.J.; et al. Macrophage IL-1 β Promotes Arteriogenesis by Autocrine STAT3- and NF-KB-Mediated Transcription of pro-Angiogenic VEGF-A. *Cell Rep.* **2022**, *38*, 110309. [[CrossRef](#)]
235. Heil, M.; Eitenmüller, I.; Schmitz-Rixen, T.; Schaper, W. Arteriogenesis versus Angiogenesis: Similarities and Differences. *J. Cell Mol. Med.* **2006**, *10*, 45. [[CrossRef](#)]
236. Wang, G.L.; Jiang, B.H.; Rue, E.A.; Semenza, G.L. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O₂ tension. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 5510–5514. [[CrossRef](#)]
237. Heil, M.; Ziegelhoeffer, T.; Wagner, S.; Fernández, B.; Helisch, A.; Martin, S.; Tribulova, S.; Kuziel, W.A.; Bachmann, G.; Schaper, W. Collateral Artery Growth (Arteriogenesis) after Experimental Arterial Occlusion Is Impaired in Mice Lacking CC-Chemokine Receptor-2. *Circ. Res.* **2004**, *94*, 671–677. [[CrossRef](#)] [[PubMed](#)]
238. Heuslein, J.L.; Meisner, J.K.; Li, X.; Song, J.; Vincentelli, H.; Leiphart, R.J.; Ames, E.G.; Blackman, B.R.; Price, R.J. Mechanisms of Amplified Arteriogenesis in Collateral Artery Segments Exposed to Flow Direction Reversal. *Arter. Thromb. Vasc. Biol.* **2015**, *35*, 2354. [[CrossRef](#)]
239. Mantovani, A.; Garlanda, C.; Locati, M. Macrophage Diversity and Polarization in Atherosclerosis: A Question of Balance. *Arter. Thromb. Vasc. Biol.* **2009**, *29*, 1419–1423. [[CrossRef](#)] [[PubMed](#)]
240. Voronov, E.; Shouval, D.S.; Krelin, Y.; Cagnano, E.; Benharoch, D.; Iwakura, Y.; Dinarello, C.A.; Apte, R.N. IL-1 Is Required for Tumor Invasiveness and Angiogenesis. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 2645. [[CrossRef](#)] [[PubMed](#)]
241. Amano, K.; Okigaki, M.; Adachi, Y.; Fujiyama, S.; Mori, Y.; Kosaki, A.; Iwasaka, T.; Matsubara, H. Mechanism for IL-1 β -Mediated Neovascularization Unmasked by IL-1 β Knock-out Mice. *J. Mol. Cell Cardiol.* **2004**, *36*, 469–480. [[CrossRef](#)]
242. De Winther, M.P.J.; Van Dijk, K.W.; Havekes, L.M.; Hofker, M.H. Macrophage Scavenger Receptor Class A: A Multifunctional Receptor in Atherosclerosis. *Arter. Thromb. Vasc. Biol.* **2000**, *20*, 290–297. [[CrossRef](#)]
243. Park, Y.M. CD36, a Scavenger Receptor Implicated in Atherosclerosis. *Exp. Mol. Med.* **2014**, *46*, e99. [[CrossRef](#)]
244. Morgan, M.J.; Liu, Z.G. Crosstalk of Reactive Oxygen Species and NF-KB Signaling. *Cell Res.* **2011**, *21*, 103–115. [[CrossRef](#)] [[PubMed](#)]
245. Xu, L.; Wang, S.; Li, B.; Sun, A.; Zou, Y.; Ge, J. A Protective Role of Ciglitazone in Ox-LDL-Induced Rat Microvascular Endothelial Cells via Modulating PPAR γ -Dependent AMPK/ENOS Pathway. *J. Cell Mol. Med.* **2015**, *19*, 92–102. [[CrossRef](#)] [[PubMed](#)]
246. Michell, B.J.; Chen, Z.P.; Tiganis, T.; Stapleton, D.; Katsis, F.; Power, D.A.; Sim, A.T.; Kemp, B.E. Coordinated Control of Endothelial Nitric-Oxide Synthase Phosphorylation by Protein Kinase C and the cAMP-Dependent Protein Kinase. *J. Biol. Chem.* **2001**, *276*, 17625–17628. [[CrossRef](#)] [[PubMed](#)]
247. Marchio, P.; Guerra-Ojeda, S.; Vila, J.M.; Aldasoro, M.; Victor, V.M.; Mauricio, M.D. Targeting Early Atherosclerosis: A Focus on Oxidative Stress and Inflammation. *Oxid. Med. Cell Longev.* **2019**, *2019*, 8563845. [[CrossRef](#)] [[PubMed](#)]
248. Chen, B.; Wei, J.; Wang, W.; Cui, G.; Zhao, Y.; Zhu, X.; Zhu, M.; Guo, W.; Yu, J. Identification of Signaling Pathways Involved in Aberrant Production of Adipokines in Adipocytes Undergoing Oxidative Stress. *Arch. Med. Res.* **2009**, *40*, 241–248. [[CrossRef](#)]
249. Chen, H.; Montagnani, M.; Funahashi, T.; Shimomura, I.; Quon, M.J. Adiponectin Stimulates Production of Nitric Oxide in Vascular Endothelial Cells. *J. Biol. Chem.* **2003**, *278*, 45021–45026. [[CrossRef](#)]
250. Motoshima, H.; Wu, X.; Mahadev, K.; Goldstein, B.J. Adiponectin Suppresses Proliferation and Superoxide Generation and Enhances ENOS Activity in Endothelial Cells Treated with Oxidized LDL. *Biochem. Biophys. Res. Commun.* **2004**, *315*, 264–271. [[CrossRef](#)]
251. Badran, A.; Nasser, S.A.; Mesmar, J.; El-Yazbi, A.F.; Bitto, A.; Fardoun, M.M.; Baydoun, E.; Eid, A.H. Reactive Oxygen Species: Modulators of Phenotypic Switch of Vascular Smooth Muscle Cells. *Int. J. Mol. Sci.* **2020**, *21*, 8764. [[CrossRef](#)]
252. Lim, S.; Quon, M.J.; Koh, K.K. Modulation of Adiponectin as a Potential Therapeutic Strategy. *Atherosclerosis* **2014**, *233*, 721–728. [[CrossRef](#)]
253. Jabłońska, A.; Neumayer, C.; Bolliger, M.; Burghuber, C.; Klinger, M.; Demyanets, S.; Nanobachvili, J.; Huk, I. Insight into the Expression of Toll-like Receptors 2 and 4 in Patients with Abdominal Aortic Aneurysm. *Mol. Biol. Rep.* **2020**, *47*, 2685–2692. [[CrossRef](#)]
254. Neumann, F.J.; Sechtem, U.; Banning, A.P.; Bonaros, N.; Bueno, H.; Bugiardini, R.; Chieffo, A.; Crea, F.; Czerny, M.; Delgado, V.; et al. 2019 ESC Guidelines for the Diagnosis and Management of Chronic Coronary Syndromes. *Eur. Heart J.* **2020**, *41*, 407–477. [[CrossRef](#)]
255. Aboyans, V.; Bauersachs, R.; Mazzolai, L.; Brodmann, M.; Palomares, J.F.R.; Debus, S.; Collet, J.P.; Drexel, H.; Espinola-Klein, C.; Lewis, B.S.; et al. Antithrombotic Therapies in Aortic and Peripheral Arterial Diseases in 2021: A Consensus Document from the ESC Working Group on Aorta and Peripheral Vascular Diseases, the ESC Working Group on Thrombosis, and the ESC Working Group on Cardiovascular Pharmacotherapy. *Eur. Heart J.* **2021**, *42*, 4013–4024. [[CrossRef](#)] [[PubMed](#)]

256. Gerhard-Herman, M.D.; Gornik, H.L.; Barrett, C.; Barshes, N.R.; Corriere, M.A.; Drachman, D.E.; Fleisher, L.A.; Fowkes, F.G.R.; Hamburg, N.M.; Kinlay, S.; et al. 2016 AHA/ACC Guideline on the Management of Patients with Lower Extremity Peripheral Artery Disease: Executive Summary: A Report of the American College of Cardiology / American Heart Association Task Force on Clinical Practice Guidelines. *Circulation* **2017**, *135*, e686–e725. [CrossRef] [PubMed]
257. Abramson, B.L.; Al-Omran, M.; Anand, S.S.; Albalawi, Z.; Coutinho, T.; de Mestral, C.; Dubois, L.; Gill, H.L.; Greco, E.; Guzman, R.; et al. Canadian Cardiovascular Society 2022 Guidelines for Peripheral Arterial Disease. *Can. J. Cardiol.* **2022**, *38*, 560–587. [CrossRef]
258. Frank, U.; Nikol, S.; Belch, J.; Boc, V.; Brodmann, M.; Carpentier, P.H.; Chraim, A.; Canning, C.; Dimakakos, E.; Gottsäter, A.; et al. ESVM Guideline on Peripheral Arterial Disease. *Vasa* **2019**, *48* (Suppl. S102), 1–80. [CrossRef]
259. El Assar, M.; Álvarez-Bustos, A.; Sosa, P.; Angulo, J.; Rodríguez-Mañas, L. Effect of Physical Activity/Exercise on Oxidative Stress and Inflammation in Muscle and Vascular Aging. *Int. J. Mol. Sci.* **2022**, *23*, 8713. [CrossRef]
260. Myette-Côté, É.; Durrer, C.; Neudorf, H.; Bammert, T.D.; Botezelli, J.D.; Johnson, J.D.; Desouza, C.A.; Little, J.P. The Effect of a Short-Term Low-Carbohydrate, High-Fat Diet with or without Postmeal Walks on Glycemic Control and Inflammation in Type 2 Diabetes: A Randomized Trial. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2018**, *315*, R1210–R1219. [CrossRef]
261. Stancu, C.; Sima, A. Statins: Mechanism of Action and Effects. *J. Cell Mol. Med.* **2001**, *5*, 378–387. [CrossRef]
262. Almeida, S.O.; Budoff, M. Effect of Statins on Atherosclerotic Plaque. *Trends Cardiovasc. Med.* **2019**, *29*, 451–455. [CrossRef]
263. Greenwood, J.; Mason, J.C. Statins and the Vascular Endothelial Inflammatory Response. *Trends Immunol.* **2007**, *28*, 88–98. [CrossRef]
264. Piechota-Polanczyk, A.; Demyanets, S.; Nykonenko, O.; Huk, I.; Mittlboeck, M.; Domenig, C.M.; Neumayer, C.; Wojta, J.; Nanobachvili, J.; Klinger, M. Decreased Tissue Levels of Cyclophilin A, a Cyclosporine a Target and Phospho-ERK1/2 in Simvastatin Patients with Abdominal Aortic Aneurysm. *Eur. J. Vasc. Endovasc. Surg.* **2013**, *45*, 682–688. [CrossRef]
265. Piechota-Polanczyk, A.; Goraca, A.; Demyanets, S.; Mittlboeck, M.; Domenig, C.; Neumayer, C.; Wojta, J.; Nanobachvili, J.; Huk, I.; Klinger, M. Simvastatin Decreases Free Radicals Formation in the Human Abdominal Aortic Aneurysm Wall via NF-KB. *Eur. J. Vasc. Endovasc. Surg.* **2012**, *44*, 133–137. [CrossRef] [PubMed]
266. Piechota-Polanczyk, A.; Demyanets, S.; Mittlboeck, M.; Hofmann, M.; Domenig, C.M.; Neumayer, C.; Wojta, J.; Klinger, M.; Nanobachvili, J.; Huk, I. The Influence of Simvastatin on NGAL, Matrix Metalloproteinases and Their Tissue Inhibitors in Human Intraluminal Thrombus and Abdominal Aortic Aneurysm Tissue. *Eur. J. Vasc. Endovasc. Surg.* **2015**, *49*, 549–555. [CrossRef] [PubMed]
267. Steiner, S.; Speidl, W.S.; Pleiner, J.; Seidinger, D.; Zorn, G.; Kaun, C.; Wojta, J.; Huber, K.; Minar, E.; Wolzt, M.; et al. Simvastatin Blunts Endotoxin-Induced Tissue Factor in Vivo. *Circulation* **2005**, *111*, 1841–1846. [CrossRef] [PubMed]
268. Ridker, P.M.; Cannon, C.P.; Morrow, D.; Rifai, N.; Rose, L.M.; McCabe, C.H.; Pfeffer, M.A.; Braunwald, E. C-Reactive Protein Levels and Outcomes after Statin Therapy. *N. Engl. J. Med.* **2005**, *352*, 8–9. [CrossRef]
269. Tawakol, A.; Fayad, Z.A.; Mogg, R.; Alon, A.; Klimas, M.T.; Dansky, H.; Subramanian, S.S.; Abdelbaky, A.; Rudd, J.H.F.; Farkouh, M.E.; et al. Intensification of Statin Therapy Results in a Rapid Reduction in Atherosclerotic Inflammation: Results of a Multicenter Fluorodeoxyglucose-Positron Emission Tomography/Computed Tomography Feasibility Study. *J. Am. Coll. Cardiol.* **2013**, *62*, 909–917. [CrossRef]
270. Ridker, P.M.; Bhatt, D.L.; Pradhan, A.D.; Glynn, R.J.; MacFadyen, J.G.; Nissen, S.E. Inflammation and Cholesterol as Predictors of Cardiovascular Events among Patients Receiving Statin Therapy: A Collaborative Analysis of Three Randomised Trials. *Lancet* **2023**, *401*, 1293–1301. [CrossRef]
271. Wang, J.; Chen, Z.; Qiu, Y.; Wu, L.; Wang, H.; Wu, L.; Zhao, L.; Xie, D. Statins Have an Anti-Inflammation in CKD Patients: A Meta-Analysis of Randomized Trials. *Biomed. Res. Int.* **2022**, *2022*, 4842699. [CrossRef]
272. Zhang, Q.X.; Zhang, H.F.; Lu, X.T.; Zhao, J.; Xu, Q. Statins Improve Asthma Symptoms by Suppressing Inflammation: A Meta-Analysis Based on RCTs. *Eur. Rev. Med. Pharmacol. Sci.* **2022**, *26*, 8401–8410. [CrossRef]
273. Valerius, N.H. In Vitro Effect of Colchicine on Neutrophil Granulocyte Locomotion. Assessment of the Effect of Colchicine on Chemotaxis, Chemokinesis and Spontaneous Motility, Using a Modified Reversible Boyden Chamber. *Acta Pathol. Microbiol. Scand. B* **1978**, *86*, 149–154.
274. Li, Z.; Davis, G.S.; Mohr, C.; Nain, M.; Gemsa, D. Inhibition of LPS-Induced Tumor Necrosis Factor-Alpha Production by Colchicine and Other Microtubule Disrupting Drugs. *Immunobiology* **1996**, *195*, 624–639. [CrossRef] [PubMed]
275. Wright, D.G.; Malawista, S.E. Mobilization and Extracellular Release of Granular Enzymes from Human Leukocytes during Phagocytosis: Inhibition by Colchicine and Cortisol but Not by Salicylate. *Arthritis Rheum.* **1973**, *16*, 749–758. [CrossRef] [PubMed]
276. Martínez, G.J.; Celermajer, D.S.; Patel, S. The NLRP3 Inflammasome and the Emerging Role of Colchicine to Inhibit Atherosclerosis-Associated Inflammation. *Atherosclerosis* **2018**, *269*, 262–271. [CrossRef] [PubMed]
277. González, L.; Bulnes, J.F.; Orellana, M.P.; Venturelli, P.M.; Rodriguez, G.M. The Role of Colchicine in Atherosclerosis: From Bench to Bedside. *Pharmaceutics* **2022**, *14*, 1395. [CrossRef] [PubMed]
278. Nidorf, S.M.; Thompson, P.L. Why Colchicine Should Be Considered for Secondary Prevention of Atherosclerosis: An Overview. *Clin. Ther.* **2019**, *41*, 41–48. [CrossRef]
279. Tardif, J.-C.; Kouz, S.; Waters, D.D.; Bertrand, O.F.; Diaz, R.; Maggioni, A.P.; Pinto, F.J.; Ibrahim, R.; Gamra, H.; Kiwan, G.S.; et al. Efficacy and Safety of Low-Dose Colchicine after Myocardial Infarction. *N. Engl. J. Med.* **2019**, *381*, 2497–2505. [CrossRef]

280. Wang, L.; Peng, Y.; Song, L.; Xia, D.; Li, C.; Li, Z.; Li, Q.; Yu, A.; Lu, C.; Wang, Y. Colchicine-Containing Nanoparticles Attenuates Acute Myocardial Infarction Injury by Inhibiting Inflammation. *Cardiovasc. Drugs Ther.* **2022**, *36*, 1075–1089. [CrossRef]
281. Chen, Y.; Zhang, H.; Chen, Y.; Li, M.; Luo, W.; Liu, Y.; Fu, Y.; Xia, H.; Xu, C.; Jiang, Y.; et al. Colchicine May Become a New Cornerstone Therapy for Coronary Artery Disease: A Meta-Analysis of Randomized Controlled Trials. *Clin. Rheumatol.* **2022**, *41*, 1873–1887. [CrossRef]
282. Bays, H.E.; Ballantyne, C.M.; Braeckman, R.A.; Stirman, W.G.; Soni, P.N. Icosapent Ethyl, a Pure Ethyl Ester of Eicosapentaenoic Acid: Effects on Circulating Markers of Inflammation from the MARINE and ANCHOR Studies. *Am. J. Cardiovasc. Drugs* **2013**, *13*, 37–46. [CrossRef]
283. Bhatt, D.L.; Steg, P.G.; Miller, M.; Brinton, E.A.; Jacobson, T.A.; Ketchum, S.B.; Doyle, R.T.; Juliano, R.A.; Jiao, L.; Granowitz, C.; et al. Cardiovascular Risk Reduction with Icosapent Ethyl for Hypertriglyceridemia. *N. Engl. J. Med.* **2019**, *380*, 11–22. [CrossRef]
284. Budoff, M.J.; Bhatt, D.L.; Kinninger, A.; Lakshmanan, S.; Muhlestein, J.B.; Le, V.T.; May, H.T.; Shaikh, K.; Shekar, C.; Roy, S.K.; et al. Effect of Icosapent Ethyl on Progression of Coronary Atherosclerosis in Patients with Elevated Triglycerides on Statin Therapy: Final Results of the EVAPORATE Trial. *Eur. Heart J.* **2020**, *41*, 3925–3932. [CrossRef]
285. Mason, R.P.; Libby, P.; Bhatt, D.L. Emerging Mechanisms of Cardiovascular Protection for the Omega-3 Fatty Acid Eicosapentaenoic Acid. *Arter. Thromb. Vasc. Biol.* **2020**, *40*, 1135–1147. [CrossRef] [PubMed]
286. Ballantyne, C.M.; Braeckman, R.A.; Bays, H.E.; Kastelein, J.J.; Otvos, J.D.; Stirman, W.G.; Doyle, R.T.; Soni, P.N.; Juliano, R.A. Effects of Icosapent Ethyl on Lipoprotein Particle Concentration and Size in Statin-Treated Patients with Persistent High Triglycerides (the ANCHOR Study). *J. Clin. Lipidol.* **2015**, *9*, 377–383. [CrossRef] [PubMed]
287. Onat, U.I.; Yıldırım, A.D.; Tufanlı, Ö.; Çimen, I.; Kocatürk, B.; Veli, Z.; Hamid, S.M.; Shimada, K.; Chen, S.; Sin, J.; et al. Intercepting the Lipid-Induced Integrated Stress Response Reduces Atherosclerosis. *J. Am. Coll. Cardiol.* **2019**, *73*, 1149–1169. [CrossRef] [PubMed]
288. Arnold, D.D.; Yalamanoglu, A.; Boyman, O. Systematic Review of Safety and Efficacy of IL-1-Targeted Biologics in Treating Immune-Mediated Disorders. *Front. Immunol.* **2022**, *13*, 888392. [CrossRef] [PubMed]
289. Ridker, P.M.; Thuren, T.; Zalewski, A.; Libby, P. Interleukin-1 β Inhibition and the Prevention of Recurrent Cardiovascular Events: Rationale and Design of the Canakinumab Anti-Inflammatory Thrombosis Outcomes Study (CANTOS). *Am. Heart J.* **2011**, *162*, 597–605. [CrossRef]
290. Ridker, P.M. From C-Reactive Protein to Interleukin-6 to Interleukin-1: Moving Upstream To Identify Novel Targets for Atheroprotection. *Circ. Res.* **2016**, *118*, 145–156. [CrossRef]
291. Ku, E.J.; Kim, B.R.; Lee, J.I.; Lee, Y.K.; Oh, T.J.; Jang, H.C.; Choi, S.H. The Anti-Atherosclerosis Effect of Anakinra, a Recombinant Human Interleukin-1 Receptor Antagonist, in Apolipoprotein E Knockout Mice. *Int. J. Mol. Sci.* **2022**, *23*, 4906. [CrossRef]
292. Ridker, P.M.; Howard, C.P.; Walter, V.; Everett, B.; Libby, P.; Hensen, J.; Thuren, T. Effects of Interleukin-1 β Inhibition with Canakinumab on Hemoglobin A1c, Lipids, C-Reactive Protein, Interleukin-6, and Fibrinogen: A Phase IIb Randomized, Placebo-Controlled Trial. *Circulation* **2012**, *126*, 2739–2748. [CrossRef]
293. Straub, R.H.; Cutolo, M. Glucocorticoids and Chronic Inflammation. *Rheumatology* **2016**, *55* (Suppl. S2), ii6–ii14. [CrossRef]
294. Cain, D.W.; Cidlowski, J.A. Immune Regulation by Glucocorticoids. *Nat. Rev. Immunol.* **2017**, *17*, 233–247. [CrossRef] [PubMed]
295. Patel, R.; Williams-Dautovich, J.; Cummins, C.L. Minireview: New Molecular Mediators of Glucocorticoid Receptor Activity in Metabolic Tissues. *Mol. Endocrinol.* **2014**, *28*, 999–1011. [CrossRef] [PubMed]
296. Cicala, M.V.; Mantero, F. Hypertension in Cushing's Syndrome: From Pathogenesis to Treatment. *Neuroendocrinology* **2010**, *92* (Suppl. S1), 44–49. [CrossRef] [PubMed]
297. Akalestou, E.; Genser, L.; Rutter, G.A. Glucocorticoid Metabolism in Obesity and Following Weight Loss. *Front. Endocrinol.* **2020**, *11*, 59. [CrossRef]
298. Arnaldi, G.; Scandali, V.M.; Tremantino, L.; Cardinaletti, M.; Appolloni, G.; Boscaro, M. Pathophysiology of Dyslipidemia in Cushing's Syndrome. *Neuroendocrinology* **2010**, *92* (Suppl. S1), 86–90. [CrossRef]
299. Coelho, M.C.A.; Santos, C.V.; Neto, L.V.; Gadelha, M.R. Adverse Effects of Glucocorticoids: Coagulopathy. *Eur. J. Endocrinol.* **2015**, *173*, M11–M21. [CrossRef]
300. Faggiano, A.; Pivonello, R.; Spiezzi, S.; De Martino, M.C.; Filippella, M.; Di Somma, C.; Lombardi, G.; Colao, A. Cardiovascular Risk Factors and Common Carotid Artery Caliber and Stiffness in Patients with Cushing's Disease during Active Disease and 1 Year after Disease Remission. *J. Clin. Endocrinol. Metab.* **2003**, *88*, 2527–2533. [CrossRef]
301. Macleod, C.; Hadoke, P.W.F.; Nixon, M. Glucocorticoids: Fuelling the Fire of Atherosclerosis or Therapeutic Extinguishers? *Int. J. Mol. Sci.* **2021**, *22*, 7622. [CrossRef]
302. Petramala, L.; Lorenzo, D.; Iannucci, G.; Concistré, A.; Zinnamosca, L.; Marinelli, C.; De Vincentis, G.; Ciardi, A.; De Toma, G.; Letizia, C. Subclinical Atherosclerosis in Patients with Cushing Syndrome: Evaluation with Carotid Intima-Media Thickness and Ankle-Brachial Index. *Endocrinol. Metab.* **2015**, *30*, 488–493. [CrossRef]
303. Souverein, P.C.; Berard, A.; Van Staa, T.P.; Cooper, C.; Egberts, A.C.G.; Leufkens, H.G.M.; Walker, B.R. Use of Oral Glucocorticoids and Risk of Cardiovascular and Cerebrovascular Disease in a Population Based Case-Control Study. *Heart* **2004**, *90*, 859–865. [CrossRef]
304. Wei, L.; MacDonald, T.M.; Walker, B.R. Taking Glucocorticoids by Prescription Is Associated with Subsequent Cardiovascular Disease. *Ann. Intern. Med.* **2004**, *141*, 764–770. [CrossRef] [PubMed]

305. Pujades-Rodriguez, M.; Morgan, A.W.; Cubbon, R.M.; Wu, J. Dose-Dependent Oral Glucocorticoid Cardiovascular Risks in People with Immune-Mediated Inflammatory Diseases: A Population-Based Cohort Study. *PLoS Med.* **2020**, *17*, e1003432. [CrossRef] [PubMed]
306. Katsiki, N.; Ferrannini, E. Anti-Inflammatory Properties of Antidiabetic Drugs: A “Promised Land” in the COVID-19 Era? *J. Diabetes Complicat.* **2020**, *34*, 107723. [CrossRef] [PubMed]
307. Vaduganathan, M.; Docherty, K.F.; Claggett, B.L.; Jhund, P.S.; de Boer, R.A.; Hernandez, A.F.; Inzucchi, S.E.; Kosiborod, M.N.; Lam, C.S.P.; Martinez, F.; et al. SGLT-2 Inhibitors in Patients with Heart Failure: A Comprehensive Meta-Analysis of Five Randomised Controlled Trials. *Lancet* **2022**, *400*, 757–767. [CrossRef] [PubMed]
308. McMurray, J.J.V.; Solomon, S.D.; Inzucchi, S.E.; Køber, L.; Kosiborod, M.N.; Martinez, F.A.; Ponikowski, P.; Sabatine, M.S.; Anand, I.S.; Bělohlávek, J.; et al. Dapagliflozin in Patients with Heart Failure and Reduced Ejection Fraction. *N. Engl. J. Med.* **2019**, *381*, 1995–2008. [CrossRef]
309. Solomon, S.D.; McMurray, J.J.V.; Claggett, B.; de Boer, R.A.; DeMets, D.; Hernandez, A.F.; Inzucchi, S.E.; Kosiborod, M.N.; Lam, C.S.P.; Martinez, F.; et al. Dapagliflozin in Heart Failure with Mildly Reduced or Preserved Ejection Fraction. *N. Engl. J. Med.* **2022**, *387*, 1089–1098. [CrossRef]
310. Packer, M.; Anker, S.D.; Butler, J.; Filippatos, G.; Pocock, S.J.; Carson, P.; Januzzi, J.; Verma, S.; Tsutsui, H.; Brueckmann, M.; et al. Cardiovascular and Renal Outcomes with Empagliflozin in Heart Failure. *N. Engl. J. Med.* **2020**, *383*, 1413–1424. [CrossRef]
311. Anker, S.D.; Butler, J.; Filippatos, G.; Ferreira, J.P.; Bocchi, E.; Böhm, M.; Brunner-La Rocca, H.-P.; Choi, D.-J.; Chopra, V.; Chuquiere-Valenzuela, E.; et al. Empagliflozin in Heart Failure with a Preserved Ejection Fraction. *N. Engl. J. Med.* **2021**, *385*, 1451–1461. [CrossRef]
312. Heidenreich, P.A.; Bozkurt, B.; Aguilar, D.; Allen, L.A.; Byun, J.J.; Colvin, M.M.; Deswal, A.; Drazner, M.H.; Dunlay, S.M.; Evers, L.R.; et al. 2022 AHA/ACC/HFSA Guideline for the Management of Heart Failure: A Report of the American College of Cardiology/American Heart Association Joint Committee on Clinical Practice Guidelines. *Circulation* **2022**, *145*, E895–E1032. [CrossRef]
313. Hou, Y.C.; Zheng, C.M.; Yen, T.H.; Lu, K.C. Molecular Mechanisms of SGLT2 Inhibitor on Cardiorenal Protection. *Int. J. Mol. Sci.* **2020**, *21*, 7833. [CrossRef]
314. Liu, Z.; Ma, X.; Ilyas, I.; Zheng, X.; Luo, S.; Little, P.J.; Kamato, D.; Sahebkar, A.; Wu, W.; Weng, J.; et al. Impact of Sodium Glucose Cotransporter 2 (SGLT2) Inhibitors on Atherosclerosis: From Pharmacology to Pre-Clinical and Clinical Therapeutics. *Theranostics* **2021**, *11*, 4502–4515. [CrossRef] [PubMed]
315. Day, E.A.; Ford, R.J.; Lu, J.H.; Lu, R.; Lundberg, L.; Desjardins, E.M.; Green, A.E.; Lally, J.S.V.; Schertzer, J.D.; Steinberg, G.R. The SGLT2 Inhibitor Canagliflozin Suppresses Lipid Synthesis and Interleukin-1 Beta in ApoE Deficient Mice. *Biochem. J.* **2020**, *477*, 2347–2361. [CrossRef] [PubMed]
316. Nasiri-Ansari, N.; Dimitriadis, G.K.; Agrogiannis, G.; Perrea, D.; Kostakis, I.D.; Kaltsas, G.; Papavassiliou, A.G.; Randeva, H.S.; Kassi, E. Canagliflozin Attenuates the Progression of Atherosclerosis and Inflammation Process in APOE Knockout Mice. *Cardiovasc. Diabetol.* **2018**, *17*, 106. [CrossRef] [PubMed]
317. Mancini, S.J.; Boyd, D.; Katwan, O.J.; Strembitska, A.; Almabrouk, T.A.; Kennedy, S.; Palmer, T.M.; Salt, I.P. Canagliflozin Inhibits Interleukin-1 β -Stimulated Cytokine and Chemokine Secretion in Vascular Endothelial Cells by AMP-Activated Protein Kinase-Dependent and -Independent Mechanisms. *Sci. Rep.* **2018**, *8*, 5276. [CrossRef]
318. Kim, S.R.; Lee, S.G.; Kim, S.H.; Kim, J.H.; Choi, E.; Cho, W.; Rim, J.H.; Hwang, I.; Lee, C.J.; Lee, M.; et al. SGLT2 Inhibition Modulates NLRP3 Inflammasome Activity via Ketones and Insulin in Diabetes with Cardiovascular Disease. *Nat. Commun.* **2020**, *11*, 2127. [CrossRef]
319. Chen, H.; Teng, D.; Xu, B.; Wang, C.; Wang, H.; Jia, W.; Gong, L.; Dong, H.; Zhong, L.; Yang, J. The SGLT2 Inhibitor Canagliflozin Reduces Atherosclerosis by Enhancing Macrophage Autophagy. *J. Cardiovasc. Transl. Res.* **2023**; *Online ahead of print*. [CrossRef]
320. Chaudhuri, A.; Ghanim, H.; Vora, M.; Sia, C.L.; Korzeniewski, K.; Dhindsa, S.; Makdissi, A.; Dandona, P. Exenatide Exerts a Potent Antiinflammatory Effect. *J. Clin. Endocrinol. Metab.* **2012**, *97*, 198–207. [CrossRef]
321. Jonik, S.; Marchel, M.; Grabowski, M.; Opolski, G.; Mazurek, T. Gastrointestinal Incretins—Glucose-Dependent Insulinotropic Polypeptide (GIP) and Glucagon-like Peptide-1 (GLP-1) beyond Pleiotropic Physiological Effects Are Involved in Pathophysiology of Atherosclerosis and Coronary Artery Disease—State of the Art. *Biology* **2022**, *11*, 288. [CrossRef]
322. Rakipovski, G.; Rolin, B.; Nøhr, J.; Klewe, I.; Frederiksen, K.S.; Augustin, R.; Hecksher-Sørensen, J.; Ingvorsen, C.; Polex-Wolf, J.; Knudsen, L.B. The GLP-1 Analogs Liraglutide and Semaglutide Reduce Atherosclerosis in ApoE $^{-/-}$ and LDLr $^{-/-}$ Mice by a Mechanism That Includes Inflammatory Pathways. *JACC Basic. Transl. Sci.* **2018**, *3*, 844–857. [CrossRef]
323. Bendotti, G.; Montefusco, L.; Lunati, M.E.; Usuelli, V.; Pastore, I.; Lazzaroni, E.; Assi, E.; Seelam, A.J.; El Essawy, B.; Jang, Y.; et al. The Anti-Inflammatory and Immunological Properties of GLP-1 Receptor Agonists. *Pharmacol. Res.* **2022**, *182*, 106320. [CrossRef]
324. Bray, J.J.H.; Foster-Davies, H.; Salem, A.; Hoole, A.L.; Obaid, D.R.; Halcox, J.P.J.; Stephens, J.W. Glucagon-like Peptide-1 Receptor Agonists Improve Biomarkers of Inflammation and Oxidative Stress: A Systematic Review and Meta-Analysis of Randomised Controlled Trials. *Diabetes Obes. Metab.* **2021**, *23*, 1806–1822. [CrossRef]
325. Bethel, M.A.; Patel, R.A.; Merrill, P.; Lokhnygina, Y.; Buse, J.B.; Mentz, R.J.; Pagidipati, N.J.; Chan, J.C.; Gustavson, S.M.; Iqbal, N.; et al. Cardiovascular Outcomes with Glucagon-like Peptide-1 Receptor Agonists in Patients with Type 2 Diabetes: A Meta-Analysis. *Lancet Diabetes Endocrinol.* **2018**, *6*, 105–113. [CrossRef]

326. Werkman, N.C.C.; Driessens, J.H.M.; Stehouwer, C.D.A.; Vestergaard, P.; Schaper, N.C.; van den Bergh, J.P.; Nielsen, J.T.H. The Use of Sodium-Glucose Co-Transporter-2 Inhibitors or Glucagon-like Peptide-1 Receptor Agonists versus Sulfonylureas and the Risk of Lower Limb Amputations: A Nation-Wide Cohort Study. *Cardiovasc. Diabetol.* **2023**, *22*, 160. [CrossRef] [PubMed]
327. Marshall, S.M. 60 Years of Metformin Use: A Glance at the Past and a Look to the Future. *Diabetologia* **2017**, *60*, 1561–1565. [CrossRef] [PubMed]
328. Campbell, J.M.; Bellman, S.M.; Stephenson, M.D.; Lisy, K. Metformin Reduces All-Cause Mortality and Diseases of Ageing Independent of Its Effect on Diabetes Control: A Systematic Review and Meta-Analysis. *Ageing Res. Rev.* **2017**, *40*, 31–44. [CrossRef] [PubMed]
329. Feng, X.; Chen, W.; Ni, X.; Little, P.J.; Xu, S.; Tang, L.; Weng, J. Metformin, Macrophage Dysfunction and Atherosclerosis. *Front. Immunol.* **2021**, *12*, 682853. [CrossRef] [PubMed]
330. Poznyak, A.V.; Litvinova, L.; Poggio, P.; Moschetta, D.; Sukhorukov, V.N.; Orekhov, A.N. From Diabetes to Atherosclerosis: Potential of Metformin for Management of Cardiovascular Disease. *Int. J. Mol. Sci.* **2022**, *23*, 9738. [CrossRef]
331. Zhao, Y.; Zhao, Y.; Tian, Y.; Zhou, Y. Metformin Suppresses Foam Cell Formation, Inflammation and Ferroptosis via the AMPK/ERK Signaling Pathway in Ox-LDL-induced THP-1 Monocytes. *Exp. Ther. Med.* **2022**, *24*, 636. [CrossRef]
332. Yang, Q.; Yuan, H.; Chen, M.; Qu, J.; Wang, H.; Yu, B.; Chen, J.; Sun, S.; Tang, X.; Ren, W. Metformin Ameliorates the Progression of Atherosclerosis via Suppressing Macrophage Infiltration and Inflammatory Responses in Rabbits. *Life Sci.* **2018**, *198*, 56–64. [CrossRef]
333. Kanigur Sultuybek, G.; Soydas, T.; Yenmis, G. NF-KB as the Mediator of Metformin’s Effect on Ageing and Ageing-Related Diseases. *Clin. Exp. Pharmacol. Physiol.* **2019**, *46*, 413–422. [CrossRef]
334. Gräni, C.; Eichhorn, C.; Bière, L.; Murthy, V.L.; Agarwal, V.; Kaneko, K.; Cuddy, S.; Aghayev, A.; Steigner, M.; Blankstein, R.; et al. Prognostic Value of Cardiac Magnetic Resonance Tissue Characterization in Risk Stratifying Patients with Suspected Myocarditis. *J. Am. Coll. Cardiol.* **2017**, *70*, 1964. [CrossRef]
335. Roussel, R.; Travert, F.; Pasquet, B.; Wilson, P.W.F.; Smith, S.C.; Goto, S.; Ravaud, P.; Marre, M.; Porath, A.; Bhatt, D.L.; et al. Metformin Use and Mortality among Patients with Diabetes and Atherothrombosis. *Arch. Intern. Med.* **2010**, *170*, 1892–1899. [CrossRef] [PubMed]
336. Meaney, E.; Vela, A.; Samaniego, V.; Meaney, A.; Asbún, J.; Zempoalteca, J.C.; Elisa, Z.N.; Emma, M.N.; Guzman, M.; Hicks, J.; et al. Metformin, Arterial Function, Intima-Media Thickness and Nitroxidation in Metabolic Syndrome: The Mefisto Study. *Clin. Exp. Pharmacol. Physiol.* **2008**, *35*, 895–903. [CrossRef] [PubMed]
337. Jadhav, S.; Ferrell, W.; Greer, I.A.; Petrie, J.R.; Cobbe, S.M.; Sattar, N. Effects of Metformin on Microvascular Function and Exercise Tolerance in Women with Angina and Normal Coronary Arteries: A Randomized, Double-Blind, Placebo-Controlled Study. *J. Am. Coll. Cardiol.* **2006**, *48*, 956–963. [CrossRef] [PubMed]
338. Preiss, D.; Lloyd, S.M.; Ford, I.; McMurray, J.J.; Holman, R.R.; Welsh, P.; Fisher, M.; Packard, C.J.; Sattar, N. Metformin for Non-Diabetic Patients with Coronary Heart Disease (the CAMERA Study): A Randomised Controlled Trial. *Lancet Diabetes Endocrinol.* **2014**, *2*, 116–124. [CrossRef]
339. Luo, F.; Das, A.; Chen, J.; Wu, P.; Li, X.; Fang, Z. Metformin in Patients with and without Diabetes: A Paradigm Shift in Cardiovascular Disease Management. *Cardiovasc. Diabetol.* **2019**, *18*, 54. [CrossRef]
340. Griffin, S.J.; Angelyn Bethel, M.; Holman, R.R.; Khunti, K.; Wareham, N.; Brierley, G.; Davies, M.; Dymond, A.; Eichenberger, R.; Evans, P.; et al. Metformin in Non-Diabetic Hyperglycaemia: The GLINT Feasibility RCT. *Health Technol. Assess.* **2018**, *22*, 1–64. [CrossRef]
341. ISRCTN—ISRCTN34875079: The Glucose Lowering in Non-Diabetic hyperglycaemia Trial (GLINT)—Glucose Lowering in Those at Risk of Diabetes. Available online: <https://www.isrctn.com/ISRCTN34875079> (accessed on 14 June 2023).
342. National Institute of Diabetes and Digestive and Kidney Diseases. Dipeptidyl Peptidase-4 Inhibitors. Available online: <https://pubmed.ncbi.nlm.nih.gov/31643671/> (accessed on 14 June 2023).
343. Cosentino, F.; Grant, P.J.; Abeyans, V.; Bailey, C.J.; Ceriello, A.; Delgado, V.; Federici, M.; Filippatos, G.; Grobbee, D.E.; Hansen, T.B.; et al. 2019 ESC Guidelines on Diabetes, Pre-Diabetes, and Cardiovascular Diseases Developed in Collaboration with the EASDThe Task Force for Diabetes, Pre-Diabetes, and Cardiovascular Diseases of the European Society of Cardiology (ESC) and the European Association for the Study of Diabetes (EASD). *Eur. Heart J.* **2020**, *41*, 255–323. [CrossRef]
344. Liu, H.; Guo, L.; Xing, J.; Li, P.; Sang, H.; Hu, X.; Du, Y.; Zhao, L.; Song, R.; Gu, H. The Protective Role of DPP4 Inhibitors in Atherosclerosis. *Eur. J. Pharmacol.* **2020**, *875*, 173037. [CrossRef]
345. Lee, D.S.; Lee, E.S.; Alam, M.M.; Jang, J.H.; Lee, H.S.; Oh, H.; Kim, Y.C.; Manzoor, Z.; Koh, Y.S.; Kang, D.G.; et al. Soluble DPP-4 up-Regulates Toll-like Receptors and Augments Inflammatory Reactions, Which Are Ameliorated by Vildagliptin or Mannose-6-Phosphate. *Metabolism* **2016**, *65*, 89–101. [CrossRef]
346. Romacho, T.; Vallejo, S.; Villalobos, L.A.; Wronkowitz, N.; Indrakusuma, I.; Sell, H.; Eckel, J.; Sanchez-Ferrer, C.F.; Peiro, C. Soluble Dipeptidyl Peptidase-4 Induces Microvascular Endothelial Dysfunction through Proteinase-Activated Receptor-2 and Thromboxane A2 Release. *J. Hypertens.* **2016**, *34*, 869–876. [CrossRef]
347. Tang, S.T.; Su, H.; Zhang, Q.; Tang, H.Q.; Wang, C.J.; Zhou, Q.; Wei, W.; Zhu, H.Q.; Wang, Y. Sitagliptin Inhibits Endothelin-1 Expression in the Aortic Endothelium of Rats with Streptozotocin-Induced Diabetes by Suppressing the Nuclear Factor-KB/I κ B α System through the Activation of AMP-Activated Protein Kinase. *Int. J. Mol. Med.* **2016**, *37*, 1558–1566. [CrossRef] [PubMed]

348. Matsubara, J.; Sugiyama, S.; Sugamura, K.; Nakamura, T.; Fujiwara, Y.; Akiyama, E.; Kurokawa, H.; Nozaki, T.; Ohba, K.; Konishi, M.; et al. A Dipeptidyl Peptidase-4 Inhibitor, Des-Fluoro-Sitagliptin, Improves Endothelial Function and Reduces Atherosclerotic Lesion Formation in Apolipoprotein E-Deficient Mice. *J. Am. Coll. Cardiol.* **2012**, *59*, 265–276. [CrossRef] [PubMed]
349. Liu, H.; Xiang, H.; Zhao, S.; Sang, H.; Lv, F.; Chen, R.; Shu, Z.; Chen, A.F.; Chen, S.; Lu, H. Vildagliptin Improves High Glucose-Induced Endothelial Mitochondrial Dysfunction via Inhibiting Mitochondrial Fission. *J. Cell Mol. Med.* **2019**, *23*, 798–810. [CrossRef] [PubMed]
350. Fadini, G.P.; Boscaro, E.; Albiero, M.; Menegazzo, L.; Frison, V.; De Kreutzenberg, S.; Agostini, C.; Tiengo, A.; Avogaro, A. The Oral Dipeptidyl Peptidase-4 Inhibitor Sitagliptin Increases Circulating Endothelial Progenitor Cells in Patients with Type 2 Diabetes: Possible Role of Stromal-Derived Factor-1alpha. *Diabetes Care* **2010**, *33*, 1607–1609. [CrossRef]
351. Fadini, G.P.; Avogaro, A. Dipeptidyl Peptidase-4 Inhibition and Vascular Repair by Mobilization of Endogenous Stem Cells in Diabetes and Beyond. *Atherosclerosis* **2013**, *229*, 23–29. [CrossRef]
352. Dai, Y.; Dai, D.; Wang, X.; Ding, Z.; Mehta, J.L. DPP-4 Inhibitors Repress NLRP3 Inflammasome and Interleukin-1beta via GLP-1 Receptor in Macrophages through Protein Kinase C Pathway. *Cardiovasc. Drugs Ther.* **2014**, *28*, 425–432. [CrossRef]
353. Cha, S.A.; Park, Y.M.; Yun, J.S.; Lim, T.S.; Song, K.H.; Yoo, K.D.; Ahn, Y.B.; Ko, S.H. A Comparison of Effects of DPP-4 Inhibitor and SGLT2 Inhibitor on Lipid Profile in Patients with Type 2 Diabetes. *Lipids Health Dis.* **2017**, *16*, 58. [CrossRef]
354. Boschmann, M.; Engeli, S.; Dobberstein, K.; Budziarek, P.; Strauss, A.; Boehnke, J.; Sweep, F.C.G.J.; Luft, F.C.; He, Y.L.; Foley, J.E.; et al. Dipeptidyl-Peptidase-IV Inhibition Augments Postprandial Lipid Mobilization and Oxidation in Type 2 Diabetic Patients. *J. Clin. Endocrinol. Metab.* **2009**, *94*, 846–852. [CrossRef]
355. Rosenstock, J.; Perkovic, V.; Johansen, O.E.; Cooper, M.E.; Kahn, S.E.; Marx, N.; Alexander, J.H.; Pencina, M.; Toto, R.D.; Wanner, C.; et al. Effect of Linagliptin vs. Placebo on Major Cardiovascular Events in Adults with Type 2 Diabetes and High Cardiovascular and Renal Risk: The CARMELINA Randomized Clinical Trial. *JAMA* **2019**, *321*, 69–79. [CrossRef]
356. Green, J.B.; Bethel, M.A.; Armstrong, P.W.; Buse, J.B.; Engel, S.S.; Garg, J.; Josse, R.; Kaufman, K.D.; Koglin, J.; Korn, S.; et al. Effect of Sitagliptin on Cardiovascular Outcomes in Type 2 Diabetes. *N. Engl. J. Med.* **2015**, *373*, 232–242. [CrossRef]
357. Scirica, B.M.; Bhatt, D.L.; Braunwald, E.; Steg, P.G.; Davidson, J.; Hirshberg, B.; Ohman, P.; Frederich, R.; Wiviott, S.D.; Hoffman, E.B.; et al. Saxagliptin and Cardiovascular Outcomes in Patients with Type 2 Diabetes Mellitus. *N. Engl. J. Med.* **2013**, *369*, 1317–1326. [CrossRef] [PubMed]
358. White, W.B.; Cannon, C.P.; Heller, S.R.; Nissen, S.E.; Bergenstal, R.M.; Bakris, G.L.; Perez, A.T.; Fleck, P.R.; Mehta, C.R.; Kupfer, S.; et al. Alogliptin after Acute Coronary Syndrome in Patients with Type 2 Diabetes. *N. Engl. J. Med.* **2013**, *369*, 1327–1335. [CrossRef] [PubMed]
359. Solomon, S.D.; de Boer, R.A.; DeMets, D.; Hernandez, A.F.; Inzucchi, S.E.; Kosiborod, M.N.; Lam, C.S.P.; Martinez, F.; Shah, S.J.; Lindholm, D.; et al. Dapagliflozin in Heart Failure with Preserved and Mildly Reduced Ejection Fraction: Rationale and Design of the DELIVER Trial. *Eur. J. Heart Fail.* **2021**, *23*, 1217–1225. [CrossRef] [PubMed]
360. Packer, M.; Butler, J.; Zannad, F.; Filippatos, G.; Ferreira, J.P.; Pocock, S.J.; Carson, P.; Anand, I.; Doehner, W.; Haass, M.; et al. Effect of Empagliflozin on Worsening Heart Failure Events in Patients with Heart Failure and Preserved Ejection Fraction: EMPEROR-Preserved Trial. *Circulation* **2021**, *144*, 1284–1294. [CrossRef]
361. Wadowski, P.P.; Weikert, C.; Pultar, J.; Lee, S.; Eichelberger, B.; Koppensteiner, R.; Lang, I.M.; Panzer, S.; Gremmel, T. Ticagrelor Inhibits Toll-Like and Protease-Activated Receptor Mediated Platelet Activation in Acute Coronary Syndromes. *Cardiovasc. Drugs Ther.* **2020**, *34*, 53–63. [CrossRef] [PubMed]
362. Panzer, B.; Wadowski, P.P.; Huber, K.; Panzer, S.; Gremmel, T. Protease-Activated Receptor-Mediated Platelet Aggregation in Patients with Type 2 Diabetes on Potent P2Y12 Inhibitors. *Diabet. Med.* **2022**, *39*, e14868. [CrossRef]
363. Wadowski, P.P.; Eichelberger, B.; Kopp, C.W.; Pultar, J.; Seidinger, D.; Koppensteiner, R.; Lang, I.M.; Panzer, S.; Gremmel, T. Disaggregation Following Agonist-Induced Platelet Activation in Patients on Dual Antiplatelet Therapy. *J. Cardiovasc. Transl. Res.* **2017**, *10*, 359–367. [CrossRef]
364. Pultar, J.; Wadowski, P.P.; Panzer, S.; Gremmel, T. Oral Antiplatelet Agents in Cardiovascular Disease. *Vasa* **2019**, *48*, 291–302. [CrossRef]
365. Li, W.; Liu, Q.; Tang, Y. Platelet to Lymphocyte Ratio in the Prediction of Adverse Outcomes after Acute Coronary Syndrome: A Meta-Analysis. *Sci. Rep.* **2017**, *7*, 40426. [CrossRef]
366. Azab, B.; Shah, N.; Akerman, M.; McGinn, J.T. Value of Platelet/Lymphocyte Ratio as a Predictor of All-Cause Mortality after Non-ST-Elevation Myocardial Infarction. *J. Thromb. Thrombolysis* **2012**, *34*, 326–334. [CrossRef]
367. Ugur, M.; Gul, M.; Bozbay, M.; Cicek, G.; Uyarel, H.; Koroglu, B.; Uluganyan, M.; Aslan, S.; Tusun, E.; Surgit, O.; et al. The Relationship between Platelet to Lymphocyte Ratio and the Clinical Outcomes in ST Elevation Myocardial Infarction Underwent Primary Coronary Intervention. *Blood Coagul. Fibrinolysis* **2014**, *25*, 806–811. [CrossRef] [PubMed]
368. Lee, S.; Hoberstorfer, T.; Wadowski, P.P.; Kopp, C.W.; Panzer, S.; Gremmel, T. Platelet-to-Lymphocyte and Neutrophil-to-Lymphocyte Ratios Predict Target Vessel Restenosis after Intrainguinal Angioplasty with Stent Implantation. *J. Clin. Med.* **2020**, *9*, 1729. [CrossRef] [PubMed]
369. Gremmel, T.; Steiner, S.; Seidinger, D.; Koppensteiner, R.; Panzer, S.; Kopp, C.W. Adenosine Diphosphate-Inducible Platelet Reactivity Shows a Pronounced Age Dependency in the Initial Phase of Antiplatelet Therapy with Clopidogrel. *J. Thromb. Haemost.* **2010**, *8*, 37–42. [CrossRef]

370. Gremmel, T.; Michelson, A.D.; Wadowski, P.P.; Pultar, J.; Weikert, C.; Tscharre, M.; Lee, S.; Panzer, S.; Frelinger, A.L. Sex-Specific Platelet Activation through Protease-Activated Receptor-1 in Patients Undergoing Cardiac Catheterization. *Atherosclerosis* **2021**, *339*, 12–19. [CrossRef] [PubMed]
371. Wadowski, P.P.; Lee, S.; Kopp, C.W.; Koppensteiner, R.; Panzer, S.; Gremmel, T. Low Levels of High-Density Lipoprotein Cholesterol Are Linked to Impaired Clopidogrel-Mediated Platelet Inhibition. *Angiology* **2018**, *69*, 786–794. [CrossRef]
372. Jäger, B.; Piackova, E.; Haller, P.M.; Andric, T.; Kahl, B.; Christ, G.; Geppert, A.; Wojta, J.; Huber, K. Increased Platelet Reactivity in Dyslipidemic Patients with Coronary Artery Disease on Dual Anti-Platelet Therapy. *Arch. Med. Sci.* **2019**, *15*, 65–71. [CrossRef]
373. Gremmel, T.; Kopp, C.W.; Seidinger, D.; Koppensteiner, R.; Panzer, S.; Sunder-Plassmann, R.; Mannhalter, C.; Steiner, S. Differential Impact of Cytochrome 2C9 Allelic Variants on Clopidogrel-Mediated Platelet Inhibition Determined by Five Different Platelet Function Tests. *Int. J. Cardiol.* **2013**, *166*, 126–131. [CrossRef]
374. Gremmel, T.; Kopp, C.W.; Moertl, D.; Seidinger, D.; Koppensteiner, R.; Panzer, S.; Mannhalter, C.; Steiner, S. Influence of Cytochrome 2C19 Allelic Variants on On-Treatment Platelet Reactivity Evaluated by Five Different Platelet Function Tests. *Thromb. Res.* **2012**, *129*, 616–622. [CrossRef]
375. Giustino, G.; Kirtane, A.J.; Baber, U.; Généreux, P.; Witzenbichler, B.; Neumann, F.J.; Weisz, G.; Maehara, A.; Rinaldi, M.J.; Metzger, C.; et al. Impact of Anemia on Platelet Reactivity and Ischemic and Bleeding Risk: From the Assessment of Dual Antiplatelet Therapy with Drug-Eluting Stents Study. *Am. J. Cardiol.* **2016**, *117*, 1877–1883. [CrossRef]
376. Wadowski, P.P.; Kopp, C.W.; Koppensteiner, R.; Lang, I.M.; Pultar, J.; Lee, S.; Weikert, C.; Panzer, S.; Gremmel, T. Decreased Platelet Inhibition by P2Y12 Receptor Blockers in Anaemia. *Eur. J. Clin. Investig.* **2018**, *48*, e12861. [CrossRef]
377. Weiss, G.; Ganz, T.; Goodnough, L.T. Anemia of Inflammation. *Blood* **2019**, *133*, 40–50. [CrossRef] [PubMed]
378. Otaki, Y.; Watanabe, T.; Takahashi, H.; Sugai, T.; Yokoyama, M.; Tamura, H.; Kato, S.; Nishiyama, S.; Arimoto, T.; Shishido, T.; et al. Impact of Iron Deficiency on Peripheral Artery Disease After Endovascular Therapy. *Circ. Rep.* **2019**, *1*, 187–195. [CrossRef] [PubMed]
379. McEvoy, J.W.; Ibrahim, K.; Kickler, T.S.; Clarke, W.A.; Hasan, R.K.; Czarny, M.J.; Keramati, A.R.; Goli, R.R.; Gratton, T.P.; Brinker, J.A.; et al. Effect of Intravenous Fentanyl on Ticagrelor Absorption and Platelet Inhibition Among Patients Undergoing Percutaneous Coronary Intervention: The PACIFY Randomized Clinical Trial (Platelet Aggregation with Ticagrelor Inhibition and Fentanyl). *Circulation* **2018**, *137*, 307–309. [CrossRef]
380. Kubica, J.; Adamski, P.; Ostrowska, M.; Sikora, J.; Kubica, J.M.; Sroka, W.D.; Stankowska, K.; Buszko, K.; Navarese, E.P.; Jilma, B.; et al. Morphine Delays and Attenuates Ticagrelor Exposure and Action in Patients with Myocardial Infarction: The Randomized, Double-Blind, Placebo-Controlled IMPRESSION Trial. *Eur. Heart J.* **2016**, *37*, 245–252. [CrossRef] [PubMed]
381. Hobl, E.L.; Reiter, B.; Schoergenhofer, C.; Schwameis, M.; Derhaschnig, U.; Lang, I.M.; Stimpfl, T.; Jilma, B. Morphine Interaction with Prasugrel: A Double-Blind, Cross-over Trial in Healthy Volunteers. *Clin. Res. Cardiol.* **2016**, *105*, 349–355. [CrossRef]
382. Hobl, E.L.; Stimpfl, T.; Ebner, J.; Schoergenhofer, C.; Derhaschnig, U.; Sunder-Plassmann, R.; Jilma-Stohlawetz, P.; Mannhalter, C.; Posch, M.; Jilma, B. Morphine Decreases Clopidogrel Concentrations and Effects: A Randomized, Double-Blind, Placebo-Controlled Trial. *J. Am. Coll. Cardiol.* **2014**, *63*, 630–635. [CrossRef]
383. Bartko, J.; Schoergenhofer, C.; Schwameis, M.; Wadowski, P.; Kubica, J.; Jilma, B.; Hobl, E.L. Morphine Interaction with Aspirin: A Double-Blind, Crossover Trial in Healthy Volunteers. *J. Pharmacol. Exp. Ther.* **2018**, *365*, 430–436. [CrossRef]
384. Masselli, E.; Pozzi, G.; Vaccarezza, M.; Mirandola, P.; Galli, D.; Vitale, M.; Carubbi, C.; Gobbi, G. ROS in Platelet Biology: Functional Aspects and Methodological Insights. *Int. J. Mol. Sci.* **2020**, *21*, 4866. [CrossRef]
385. Falconer, D.; Papageorgiou, N.; Salem, K.; Lim, W.Y.; Katsargyris, A.; Avgerinos, E.; Tousoulis, D. Nitric Oxide Donors for Peripheral Artery Disease. *Curr. Opin. Pharmacol.* **2018**, *39*, 77–85. [CrossRef]
386. Park, S.Y.; Pekas, E.J.; Headid, R.J.; Son, W.M.; Wooden, T.K.; Song, J.; Layec, G.; Yadav, S.K.; Mishra, P.K.; Pipinos, I.I. Acute Mitochondrial Antioxidant Intake Improves Endothelial Function, Antioxidant Enzyme Activity, and Exercise Tolerance in Patients with Peripheral Artery Disease. *Am. J. Physiol. Heart Circ. Physiol.* **2020**, *319*, H456–H467. [CrossRef]
387. Gorabi, A.M.; Kiaie, N.; Hajighasemi, S.; Banach, M.; Penson, P.E.; Jamialahmadi, T.; Sahebkar, A. Statin-Induced Nitric Oxide Signaling: Mechanisms and Therapeutic Implications. *J. Clin. Med.* **2019**, *8*, 2051. [CrossRef]
388. Durante, W. Targeting Heme Oxygenase-1 in Vascular Disease. *Curr. Drug Targets* **2010**, *11*, 1504–1516. [CrossRef] [PubMed]
389. Dulak, J.; Deshane, J.; Jozkowicz, A.; Agarwal, A. Heme Oxygenase-1 and Carbon Monoxide in Vascular Pathobiology: Focus on Angiogenesis. *Circulation* **2008**, *117*, 231–241. [CrossRef] [PubMed]
390. Lee, T.S.; Chang, C.C.; Zhu, Y.; Shyy, J.Y.J. Simvastatin Induces Heme Oxygenase-1: A Novel Mechanism of Vessel Protection. *Circulation* **2004**, *110*, 1296–1302. [CrossRef]
391. Piechota-Polanczyk, A.; Kopacz, A.; Kłoska, D.; Zagrapan, B.; Neumayer, C.; Grochot-Przeczek, A.; Huk, I.; Brostjan, C.; Dulak, J.; Jozkowicz, A. Simvastatin Treatment Upregulates HO-1 in Patients with Abdominal Aortic Aneurysm but Independently of Nrf2. *Oxid. Med. Cell Longev.* **2018**, *2018*, 2028936. [CrossRef] [PubMed]
392. Andreas, M.; Oeser, C.; Kainz, F.M.; Shabani, S.; Aref, T.; Bilban, M.; Messner, B.; Heidtmann, J.; Laufer, G.; Kocher, A.; et al. Intravenous Heme Arginate Induces HO-1 (Heme Oxygenase-1) in the Human Heart. *Arter. Thromb. Vasc. Biol.* **2018**, *38*, 2755–2762. [CrossRef] [PubMed]
393. Doberer, D.; Haschemi, A.; Andreas, M.; Zapf, T.C.; Clive, B.; Jeitler, M.; Heinzl, H.; Wagner, O.; Wolzt, M.; Bilban, M. Haem Arginate Infusion Stimulates Haem Oxygenase-1 Expression in Healthy Subjects. *Br. J. Pharmacol.* **2010**, *161*, 1751–1762. [CrossRef]

394. Andreas, M.; Schmid, A.I.; Doberer, D.; Schewzow, K.; Weisshaar, S.; Heinze, G.; Bilban, M.; Moser, E.; Wolzt, M. Heme Arginate Improves Reperfusion Patterns after Ischemia: A Randomized, Placebo-Controlled Trial in Healthy Male Subjects. *J. Cardiovasc. Magn. Reson.* **2012**, *14*, 55. [[CrossRef](#)]
395. Heusch, G. Myocardial Ischaemia-Reperfusion Injury and Cardioprotection in Perspective. *Nat. Rev. Cardiol.* **2020**, *17*, 773–789. [[CrossRef](#)]
396. Andreas, M.; Schmid, A.I.; Keilani, M.; Doberer, D.; Bartko, J.; Crevenna, R.; Moser, E.; Wolzt, M. Effect of Ischemic Preconditioning in Skeletal Muscle Measured by Functional Magnetic Resonance Imaging and Spectroscopy: A Randomized Crossover Trial. *J. Cardiovasc. Magn. Reson.* **2011**, *13*, 32. [[CrossRef](#)]
397. Hentia, C.; Rizzato, A.; Camporesi, E.; Yang, Z.; Muntean, D.M.; Săndesc, D.; Bosco, G. An Overview of Protective Strategies against Ischemia/Reperfusion Injury: The Role of Hyperbaric Oxygen Preconditioning. *Brain Behav.* **2018**, *8*, e00959. [[CrossRef](#)] [[PubMed](#)]
398. Rout, A.; Tantry, U.S.; Novakovic, M.; Sukhi, A.; Gurbel, P.A. Targeted Pharmacotherapy for Ischemia Reperfusion Injury in Acute Myocardial Infarction. *Expert. Opin. Pharmacother.* **2020**, *21*, 1851–1865. [[CrossRef](#)] [[PubMed](#)]
399. Belcher, D.A.; Williams, A.T.; Munoz, C.J.; Muller, C.R.; Walser, C.; Palmer, A.F.; Cabrales, P. Attenuating Ischemia-Reperfusion Injury with Polymerized Albumin. *J. Appl. Physiol.* **2022**, *132*, 489–496. [[CrossRef](#)] [[PubMed](#)]
400. Merchant, S.H.; Gurule, D.M.; Larson, R.S. Amelioration of Ischemia-Reperfusion Injury with Cyclic Peptide Blockade of ICAM-1. *Am. J. Physiol. Heart Circ. Physiol.* **2003**, *284*, H1260–H1268. [[CrossRef](#)]
401. O'Donnell, M.E.; Badger, S.A.; Sharif, M.A.; Makar, R.R.; McEneny, J.; Young, I.S.; Lee, B.; Soong, C.V. The Effects of Cilostazol on Exercise-Induced Ischaemia-Reperfusion Injury in Patients with Peripheral Arterial Disease. *Eur. J. Vasc. Endovasc. Surg.* **2009**, *37*, 326–335. [[CrossRef](#)] [[PubMed](#)]
402. Zhong, L.; Simard, M.J.; Huot, J. Endothelial MicroRNAs Regulating the NF-KB Pathway and Cell Adhesion Molecules during Inflammation. *FASEB J.* **2018**, *32*, 4070–4084. [[CrossRef](#)]
403. Perkins, L.A.; Anderson, C.J.; Novelli, E.M. Targeting P-Selectin Adhesion Molecule in Molecular Imaging: P-Selectin Expression as a Valuable Imaging Biomarker of Inflammation in Cardiovascular Disease. *J. Nucl. Med.* **2019**, *60*, 1691–1697. [[CrossRef](#)]
404. Wang, Y.; Wang, X.; Sun, M.; Zhang, Z.; Cao, H.; Chen, X. NF-KB Activity-Dependent P-Selectin Involved in Ox-LDL-Induced Foam Cell Formation in U937 Cell. *Biochem. Biophys. Res. Commun.* **2011**, *411*, 543–548. [[CrossRef](#)]
405. Chao, T.H.; Tseng, S.Y.; Chen, I.C.; Tsai, Y.S.; Huang, Y.Y.; Liu, P.Y.; Ou, H.Y.; Li, Y.H.; Wu, H.L.; Cho, C.L.; et al. Cilostazol Enhances Mobilization and Proliferation of Endothelial Progenitor Cells and Collateral Formation by Modifying Vasculo-Angiogenic Biomarkers in Peripheral Arterial Disease. *Int. J. Cardiol.* **2014**, *172*, e371–e374. [[CrossRef](#)]
406. Shishehbor, M.H.; Rundback, J.; Bunte, M.; Hammad, T.A.; Miller, L.; Patel, P.D.; Sadanandan, S.; Fitzgerald, M.; Pastore, J.; Kashyap, V.; et al. SDF-1 Plasmid Treatment for Patients with Peripheral Artery Disease (STOP-PAD): Randomized, Double-Blind, Placebo-Controlled Clinical Trial. *Vasc. Med.* **2019**, *24*, 200–207. [[CrossRef](#)]
407. Zhang, X.; Xie, Y.W.; Nasjletti, A.; Xu, X.; Wolin, M.S.; Hintze, T.H. ACE Inhibitors Promote Nitric Oxide Accumulation to Modulate Myocardial Oxygen Consumption. *Circulation* **1997**, *95*, 176–182. [[CrossRef](#)]
408. Steven, S.; Oelze, M.; Hanf, A.; Kröller-Schön, S.; Kashani, F.; Roohani, S.; Welschof, P.; Kopp, M.; Gödtel-Armbrust, U.; Xia, N.; et al. The SGLT2 Inhibitor Empagliflozin Improves the Primary Diabetic Complications in ZDF Rats. *Redox Biol.* **2017**, *13*, 370–385. [[CrossRef](#)] [[PubMed](#)]
409. Uthman, L.; Homayr, A.; Juni, R.P.; Spin, E.L.; Kerindongo, R.; Boomsma, M.; Hollmanna Benedikt Preckel, M.W.; Koolwijk, P.; Van Hinsbergh, V.W.M.; Zuurbier, C.J.; et al. Empagliflozin and Dapagliflozin Reduce ROS Generation and Restore NO Bioavailability in Tumor Necrosis Factor α -Stimulated Human Coronary Arterial Endothelial Cells. *Cell Physiol. Biochem.* **2019**, *53*, 865–886. [[CrossRef](#)] [[PubMed](#)]
410. Wang, C.; Li, L.; Liu, S.; Liao, G.; Li, L.; Chen, Y.; Cheng, J.; Lu, Y.; Liu, J. GLP-1 Receptor Agonist Ameliorates Obesity-Induced Chronic Kidney Injury via Restoring Renal Metabolism Homeostasis. *PLoS ONE* **2018**, *13*, e0193473. [[CrossRef](#)] [[PubMed](#)]
411. Herat, L.Y.; Ward, N.C.; Magno, A.L.; Rakoczy, E.P.; Kiuchi, M.G.; Schlaich, M.P.; Matthews, V.B. Sodium Glucose Co-Transporter 2 Inhibition Reduces Succinate Levels in Diabetic Mice. *World J. Gastroenterol.* **2020**, *26*, 3225–3235. [[CrossRef](#)]
412. Van Royen, N.; Schirmer, S.H.; Atasever, B.; Behrens, C.Y.H.; Ubbink, D.; Buschmann, E.E.; Voskuil, M.; Bot, P.; Hoefer, I.; Schlingemann, R.O.; et al. START Trial: A Pilot Study on STimulation of ARTeriogenesis Using Subcutaneous Application of Granulocyte-Macrophage Colony-Stimulating Factor as a New Treatment for Peripheral Vascular Disease. *Circulation* **2005**, *112*, 1040–1046. [[CrossRef](#)]
413. Cooper, L.T.; Hiatt, W.R.; Creager, M.A.; Regensteiner, J.G.; Casscells, W.; Isner, J.M.; Cooke, J.P.; Hirsch, A.T. Proteinuria in a Placebo-Controlled Study of Basic Fibroblast Growth Factor for Intermittent Claudication. *Vasc. Med.* **2001**, *6*, 235–239. [[CrossRef](#)]
414. Zachman, A.L.; Wang, X.; Tucker-Schwartz, J.M.; Fitzpatrick, S.T.; Lee, S.H.; Guelcher, S.A.; Skala, M.C.; Sung, H.J. Uncoupling Angiogenesis and Inflammation in Peripheral Artery Disease with Therapeutic Peptide-Loaded Microgels. *Biomaterials* **2014**, *35*, 9635. [[CrossRef](#)]
415. Caolo, V.; Vries, M.; Zupancich, J.; Houben, M.; Mihov, G.; Wagenaar, A.; Swennen, G.; Nossent, Y.; Quax, P.; Suylen, D.; et al. CXCL1 Microspheres: A Novel Tool to Stimulate Arteriogenesis. *Drug Deliv.* **2016**, *23*, 2919–2926. [[CrossRef](#)]
416. Chang, T.T.; Lin, L.Y.; Chen, J.W. Inhibition of Macrophage Inflammatory Protein-1 β Improves Endothelial Progenitor Cell Function and Ischemia-Induced Angiogenesis in Diabetes. *Angiogenesis* **2019**, *22*, 53–65. [[CrossRef](#)]

417. Cooke, J.P. Therapeutic Transdifferentiation: A Novel Approach for Vascular Disease. *Circ. Res.* **2013**, *112*, 748–750. [CrossRef] [PubMed]
418. Sayed, N.; Wong, W.T.; Ospino, F.; Meng, S.; Lee, J.; Jha, A.; Dexheimer, P.; Aronow, B.J.; Cooke, J.P. Transdifferentiation of Human Fibroblasts to Endothelial Cells Role of Innate Immunity. *Circulation* **2015**, *131*, 300–309. [CrossRef] [PubMed]
419. Tan, R.P.; Ryder, I.; Yang, N.; Lam, Y.T.; Santos, M.; Michael, P.L.; Robinson, D.A.; Ng, M.K.; Wise, S.G. Macrophage Polarization as a Novel Therapeutic Target for Endovascular Intervention in Peripheral Artery Disease. *JACC Basic. Transl. Sci.* **2021**, *6*, 693. [CrossRef] [PubMed]
420. Schlager, O.; Giurgea, A.; Schuhfried, O.; Seidinger, D.; Hammer, A.; Gröger, M.; Fialka-Moser, V.; Gschwandtner, M.; Koppensteiner, R.; Steiner, S. Exercise Training Increases Endothelial Progenitor Cells and Decreases Asymmetric Dimethylarginine in Peripheral Arterial Disease: A Randomized Controlled Trial. *Atherosclerosis* **2011**, *217*, 240–248. [CrossRef] [PubMed]
421. Wang, Y.; Xu, D. Effects of Aerobic Exercise on Lipids and Lipoproteins. *Lipids Health Dis.* **2017**, *16*, 132. [CrossRef]
422. Sampath Kumar, A.; Maiya, A.G.; Shastry, B.A.; Vaishali, K.; Ravishankar, N.; Hazari, A.; Gundmi, S.; Jadhav, R. Exercise and Insulin Resistance in Type 2 Diabetes Mellitus: A Systematic Review and Meta-Analysis. *Ann. Phys. Rehabil. Med.* **2019**, *62*, 98–103. [CrossRef]
423. Sharman, J.E.; La Gerche, A.; Coombes, J.S. Exercise and Cardiovascular Risk in Patients with Hypertension. *Am. J. Hypertens.* **2015**, *28*, 147–158. [CrossRef]
424. Palmefors, H.; DuttaRoy, S.; Rundqvist, B.; Börjesson, M. The Effect of Physical Activity or Exercise on Key Biomarkers in Atherosclerosis—A Systematic Review. *Atherosclerosis* **2014**, *235*, 150–161. [CrossRef]
425. Carvalho de Arruda Veiga, E.; Ferreira Levy, R.; Sales Bocalini, D.; Maria Soares Junior, J.; Chada Baracat, E.; Carvalho Cavalli, R.; dos Santos, L. Exercise Training and Experimental Myocardial Ischemia and Reperfusion: A Systematic Review and Meta-Analysis. *IJC Heart Vasc.* **2023**, *46*, 101214. [CrossRef]
426. Quindry, J.C.; Franklin, B.A. Exercise Preconditioning as a Cardioprotective Phenotype. *Am. J. Cardiol.* **2021**, *148*, 8–15. [CrossRef]
427. Wong, B.W.; Marsch, E.; Treps, L.; Baes, M.; Carmeliet, P. Endothelial Cell Metabolism in Health and Disease: Impact of Hypoxia. *EMBO J.* **2017**, *36*, 2187–2203. [CrossRef] [PubMed]
428. De Vries, M.R.; Quax, P.H.A. Plaque Angiogenesis and Its Relation to Inflammation and Atherosclerotic Plaque Destabilization. *Curr. Opin. Lipidol.* **2016**, *27*, 499–506. [CrossRef] [PubMed]
429. Xu, Y.; An, X.; Guo, X.; Habetsion, T.G.; Wang, Y.; Xu, X.; Kandala, S.; Li, Q.; Li, H.; Zhang, C.; et al. Endothelial PFKFB3 Plays a Critical Role in Angiogenesis. *Arter. Thromb. Vasc. Biol.* **2014**, *34*, 1231–1239. [CrossRef] [PubMed]
430. Lindstedt, K.A.; Mäyränpää, M.I.; Kovánen, P.T. Mast Cells in Vulnerable Atherosclerotic Plaques—A View to a Kill. *J. Cell Mol. Med.* **2007**, *11*, 739–758. [CrossRef]
431. Döring, Y.; Soehnlein, O.; Weber, C. Neutrophil Extracellular Traps in Atherosclerosis and Atherothrombosis. *Circ. Res.* **2017**, *120*, 736–743. [CrossRef]
432. Wadowski, P.P.; Steinlechner, B.; Zimpfer, D.; Schläglhofer, T.; Schima, H.; Hülsmann, M.; Lang, I.M.; Gremmel, T.; Koppensteiner, R.; Zehetmayer, S.; et al. Functional Capillary Impairment in Patients with Ventricular Assist Devices. *Sci. Rep.* **2019**, *9*, 5909. [CrossRef]
433. Wadowski, P.P.; Hülsmann, M.; Schörgerhofer, C.; Lang, I.M.; Wurm, R.; Gremmel, T.; Koppensteiner, R.; Steinlechner, B.; Schwameis, M.; Jilma, B. Sublingual Functional Capillary Rarefaction in Chronic Heart Failure. *Eur. J. Clin. Investig.* **2018**, *48*, e12869. [CrossRef]
434. Wadowski, P.P.; Schörgerhofer, C.; Rieder, T.; Ertl, S.; Pultar, J.; Serles, W.; Sycha, T.; Mayer, F.; Koppensteiner, R.; Gremmel, T.; et al. Microvascular Rarefaction in Patients with Cerebrovascular Events. *Microvasc. Res.* **2022**, *140*, 104300. [CrossRef]
435. Pultar, J.; Ertl, S.; Weikert, C.; Gremmel, T.; Kopp, C.; Mitteregger, M.; Cenan, E.; Jilma, B.; Koppensteiner, R.; Wadowski, P. Systemic Capillary Rarefaction in Patients with Peripheral Arterial Disease. *Wien. Klin. Wochenschr.* **2022**, *134*, 211–227. [CrossRef]
436. Criqui, M.H.; Aboyans, V. Epidemiology of Peripheral Artery Disease. *Circ. Res.* **2015**, *116*, 1509–1526. [CrossRef]
437. Welten, G.M.; Schouten, O.; Hoeks, S.E.; Chonchol, M.; Vidakovic, R.; van Domburg, R.T.; Bax, J.J.; van Sambeek, M.R.; Poldermans, D. Long-Term Prognosis of Patients With Peripheral Arterial Disease: A Comparison in Patients With Coronary Artery Disease. *J. Am. Coll. Cardiol.* **2008**, *51*, 1588–1596. [CrossRef] [PubMed]
438. Abdoli, S.; Katz, S.; Ochoa, C. Long-Term Patency and Clinical Outcomes of Nitinol Stenting for Femoropopliteal Atherosclerotic Disease. *Ann. Vasc. Surg.* **2020**, *66*, 566–572. [CrossRef]
439. Kaplovitch, E.; Eikelboom, J.W.; Dyal, L.; Aboyans, V.; Abola, M.T.; Verhamme, P.; Avezum, A.; Fox, K.A.A.; Berkowitz, S.D.; Bangdiwala, S.I.; et al. Rivaroxaban and Aspirin in Patients With Symptomatic Lower Extremity Peripheral Artery Disease: A Subanalysis of the COMPASS Randomized Clinical Trial. *JAMA Cardiol.* **2021**, *6*, 21–29. [CrossRef] [PubMed]
440. Zanini, G.; Selleri, V.; Roncati, L.; Coppi, F.; Nasi, M.; Farinetti, A.; Manenti, A.; Pinti, M.; Mattioli, A.V. Vascular “Long COVID”: A New Vessel Disease? *Angiology*, **2023**; *Online ahead of print*. [CrossRef]
441. Wadowski, P.P.; Piechota-Polańczyk, A.; Andreas, M.; Kopp, C.W. Cardiovascular Disease Management in the Context of Global Crisis. *Int. J. Environ. Res. Public Health* **2022**, *20*, 689. [CrossRef] [PubMed]
442. Panagiotides, N.G.; Zimprich, F.; Machold, K.; Schlager, O.; Müller, M.; Ertl, S.; Löffler-Stastka, H.; Koppensteiner, R.; Wadowski, P.P. A Case of Autoimmune Small Fiber Neuropathy as Possible Post COVID Sequelae. *Int. J. Environ. Res. Public Health* **2023**, *20*, 4918. [CrossRef]

443. Yue, C.; Zhang, C.; Ying, C.; Jiang, H. Reduced Serum Cholinesterase Is an Independent Risk Factor for All-Cause Mortality in the Pediatric Intensive Care Unit. *Front. Nutr.* **2022**, *9*, 809449. [[CrossRef](#)]
444. Shao, X.; Yang, L.; Hu, K.; Shen, R.; Ye, Q.; Yuan, X.; Zhao, Q.; Shen, J. Serum Cholinesterases, a Novel Marker of Clinical Activity in Inflammatory Bowel Disease: A Retrospective Case-Control Study. *Mediat. Inflamm.* **2020**, *2020*, 4694090. [[CrossRef](#)]
445. Gremmel, T.; Wadowski, P.P.; Mueller, M.; Kopp, C.W.; Koppensteiner, R.; Panzer, S. Serum Cholinesterase Levels Are Associated With 2-Year Ischemic Outcomes After Angioplasty and Stenting for Peripheral Artery Disease. *J. Endovasc. Ther.* **2016**, *23*, 738–743. [[CrossRef](#)]
446. Turk, B.R.; Gschwandtner, M.E.; Mauerhofer, M.; Löffler-Stastka, H. Can We Clinically Recognize a Vascular Depression? The Role of Personality in an Expanded Threshold Model. *Medicine* **2015**, *94*, e743. [[CrossRef](#)]
447. Balin, M.; Kivrak, T. Effect of Repeated Remote Ischemic Preconditioning on Peripheral Arterial Disease in Patients Suffering from Intermittent Claudication. *Cardiovasc. Ther.* **2019**, *2019*, 9592378. [[CrossRef](#)]
448. Poznyak, A.V.; Bharadwaj, D.; Prasad, G.; Grechko, A.V.; Sazonova, M.A.; Orekhov, A.N. Renin-Angiotensin System in Pathogenesis of Atherosclerosis and Treatment of CVD. *Int. J. Mol. Sci.* **2021**, *22*, 6702. [[CrossRef](#)]
449. Dardano, A.; Miccoli, R.; Bianchi, C.; Daniele, G.; Del Prato, S. Invited Review. Series: Implications of the Recent CVOTs in Type 2 Diabetes: Which Patients for GLP-1RA or SGLT-2 Inhibitor? *Diabetes Res. Clin. Pract.* **2020**, *162*, 108112. [[CrossRef](#)]
450. Jaipersad, A.S.; Lip, G.Y.H.; Silverman, S.; Shantsila, E. The Role of Monocytes in Angiogenesis and Atherosclerosis. *J. Am. Coll. Cardiol.* **2014**, *63*, A22. [[CrossRef](#)]
451. Hurley, D.M.; Williams, E.R.; Cross, J.M.; Riedinger, B.R.; Meyer, R.A.; Abela, G.S.; Slade, J.M. Aerobic Exercise Improves Microvascular Function in Older Adults. *Med. Sci. Sports Exerc.* **2019**, *51*, 773–781. [[CrossRef](#)]
452. Steiner, S.; Niessner, A.; Ziegler, S.; Richter, B.; Seidinger, D.; Pleiner, J.; Penka, M.; Wolzt, M.; Huber, K.; Wojta, J.; et al. Endurance Training Increases the Number of Endothelial Progenitor Cells in Patients with Cardiovascular Risk and Coronary Artery Disease. *Atherosclerosis* **2005**, *181*, 305–310. [[CrossRef](#)]

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