

Review

# Serotonergic Modulation of Neurovascular Transmission: A Focus on Prejunctional 5-HT Receptors/Mechanisms

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**Abstract:** 5-Hydroxytryptamine (5-HT), or serotonin, plays a crucial role as a neuromodulator and/or neurotransmitter of several nervous system functions. Its actions are complex, and depend on multiple factors, including the type of effector or receptor activated. Briefly, 5-HT can activate: (i) metabotropic (G-protein-coupled) receptors to promote inhibition (5-HT<sub>1</sub>, 5-HT<sub>5</sub>) or activation (5-HT<sub>4</sub>, 5-HT<sub>6</sub>, 5-HT<sub>7</sub>) of adenylyl cyclase, as well as activation (5-HT<sub>2</sub>) of phospholipase C; and (ii) ionotropic receptor (5-HT<sub>3</sub>), a ligand-gated Na<sup>+</sup>/K<sup>+</sup> channel. Regarding blood pressure regulation (and beyond the intricacy of central 5-HT effects), this monoamine also exerts direct postjunctional (on vascular smooth muscle and endothelium) or indirect prejunctional (on autonomic and sensory perivascular nerves) effects. At the prejunctional level, 5-HT can facilitate or preclude the release of autonomic (e.g., noradrenaline and acetylcholine) or sensory (e.g., calcitonin gene-related peptide) neurotransmitters facilitating hypertensive or hypotensive effects. Hence, we cannot formulate a specific impact of 5-HT on blood pressure level, since an increase or decrease in neurotransmitter release would be favoured, depending on the type of prejunctional receptor involved. This review summarizes and discusses the current knowledge on the prejunctional mechanisms involved in blood pressure regulation by 5-HT and its impact on some vascular-related diseases.

**Keywords:** serotonin; CGRP; blood pressure; migraine; hypertension



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## 1. Introduction

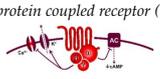
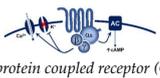
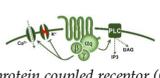
Among all biogenic monoamines, serotonin (5-hydroxytryptamine; 5-HT) stands out for its complex effects, the participation of a wide variety of receptors (which include the 5-HT<sub>1</sub>, 5-HT<sub>2</sub>, 5-HT<sub>3</sub>, 5-HT<sub>4</sub>, 5-HT<sub>5</sub>, 5-HT<sub>6</sub>, and 5-HT<sub>7</sub> receptors), and its extensive distribution in vertebrates and invertebrates [1]. In mammals, 5-HT is mainly synthesised in enterochromaffin cells (~90%) and in serotonergic neurons of the brain (1–2%) [2]. Indeed, this monoamine is predominantly found in platelets, enterochromaffin cells and in the central nervous system (CNS), but in many cases, its physiological role remains elusive [1,3]. Fortunately, with the progressive development of agonists and antagonists that act selectively on 5-HT receptors, many functions of 5-HT in the CNS and in the periphery have been discovered [1,3].

### 1.1. A Summary on 5-HT Receptors

This review will not document historical aspects of 5-HT research, discovery or 5-HT receptors. However, published research on the mechanisms involved in the effects of 5-HT (even long before its identification as 5-HT) has accumulated over 130 years [1,3–5].

As summarized in Table 1, with the conjunction of structural, transductional, and operational (pharmacological) criteria, 5-HT receptors have been classified into seven receptor types (5-HT<sub>1</sub>–5-HT<sub>7</sub>) that can be grouped into: (i) six metabotropic (G-protein-coupled) receptors, namely: the 5-HT<sub>1</sub> (further subdivided into the 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub>, 5-HT<sub>1D</sub>, 5-HT<sub>1e</sub> and 5-HT<sub>1F</sub> subtypes), 5-HT<sub>2</sub> (further subdivided into the 5-HT<sub>2A</sub>, 5-HT<sub>2B</sub> and 5-HT<sub>2C</sub> subtypes), 5-HT<sub>4</sub>, 5-HT<sub>5</sub> (further subdivided into the 5-HT<sub>5A</sub> and 5-HT<sub>5B</sub> subtypes), 5-HT<sub>6</sub> and 5-HT<sub>7</sub> receptor types; and (ii) one ligand-gated ion channel represented by the ionotropic 5-HT<sub>3</sub> receptor type [1,3–5]. The corresponding subtypes of the 5-HT<sub>1</sub>, 5-HT<sub>2</sub>, and 5-HT<sub>5</sub> receptor types share similar structural and transductional properties, but display very different pharmacological profiles.

**Table 1.** Classification of 5-HT receptors <sup>a</sup>.

5-HT Receptor	Receptor Subtype	Agonists	Antagonists	Some Functions	Canonical Transduction System	
5-HT <sub>1</sub>	5-HT <sub>1A</sub>	8-OH-DPAT	WAY 100635	Central hypotension		
	5-HT <sub>1B</sub>	Sumatriptan	SB224289	Vasoconstriction, sympatho-inhibition		
	5-HT <sub>1D</sub>	CP-93,129 (rodents) PNU-109291 PNU-142633	BRL15572	Autoreceptor, sympatho-inhibition		
	5-HT <sub>1e</sub> *	5-HT >> 5-CT, LY334370	Methiothepin (non-selective)	Unknown		
	5-HT <sub>F</sub>	LY344864, lasmiditan, LY334370	Methysergide (non-selective)	(–) Trigeminal system		
5-HT <sub>5</sub>	5-HT <sub>5A</sub> 5-HT <sub>5B</sub> *	5-HT, ergotamine 5-CT (non-selective)	SB699551 Unknown	Cardiac sympatho-inhibition in rats Unknown		
5-HT <sub>4</sub>	-	Renzapride, BIMU8, ML10302, SC53116	GR 113808, SB204070	(+) Neuronal activity, vasodilatation, tachycardia in pigs and humans		
5-HT <sub>6</sub>	-	5-MeO-T ≥ 5-HT SB357134, SB271046	Ro 630563	Memory, not involved in cardiovascular regulation		
5-HT <sub>7</sub>	-	5-CT >> 5-HT AS-19	SB269970, SB258719	Circadian rhythm, vasodila- tion, tachycardia in cats		
5-HT <sub>2</sub>	5-HT <sub>2A</sub>	DOI, DOB	MDL100907	Vasoconstriction, platelet aggregation		
	5-HT <sub>2B</sub>	α-methyl-5-HT DOI, BW723C86	Ketanserin SB204741	Vasoconstriction, release of NO		
	5-HT <sub>2C</sub>	α-methyl-5-HT	DOI, RS-127445	RS-127445		CSF production
		α-methyl-5-HT	DOI, Ro 60-0175	SB242084 RS-102221		
5-HT <sub>3</sub>	Pentameric ion channel **	Phenylbiguanide 2-methyl-5-HT	Tropisetron, Granisetron MDL-72222	(+) Neuronal activity, reflex bradycardia		

Modified from Villalón [3]. AS-19, (2S)-(+)-5-(1,3,5-Trimethylpyrazol-4-yl)-2-(dimethylamino)tetralin; CNS, central nervous system; CSF, cerebrospinal fluid; LSD, lysergic acid diethylamide; 5-MeOT, 5-methoxytryptamine; 5-CT, 5-carboxamidotryptamine; DOI, 1-(2,5-dimethoxy-4-iodophenyl)-2-aminopropane; NO, nitric oxide; (–), inhibits; (+), stimulates. \* Lowercase is used to denote a receptor with unknown functional roles in native cells or tissues. \*\* Five known subunits have been described (5-HT<sub>3A</sub>–5-HT<sub>3E</sub>) forming homomeric or heteromeric complexes. At least two subunits of 5-HT<sub>3A</sub> type are required to form a functional ion channel. <sup>a</sup> The pharmacological profile of each 5-HT receptor type is identified by applying inclusion and exclusion criteria, as explained in Section 1.1.

Some agonists and antagonists employed to identify the pharmacological profile of each 5-HT receptor type are shown in Table 1. As previously established [1,3–5], the pharmacological identification of a specific 5-HT receptor type is based on the application of (i) inclusion criteria (i.e., selective agonists for this receptor mimic the effects of 5-HT, while selective antagonists for this receptor produce a blockade of the effects of 5-HT and the corresponding agonist); and (ii) exclusion criteria (i.e., agonists and antagonists for the other 5-HT receptors—and sometimes even for receptors unrelated to 5-HT—are inactive) (see Table 1).

This knowledge (i) has helped to establish the role of 5-HT receptors in several diseases, including anxiety, depression, schizophrenia, drug addiction, cardiovascular pathologies (e.g., systemic, pulmonary and portal hypertension), cardiac disorders, migraine, etc.; and (ii) has led to the development of agonists and antagonists at 5-HT receptors for the therapeutic treatment of these—and other—diseases [1,3–8].

## 1.2. An Overview of the Effects of 5-HT on the Cardiovascular System

As previously described in other reviews dealing with 5-HT and the cardiovascular system [3,6–9], the cardiovascular effects of 5-HT are complex and include bradycardia/tachycardia, hypotension/hypertension, and vasodilatation/vasoconstriction. This

complexity of effects is due to (i) the capability of 5-HT to interact at various levels, including the heart and blood vessels, as well as the central and peripheral (autonomic and sensory) nervous systems; and (ii) the involvement of serotonin 5-HT<sub>1</sub>, 5-HT<sub>2</sub>, 5-HT<sub>3</sub>, 5-HT<sub>4</sub>, 5-HT<sub>5A</sub>, and 5-HT<sub>7</sub> receptors, as well as a tyramine-like action or unidentified mechanisms, depending on the species and the experimental conditions [3,6–9]. Interestingly, the 5-HT<sub>6</sub> receptor is not involved in the cardiovascular effects of 5-HT [3,8].

### 1.3. The Specific Interactions of 5-HT at Peripheral and Central Levels to Induce Cardiovascular Effects

#### 1.3.1. Sensory Afferents

Overall, an intravenous (i.v.) bolus injection of 5-HT in anaesthetised animals results in a reflex bradycardia and hypotension by stimulating 5-HT<sub>3</sub> receptors on vagal sensory afferents [3]. These neuronal 5-HT<sub>3</sub> receptors were identified using selective agonists and antagonists (see Table 1).

#### 1.3.2. Sympathetic Ganglia

It has been shown that i.v. 5-HT can stimulate and/or inhibit the sympathetic ganglia producing stimulation or inhibition of the sympathetic drive, and this results in changes in blood pressure and heart rate [3]. Moreover, the hyperpolarization of sympathetic ganglia produced by 5-HT is caused by the activation of 5-HT<sub>1A</sub> receptors in rats; these 5-HT<sub>1A</sub> receptors were identified by using selective agonists and antagonists (see Table 1).

#### 1.3.3. Cardiac Effects of 5-HT

Central or i.v. administration of 5-HT may produce bradycardia and/or tachycardia, and the 5-HT receptors involved in these effects have been identified by using some of the agonists and antagonists shown in Table 1 [3,6].

Overall, two central 5-HT receptors regulate cardiovascular function: 5-HT<sub>1A</sub> receptors (generally inhibiting the sympathetic drive) and 5-HT<sub>2</sub> receptors (largely stimulating the sympathetic drive) [3,10,11]; some of the agonists and antagonists used to identify these receptors (with the inclusion and exclusion criteria described in Section 1.1) are shown in Table 1. Admittedly, central administration of 5-HT elicits complex and contradictory cardiac effects which depend on, among other factors, the species, the exact site of central application, the drug used, and the dose employed [3,10,11]. In contrast, the bradycardia or tachycardia produced by i.v. administration of 5-HT is more controllable and consistent (see below) in view of the implied simplicity of the procedure.

#### Bradycardia

I.v. administration of 5-HT in intact animals results in a pronounced and transient bradycardia that is abolished after ganglion blockade, vagotomy, atropine, spinal section, or 5-HT<sub>3</sub> receptor antagonists [3,6]. This response involves the Bezold–Jarisch reflex, originating from the depolarization of afferent cardiac sensory neurons via activation of 5-HT<sub>3</sub> receptors [3,6]. Furthermore, 5-HT can also produce bradycardia by (i) a cardiac sympatho-inhibition via activation of prejunctional 5-HT<sub>1B</sub>, 5-HT<sub>1D</sub> and 5-HT<sub>5A</sub> receptors in pithed rats [3,12,13]; or (ii) a cardiac vagal stimulation via activation of 5-HT<sub>3</sub> receptors on parasympathetic ganglia and postganglionic vagal nerves in rabbits [3,6] (see Table 1 for pharmacological tools).

#### Tachycardia

I.v. administration of 5-HT in vagotomised animals induces a tachycardic effect that may be mediated by a wide variety of receptors/mechanisms, depending on the species and the experimental conditions [3,6]. These receptors/mechanisms include: (i) a tyramine-like action in spinal guinea pigs; (ii) direct stimulation of 5-HT<sub>2A</sub> receptors on the cardiac pacemaker in reserpinized pithed rats; (iii) activation of 5-HT<sub>3</sub> receptors on cardiac sympathetic neurons in the perfused heart of a rabbit, resulting in noradrenaline release and cardiac stimulation; (iv) activation of 5-HT<sub>3</sub> receptors on a calcitonin gene-related

peptide (CGRP)-containing sensory neurons in an isolated guinea pig atrium, resulting in CGRP release and cardiac stimulation; (v) direct stimulation of 5-HT<sub>3</sub> receptors on a cardiac pacemaker in conscious dogs; (vi) direct stimulation of 5-HT<sub>4</sub> receptors on a cardiac pacemaker in healthy anaesthetized pigs (which is also involved in the positive inotropic effects of 5-HT in isolated human atria and in rats with chronic heart failure); (vii) direct stimulation of 5-HT<sub>7</sub> receptors on a cardiac pacemaker in spinal cats; and (viii) unidentified mechanisms in the isolated hearts of certain lamellibranch and gastropod species (including *Mercenaria mercenaria*, *Patella vulgata*, *Tapes watlingi*, *Helix aspersa*, *Aplysia* spp., etc.). These receptors were pharmacologically identified using selective agonists and antagonists for each type, and the inclusion and exclusion criteria explained in Section 1.1. (see Table 1).

#### 1.3.4. Vascular and Blood Pressure Effects of 5-HT

As explained in other reviews [3,7,8], i.v. administration of 5-HT results in a triphasic effect on arterial blood pressure, consisting of an initial transient vasodepressor effect followed by a vasopressor effect, and then a late long-lasting vasodepressor effect.

##### Initial Transient Vasodepressor Effect

This response results from an abrupt bradycardia (and the consequent decrease in cardiac output) following stimulation of 5-HT<sub>3</sub> receptors on afferent cardiac vagal afferents (i.e., the Bezold–Jarisch reflex; see above and Table 1).

##### Vasopressor Effect

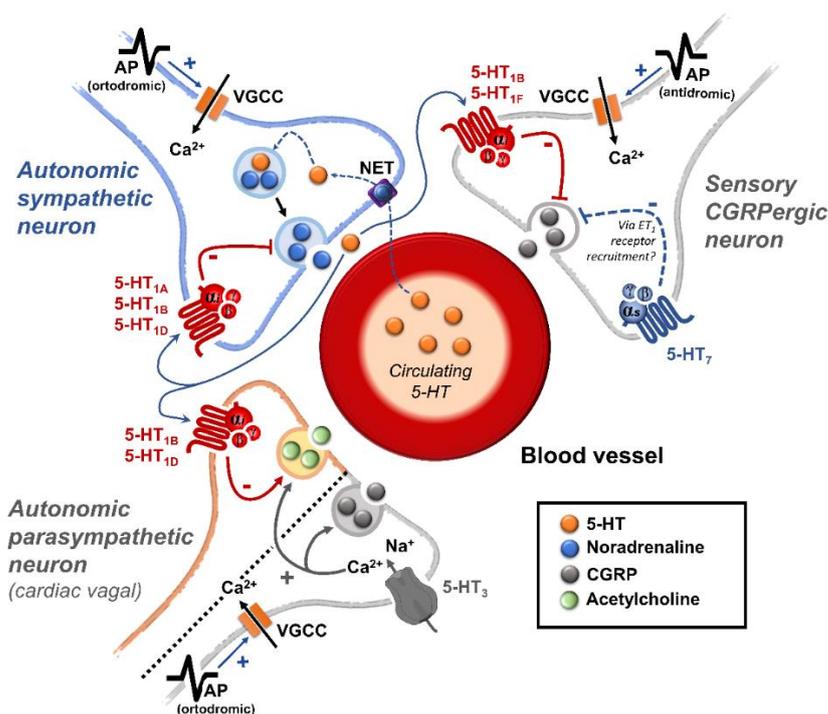
This effect (which varies quantitatively, depending on the species and the experimental conditions) involves the activation of vascular 5-HT<sub>2</sub> receptors in resistance blood vessels (resulting in peripheral vasoconstriction). It is worth noting that a release of catecholamines by activation of 5-HT<sub>2</sub> receptors in the adrenal medulla also plays a role in dogs, whereas activation of 5-HT<sub>1B</sub> receptors produces vasoconstriction in cranial and carotid arteries in humans, pigs and dogs [3]. Interestingly, 5-HT<sub>1B</sub> and 5-HT<sub>2</sub> receptors elicit vasoconstriction in the internal carotid bed of anaesthetised dogs, while 5-HT directly activates, in vitro,  $\alpha$ -adrenoceptors in rabbit ears and external carotid arteries [3]. Some of the agonists and antagonists used to identify these receptors (applying the inclusion and exclusion criteria defined in Section 1.1) are shown in Table 1.

##### Late Long-Lasting Vasodepressor Effect

This effect predominantly involves the activation of muscrotropic 5-HT<sub>7</sub> receptors [3,7,8], although several receptors/mechanisms may play a role, depending on the experimental conditions. These receptors/mechanisms may include:

(i) *Direct vasodilatation.* The direct vasodilatation to 5-HT involves 5-HT<sub>7</sub> receptors in a wide variety of blood vessels under different experimental conditions [3,6–8]. Some of the agonists and antagonists used to identify these receptors (applying the aforementioned inclusion and exclusion criteria) are shown in Table 1. Moreover, in the blood vessels where 5-HT<sub>7</sub> receptors produce vasodilatation and 5-HT<sub>2</sub>/5-HT<sub>1B</sub> receptors produce vasoconstriction, the final effect of 5-HT would depend on the pre-existing vascular tone, the dose employed, and the proportions in which these receptors are distributed [3].

(ii) *Prejunctional inhibition of perivascular sympathetic neurons.* The prejunctional inhibition induced by 5-HT and related agonists on perivascular sympathetic neurons has been confirmed in vitro and in vivo in many blood vessels [3]. This vascular sympatho-inhibition, generally mediated by 5-HT<sub>1</sub> receptors, may involve the 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub> and/or 5-HT<sub>1D</sub> receptor subtypes, depending on the vascular bed under study, the species, and the experimental conditions [3]. Interestingly, sympatho-inhibitory 5-HT<sub>7</sub> receptors could also be involved when rats are chronically pretreated with the 5-HT<sub>2</sub> receptor antagonist sarpogrelate [3,14]. These receptors were pharmacologically identified by applying the inclusion and exclusion criteria explained in Section 1.1 (see Table 1 and Figure 1).



**Figure 1.** Prejunctional 5-HT receptors are involved in the inhibition of postganglionic autonomic and sensory CGRPergic function at the vascular level. Generally, 5-HT can inhibit the release of noradrenaline, acetylcholine, and CGRP via activation of the 5-HT<sub>1</sub> receptor family (coupled to G<sub>i/o</sub> proteins; this figure shows the corresponding G<sub>α/β/γ</sub> subunits). In the case of the parasympathetic outflow, activation of 5-HT<sub>3</sub> (ligand-gated ion channel) receptors favours the release of acetylcholine. Furthermore, in sensory CGRPergic neurons, prejunctional activation of 5-HT<sub>7</sub> receptors seems to recruit the endothelin system (via an unknown pathway), favouring the activation of the ET<sub>1</sub> receptor and promoting inhibition of CGRP release. Interestingly, (i) in some isolated cases, activation of prejunctional 5-HT<sub>3</sub> receptors on parasympathetic fibres facilitates the release of CGRP; and (ii) circulating 5-HT can be recaptured via NET, and subsequently vesiculated and released upon electrical stimulation of the sympathetic outflow. See text for details. AP: action potential; NET: noradrenaline transporters; VGCC: voltage-gated ion channels.

(iii) *Endothelium-dependent vasodilatation.* In isolated blood vessels of several species without a functional endothelium, the vasodilatation to 5-HT is attenuated, while the vasoconstriction is augmented [3]. This vasodilatation, involving endothelial release of nitric oxide (NO), is mainly mediated by 5-HT<sub>1</sub> receptors [3]. Interestingly, in porcine blood vessels, the 5-HT-induced endothelium-dependent vasodilatation involves (i) 5-HT<sub>1B/1D</sub> receptors in coronary arteries; or (ii) 5-HT<sub>2B</sub> receptors in pulmonary arteries (see Table 1).

(iv) *Actions in the CNS.* Central administration of 5-HT may produce vasodepressor, vasopressor or biphasic effects, depending on the exact site of application, dose employed, depth of anaesthesia, the species used, etc. [3]. As previously reviewed [3,11], the cardiovascular regulation by central 5-HT neurons involves (i) 5-HT<sub>1A</sub> receptors (associated with sympatho-inhibition, hypotension, and bradycardia); and (ii) 5-HT<sub>2</sub> receptors (associated with sympatho-excitation and hypertension). Indeed, when directly applied in the CNS, 5-HT may produce both sympatho-inhibition and cardiac-vagal stimulation via 5-HT<sub>1A</sub> receptors [10,15]. In fact, psychiatric conditions that involve alterations in the serotonergic limbic components are usually accompanied by an autonomic imbalance; for example, posttraumatic stress disorder includes clinical manifestations such as cardiac arrhythmia, tachycardia, high blood pressure, etc. [16,17]. Moreover, anxiety correlates strongly with adrenaline levels in a positive direction [18], while aberrations in the autonomic nervous system (ANS) have been reported in patients with depression or other mood alterations [19].

Hence, central 5-HT is a powerful modulator of the ANS whose complex mechanisms fall beyond the scope of the present review. Interestingly, brain 5-HT can cross the blood–brain barrier via the 5-HT reuptake transporter (SERT) in endothelial cells and, consequently, can reach systemic circulation [20].

### 1.3.5. Receptor-Independent Actions of 5-HT

Apart from the above cardiovascular effects of 5-HT mediated by 5-HT receptors, other studies suggest that 5-HT can also play cardiovascular (patho)physiological roles independent of 5-HT receptor activation [3]. For example, (i) rats pretreated with fluoxetine (a SERT inhibitor) were protected from monocrotaline-induced pulmonary hypertension [21]; and (ii) 5-HT uptake can “serotonylate” proteins by transglutaminase-2 [22], a mechanism involved in the mitogenic and profibrotic effects of 5-HT without receptor activation [23].

## 2. Peripheral Autonomic Nervous System and Prejunctional 5-HT Receptors

### 2.1. An Overview of the Peripheral Actions of 5-HT Regulating the Vascular Function

Although 5-HT modulates the ANS at the central level [3,10,11], presynaptic and pre-junctional mechanisms by which 5-HT controls perivascular cholinergic and adrenergic outflows are relevant. Indeed, mutant mice lacking the SERT gene showed increased plasma noradrenaline levels after immobilization [24]; this suggests that, during stressful situations, peripheral 5-HT reuptake may be an essential mechanism in the systemic catecholaminergic modulation by 5-HT. Nevertheless, acute and systemic administration of SERT inhibitors may produce sympathetic inhibition (mainly by central mechanisms) [25].

Certainly, both SERT and 5-HT receptors are expressed in rodent adrenal glands, particularly in chromaffin cells [26], and 5-HT is involved in the development of the adrenal medulla [27]. Moreover, the number of adrenal chromaffin cells in mice embryos seems to be controlled by 5-HT<sub>3</sub> receptors expressed in their Schwann cell precursors [28]. Hence, 5-HT modulates adrenal chromaffin cells since their early development and, probably, during the rest of their lifetime.

Interestingly, when considering the distribution of SERTs in the adrenal chromaffin cells population, 5-HT seems to be strategically taken up by cells that exert an autocrine/paracrine modulation on the rest of chromaffin cells that release several vaso-contractile mediators to the systemic circulation; these include adrenaline (~79%), noradrenaline (~18%), and other mediators (~1–3%) during a sympathetic fight/flight situation induced by fear, stress, exercise, or conflict [26,29]. In this manner, the adrenal chromaffin release is controlled both neurogenically (by the ANS) and non-neurogenically (by several mediators, including 5-HT) [26,29].

On the other hand, it is noteworthy that adrenal chromaffin cells do not synthesize 5-HT by themselves [30,31], but they can take up 5-HT via their high expression of SERTs [26,31]. Moreover, activation of 5-HT<sub>1A</sub> receptors decreased adrenal chromaffin release [19,30]. Hence, 5-HT may act as a neuroendocrine tool to modulate (negatively) catecholamines release after stressful events via 5-HT<sub>1</sub> receptors. In addition, the autonomic control of perivascular sympathetic nerves is strategically organized to exert a local modulation of blood vessel sections, or even complete vascular beds [32]; hence, these sympathetic nerves form a complex varicose network that surrounds the blood vessels at the level of the adventitia layer in close proximity with the smooth muscle cells. However, neurotransmitters can diffuse and reach the endothelium [33]; this opens the possibility for a highly specific modulation by 5-HT of each blood vessel layer, namely, tunica intima, tunica media, and tunica externa (also called tunica adventitia).

The parasympathetic branch of the ANS innervates only cerebral vascular beds, whereas it does not innervate peripheral resistance blood vessels [34,35]. Particularly, intracerebral posterior blood vessels are richly innervated by parasympathetic fibres that seem to exert an essential control of blood flow in the polygon of Willis [36]. In peripheral blood vessels, vagal parasympathetic molecules (mainly acetylcholine) may be released systemically and reach the endothelium, exerting vasorelaxant neuroendocrine actions [33].

In short, both sympathetic and vagal parasympathetic varicosities express 5-HT receptors [37,38]. Thus, 5-HT may modulate sympathetic and parasympathetic perivascular nerves, and exert direct vascular actions [39,40], as described in Section 1.

## 2.2. The Role of Prejunctional 5-HT Receptors

There are several sources of 5-HT that may contribute to the modulation of perivascular autonomic and sensory nerve terminals; these include (i) the systemic circulation, where 5-HT is transported via blood platelets and released upon activation [41,42]; (ii) chromaffin cells of the adrenal medulla [26,30]; (iii) enterochromaffin gastrointestinal cells [43]; (iv) a subgroup of trigeminal C-fibres which store 5-HT [44]; and (v) cortical terminals from raphe neurons [20].

In the parasympathetic branch, the sphenopalatine ganglion (SPG) positively regulates cerebral blood flow; interestingly, more than 96% of the SPG body cells express 5-HT receptors [45]. Hence, 5-HT may be seen as a ubiquitous autonomic modulator.

5-HT<sub>1/2/3</sub> receptors are highly active during motor, sensory, and autonomic neuron development [46]. Thus, these receptors may maintain homeostatic functions in the motor, sensory, and autonomic neurons of the developed organism. According to their transduction systems, 5-HT<sub>1</sub> receptors are mainly involved in sympathetic inhibition, whereas 5-HT<sub>2</sub> and 5-HT<sub>3</sub> receptors may facilitate parasympathetic outflow (see below).

### 2.2.1. The 5-HT Receptors Inhibiting the Autonomic Outflow

Apart from its central sites of action, 5-HT can inhibit the tachycardia induced by sympathetic electrical stimulation, but not the one induced by exogenous noradrenaline [47]. This finding revealed the existence of a 5-HT-induced cardiac sympatho-inhibition at the prejunctional level (Figure 1).

5-HT<sub>1</sub> receptors are widely expressed in perivascular and cardiac sympathetic nerve endings; their activation is linked to cardiovascular sympathetic inhibition [48–51]. Specifically, selective stimulation of 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub>, and 5-HT<sub>1D</sub> receptors produced inhibition of the sympathetic vasopressor outflow in pithed rats [38,52]. This is a useful experimental model to pharmacologically study the prejunctional modulation of sympathetic nerve endings, since the CNS is not functional and, consequently, central compensatory cardiovascular reflexes are excluded [3]. Furthermore, in vitro experiments in the human atrium have shown that sympathetic nerves express 5-HT<sub>1D</sub> receptors, which mediate sympathetic inhibition [53]. Similarly, in pithed rats, the sympathetic cardioaccelerator outflow is inhibited by 5-HT<sub>1B/1D</sub> receptor activation [12].

### 2.2.2. The 5-HT Receptors as Facilitators of the Autonomic Outflow

In pithed rats, the bradycardia induced by vagal electrical stimulation may be increased by 5-HT during the blockade of 5-HT<sub>1/2</sub> receptors and by selective 5-HT<sub>3</sub> receptor agonists [54]. In contrast, activation of 5-HT<sub>2</sub> receptors inhibited this bradycardia induced by vagal electrical stimulation [54]. These findings suggest a dual role for 5-HT receptors in the cardiac parasympathetic outflow. Interestingly, in cerebral blood vessels, most SPG parasympathetic neurons (i) highly express 5-HT<sub>3A</sub> > 5-HT<sub>3B</sub> receptors; (ii) slightly express 5-HT<sub>2B</sub> > 5-HT<sub>2A</sub> > 5-HT<sub>1B</sub> receptors; and (iii) practically lack the expression of 5-HT<sub>1A</sub>, 5-HT<sub>1D</sub>, 5-HT<sub>1F</sub>, 5-HT<sub>2C</sub>, 5-HT<sub>4</sub>, 5-HT<sub>5A</sub>, 5-HT<sub>5B</sub>, 5-HT<sub>6</sub> and 5-HT<sub>7</sub> receptors [45]. In view that the 5-HT<sub>3</sub> receptor forms a ligand-gated Na<sup>+</sup>/K<sup>+</sup> channel, its parasympathetic expression in SPG neurons leads to a release of acetylcholine, NO, and vasoactive intestinal polypeptide (VIP), which, in turn, results in vasodilatation [55]. On the other hand, it remains unclear whether major cerebral blood vessels are rich in 5-HT receptors [56].

## 2.3. Clinical Relevance and Therapeutic Potential

The serotonergic negative modulation of sympathetic cardiovascular activity by 5-HT<sub>1A/1B/1D</sub> receptors may be achieved endogenously through platelet activation by catecholamines [42]. Indeed, a recent clinical study exposed 79 healthy male and female

volunteers to tryptophan enhancement, and 85 others to tryptophan depletion conditions, to analyse adrenaline and noradrenaline plasma levels [57]. Participants from the tryptophan enhancement condition showed a clear increment in plasma adrenaline, while noradrenaline decreased. Interestingly, this depletion condition slightly increased adrenaline and noradrenaline levels compared with baseline [57], suggesting that some preclinical findings are also observed clinically. Hence, 5-HT<sub>1</sub> receptors on perivascular fibres represent therapeutic targets to decrease sympathetic noradrenaline release.

On the other hand, a meta-analysis with cancer and cancer-depressed patients [58] concluded that management of stress-linked-emotions (which include serotonergic alterations) is a crucial element to (i) prevent comorbidities related to disruption of endocrine and autonomic (sympathetic) nervous system homeostasis; and (ii) improve survival time in these patients. Hence, stabilizing 5-HT levels may be a strategy to prevent autonomic disorders as comorbidities in diseases with associated high emotional stress.

It is important to keep in mind that some blood vessels (e.g., those from the coronary and carotid vascular beds) express the 5-HT<sub>1</sub> receptor in the smooth muscle layer, whose activation produces vasoconstriction [59]. These receptors are directly and/or indirectly involved in the pharmacotherapy of migraine, and some of the prophylactic (e.g., methysergide) and acute (e.g., triptans and ergots) antimigraine drugs interact with these receptors [60–62]. Hence, as a well-founded concern, direct vascular effects should be considered in any strategy that modifies 5-HT levels.

### 3. Sensory CGRPergic Perivascular Nerves and Prejunctional 5-HT Receptors

#### 3.1. The Sensory Perivascular CGRPergic Neurons as an Intrinsic Modulator of Vascular Tone

In general, CGRP is a potent vasodilator that can be released by capsaicin [63]; hence, CGRP release is associated with the activation of TRPV1 receptors on sensory nerves [64,65]. Nevertheless, the role of other TRP ion channels (e.g., TRPA1, TRPM3) located on nociceptors that induce the release of CGRP, has also been documented [66]. It is noteworthy that sensory nerves, which originate from the spinal cord [67], can exert (i) afferent actions [67]; and (ii) efferent actions via local (axonal) or central (dorsal root) reflexes [9]. In contrast to the efferent autonomic perivascular innervation from the spinal ventral horn, the sensory afferent fibres arrive at the spinal dorsal horn conveying information from the periphery to the spinal cord [9].

The relevance of the sensory nervous system (particularly CGRP release) as an intrinsic modulator of vascular tone was elegantly demonstrated by a series of *in situ* and *in vivo* experiments led by the group of Kawasaki in the early 90s. Indeed, they showed that, after pharmacological blockade of autonomic function, electrical stimulation of perivascular sensory nerves resulted in a vasodilator action mediated by CGRP release (blocked by CGRP<sub>(8–37)</sub>, a CGRP receptor antagonist), which was insensitive to blockade of  $\beta$ -adrenergic, muscarinic and histaminergic receptors [68–71]. More recently, our group has shown in pithed rats that, after CGRP receptor blockade with olcegepant, not only are the neurogenic and non-neurogenic vasodepressor responses to CGRP precluded, but a potentiation of the noradrenergic vasopressor responses is also unmasked [72]. Together, these data demonstrate that selective stimulation of perivascular sensory nerves results in CGRP release at the prejunctional level, activating CGRP receptors and evoking vasodilation. Current data strongly support the notion that CGRPergic sensory transmission modulates vascular tone via smooth muscle or endothelial mechanisms [73,74].

At the prejunctional level, several mechanisms have been reported to impact the sensory release of CGRP. One of the first lines of evidence suggesting that prejunctional heteroreceptors in sensory nerves modulate CGRP release was observed in experiments performed in the mesenteric vascular beds [69]. Briefly, Kawasaki et al. [69] showed that the vasodilation induced by periarterial nerve stimulation is smaller in vascular beds precontracted with noradrenaline (the endogenous ligand; non-selective  $\alpha_{1/2}$ - and  $\beta$ -adrenergic agonist) than in those precontracted with methoxamine (a selective  $\alpha_1$ -adrenoceptor agonist); this finding correlated with activation of  $\alpha_2$ -adrenoceptor activation [68]. These data

suggest that the sympathetic perivascular outflow induces a direct vasoconstrictor effect mediated by vascular activation of  $\alpha_{1/2}$ -adrenoceptors, and an indirect action by inhibiting the vasodilator function of sensory perivascular fibres. Furthermore, since  $\alpha_2$ -adrenoceptors are divided into three functional subtypes ( $\alpha_{2A/2B/2C}$ ), further pharmacological analysis in pithed rats showed that a fine-tuning of the perivascular sensory release of CGRP at the systemic level exists by selective activation of  $\alpha_{2A/2C}$ -adrenoceptors [75]. In this regard, several other prejunctional heteroreceptors facilitating (e.g., TRPV<sub>1</sub>) or inhibiting (e.g.,  $\mu$ -opioid, D<sub>2</sub>-like, CB<sub>1</sub>, H<sub>3</sub>, P2Y<sub>1/13</sub>, and 5-HT<sub>1</sub> receptors) CGRPergic neurovascular transmission have been described (for references see [9]).

It is worth noting that the potential relevance of serotonergic transmission modulating the perivascular sensory CGRPergic outflow has been established in the last 15 years [76,77]. In the case of 5-HT receptors modulating perivascular CGRPergic transmission, special attention has been paid in the context of migraine pathophysiology and pharmacotherapy. Indeed, triptans such as sumatriptan, which is a 5-HT<sub>1B/1D/1F</sub> receptor agonist considered the gold standard in acute migraine treatment, aborts migraine attacks by producing (i) direct vasoconstriction of intracranial and extracranial arteries; and (ii) inhibition of CGRP release at the trigeminal level and on perivascular sensory nerves [60,78].

### 3.2. Prejunctional 5-HT Receptors as Inhibitors of the Perivascular Sensory CGRPergic Outflow

As mentioned above, triptans and ergots (agonists at 5-HT<sub>1</sub> receptors) can prejunctionally inhibit the trigeminal release of CGRP [79,80]. Indeed, the first evidence about the role of 5-HT<sub>1</sub> receptors as inhibitors of CGRPergic transmission derived from pharmacological research on the mechanisms involved in the therapeutic effects of acute antimigraine drugs [81–84]. Admittedly, the discussion on the relevance of serotonergic mechanisms modulating CGRPergic outflow in the context of antimigraine therapy (i.e., at trigeminovascular level) falls beyond the scope of the present review, and several excellent reviews on this topic have been published elsewhere (see refs. [60,79,85–87]).

Considering that triptans and ergots are associated with cardiovascular side effects [60], a study in pithed rats demonstrated that acute (rather than prophylactic) antimigraine drugs are capable of inhibiting the perivascular sensory CGRPergic outflow at the systemic level, via prejunctional mechanisms [88]. Specifically, the pithed rat model was used to analyse vascular and prejunctional mechanisms excluding the influence of any central compensatory reflex mechanisms. Under these experimental conditions, in animals infused with hexamethonium (a sympathetic ganglionic blocker) and methoxamine (an  $\alpha_1$ -adrenoceptor agonist to induce sustained systemic vasoconstriction), the treatment with sumatriptan, ergotamine, or dihydroergotamine inhibited the vasodepressor responses elicited by electrical stimulation of the T<sub>9</sub>–T<sub>12</sub> spinal cord segments (an effect associated with inhibition of CGRP release from perivascular sensory nerves [88]).

The above data strongly support the hypothesis that 5-HT receptors on perivascular sensory nerve endings modulate CGRP release in the vascular system (such as at the trigeminovascular level) (Figure 1). Certainly, molecular biology evidence at the dorsal root ganglion level has suggested that mRNA expression correlates with 5-HT<sub>1B</sub> and 5-HT<sub>1F</sub>, but not with 5-HT<sub>1A</sub> or 5-HT<sub>1D</sub>, receptors [89]. In this regard, further functional pharmacological experiments using the pithed rat model showed that the selective 5-HT<sub>1B</sub> receptor agonist, CP-93,129, selectively inhibits the neurogenic CGRPergic vasodepressor responses via prejunctional sensory mechanisms [77]. Likewise, some data suggest that trigeminal activation of prejunctional 5-HT<sub>1B</sub> receptors (by sumatriptan or donitriptan) inhibits the external carotid vasodilation induced by capsaicin [90,91], highlighting the relevance of this receptor subtype in the modulation of CGRP release. Furthermore, as discussed by Rubio-Beltrán et al. [92], since 5-HT<sub>1F</sub> receptors have been found on sensory nerves, the role of these receptors in the modulation of CGRP release is suggested. Indeed, lasmiditan (a selective 5-HT<sub>1F</sub> receptor agonist) can prejunctionally inhibit CGRP release not only at the central (trigeminal) level, but also at the peripheral (meninges) level [80].

In view that sumatriptan is a non-selective 5-HT<sub>1A/1B/1D/1F</sub> receptor agonist, the role of these receptor subtypes was also analysed in the inhibition of the vasodepressor sensory CGRPergic outflow in pithed rats [76]. The data using selective agonists and antagonists for each 5-HT<sub>1</sub> receptor subtype (see Table 1) (i) corroborated the relevance of 5-HT<sub>1B</sub> receptors, and further showed that activation of prejunctional 5-HT<sub>1F</sub> receptors inhibited CGRP release; and (ii) excluded the role of 5-HT<sub>1A</sub> and 5-HT<sub>1D</sub> receptors [76]. It is noteworthy that prejunctional 5-HT<sub>1D</sub> receptors have also been suggested to inhibit the trigeminal release of CGRP [93]; however, PNU-142633 (a selective 5-HT<sub>1D</sub> receptor agonist; see Table 1) was not effective in the acute treatment of migraine [94], a neurovascular disorder where trigeminal release of CGRP plays a key role [85–87].

The ergots ergotamine and dihydroergotamine can also inhibit the perivascular sensory CGRPergic outflow [60]. Nevertheless, their pharmacology is much more complex, since these compounds display an affinity for all 5-HT (except 5-HT<sub>3</sub>) receptors [3–5] and also interact with dopaminergic and noradrenergic receptors [3]. Indeed, a detailed pharmacological analysis showed that, apart from prejunctional 5-HT<sub>1B/1F</sub> receptors, these ergots activate prejunctional D<sub>2</sub>-like and  $\alpha_2$ -adrenergic receptors to inhibit the vasodepressor sensory CGRPergic outflow [60,95].

Regarding transductional mechanisms, the 5-HT<sub>1</sub> receptor family is coupled to G<sub>i/o</sub> proteins [1] which, in turn, (i) via the G $\alpha$  subunit reduce the activity of adenylate cyclase, diminishing intracellular cAMP levels and consequently inhibiting the activity of protein kinase A; and (ii) via the G $\beta/\gamma$  subunits increase the activity of K<sup>+</sup> channels. Both mechanisms are intrinsically associated with the inhibition of neurotransmitter release [96]. From this perspective, the fact that AS-19 (a 5-HT<sub>7</sub> receptor agonist; Table 1) activated prejunctional 5-HT<sub>7</sub> receptors to inhibit the rat vasodepressor sensory CGRPergic outflow was counterintuitive [97] since this receptor is positively coupled to G<sub>s</sub> proteins [1]. Hence, one would have expected facilitation rather than inhibition of this sensory CGRPergic outflow. Nonetheless, the possibility exists that this 5-HT<sub>7</sub> receptor-induced sensory inhibition may involve (i) an ATP-dependent K<sup>+</sup> channel-mediated hyperpolarization sensitive to glibenclamide [97], as reported for the 5-HT-induced inhibition of the contractile and electrical activities in the guinea pig mesenteric bed [98]; and (ii) the endothelin pathway, as this response was blocked by sulfisoxazole [97], an endothelin ET<sub>A</sub> receptor antagonist [99]. Indeed, endothelin-1 can inhibit the neuroeffector transmission in smooth muscle [100], as well as the capsaicin-induced CGRP release [101]. Hence, the prejunctional 5-HT<sub>7</sub> receptor seems to promote endothelin-1 secretion, which inhibits CGRP release.

Moreover, activation of the 5-HT<sub>7</sub> receptor at the spinal cord level results in an antinociceptive action whereas, at the peripheral level, it enhances capsaicin-induced sensitization [102]. Clearly, the location of this receptor is the determining factor in its effects.

Interestingly, molecular expression analysis of 5-HT receptors in dorsal root ganglion neurons showed that, apart from 5-HT<sub>1B</sub>, 5-HT<sub>1F</sub>, and 5-HT<sub>7</sub> receptors, 5-HT<sub>2A</sub>, 5-HT<sub>2C</sub>, 5-HT<sub>3</sub>, 5-HT<sub>5A</sub>, 5-HT<sub>5B</sub>, and 5-HT<sub>6</sub> receptors can also be found in this type of cells [89,103,104]. Although the functional role of the latter receptor (sub)types in sensory vascular neurotransmission has not yet been reported, experiments exploring nociception showed that activation of 5-HT<sub>2A</sub> or 5-HT<sub>4</sub> receptors enhances CGRPergic transmission [105,106], while activation of 5-HT<sub>5A</sub> receptors have the opposite effect [107]. Furthermore, in guinea pig isolated cardiac atria, 5-HT favours CGRP release via sensory 5-HT<sub>3</sub> receptor activation, leading to a positive inotropic response [108]. However, it must be highlighted that these findings do not necessarily imply that similar results can be obtained at the vascular sensory neuroeffector level, as illustrated above with the case of the 5-HT<sub>7</sub> receptor.

### 3.3. Clinical Relevance

Apart from the well-established therapeutic relevance of acute antimigraine serotonergic drugs, which inhibit trigeminal CGRPergic transmission by 5-HT<sub>1B/1D/1F</sub> receptor activation [92], little attention has been paid to the interaction between 5-HT and the perivascular sensory nerves modulating systemic vascular responses (i.e., changes in ar-

terial blood pressure). Admittedly, this is partly because there is no consensus on the pivotal role of CGRP in maintaining blood pressure [73,109]. In addition, since the pharmacology of serotonergic transmission is complex at the peripheral and central levels, the (cardio)vascular effects resulting from activation of the different 5-HT receptors at both levels are hard to explain [3,7,8].

Regarding perivascular CGRPergic transmission on the systemic vasculature, some findings seem to exclude the relevance of this neuropeptide in regulating blood pressure, since acute CGRP receptor blockade in anaesthetized rats does not significantly impact blood pressure levels [74,109]. Accordingly, resting blood pressure is not affected in transgenic mice lacking the CGRP receptor [110]; conversely, continuous recording of blood pressure (in CGRP receptor KO mice) showed that this parameter is globally increased by an enhancement of the sympathetic autonomic function [111]. Indeed, in pithed rats (in which the CNS is not functional), acute pharmacological blockade of the CGRP receptor with olcegepant not only inhibits the vasodepressor sensory CGRPergic outflow, but also enhanced the sympathetic vasopressor responses [72]. These data may imply that continuous blockade of CGRPergic vascular transmission with olcegepant (or any other CGRP antagonist) could favour a hypertensive state [72]. Although this effect was apparently absent in clinical trials [112], real-world studies now suggest that the use of CGRP (receptor) blocking medications may increase blood pressure [113]. As elsewhere discussed [73,114,115], CGRP may play a physiological protective role in the cardiovascular system; nevertheless, the relevance of CGRPergic transmission in blood pressure regulation is only unmasked in cardiovascular pathologies [116].

Certainly, a decrease in CGRP levels has been observed in spontaneously hypertensive rats and in humans with essential hypertension [117,118]; in addition, it has been suggested that a diminution of the perivascular CGRPergic innervation may play a role in the development of this pathology [119]. Furthermore, within the bounds of 5-HT receptors, it is to be highlighted that, globally, 5-HT produces vasopressor responses by activation of vascular 5-HT<sub>2A</sub> receptors [3,8]; however, under vascular damage conditions (e.g., in hypertension), the vasculature is more sensitive to 5-HT to cause contraction [8]. Thus, apart from an enhanced 5-HT-induced vasoconstriction in hypertensive subjects [8], we suggest that the release of CGRP in these subjects may be diminished by activation of prejunctional 5-HT<sub>1B/1F</sub> and 5-HT<sub>7</sub> receptors, favouring a pro-hypertensive state.

#### 4. Perspectives and Some Future Directions

Physiologically, blood pressure is regulated by changes in peripheral vascular tone (caused by resistance blood vessels) and cardiac output, and these parameters are homeostatically maintained by neuronal, humoral, and local mechanisms [120]. When considering the neurovascular junction, it is well known that vascular tone is modulated by (i) autonomic sympathetic nerves, which produce vasoconstriction by noradrenaline release [120,121]; and (ii) primary sensory nerves, which produce vasodilatation by neuropeptides release, mainly CGRP [69,70,122–125]. Several mechanisms exist at the neurovascular junction to modulate the neuronal outflow to the blood vessels; one of these mechanisms is the serotonergic transmission. Certainly, 5-HT can generally modulate the autonomic and sensory outflows via prejunctional receptors (see Figure 1) to regulate blood pressure.

From a global perspective, 5-HT is an amphibaric agent, and its actions on haemodynamic parameters are complex, depending on the experimental conditions [3,7]. This may be explained in terms of the numerous sites of action to 5-HT, which include: (i) the CNS; (ii) autonomic ganglia; (iii) perivascular autonomic and sensory nerve terminals; (iv) endothelial cells; (v) smooth muscle cells; and (vi) the heart [3,37,38,46,126,127]. Consequently, rather than assigning a single cardiovascular function to 5-HT, it is clear that 5-HT exerts multiple cardiovascular actions, which may be further complicated by pathophysiological states including, but not limited to, hypertension [3,7], depression [128], and pain [128]. Thus, when studying the effects of 5-HT, it is imperative to be open-minded about its possible pleiotropic actions.

On this basis, future research should focus, among other approaches, on the significance of interspecies differences, the (patho)physiological conditions that may affect the function of 5-HT and its receptors/transporters, as well as interindividual differences caused by gender, ethnic background, body mass index, age, etc. For example, 17 $\beta$ -oestradiol may modulate the SERT and 5-HT metabolism in the brain [129], and it is well known that the correct function of the serotonergic system critically depends on the function of the SERT [130]; hence, this finding may be relevant in perimenopausal women.

Furthermore, together with the seminal discovery that 5-HT is a vasoconstrictor [131], the fact that plasma 5-HT levels are increased in hypertension favoured the hypothesis about the hypertensive action of 5-HT [132]. However, a chronic 5-HT infusion induced a fall in blood pressure [133], probably via an increase in endothelial NOs system activity [134]. This effect is clearly dependent on SERT function, as SERT knock-out mice are less prone to the chronic 5-HT-induced drop in blood pressure [134]. Interestingly, some clinical studies have shown that chronic inhibition of 5-HT reuptake by SERT inhibitors reduced the risk of a myocardial infarction [135], but an increase in blood pressure was recorded at night [136]. In this regard, in dogs, chronic oral treatment with fluoxetine does not affect mean arterial blood pressure (measured at diurnal time), but the capsaicin-induced trigeminal CGRP release (resulting in vasodilatation) is diminished at the craniovascular (external carotid) level [137]; this finding may help explain the mechanism of action of some SERT inhibitors to treat migraine.

Moreover, other pathophysiological states (apart from hypertension, depression and pain) may affect the actions of the serotonergic system, as exemplified by a recent study describing that the SERT is negatively associated with body mass index after glucose loading [138]; this finding highlights the importance of a holistic approach, taking factors such as body weight and/or obesity into account when investigating the serotonergic system.

In addition to the above differential role of SERT in various conditions, the 5-HT receptor function may also have specific relevance in several circumstances. For example, the 5-HT<sub>1F</sub> receptor has been described to promote, in rodents, mitochondrial biogenesis and recovery from acute kidney injury [139], as well as spinal cord injury [140], which might potentially also differ between sexes [140]. With these preclinical findings, it would be tempting to suggest in a clinical setting (including medical emergencies) that patients with kidney and spinal cord injuries could be treated with the selective 5-HT<sub>1F</sub> receptor agonist lasmiditan; this is a Federal Drug Administration-approved antimigraine drug that prejunctionally inhibits the trigeminal release of CGRP [80,92].

## 5. Conclusions

As evident from the present review, in addition to the pathophysiological relevance of the SERT in the receptor-independent intracellular actions of 5-HT [23], this monoamine induces a plethora of complex, and sometimes opposing, actions in the cardiovascular system. These cardiovascular actions of 5-HT may be even further complicated by pathophysiological states [3,7,9,18,19,128], and also by 5-HT receptor intracellular signalling (i.e., the impact of biased agonists) [141]. Admittedly, it is challenging to unravel the effects of these conditions on the functioning of 5-HT and its receptors as well as transporters. Despite its complex action, research on human differentiated tissues obtained from induced pluripotent stem cells (iPSCs) may be a valuable tool for the study of rare diseases and the influence of different (culture) conditions. Because of the many modulating roles of 5-HT in other systems, targeting specific 5-HT receptors may provide valuable novel therapeutic avenues, besides its currently known therapeutic applications [1,3–8]. Accordingly, further understanding of the actions of 5-HT under different conditions will provide new insights and therapeutic treatment possibilities, with the serotonergic system as a pharmacological target.

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## References

1. Barnes, N.M.; Ahern, G.P.; Becamel, C.; Bockaert, J.; Camilleri, M.; Chaumont-Dubel, S.; Claeysen, S.; Cunningham, K.A.; Fone, K.C.; Gershon, M.; et al. International Union of Basic and Clinical Pharmacology. CX. Classification of Receptors for 5-hydroxytryptamine; Pharmacology and Function. *Pharmacol. Rev.* **2021**, *73*, 310–520. [[CrossRef](#)] [[PubMed](#)]
2. Lv, J.; Liu, F. The Role of Serotonin beyond the Central Nervous System during Embryogenesis. *Front. Cell Neurosci.* **2017**, *11*, 74. [[CrossRef](#)] [[PubMed](#)]
3. Villalón, C.M. The role of serotonin receptors in the control of cardiovascular function. In *The Serotonin System*; Tricklebank, M.D., Daly, E., Eds.; Academic Press: Cambridge, MA, USA, 2019; Chapter 3; pp. 45–61.
4. Hoyer, D.; Clarke, D.E.; Fozard, J.R.; Hartig, P.R.; Martin, G.R.; Mylecharane, E.J.; Saxena, P.R.; Humphrey, P.P. International Union of Pharmacology classification of receptors for 5-hydroxytryptamine (Serotonin). *Pharmacol. Rev.* **1994**, *46*, 157–203. [[PubMed](#)]
5. Hoyer, D.; Hannon, J.P.; Martin, G.R. Molecular, pharmacological and functional diversity of 5-HT receptors. *Pharmacol. Biochem. Behav.* **2002**, *71*, 533–554. [[CrossRef](#)]
6. Kaumann, A.J.; Levy, F.O. 5-hydroxytryptamine receptors in the human cardiovascular system. *Pharmacol. Ther.* **2006**, *111*, 674–706. [[CrossRef](#)]
7. Watts, S.W.; Davis, R.P. 5-hydroxytryptamine receptors in systemic hypertension: An arterial focus. *Cardiovasc. Ther.* **2011**, *29*, 54–67. [[CrossRef](#)]
8. Watts, S.W.; Morrison, S.F.; Davis, R.P.; Barman, S.M. Serotonin and blood pressure regulation. *Pharmacol. Rev.* **2012**, *64*, 359–388. [[CrossRef](#)]
9. González-Hernández, A.; Marichal-Cancino, B.A.; Lozano-Cuenca, J.; López-Canales, J.S.; Muñoz-Islas, E.; Ramírez-Rosas, M.B.; Villalón, C.M. Heteroreceptors Modulating CGRP Release at Neurovascular Junction: Potential Therapeutic Implications on Some Vascular-Related Diseases. *Biomed. Res. Int.* **2016**, *2016*, 2056786. [[CrossRef](#)]
10. Ramage, A.G. Influence of 5-HT<sub>1A</sub> receptor agonists on sympathetic and parasympathetic nerve activity. *J. Cardiovasc. Pharmacol.* **1990**, *15* (Suppl. S7), S75–S85. [[CrossRef](#)]
11. Ramage, A.G. Central cardiovascular regulation and 5-hydroxytryptamine receptors. *Brain Res. Bull.* **2001**, *56*, 425–439. [[CrossRef](#)]
12. Sánchez-Lopez, A.; Centurión, D.; Vázquez, E.; Arulmani, U.; Saxena, P.R.; Villalón, C.M. Pharmacological profile of the 5-HT-induced inhibition of cardioaccelerator sympathetic outflow in pithed rats: Correlation with 5-HT<sub>1</sub> and putative 5-HT<sub>5A/5B</sub> receptors. *Br. J. Pharmacol.* **2003**, *140*, 725–735. [[CrossRef](#)]
13. García-Pedraza, J.; Hernández-Abreu, O.; García, M.; Morán, A.; Villalón, C.M. Chronic 5-HT<sub>2</sub> receptor blockade unmasks the role of 5-HT<sub>1F</sub> receptors in the inhibition of rat cardioaccelerator sympathetic outflow. *Can. J. Physiol. Pharmacol.* **2018**, *96*, 328–336. [[CrossRef](#)]
14. García-Pedraza, J.; García, M.; Martín, M.L.; Gómez-Escudero, J.; Rodríguez-Barbero, A.; Román, L.S.; Morán, A. Peripheral 5-HT<sub>1D</sub> and 5-HT<sub>7</sub> serotonergic receptors modulate sympathetic neurotransmission in chronic sarpgogrelate treated rats. *Eur. J. Pharmacol.* **2013**, *714*, 65–73. [[CrossRef](#)]
15. Dabiré, H. Central 5-hydroxytryptamine (5-HT) receptors in blood pressure regulation. *Thérapie* **1991**, *46*, 421–429.
16. Bedi, U.S.; Arora, R. Cardiovascular manifestations of posttraumatic stress disorder. *J. Natl. Med. Assoc.* **2007**, *99*, 642–649.
17. Tania, V.; Catherine, V. Roles of the Serotonergic System in Coping with Traumatic Stress. In *Serotonin and the CNS*; Berend, O., Ed.; IntechOpen: Rijeka, Croatia, 2021; pp. 1–5.

18. Paine, N.J.; Watkins, L.L.; Blumenthal, J.A.; Kuhn, C.M.; Sherwood, A. Association of depressive and anxiety symptoms with 24-hour urinary catecholamines in individuals with untreated high blood pressure. *Psychosom. Med.* **2015**, *77*, 136–144. [[CrossRef](#)]
19. Brindley, R.L.; Bauer, M.B.; Blakely, R.D.; Currie, K.P.M. An interplay between the serotonin transporter (SERT) and 5-HT receptors controls stimulus-secretion coupling in sympathoadrenal chromaffin cells. *Neuropharmacology* **2016**, *110*, 438–448. [[CrossRef](#)]
20. Nakatani, Y.; Sato-Suzuki, I.; Tsujino, N.; Nakasato, A.; Seki, Y.; Fumoto, M.; Arita, H. Augmented brain 5-HT crosses the blood-brain barrier through the 5-HT transporter in rat. *Eur. J. Neurosci.* **2008**, *27*, 2466–2472. [[CrossRef](#)]
21. Wang, H.M.; Wang, Y.; Liu, M.; Bai, Y.; Zhang, X.H.; Sun, Y.X.; Wang, H.L. Fluoxetine inhibits monocrotaline-induced pulmonary arterial remodeling involved in inhibition of RhoA-Rho kinase and Akt signalling pathways in rats. *Can. J. Physiol. Pharmacol.* **2012**, *90*, 1506–1515. [[CrossRef](#)]
22. Lin, J.C.; Chou, C.C.; Tu, Z.; Yeh, L.F.; Wu, S.C.; Khoo, K.H.; Lin, C.H. Characterization of protein serotonylation via bioorthogonal labeling and enrichment. *J. Proteome Res.* **2014**, *13*, 3523–3529. [[CrossRef](#)]
23. Penumatsa, K.C.; Fanburg, B.L. Transglutaminase 2-mediated serotonylation in pulmonary hypertension. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2014**, *306*, L309–L315. [[CrossRef](#)] [[PubMed](#)]
24. Tjurmina, O.A.; Armando, I.; Saavedra, J.M.; Goldstein, D.S.; Murphy, D.L. Exaggerated adrenomedullary response to immobilization in mice with targeted disruption of the serotonin transporter gene. *Endocrinology* **2002**, *143*, 4520–4526. [[CrossRef](#)] [[PubMed](#)]
25. Tiradentes, R.V.; Pires, J.G.; Silva, N.F.; Ramage, A.G.; Santuzzi, C.H.; Futuro Neto, H.A. Effects of acute administration of selective serotonin reuptake inhibitors on sympathetic nerve activity. *Braz. J. Med. Biol.* **2014**, *47*, 554–559. [[CrossRef](#)] [[PubMed](#)]
26. Schroeter, S.; Levey, A.I.; Blakely, R.D. Polarized expression of the antidepressant-sensitive serotonin transporter in epinephrine-synthesizing chromaffin cells of the rat adrenal gland. *Mol. Cell. Neurosci.* **1997**, *9*, 170–184. [[CrossRef](#)]
27. Furlan, A.; Dyachuk, V.; Kastrić, M.E.; Calvo-Enrique, L.; Abdo, H.; Hadjab, S.; Chontorotzea, T.; Akkuratova, N.; Usoskin, D.; Kamenev, D.; et al. Multipotent peripheral glial cells generate neuroendocrine cells of the adrenal medulla. *Science* **2017**, *357*, eaal3753. [[CrossRef](#)]
28. Kameneva, P.; Melnikova, V.I.; Kastrić, M.E.; Kurtova, A.; Kryukov, E.; Murtazina, A.; Faure, L.; Poverennaya, I.; Artemov, A.V.; Kalinina, T.S.; et al. Serotonin limits generation of chromaffin cells during adrenal organ development. *Nat. Commun.* **2022**, *13*, 2901. [[CrossRef](#)]
29. Carbone, E.; Borges, R.; Eiden, L.E.; García, A.G.; Hernández-Cruz, A. Chromaffin Cells of the Adrenal Medulla: Physiology, Pharmacology, and Disease. *Compr. Physiol.* **2019**, *9*, 1443–1502. [[CrossRef](#)]
30. Brindley, R.L.; Bauer, M.B.; Blakely, R.D.; Currie, K.P.M. Serotonin and Serotonin Transporters in the Adrenal Medulla: A Potential Hub for Modulation of the Sympathetic Stress Response. *ACS Chem. Neurosci.* **2017**, *8*, 943–954. [[CrossRef](#)]
31. Linder, A.E.; Beggs, K.M.; Burnett, R.J.; Watts, S.W. Body distribution of infused serotonin in rats. *Clin. Exp. Pharmacol. Physiol.* **2009**, *36*, 599–601. [[CrossRef](#)]
32. Kobayashi, S.; Tsukahara, S.; Sugita, K.; Nagata, T. Adrenergic and cholinergic innervation of rat cerebral arteries. Consecutive demonstration on whole mount preparations. *Histochemistry* **1981**, *70*, 129–138. [[CrossRef](#)]
33. Sheng, Y.; Zhu, L. The crosstalk between autonomic nervous system and blood vessels. *Int. J. Physiol. Pathophysiol. Pharmacol.* **2018**, *10*, 17–28.
34. Koep, J.L.; Taylor, C.E.; Coombes, J.S.; Bond, B.; Ainslie, P.N.; Bailey, T.G. Autonomic control of cerebral blood flow: Fundamental comparisons between peripheral and cerebrovascular circulations in humans. *J. Physiol.* **2022**, *600*, 15–39. [[CrossRef](#)]
35. Suzuki, N.; Hardebo, J.E. The cerebrovascular parasympathetic innervation. *Cerebrovasc. Brain Metab. Rev.* **1993**, *5*, 33–46.
36. Roloff, E.V.L.; Tomiak-Baquero, A.M.; Kasparov, S.; Paton, J.F.R. Parasympathetic innervation of vertebrobasilar arteries: Is this a potential clinical target? *J. Physiol.* **2016**, *594*, 6463–6485. [[CrossRef](#)]
37. Miller, K.E.; Salvatierra, A.T. Apposition of enkephalin- and neurotensin-immunoreactive neurons by serotonin-immunoreactive varicosities in the rat spinal cord. *Neuroscience* **1998**, *85*, 837–846. [[CrossRef](#)]
38. Villalón, C.M.; Centurión, D.; Rabelo, G.; de Vries, P.; Saxena, P.R.; Sánchez-López, A. The 5-HT<sub>1</sub>-like receptors mediating inhibition of sympathetic vasopressor outflow in the pithed rat: Operational correlation with the 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub> and 5-HT<sub>1D</sub> subtypes. *Br. J. Pharmacol.* **1998**, *124*, 1001–1011. [[CrossRef](#)]
39. Rapport, M.M.; Green, A.A.; Page, I.H. Serum vasoconstrictor, serotonin; isolation and characterization. *J. Biol. Chem.* **1948**, *176*, 1243–1251. [[CrossRef](#)]
40. Rapport, M.M.; Green, A.A.; Page, I.H. Partial purification of the vasoconstrictor in beef serum. *J. Biol. Chem.* **1948**, *174*, 735–741. [[CrossRef](#)]
41. Herr, N.; Bode, C.; Duerschmied, D. The Effects of Serotonin in Immune Cells. *Front. Cardiovasc. Med.* **2017**, *4*, 48. [[CrossRef](#)]
42. Palermo, A.; del Rosso, G.; Costantini, C.; Bertalero, P.; Rizzi, S.; Libretti, A. Platelet content of serotonin and response to stress. *J. Hypertens. Suppl.* **1986**, *4*, S43–S45.
43. Teff, K.L.; Young, S.N. Effects of carbohydrate and protein administration on rat tryptophan and 5-hydroxytryptamine: Differential effects on the brain, intestine, pineal, and pancreas. *Can. J. Physiol. Pharmacol.* **1988**, *66*, 683–688. [[CrossRef](#)] [[PubMed](#)]
44. Edvinsson, J.C.A.; Maddahi, A.; Christiansen, I.M.; Reducha, P.V.; Warfvinge, K.; Sheykhzade, M.; Edvinsson, L.; Haanes, K.A. Lasmiditan and 5-Hydroxytryptamine in the rat trigeminal system; expression, release and interactions with 5-HT<sub>1</sub> receptors. *J. Headache Pain* **2022**, *23*, 26. [[CrossRef](#)] [[PubMed](#)]
45. Ishida, Y.; Sugiura, Y.; Magome, T.; Kamakura, T.; Takimoto, Y.; Hanada, Y.; Kitayama, K.; Nakamura, Y.; Shimada, S.; Ohta, N.; et al. Expression Analysis of Serotonin Receptors, Serotonin Transporter and L-Amino Acid Decarboxylase in the Mouse Sphenopalatine Ganglion by RT-PCR, Northern Blot Analysis and In Situ Hybridization. *Neuroscience* **2019**, *411*, 23–36. [[CrossRef](#)] [[PubMed](#)]

46. Punda, H.; Mardesic, S.; Filipovic, N.; Kosovic, I.; Benzon, B.; Ogorevc, M.; Bocina, I.; Kolic, K.; Vukojevic, K.; Saraga-Babic, M. Expression Pattern of 5-HT (Serotonin) Receptors during Normal Development of the Human Spinal Cord and Ganglia and in Fetus with Cervical Spina Bifida. *Int. J. Mol. Sci.* **2021**, *22*, 7320. [[CrossRef](#)] [[PubMed](#)]
47. Kimura, T.; Satoh, S. Presynaptic inhibition by serotonin of cardiac sympathetic transmission in dogs. *Clin. Exp. Pharmacol. Physiol.* **1983**, *10*, 535–542. [[CrossRef](#)]
48. García-Pedraza, J.; Hernández-Abreu, O.; Morán, A.; Carretero, J.; García-Domingo, M.; Villalón, C.M. Role of peripheral 5-HT(5A) receptors in 5-HT-induced cardiac sympatho-inhibition in type 1 diabetic rats. *Sci. Rep.* **2020**, *10*, 19358. [[CrossRef](#)]
49. Morán, A.; Fernández, M.M.; Velasco, C.; Martín, M.L.; San Román, L. Characterization of prejunctional 5-HT<sub>1</sub> receptors that mediate the inhibition of pressor effects elicited by sympathetic stimulation in the pithed rat. *Br. J. Pharmacol.* **1998**, *123*, 1205–1213. [[CrossRef](#)]
50. Pilowsky, P.M. Chapter 16—Serotonin in Central Cardiovascular Regulation: Ex Uno Plura (From One Comes Many). In *Serotonin*; Pilowsky, P.M., Ed.; Academic Press: Cambridge, MA, USA, 2019; pp. 335–347.
51. Morán, A.; Velasco, C.; Salvador, T.; Martín, M.L.; San Román, L. Inhibitory 5-hydroxytryptamine receptors involved in pressor effects obtained by stimulation of sympathetic outflow from spinal cord in pithed rats. *Br. J. Pharmacol.* **1994**, *113*, 1358–1362. [[CrossRef](#)]
52. Villamil-Hernández, M.T.; Alcántara-Vázquez, O.; Sánchez-López, A.; Gutiérrez-Lara, E.J.; Centurión, D. Pharmacological evidence that 5-HT<sub>1A/1B/1D</sub>,  $\alpha$ <sub>2</sub>-adrenoceptors and D<sub>2</sub>-like receptors mediate ergotamine-induced inhibition of the vasopressor sympathetic outflow in pithed rats. *Eur. J. Pharmacol.* **2014**, *740*, 512–521. [[CrossRef](#)]
53. Molderings, G.J.; Frölich, D.; Likungu, J.; Göthert, M. Inhibition of noradrenaline release via presynaptic 5-HT<sub>1D</sub> alpha receptors in human atrium. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1996**, *353*, 272–280. [[CrossRef](#)]
54. Morán, A.; Velasco, C.; Martín, M.L.; San Román, L. Pharmacological characterization of 5-HT receptors in parasympathetic innervation of rat heart. *Eur. J. Pharmacol.* **1994**, *252*, 161–166. [[CrossRef](#)]
55. Lee, T.J.; Liu, J.; Evans, M.S. Cholinergic-nitric transmitter mechanisms in the cerebral circulation. *Microsc. Res. Tech.* **2001**, *53*, 119–128. [[CrossRef](#)]
56. Jackowski, A.; Crockard, A.; Burnstock, G. 5-Hydroxytryptamine demonstrated immunohistochemically in rat cerebrovascular nerves largely represents 5-hydroxytryptamine uptake into sympathetic nerve fibres. *Neuroscience* **1989**, *29*, 453–462. [[CrossRef](#)]
57. Boyle, S.H.; Brummett, B.H.; Kuhn, C.M.; Barefoot, J.C.; Siegler, I.C.; Williams, R.B.; Georgiades, A. The Effects of Tryptophan Enhancement and Depletion on Plasma Catecholamine Levels in Healthy Individuals. *Psychosom. Med.* **2019**, *81*, 34–40. [[CrossRef](#)]
58. Švec, J.; Švec, P.; Bencová, V.; Krčméry, V. Anxio-depressive Syndrome—Biopsychosocial Model of Supportive Care. *Klin. Onkol.* **2015**, *28*, 177–182. [[CrossRef](#)]
59. Razzaque, Z.; Pickard, J.D.; Ma, Q.P.; Shaw, D.; Morrison, K.; Wang, T.; Longmore, J. 5-HT<sub>1B</sub>-receptors and vascular reactivity in human isolated blood vessels: Assessment of the potential craniovascular selectivity of sumatriptan. *Br. J. Clin. Pharmacol.* **2002**, *53*, 266–274. [[CrossRef](#)]
60. Gonzalez-Hernandez, A.; Marichal-Cancino, B.A.; MaassenVanDenBrink, A.; Villalón, C.M. Side effects associated with current and prospective antimigraine pharmacotherapies. *Expert. Opin. Drug. Metab. Toxicol.* **2018**, *14*, 25–41. [[CrossRef](#)]
61. Marichal-Cancino, B.A.; González-Hernández, A.; Guerrero-Alba, R.; Medina-Santillán, R.; Villalón, C.M. A critical review of the neurovascular nature of migraine and the main mechanisms of action of prophylactic antimigraine medications. *Expert. Rev. Neurother.* **2021**, *21*, 1035–1050. [[CrossRef](#)]
62. Villalón, C.M.; Sánchez-López, A.; Centurión, D. Operational characteristics of the 5-HT<sub>1</sub>-like receptors mediating external carotid vasoconstriction in vagosympathetomized dogs. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1996**, *354*, 550–556. [[CrossRef](#)]
63. Zaidi, M.; Bevis, P.J.; Girgis, S.I.; Lynch, C.; Stevenson, J.C.; MacIntyre, I. Circulating CGRP comes from the perivascular nerves. *Eur. J. Pharmacol.* **1985**, *117*, 283–284. [[CrossRef](#)]
64. Escott, K.J.; Connor, H.E.; Brain, S.D.; Beattie, D.T. The involvement of calcitonin gene-related peptide (CGRP) and substance P in feline pial artery diameter responses evoked by capsaicin. *Neuropeptides* **1995**, *29*, 129–135. [[CrossRef](#)] [[PubMed](#)]
65. Van der Schueren, B.J.; de Hoon, J.N.; Vanmolkot, F.H.; Van Hecken, A.; Depre, M.; Kane, S.A.; De Lepeleire, I.; Sinclair, S.R. Reproducibility of the capsaicin-induced dermal blood flow response as assessed by laser Doppler perfusion imaging. *Br. J. Clin. Pharmacol.* **2007**, *64*, 580–590. [[CrossRef](#)] [[PubMed](#)]
66. Dux, M.; Rosta, J.; Messlinger, K. TRP Channels in the Focus of Trigeminal Nociceptor Sensitization Contributing to Primary Headaches. *Int. J. Mol. Sci.* **2020**, *21*, 342. [[CrossRef](#)]
67. Julius, D.; Basbaum, A.I. Molecular mechanisms of nociception. *Nature* **2001**, *413*, 203–210. [[CrossRef](#)]
68. Kawasaki, H.; Nuki, C.; Saito, A.; Takasaki, K. Adrenergic modulation of calcitonin gene-related peptide (CGRP)-containing nerve-mediated vasodilation in the rat mesenteric resistance vessel. *Brain Res.* **1990**, *506*, 287–290. [[CrossRef](#)]
69. Kawasaki, H.; Takasaki, K.; Saito, A.; Goto, K. Calcitonin gene-related peptide acts as a novel vasodilator neurotransmitter in mesenteric resistance vessels of the rat. *Nature* **1988**, *335*, 164–167. [[CrossRef](#)]
70. Taguchi, T.; Kawasaki, H.; Imamura, T.; Takasaki, K. Endogenous calcitonin gene-related peptide mediates nonadrenergic noncholinergic depressor response to spinal cord stimulation in the pithed rat. *Circ. Res.* **1992**, *71*, 357–364. [[CrossRef](#)]
71. Kawasaki, H.; Saito, A.; Takasaki, K. Age-related decrease of calcitonin gene-related peptide-containing vasodilator innervation in the mesenteric resistance vessel of the spontaneously hypertensive rat. *Circ. Res.* **1990**, *67*, 733–743. [[CrossRef](#)]
72. Avilés-Rosas, V.H.; Rivera-Mancilla, E.; Marichal-Cancino, B.A.; Manrique-Maldonado, G.; Altamirano-Espinoza, A.H.; Maassen Van Den Brink, A.; Villalón, C.M. Olcegepant blocks neurogenic and non-neurogenic CGRPergic vasodepressor responses and facilitates noradrenergic vasopressor responses in pithed rats. *Br. J. Pharmacol.* **2017**, *174*, 2001–2014. [[CrossRef](#)]

73. Russell, F.A.; King, R.; Smillie, S.J.; Kodji, X.; Brain, S.D. Calcitonin gene-related peptide: Physiology and pathophysiology. *Physiol. Rev.* **2014**, *94*, 1099–1142. [[CrossRef](#)]
74. Russo, A.F.; Hay, D.L. CGRP physiology, pharmacology, and therapeutic targets: Migraine and beyond. *Physiol. Rev.* **2023**, *103*, 1565–1644. [[CrossRef](#)]
75. Villalón, C.M.; Albarrán-Juárez, J.A.; Lozano-Cuenca, J.; Pertz, H.H.; Görnemann, T.; Centurión, D. Pharmacological profile of the clonidine-induced inhibition of vasodepressor sensory outflow in pithed rats: Correlation with alpha(2A/2C)-adrenoceptors. *Br. J. Pharmacol.* **2008**, *154*, 51–59. [[CrossRef](#)]
76. González-Hernández, A.; Manrique-Maldonado, G.; Lozano-Cuenca, J.; Muñoz-Islas, E.; Centurión, D.; Maassen VanDenBrink, A.; Villalón, C.M. The 5-HT(1) receptors inhibiting the rat vasodepressor sensory CGRPergic outflow: Further involvement of 5-HT(1F), but not 5-HT(1A) or 5-HT(1D), subtypes. *Eur. J. Pharmacol.* **2011**, *659*, 233–243. [[CrossRef](#)]
77. González-Hernández, A.; Muñoz-Islas, E.; Lozano-Cuenca, J.; Ramírez-Rosas, M.B.; Sánchez-López, A.; Centurión, D.; Ramírez-San Juan, E.; Villalón, C.M. Activation of 5-HT1B receptors inhibits the vasodepressor sensory CGRPergic outflow in pithed rats. *Eur. J. Pharmacol.* **2010**, *637*, 131–137. [[CrossRef](#)]
78. Ibrahimi, K.; Danser, A.; Terwindt, G.M.; van den Meiracker, A.H.; MaassenVanDenBrink, A. A human trigeminovascular biomarker for antimigraine drugs: A randomised, double-blind, placebo-controlled, crossover trial with sumatriptan. *Cephalalgia* **2017**, *37*, 94–98. [[CrossRef](#)]
79. Benemei, S.; Cortese, F.; Labastida-Ramírez, A.; Marchese, F.; Pellesi, L.; Romoli, M.; Vollesen, A.L.; Lampl, C.; Ashina, M. Triptans and CGRP blockade—Impact on the cranial vasculature. *J. Headache Pain* **2017**, *18*, 103. [[CrossRef](#)]
80. Labastida-Ramírez, A.; Rubio-Beltrán, E.; Haanes, K.A.; Chan, K.Y.; Garrelds, I.M.; Johnson, K.W.; Danser, A.H.J.; Villalón, C.M.; MaassenVanDenBrink, A. Lasmiditan inhibits calcitonin gene-related peptide release in the rodent trigeminovascular system. *Pain* **2020**, *161*, 1092–1099. [[CrossRef](#)]
81. Buzzi, M.G.; Carter, W.B.; Shimizu, T.; Heath, H., 3rd; Moskowitz, M.A. Dihydroergotamine and sumatriptan attenuate levels of CGRP in plasma in rat superior sagittal sinus during electrical stimulation of the trigeminal ganglion. *Neuropharmacology* **1991**, *30*, 1193–1200. [[CrossRef](#)]
82. Gupta, S.; Akerman, S.; van den Maagdenberg, A.M.; Saxena, P.R.; Goadsby, P.J.; van den Brink, A.M. Intravital microscopy on a closed cranial window in mice: A model to study trigeminovascular mechanisms involved in migraine. *Cephalalgia* **2006**, *26*, 1294–1303. [[CrossRef](#)]
83. Limmroth, V.; Katsarava, Z.; Liedert, B.; Guehring, H.; Schmitz, K.; Diener, H.C.; Michel, M.C. An in vivo rat model to study calcitonin gene related peptide release following activation of the trigeminal vascular system. *Pain* **2001**, *92*, 101–106. [[CrossRef](#)]
84. Williamson, D.J.; Hargreaves, R.J.; Hill, R.G.; Shephard, S.L. Sumatriptan inhibits neurogenic vasodilation of dural blood vessels in the anaesthetized rat-intravital microscope studies. *Cephalalgia* **1997**, *17*, 525–531. [[CrossRef](#)] [[PubMed](#)]
85. Ashina, M.; Hansen, J.M.; Do, T.P.; Melo-Carrillo, A.; Burstein, R.; Moskowitz, M.A. Migraine and the trigeminovascular system—40 years and counting. *Lancet Neurol.* **2019**, *18*, 795–804. [[CrossRef](#)] [[PubMed](#)]
86. Goadsby, P.J.; Holland, P.R.; Martins-Oliveira, M.; Hoffmann, J.; Schankin, C.; Akerman, S. Pathophysiology of Migraine: A Disorder of Sensory Processing. *Physiol. Rev.* **2017**, *97*, 553–622. [[CrossRef](#)] [[PubMed](#)]
87. Moreno-Ajona, D.; Chan, C.; Villar-Martínez, M.D.; Goadsby, P.J. Targeting CGRP and 5-HT(1F) Receptors for the Acute Therapy of Migraine: A Literature Review. *Headache* **2019**, *59* (Suppl. S2), 3–19. [[CrossRef](#)]
88. Lozano-Cuenca, J.; González-Hernández, A.; Muñoz-Islas, E.; Sánchez-López, A.; Centurión, D.; Cobos-Puc, L.E.; Villalón, C.M. Effect of some acute and prophylactic antimigraine drugs on the vasodepressor sensory CGRPergic outflow in pithed rats. *Life Sci.* **2009**, *84*, 125–131. [[CrossRef](#)]
89. Nicholson, R.; Small, J.; Dixon, A.K.; Spanswick, D.; Lee, K. Serotonin receptor mRNA expression in rat dorsal root ganglion neurons. *Neurosci. Lett.* **2003**, *337*, 119–122. [[CrossRef](#)]
90. Muñoz-Islas, E.; Gupta, S.; Jiménez-Mena, L.R.; Lozano-Cuenca, J.; Sánchez-López, A.; Centurión, D.; Mehrotra, S.; MaassenVanDenBrink, A.; Villalón, C.M. Donitriptan, but not sumatriptan, inhibits capsaicin-induced canine external carotid vasodilatation via 5-HT1B rather than 5-HT1D receptors. *Br. J. Pharmacol.* **2006**, *149*, 82–91. [[CrossRef](#)]
91. Muñoz-Islas, E.; Lozano-Cuenca, J.; González-Hernández, A.; Ramírez-Rosas, M.B.; Sánchez-López, A.; Centurión, D.; MaassenVanDenBrink, A.; Villalón, C.M. Spinal sumatriptan inhibits capsaicin-induced canine external carotid vasodilatation via 5-HT1B rather than 5-HT1D receptors. *Eur. J. Pharmacol.* **2009**, *615*, 133–138. [[CrossRef](#)]
92. Rubio-Beltrán, E.; Labastida-Ramírez, A.; Villalón, C.M.; MaassenVanDenBrink, A. Is selective 5-HT(1F) receptor agonism an entity apart from that of the triptans in antimigraine therapy? *Pharmacol. Ther.* **2018**, *186*, 88–97. [[CrossRef](#)]
93. Fujii, H.; Takatori, S.; Zamami, Y.; Hashikawa-Hobara, N.; Miyake, N.; Tangsucharit, P.; Mio, M.; Kawasaki, H. Adrenergic stimulation-released 5-HT stored in adrenergic nerves inhibits CGRPergic nerve-mediated vasodilatation in rat mesenteric resistance arteries. *Br. J. Pharmacol.* **2012**, *166*, 2084–2094. [[CrossRef](#)]
94. Gomez-Mancilla, B.; Cutler, N.R.; Leibowitz, M.T.; Spierings, E.L.; Klapper, J.A.; Diamond, S.; Goldstein, J.; Smith, T.; Couch, J.R.; Fleishaker, J.; et al. Safety and efficacy of PNU-142633, a selective 5-HT1D agonist, in patients with acute migraine. *Cephalalgia* **2001**, *21*, 727–732. [[CrossRef](#)]
95. González-Hernández, A.; Marichal-Cancino, B.A.; Lozano-Cuenca, J.; MaassenVanDenBrink, A.; Villalón, C.M. Functional Characterization of the Prejunctional Receptors Mediating the Inhibition by Ergotamine of the Rat Perivascular Sensory Peptidergic Drive. *ACS Chem. Neurosci.* **2019**, *10*, 3173–3182. [[CrossRef](#)]

96. de Jong, A.P.; Verhage, M. Presynaptic signal transduction pathways that modulate synaptic transmission. *Curr. Opin. Neurobiol.* **2009**, *19*, 245–253. [[CrossRef](#)]
97. Cuesta, C.; García-Pedraza, J.; García, M.; Villalón, C.M.; Morán, A. Role of 5-HT<sub>7</sub> receptors in the inhibition of the vasodepressor sensory CGRPergic outflow in pithed rats. *Vasc. Pharmacol.* **2014**, *63*, 4–12. [[CrossRef](#)]
98. Chan, A.K.; von der Weid, P.Y. 5-HT decreases contractile and electrical activities in lymphatic vessels of the guinea-pig mesentery: Role of 5-HT<sub>7</sub>-receptors. *Br. J. Pharmacol.* **2003**, *139*, 243–254. [[CrossRef](#)]
99. Chan, M.F.; Okun, I.; Stavros, F.L.; Hwang, E.; Wolff, M.E.; Balaji, V.N. Identification of a new class of ETA selective endothelin antagonists by pharmacophore directed screening. *Biochem. Biophys. Res. Commun.* **1994**, *201*, 228–234. [[CrossRef](#)]
100. Wiklund, N.P.; Wiklund, C.U.; Cederqvist, B.; Ohlén, A.; Hedqvist, P.; Gustafsson, L.E. Endothelin modulation of neuroeffector transmission in smooth muscle. *J. Cardiovasc. Pharmacol.* **1991**, *17* (Suppl. S7), S335–S339. [[CrossRef](#)]
101. Filippelli, A.; Falciani, M.; Piucci, B.; D'Amico, M.; D'Agostino, B.; Filippelli, W.; Rossi, F. Endothelin-1 affects capsaicin-evoked release of neuropeptides from rat vas deferens. *Eur. J. Pharmacol.* **1999**, *364*, 183–191. [[CrossRef](#)]
102. Brenchat, A.; Zamanillo, D.; Hamon, M.; Romero, L.; Vela, J.M. Role of peripheral versus spinal 5-HT(7) receptors in the modulation of pain undersensitizing conditions. *Eur. J. Pain.* **2012**, *16*, 72–81. [[CrossRef](#)]
103. Chen, J.J.; Vasko, M.R.; Wu, X.; Staeva, T.P.; Baez, M.; Zgombick, J.M.; Nelson, D.L. Multiple subtypes of serotonin receptors are expressed in rat sensory neurons in culture. *J. Pharmacol. Exp. Ther.* **1998**, *287*, 1119–1127.
104. Pierce, P.A.; Xie, G.X.; Meuser, T.; Peroutka, S.J. 5-Hydroxytryptamine receptor subtype messenger RNAs in human dorsal root ganglia: A polymerase chain reaction study. *Neuroscience* **1997**, *81*, 813–819. [[CrossRef](#)] [[PubMed](#)]
105. Sugiuar, T.; Bielefeldt, K.; Gebhart, G.F. TRPV1 function in mouse colon sensory neurons is enhanced by metabotropic 5-hydroxytryptamine receptor activation. *J. Neurosci.* **2004**, *24*, 9521–9530. [[CrossRef](#)] [[PubMed](#)]
106. Wang, D.; Chen, T.; Gao, Y.; Quirion, R.; Hong, Y. Inhibition of SNL-induced upregulation of CGRP and NPY in the spinal cord and dorsal root ganglia by the 5-HT(2A) receptor antagonist ketanserin in rats. *Pharmacol. Biochem. Behav.* **2012**, *101*, 379–386. [[CrossRef](#)] [[PubMed](#)]
107. Muñoz-Islas, E.; Vidal-Cantú, G.C.; Bravo-Hernández, M.; Cervantes-Durán, C.; Quiñonez-Bastidas, G.N.; Pineda-Farias, J.B.; Barragán-Iglesias, P.; Granados-Soto, V. Spinal 5-HT<sub>5A</sub> receptors mediate 5-HT-induced antinociception in several pain models in rats. *Pharmacol. Biochem. Behav.* **2014**, *120*, 25–32. [[CrossRef](#)] [[PubMed](#)]
108. Tramontana, M.; Giuliani, S.; Del Bianco, E.; Lecci, A.; Maggi, C.A.; Evangelista, S.; Geppetti, P. Effects of capsaicin and 5-HT<sub>3</sub> antagonists on 5-hydroxytryptamine-evoked release of calcitonin gene-related peptide in the guinea-pig heart. *Br. J. Pharmacol.* **1993**, *108*, 431–435. [[CrossRef](#)]
109. Smillie, S.J.; Brain, S.D. Calcitonin gene-related peptide (CGRP) and its role in hypertension. *Neuropeptides* **2011**, *45*, 93–104. [[CrossRef](#)]
110. Lu, J.T.; Son, Y.J.; Lee, J.; Jetton, T.L.; Shiota, M.; Moscoso, L.; Niswender, K.D.; Loewy, A.D.; Magnuson, M.A.; Sanes, J.R.; et al. Mice lacking alpha-calcitonin gene-related peptide exhibit normal cardiovascular regulation and neuromuscular development. *Mol. Cell. Neurosci.* **1999**, *14*, 99–120. [[CrossRef](#)]
111. Mai, T.H.; Wu, J.; Diedrich, A.; Garland, E.M.; Robertson, D. Calcitonin gene-related peptide (CGRP) in autonomic cardiovascular regulation and vascular structure. *J. Am. Soc. Hypertens. JASH* **2014**, *8*, 286–296. [[CrossRef](#)]
112. Kudrow, D.; Pascual, J.; Winner, P.K.; Dodick, D.W.; Tepper, S.J.; Reuter, U.; Hong, F.; Klatt, J.; Zhang, F.; Cheng, S.; et al. Vascular safety of erenumab for migraine prevention. *Neurology* **2020**, *94*, e497–e510. [[CrossRef](#)]
113. de Vries Lentsch, S.; van der Arend, B.W.H.; Maassen VanDenBrink, A.; Terwindt, G.M. Blood Pressure in Patients with Migraine Treated with Monoclonal Anti-CGRP (Receptor) Antibodies: A Prospective Follow-up Study. *Neurology* **2022**, *99*, e1897–e1904. [[CrossRef](#)]
114. Kumar, A.; Potts, J.D.; DiPette, D.J. Protective Role of  $\alpha$ -Calcitonin Gene-Related Peptide in Cardiovascular Diseases. *Front. Physiol.* **2019**, *10*, 821. [[CrossRef](#)]
115. Kumar, A.; Williamson, M.; Hess, A.; DiPette, D.J.; Potts, J.D. Alpha-Calcitonin Gene Related Peptide: New Therapeutic Strategies for the Treatment and Prevention of Cardiovascular Disease and Migraine. *Front. Physiol.* **2022**, *13*, 826122. [[CrossRef](#)]
116. MaassenVanDenBrink, A.; Terwindt, G.M.; van den Maagdenberg, A. Calcitonin gene-related peptide (receptor) antibodies: An exciting avenue for migraine treatment. *Genome Med.* **2018**, *10*, 10. [[CrossRef](#)]
117. Shi, X.Y.; Yang, Y.; Zhao, Y.T. Plasma calcitonin gene-related peptide (CGRP) level in patients with essential hypertension. *Zhonghua Nei Ke Za Zhi* **1990**, *29*, 616–618.
118. Xu, D.; Wang, X.A.; Wang, J.P. Calcitonin gene-related peptide (CGRP) in normotensive and spontaneously hypertensive rats. *Peptides* **1989**, *17*, 174–177. [[CrossRef](#)]
119. Watson, R.E.; Supowit, S.C.; Zhao, H.; Katki, K.A.; Dipette, D.J. Role of sensory nervous system vasoactive peptides in hypertension. *Braz. J. Med. Biol. Res.* **2002**, *35*, 1033–1045. [[CrossRef](#)]
120. Guyton, A.C.; Coleman, T.G.; Cowley, A.V., Jr.; Scheel, K.W.; Manning, R.D., Jr.; Norman, R.A., Jr. Arterial pressure regulation. Overriding dominance of the kidneys in long-term regulation and in hypertension. *Am. J. Med.* **1972**, *52*, 584–594. [[CrossRef](#)]
121. Hoffman, B.B. Catecholamines, sympathomimetic drugs, and adrenergic receptor antagonists. In *Goodman & Gilman's The Pharmacological Basis of Therapeutics*, 10th ed.; Hardman, J.G., Laurence, L., Goodman Gilman, A., Eds.; McGraw-Hill: New York, NY, USA, 2001; pp. 215–268.
122. Gardiner, S.M.; Compton, A.M.; Bennett, T. Regional hemodynamic effects of calcitonin gene-related peptide. *Am. J. Physiol.* **1989**, *256*, R332–R338. [[CrossRef](#)]

123. Han, S.P.; Naes, L.; Westfall, T.C. Calcitonin gene-related peptide is the endogenous mediator of nonadrenergic-noncholinergic vasodilation in rat mesentery. *J. Pharmacol. Exp. Ther.* **1990**, *255*, 423–428.
124. Han, S.P.; Naes, L.; Westfall, T.C. Inhibition of periarterial nerve stimulation-induced vasodilation of the mesenteric arterial bed by CGRP (8–37) and CGRP receptor desensitization. *Biochem. Biophys. Res. Commun.* **1990**, *168*, 786–791. [[CrossRef](#)]
125. Holzer, P. Local effector functions of capsaicin-sensitive sensory nerve endings: Involvement of tachykinins, calcitonin gene-related peptide and other neuropeptides. *Neuroscience* **1988**, *24*, 739–768. [[CrossRef](#)] [[PubMed](#)]
126. Machida, T.; Iizuka, K.; Hirafuji, M. 5-hydroxytryptamine and its receptors in systemic vascular walls. *Biol. Pharm. Bull.* **2013**, *36*, 1416–1419. [[CrossRef](#)] [[PubMed](#)]
127. Sumner, M.J. Characterization of the 5-HT receptor mediating endothelium-dependent relaxation in porcine vena cava. *Br. J. Pharmacol.* **1991**, *102*, 938–942. [[CrossRef](#)] [[PubMed](#)]
128. Rivasi, G.; Menale, S.; Turrin, G.; Coscarelli, A.; Giordano, A.; Ungar, A. The Effects of Pain and Analgesic Medications on Blood Pressure. *Curr. Hypertens. Rep.* **2022**, *24*, 385–394. [[CrossRef](#)]
129. Sánchez, M.G.; Morissette, M.; Di Paolo, T. Oestradiol Modulation of Serotonin Reuptake Transporter and Serotonin Metabolism in the Brain of Monkeys. *J. Neuroendocrinol.* **2013**, *25*, 560–569. [[CrossRef](#)]
130. Soslau, G. Cardiovascular serotonergic system: Evolution, receptors, transporter, and function. *J. Exp. Zool. A Ecol. Integr. Physiol.* **2022**, *337*, 115–127. [[CrossRef](#)]
131. Erspamer, V.; Asero, B. Identification of enteramine, the specific hormone of the enterochromaffin cell system, as 5-hydroxytryptamine. *Nature* **1952**, *169*, 800–801. [[CrossRef](#)]
132. Fetkovska, N.; Pletscher, A.; Ferracin, F.; Amstein, R.; Buhler, F.R. Impaired uptake of 5 hydroxytryptamine platelet in essential hypertension: Clinical relevance. *Cardiovasc. Drugs Ther.* **1990**, *4* (Suppl. S1), 105–109. [[CrossRef](#)]
133. Diaz, J.; Ni, W.; Thompson, J.; King, A.; Fink, G.D.; Watts, S.W. 5-Hydroxytryptamine lowers blood pressure in normotensive and hypertensive rats. *J. Pharmacol. Exp. Ther.* **2008**, *325*, 1031–1038. [[CrossRef](#)]
134. Patrick Davis, R.; Linder, A.E.; Watts, S.W. Lack of the serotonin transporter (SERT) reduces the ability of 5-hydroxytryptamine to lower blood pressure. *Naunyn Schmiedebergs Arch. Pharmacol.* **2011**, *383*, 543–546. [[CrossRef](#)]
135. Karlsen, H.R.; Løchen, M.L.; Langvik, E. Antidepressant Use and Risk of Myocardial Infarction: A Longitudinal Investigation of Sex-Specific Associations in the HUNT Study. *Psychosom. Med.* **2023**, *85*, 26–33. [[CrossRef](#)]
136. Niazi, S.K.; Memon, S.H.; Lesser, E.R.; Brennan, E.; Aslam, N. Assessment of psychiatric comorbidities and serotonergic or noradrenergic medication use on blood pressure using 24-hour ambulatory blood pressure monitoring. *J. Clin. Hypertens.* **2021**, *23*, 1599–1607. [[CrossRef](#)]
137. Muñoz-Islas, E.; González-Hernández, A.; Lozano-Cuenca, J.; Ramírez-Rosas, M.B.; Medina-Santillán, R.; Centurión, D.; Maassen-VanDenBrink, A.; Villalón, C.M. Inhibitory effect of chronic oral treatment with fluoxetine on capsaicin-induced external carotid vasodilatation in anaesthetised dogs. *Cephalalgia* **2015**, *35*, 1041–1053. [[CrossRef](#)]
138. Pak, K.; Kim, K.; Seo, S.; Lee, M.J.; Kim, I.J. Serotonin transporter is negatively associated with body mass index after glucose loading in humans. *Brain Imaging Behav.* **2022**, *16*, 1246–1251. [[CrossRef](#)]
139. Garrett, S.M.; Whitaker, R.M.; Beeson, C.C.; Schnellmann, R.G. Agonism of the 5-hydroxytryptamine 1F receptor promotes mitochondrial biogenesis and recovery from acute kidney injury. *J. Pharmacol. Exp. Ther.* **2014**, *350*, 257–264. [[CrossRef](#)]
140. Simmons, E.C.; Scholpa, N.E.; Cleveland, K.H.; Schnellmann, R.G. 5-hydroxytryptamine 1F Receptor Agonist Induces Mitochondrial Biogenesis and Promotes Recovery from Spinal Cord Injury. *J. Pharmacol. Exp. Ther.* **2020**, *372*, 216–223. [[CrossRef](#)]
141. Newman-Tancredi, A.; Depoortère, R.Y.; Kleven, M.S.; Kołaczkowski, M.; Zimmer, L. Translating biased agonists from molecules to medications: Serotonin 5-HT<sub>1A</sub> receptor functional selectivity for CNS disorders. *Pharmacol. Ther.* **2022**, *229*, 107937. [[CrossRef](#)]

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