

Article

Effects of the Co-Overexpression of the *BCL* and *BDNF* Genes on the Gamma-Aminobutyric Acid-Ergic Differentiation of Wharton's-Jelly-Derived Mesenchymal Stem Cells

Paulina Borkowska *, Julia Morys, Aleksandra Zielinska and Jan Kowalski

Department of Medical Genetics, Medical University of Silesia, 41-200 Sosnowiec, Poland

* Correspondence: pborkowska@sum.edu.pl; Tel.: +48-32-364-1216

Abstract: One of the problems with using MSCs (mesenchymal stem cells) to treat different neurodegenerative diseases of the central nervous system is their low ability to spontaneously differentiate into functional neurons. The aim of this study was to investigate how the co-overexpression of the *BCL* and *BDNF* genes affects the ability of genetically modified MSCs to differentiate into GABA-ergic neurons. A co-overexpression of two genes was performed, one of which, *BCL*, was supposed to increase the resistance of the cells to the toxic agents in the brain environment. The second one, *BDNF*, was supposed to direct the cells onto the neuronal differentiation pathway. As a result, the co-overexpression of both *BCL2 + BDNF* and *BCLXL + BDNF* caused an increase in the *MAP2* gene expression level (a marker of the neuronal pathway) and the *SYP* gene that is associated with synaptogenesis. In both cases, approximately 18% of the genetically modified and then differentiated cells exhibited the presence of the *GAD* protein, which is characteristic of GABA-ergic neurons. Despite the presence of *GAD*, after both modifications, only the *BCL2* and *BDNF* co-overexpression correlated with the ability of the modified cells to release gamma-aminobutyric acid (GABA) after depolarization. Our study identified a novel model of genetically engineered MSCs that can be used as a tool to deliver the antiapoptotic proteins (*BCL*) and neurotrophic factor (*BDNF*) directly into the brain microenvironment. Additionally, in the investigated model, the genetically modified MSCs could easily differentiate into functional GABA-ergic neurons and, moreover, due to the secreted *BCL* and *BDNF*, promote endogenous neuronal growth and encourage synaptic connections between neurons.

Keywords: mesenchymal stem cells; co-overexpression; *BCL2*; *BCLXL*; *BDNF*; GABA-ergic neurons



Citation: Borkowska, P.; Morys, J.; Zielinska, A.; Kowalski, J. Effects of the Co-Overexpression of the *BCL* and *BDNF* Genes on the Gamma-Aminobutyric Acid-Ergic Differentiation of Wharton's-Jelly-Derived Mesenchymal Stem Cells. *Biomedicines* **2023**, *11*, 1751. <https://doi.org/10.3390/biomedicines11061751>

Academic Editors: Anita Lyssek-Borón and Benjamin Grabarek

Received: 29 May 2023

Revised: 15 June 2023

Accepted: 16 June 2023

Published: 18 June 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Genetically modified mesenchymal stem cells seem to be worth investigating in terms of their possible use in transplants in patients suffering from neurodegenerative diseases [1–3]. The basic features of cells, such as their multilineage potential [4,5], low tumorigenic potential and low level of MHC (major histocompatibility complex) protein expression [6], can be enriched by the features that are acquired from genetic modifications. The main problem inherent in using unmodified mesenchymal stem cells is their negligible survivability after the transplantation procedure [7–9]. Poor environmental conditions in the brain, such as insufficient nutrition and oxygen-free radical toxicity, limit the efficacy of BMSC therapy [10]. Another problem seems to be their poor ability to spontaneously differentiate, especially into the cells that originate from a germ layer other than themselves [11,12]. Thus, the hypothesis of the co-overexpression of genes was created, which has a chance to modify both of these features. To investigate this hypothesis, two genes were selected: one from the *BCL* (B-cell lymphoma) gene family, which was expected to increase cell survival and resistance to toxic factors, and the other—*BDNF* (brain-derived neurotrophic factor)—whose overexpression should increase their ability to differentiate

into the progenitors of nerve cells or mature cells or different types of neurons. Previous studies have shown that the co-overexpression of the *BCL2* and *BDNF* or *BCLXL* and *BDNF* genes in an analogous transduction system increases cell survival under toxic conditions in an in vitro culture [13,14] and increases their ability to differentiate into cells that exhibit a cholinergic and/or dopaminergic phenotype [13].

Because neurodegenerative diseases are rarely caused by a deficit of one type of cells in one brain structure, and it seems ideal to optimize the genetic modification of MSCs in such a way that it would be possible to transplant cells that have a wide potential for differentiation [8]. Demonstrating the influence of the co-overexpression of *BCL2* and *BDNF* as well as *BCLXL* and *BDNF* on the possibility of obtaining several types of nerve cells in a culture would support the fact that the proposed modification scheme could be tested in vivo in various types of diseases because, as a result of the modification, cells with increased resistance to toxic conditions [13,14], which are also progenitors of nerve cells, would be obtained. Performing the in vitro transduction and then transplantation of freshly modified cells would subject them to the microenvironment of the brain, which would define their final path of differentiation depending on what cells are needed [15]. How the co-overexpression of *BCL2* and *BDNF* or *BCLXL* and *BDNF* genes affects the ability to differentiate into GABA-ergic cells remains unexplored and, therefore, this analysis was the aim of this study.

2. Materials and Methods

2.1. Isolating, Culturing and Characterizing the WJ-MSC

The study was approved by the Bioethical Committee of the Medical University of Silesia in Katowice (Resolution No. KNW/0022/KB/195/14). The participants (mothers) were informed of the procedure and gave their written consent for the use of the umbilical cords.

The WJ-MSCs (Wharton-Jelly-derived mesenchymal stem cells) were isolated from human umbilical cords, which were diced into fragments and placed on culture dishes containing a Mesenchymal Stem Cell Expansion Medium (1X) (#SCM015; Sigma Aldrich®, Merck®, St. Louis, MO, USA) that had been supplemented with a 1% Antibiotic Antimycotic Solution (#P06-07300; PAN-Biotech™, Am Gewerbepark, Aidenbach, Germany). A seven-day expansion was performed in a humidified incubator at 37 °C with 5% CO₂. During the process, the WJ-MSCs migrated from the tissue fragments to the bottom of the culture dishes. After seven days, the tissue fragments were removed. The WJ-MSCs were passaged for the first time and put into the culture flasks. The cells were treated with 0.05% trypsin with 0.02% EDTA in phosphate-buffered saline (PBS) without calcium and magnesium (#PCS-999-003; ATCC®, University Blvd, Manassas, VA, USA) for three min. in order to form single cells. After that, to perform a rapid inactivation, a trypsin neutralizing solution (5% FBS (fetal bovine serum) in phosphate-buffered saline without calcium and magnesium) (#PCS-999-004; ATCC®, University Blvd, Manassas, VA, USA) was used. The volume of trypsin and trypsin inactivator was used in a 1:1 ratio. During the entire experiment or until banking, the cells were cultured in a mesenchymal stem cell basal medium (#PCS-500-030; ATCC®, University Blvd, Manassas, VA, USA) that had been supplemented with a mesenchymal stem cell growth kit for adipose- and umbilical-derived MSC-low serum (#PCS-500-040; ATCC®, University Blvd, Manassas, VA, USA). The final concentration for each component in a complete mesenchymal stem cell growth medium was as follows: rh FGF basic: 5 ng/mL; rh FGF acidic: 5 ng/mL; rh EGF: 5 ng/mL; fetal bovine serum: 2% and L-Alanyl-L-Glutamine: 2.4 mM. Then, the cells at an 80% confluence were passaged and cryopreserved to be left as a WJ-MSC bank for further use.

The homogeneity of the WJ-MSCs was quantified using flow cytometry; the presence of CD73, CD90, CD34, CD11b, CD19, CD45 and HLA-DR (human leukocyte antigen—DR isotype) was also determined. The samples were prepared by trypsinizing the cells with 0.05% trypsin/EDTA (#P10-029500; PAN-Biotech™, Am Gewerbepark, Aidenbach, Germany) and suspending them in phosphate-buffered saline (DPBS) (#P04-36500; PAN-

Biotech™, Am Gewerbepark, Aid-enbach, Germany). The suspensions prepared in this way were then analyzed via flow cytometry (BD FACSAria II, Qume Dr., San Jose, CA, USA; BD FACSDiva Software V6.1.2), seeking the areal markers presented by the WJ-MSCs [16]. The WJ-MSCs were also characterized based on their differentiation capacity toward adipocytes and osteocytes. For adipogenic and osteogenic differentiation, the following conditions were provided: a culture of WJ-MSCs in StemMACS AdipoDiff Media (#130-091-677; Miltenyi Biotec© Teterow, Germany) for over 21 days and detection with the use of Oil Red O (#O9755-256; Sigma Aldrich®, Merck®, St. Louis, MO, USA) for adipocytes, and a cell culture in StemMACS OsteoDiff Media (#130-091-678; Miltenyi Biotec© Teterow, Germany) for the same time period and detection with the use of alkaline phosphatase using a 5-bromo-4-chloro-3-indolyl phosphate/Nitro blue tetrazolium (BCIP/NBT) substrate (#B5655-5TAB; Sigma Aldrich®, Merck®, St. Louis, MO, USA) for osteocytes. All of the experimental measures taken with the mentioned reagents were conducted according to the instructions provided by the manufacturer. The control probe was not treated with any additional factors and consisted of the WJ-MSCs cultured in DMEM/F12 medium (#P04-41250; PAN-Biotech™, Am Gewerbepark, Aidenbach, Germany) supplemented with 15% FBS (#P30-8500; PAN-Biotech™, Am Gewerbepark, Aidenbach, Germany) and 1% Antibiotic Solution [16].

2.2. Lentiviral Transduction and Neuronal Differentiation

The lentivirus production protocol started with culturing the HEK 293T cells in 25 cm³ culture flasks in a proper medium that consisted of DMEM (#P04-05550; PAN-Biotech™, Am Gewerbepark, Aidenbach, Germany) medium supplemented with 10% FBS (#P04-03590; PAN-Biotech™, Am Gewerbepark, Aidenbach, Germany) and 1% Antibiotic Solution (#P06-07300; PAN-Biotech™, Am Gewerbepark, Aidenbach, Germany) at 37 °C in 5% CO₂ conditions. After achieving the 80% confluence in culture flasks, the cells were trypsinized and transferred onto 10 cm Ø Petri dishes for culture under humidified conditions, as mentioned previously. Subsequently, the 20 µg of the respective plasmid (Figure 1) plus 6 µg of pMD2.G (pMD2.G from Didier Trono (Addgene plasmid #12259; <http://n2t.net/addgene:12259>, accessed on 28 May 2023; RRID:Addgene_12259) and 15 µg psPAX (psPAX2 from Didier Trono (Addgene plasmid #12260; <http://n2t.net/addgene:12260>, accessed on 28 May 2023; RRID:Addgene_12260) plasmids were added. The supernatants containing lentiviral vectors were collected from each Petri dish with a sterile Pasteur pipette, infiltrated with the use of Corning® 150 mL Vacuum Filter/Storage Bottle System (#431153; Corning®, Glendale, AZ, USA) and diluted using the Lenti-X Concentrator (#631231; TaKaRa Bio©; Kyoto, Japan). Centrifuged lentiviruses were aliquoted and stored at −70 °C. These stocks were optimized on MSC cultures by performing the transduction and checking the amount of positively transduced cells with the use of flow cytometry. The coding sequences of *BCL2* and *BCLXL* were cloned to LeGO-iG2 (Figure 1a), and the coding sequence of *BDNF* was cloned to LeGO-iT2 backbones (Figure 1b). As a result, the plasmids were obtained as follows: LeGO-iG2-Bcl-2 (Figure 1c), LeGO-iG2-Bcl-XL (Figure 1d) and LeGO-iT2-BDNF (Figure 1e). Boris Fehse kindly provided the LeGO-iG2 (Addgene plasmid # 27341; <http://n2t.net/addgene:27341>, accessed on 28 May 2023; RRID:Addgene_27341) and LeGO-iT2 (Addgene plasmid # 27343; <http://n2t.net/addgene:27343>, accessed on 28 May 2023; RRID:Addgene_27343) [17].

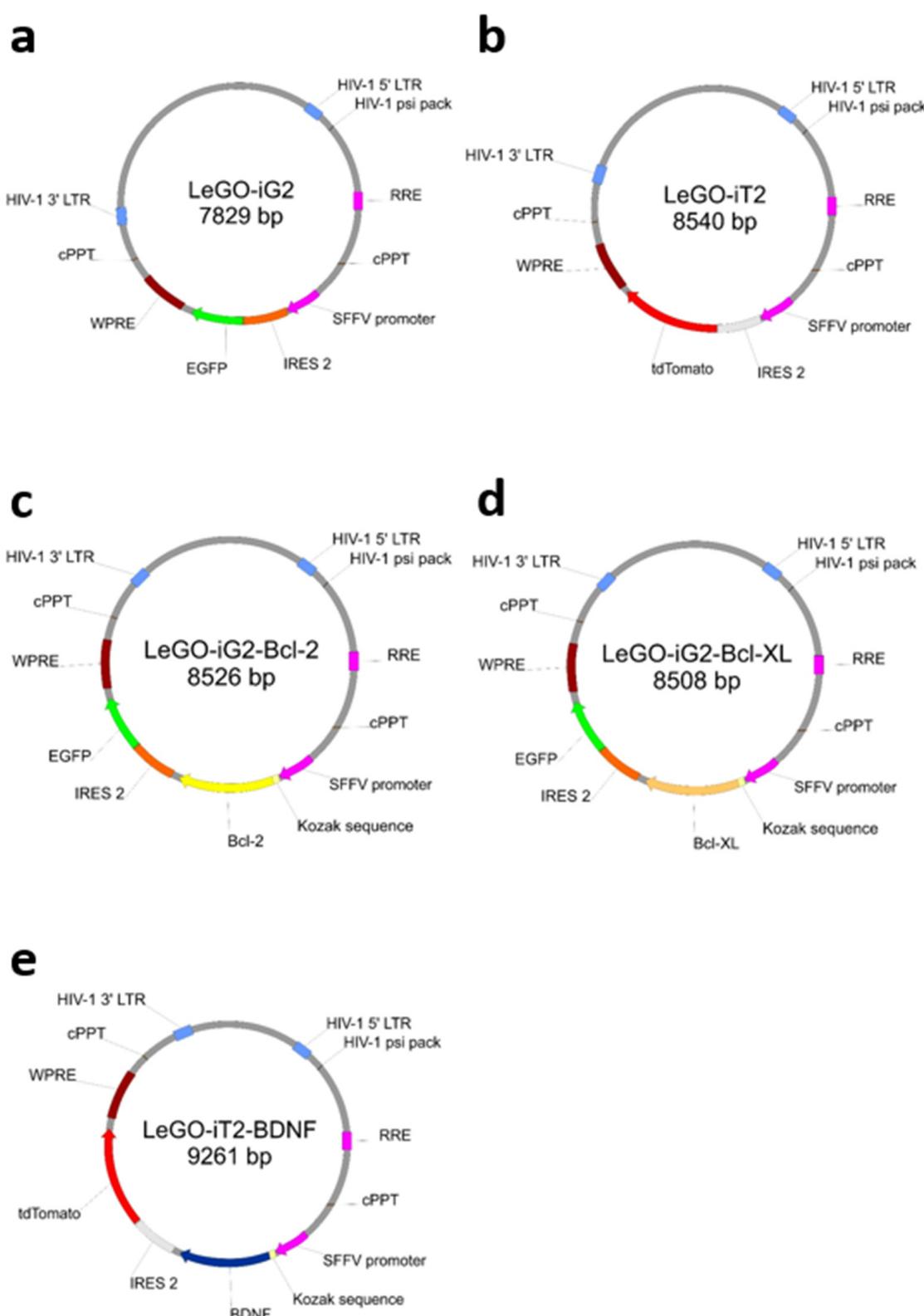


Figure 1. The simplified plasmid maps that were used to prepare the lentivirus. The lentiviral backbone plasmids contained the coding sequence of green fluorescence protein (*EGFP*) (a) or coding sequence of red fluorescence protein (*tdTomato*) (b). Into backbone “a”, coding sequence of *BCL2* gene (c) or *BCLXL* gene (d), and into backbone “b”, coding sequence of *BDNF* gene (e), were cloned to overproduced *BCL2*, *BCLXL* or *BDNF* proteins.

The day before the transduction, 100,000 WJ-MSCs per 35 mm Ø dish were plated in order to achieve a 70–80% confluence immediately prior to the transduction. An 18 h incubation of the preconfluent cells with an optimal dilution of a virus in an Opti-MEM I Reduced Serum Medium (#31985070; Gibco®, Thermo Fischer Scientific®, Grand Island, NE, USA) in the presence of 5 µg/mL polybrene (without any antibiotics) was followed by a 12-day neuronal differentiation with a medium containing Neurobasal PLUS (#A3582901; Gibco®, Thermo Fischer Scientific®, Grand Island, NE, USA) and a B27 PLUS supplement (#A3582801; Gibco®, Thermo Fischer Scientific®, Grand Island, NE, USA).

2.3. RT-qPCR

The differentiated cells were used to extract the total cellular RNA using NucleoZOL (#740404.200; Macherey-Nagel GmbH & Co. KG, Richterstr., Berlin, Germany) according to the manufacturer's protocol. Spectrophotometry was used to determine the quality and concentration of the RNA. The primers that were used for the selected genes were purchased from Sigma Aldrich (#KiCqStart SYBR Green Primers; Sigma Aldrich®, Merck®, St. Louis, MO, USA). *RPS17* was used as the reference gene. The sequences of the primers that were used were as follows: *RPS17* (P_F 5' CCATTATCCCCAGCAAAAG 3'; P_R 5' GAGACCTCAGGAACATAATTG 3'; Primer Pair ID H_RPS17_1), *SYP* (P_F 5' CCTTCGGTATTGTTCAAAG 3'; P_R 5' TTTGACTAGGTGGTTAAGGAG 3'; Primer Pair ID H_SYP_1), *GAD* (P_F 5' GGTTGGATATGGTGGATTAG 3'; P_R 5' GGAGCAATTTCATAG-GTGAAC 3'; Primer Pair ID H_GAD_2), *BAX* (P_F 5' AACTGGACAGTAACATGGAG 3'; P_R 5' TTGCTGGCAAAGTAGAAAAG 3'; *MAP2* (P_F 5' GAAGATTACTTACAGCCTCG 3'; P_R 5' GGTAAAGTTTACTTGTCTCTGG 3'; Primer Pair ID H_MAP2_2). A GoTaq 1-Step RT-qPCR System (#A6020; Promega®, Woods Hollow Rd., Madison, WI, USA) was used to perform the one-step RT-qPCR in triplicate. A 10 µL reaction with 15 ng of total RNA and a 0.2 µM final primer concentration for each forward and reverse primer was used to perform the RT-qPCR in a C1000 Touch Thermal Cycler equipped with a CFX96 Real-Time System (Bio-Rad CFX Manager, Alfred Nobel Dr., Hercules, CA, USA; Software Version 3.1). The RT-qPCR procedure comprised a 15 min RT reaction at 37 °C, a 10 min PCR activation at 95 °C and then 40 cycles of a 10 s denaturation at 95 °C, a 30 s annealing at 60 °C at the melting temperature of the lowest primers' pair and a 30 s PCR extension at 72 °C. Lastly, a melting curve analysis was performed in order to confirm the RT-qPCR specificity. Each run of the RT-qPCR included negative controls that had no total RNA. A Bio-Rad CFX96 Real-Time System (Bio-Rad CFX Manager, Alfred Nobel Dr., Hercules, CA, USA; Software Version 3.1) was used to automatically determine the Ct value.

2.4. Evaluating the GAD67 Protein Expression

Indirect labeling with a specific anti-GAD67 antibody was used to identify the GAD67 protein-positive cells. After the differentiation, the cells were washed twice in PBS and fixed in PBS with 4% paraformaldehyde (#158127; Sigma Aldrich®, Merck®, St. Louis, MO, USA). The cells were then washed three times in PBS with 1% BSA (#A9418; Sigma Aldrich®, Merck®, St. Louis, MO, USA), permeabilized with 0.3% Triton X-100 (#T8787; Sigma Aldrich®, Merck®, St. Louis, MO, USA)/1% BSA/PBS with 10% normal goat serum (#005-000-121; Jackson ImmunoResearch®, St. Thomas' Place, Ely, UK) for 45 min and incubated with rabbit anti-GAD67 antibody (dilution 1:250; # GTX101881 GeneTex®, Alton Pkwy., Irvine, CA, USA) in 1% BSA/PBS with 10% goat serum overnight at 4 °C. The cells were washed three times in PBS with 1% BSA and incubated with goat anti-rabbit IgG secondary antibody (DY405) (dilution 1:1000; # LS-C355899; LifeSpan BioSciences®, Shaker Rd., Suites Shirley, MA, USA) in 1% BSA/PBS for 1 h in the dark at room temperature. In order to determine any non-specific binding, similar staining was performed using a rabbit IgG isotype control (dilution 1:625; # bs-0295P; Bioss Antibodies®, Tradecenter Ste., Woburn, MA, USA). Flow cytometry (BD FACSAria II, Qume Dr., San Jose, CA, USA; BD FACSDiva Software V6.1.2) and fluorescence microscope (BX60, Olympus®, Shinjuku, Tokyo, Japan; cellSense Standard Software V3.2) were used to analyze the staining.

2.5. Gamma-Aminobutyric Acid Release Analysis

The procedure used to perform the assay was previously published [13]. The neurotransmitter levels were obtained and quantified using an ELISA (enzyme-linked immunosorbent assay) kit (#CEA900Ge; Cloud-Clone Corp.©, W. Fernhurst Dr., Katy, TX, USA) according to the manufacturer's instructions. An ELX 800 IU automated Microplate Reader (Bio-Tek Instruments; Gene 5 Software V 3.02) was used to read the absorbance at 450 nm. A quadratic log-log curve fit was used to analyze the results. The supernatants (low and high K⁺ = before and after the GABA release) were analyzed for each well. In order to minimize the effect of the differences in the number of cells in the wells, the amount of GABA that had been produced per well was calculated based on the difference between the amount of GABA in a high K⁺ supernatant and the amount of GABA in a low K⁺ supernatant.

2.6. Statistical Analysis

All of the collected data were compiled using Microsoft® Excel 2023 V16.70 spreadsheets (Microsoft Corporation, Redmond, WA, USA) and statistically analyzed using GraphPad® Prism 8.0 software (Graphpad Software Inc., San Diego, CA, USA). First, a column analysis (Shapiro–Wilk normality test) was performed to determine whether there was a Gaussian distribution of the data. Next, the data were analyzed using the ordinary one-way ANOVA test followed by a Tukey multiple comparison test. For all of the tests, $p < 0.05$ was considered to be statistically significant. Reproducible results were obtained from six or more samples. The data are presented as the mean \pm SD. The qRT-PCR results are presented as the fold change ($2^{-\Delta\Delta CT}$).

3. Results

3.1. The Co-Overexpression of BCL and BDNF Affects the Expression Level of the Neuronal Pathway Genes

The investigated co-overexpression of BCL and BDNF resulted in an increase in the gene expression level of the neuronal pathway genes in the culture of genetically modified MSCs. It was observed that the co-overexpression of BCL2 and BDNF caused an increase in the expression of MAP2 (Figure 2a, iG2 + iT2 vs. BCL2 + BDNF) and GAD (Figure 2c, iG2 + iT2 vs. BCL2 + BDNF: $p < 0.01$) and a ten-fold increase in the SYP gene level (Figure 2b, iG2 + iT2 vs. BCL2 + BDNF: $p < 0.01$). At the same time, it was observed that the co-overexpression of BCL2 and BDNF changed the BAX gene expression level in accordance with the control group (Figure 2d, Control vs. BCL2 + BDNF: $p < 0.001$). In turn, because of the co-overexpression of BCLXL and BDNF, the differences between the investigated groups were observed in accordance with GAD and BAX expression levels (Figure 2c,d; Control vs. BCLXL + BDNF: $p < 0.05$).

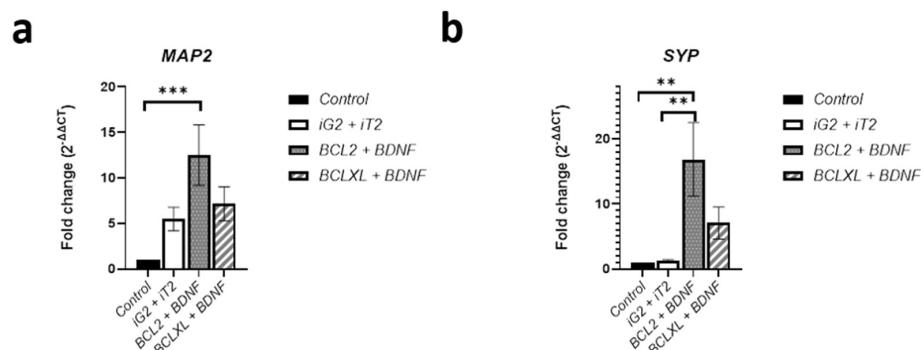


Figure 2. Cont.

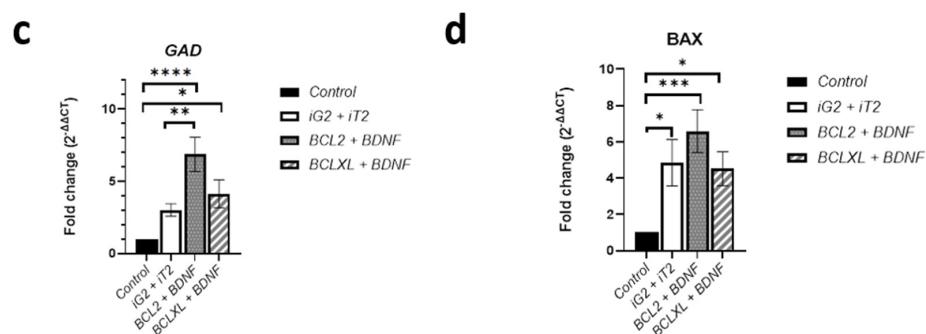


Figure 2. The qRT-PCR analysis is presented as the fold change ($2^{-\Delta\Delta CT}$) in the level of MAP2 (a); SYP (b); GAD (c) and BAX (d) expression, which was normalized to the RPS17 reference gene. The points represent the mean value \pm SD ($n = 9$; three independent experiments). Statistically significant * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; **** $p < 0.0001$; one-way ANOVA test followed by a post hoc Tukey's test.

3.2. The Co-Overexpression of BCL and BDNF Affects the GAD67 Protein Expression Level

The investigated co-overexpression of BCL and BDNF resulted in an increase in the GAD67 protein expression level in the culture of genetically modified MSC. It was observed that both the co-overexpression of BCL2 and BDNF (Figure 3d vs. Figure 3f; Figure 4b vs. Figures 4c and 5, iG2 + iT2 vs. BCL2 + BDNF: $p < 0.01$) and the co-overexpression of BCLXL and BDNF (Figure 3d vs. Figure 3h; Figure 4b vs. Figures 4d and 5, iG2 + iT2 vs. BCLXL + BDNF: $p < 0.001$) increased the expression of the GAD67 protein.

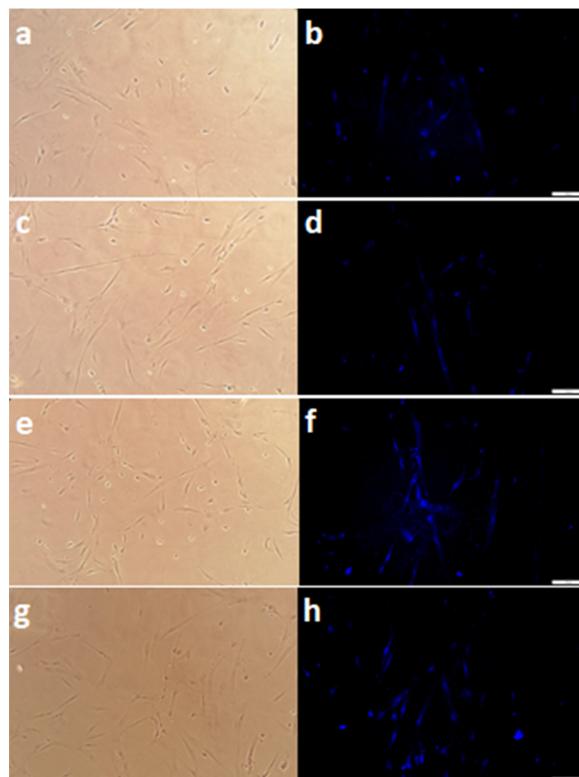


Figure 3. GABA-ergic neurons after co-overexpression and 12 days in differentiation medium. The GAD67 positive cells (blue) had a neuron-typical spindle shape. (a,b) Control. (c,d) iG2 and iT2 (Empty vectors) co-overexpression (transduction with vectors presented in Figure 1a,b). (e,f) BCL2 and BDNF co-overexpression (transduction with vectors presented in Figure 1c,e). (g,h) the BCLXL and BDNF co-overexpression (transduction with vectors presented in Figure 1d,e).

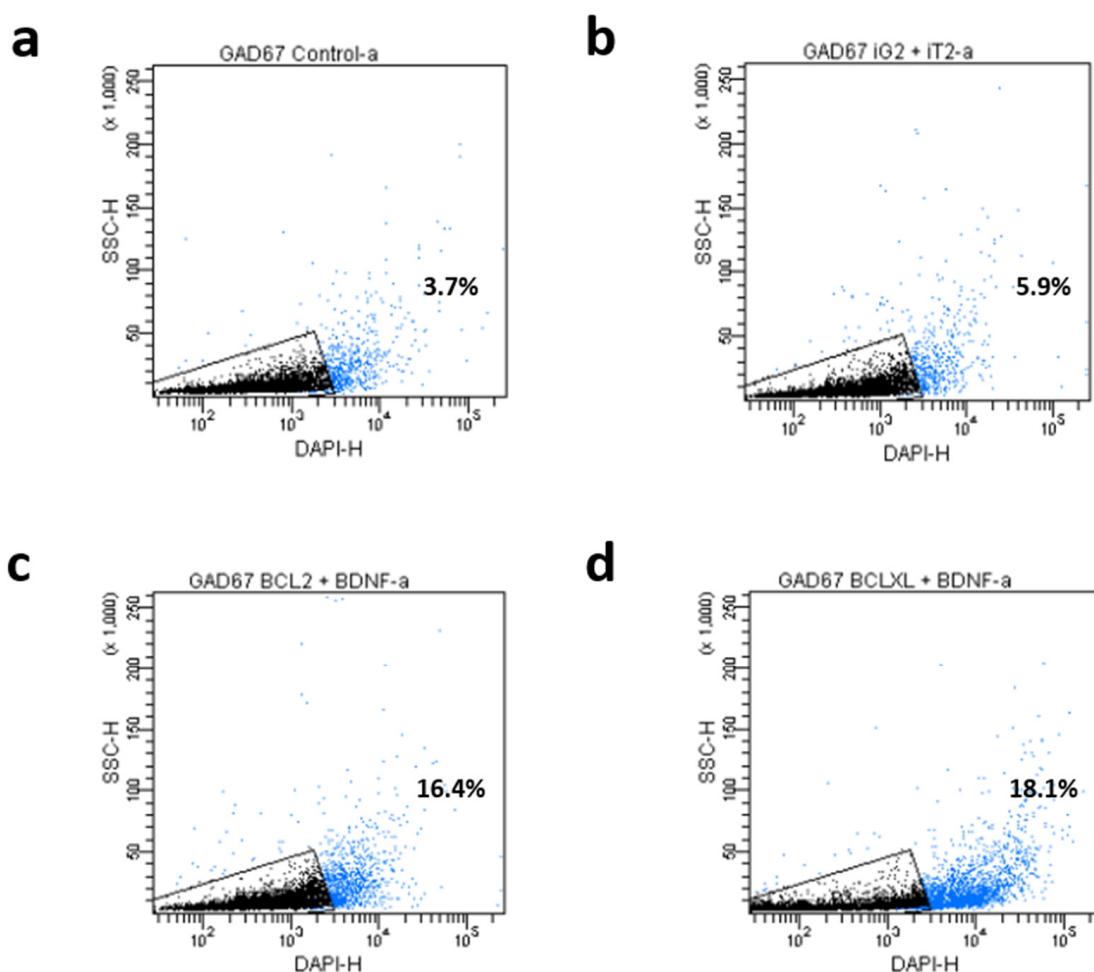


Figure 4. Flow cytometric analysis for the expression of GABA-ergic neural-specific proteins (GAD67) after 12 days of differentiation. The percentage values represent the mean value ($n = 6$; three independent experiments); (a) Control; (b) iG2 and iT2 (Empty vectors) co-overexpression (transduction with vectors presented in Figure 1a,b); (c) BCL2 and BDNF co-overexpression (transduction with vectors presented in Figure 1c,e); (d) BCLXL and BDNF co-overexpression (transduction with vectors presented in Figure 1d,e). A total of 10,000 cells were counted and analyzed.

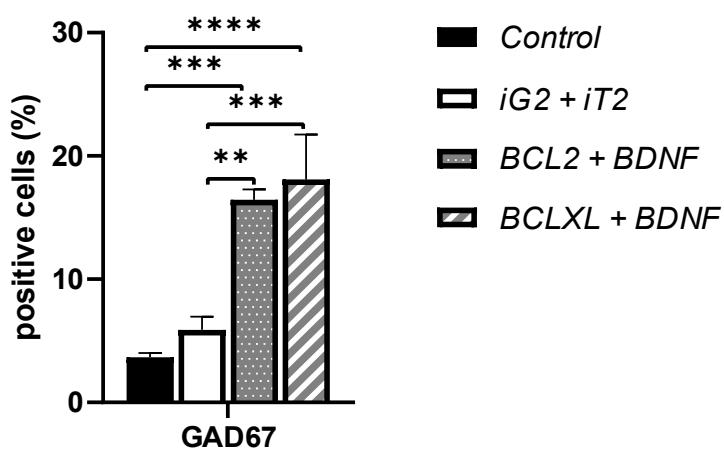


Figure 5. The number of GAD67 positive cells. The calculations were based on both flow cytometry analysis and immunofluorescence. A total of 10,000 cells were counted and analyzed. The points represent the mean value \pm SD ($n = 6$; three independent experiments). Statistically significant
 $^{**} p < 0.01$; $^{***} p < 0.01$; $^{****} p < 0.0001$ one-way ANOVA test followed by a post hoc Tukey's test.

In the cultures with an increased expression of the BCL2 and BDNF, 16% of the cells, and in cultures with an increased expression of the BCLXL and BDNF, 18% of the cells expressed the GAD67 protein that is characteristic of the GABA-ergic neurons, while in the control samples, only 4%, and in samples that had been transduced with empty vectors, only 6% of the cells exhibited the presence of GAD67. This result was confirmed using two independent methods: flow cytometry and immunofluorescence. A total of 10,000 cells were counted and analyzed.

3.3. The Co-Overexpression of BCL and BDNF Affects the Release of Gamma-Aminobutyric Acid

It was demonstrated that only the overexpression of the BCL2 and BDNF increased the release of gamma-aminobutyric acid as a result of depolarization. In the samples with an overexpression of the BCL2 and BDNF, after the differentiation process, the basic level of the release of GABA within 30 min. was $3.64 \text{ pg/mL} \pm 0.76$. As a result, a 30 min. depolarization increased the level of the GABA release to $3.87 \text{ pg/mL} \pm 0.15$. The amount of neurotransmitters that were produced increased by 0.23 pg/mL . For all of the other groups, including the cells with an overexpression of the BCLXL and BDNF, there was no increase in the GABA secretion as a result of depolarization.

4. Discussion

Huntington's disease (HD) is an inherited neurodegenerative genetic disorder that leads to the onset of motor, neuropsychiatric and cognitive disturbances. HD is characterized by the loss of GABA-ergic neurons. In order to use genetically modified MSCs as a cell therapy in HD, it seemed to be necessary to investigate the effect of the gene co-overexpression on the ability of MSCs to differentiate into GABA-ergic neurons. In previous studies, the effect of the co-overexpression of the *BCL2* and *BDNF* genes on the ability of MSC to differentiate into cholinergic neurons was demonstrated [13]. It was also shown that the co-overexpression of both *BCL2 + BDNF* and *BCLXL + BDNF* increases the ability of cells to survive under toxic conditions [13,14]. This confirmed that the increased viability of genetically modified MSCs is a significant advantage in the context of using these cells to treat HD. The role of ferroptosis in the etiopathogenesis of neurodegenerative diseases, such as HD, Parkinson's disease and Alzheimer's disease, is well known [18]. There are reports that stress the role of oxidative damage in the etiology of degeneration and neuronal damage in patients with HD [19]. The increased survival rate of genetically modified MSCs, generated via the co-overexpression of *BCL2 + BDNF* and *BCLXL + BDNF*, increases the likelihood of transplant survival after a stroke procedure, such as transplantation. It is also important that the environment into which the transplant is introduced is a toxic environment with an increased amount of free radicals, with β -catenin in its phosphorylated form [20] and with factors that direct cells onto the ferroptosis path. Under such conditions, the demonstrated increased ability to survive under toxic conditions would be extremely desirable. Because it is important that the increased survival is not a permanent state not being subjected to control mechanisms, the increased level of the *BAX* gene expression that was demonstrated enabled us to conclude that the mechanisms that regulate cell death were not permanently silenced.

MAP2 is an early neuronal marker that is commonly used to indicate these cells' ability to differentiate into neurons. The generally higher expression of the mature neuronal marker *MAP2* compared with the glial markers at the mRNA and protein levels suggest an enhanced potential for MSC neuronal differentiation [21]. In our study, we observed that especially the co-overexpression of *BCL2* and *BDNF* increased the *MAP2* gene expression level. Other authors have proved that the induction of the neural markers GFAP, NF, MAP2, and O4 and a decrease in the expression of the immature neural markers β -tubulin III, Nestin and NG2, co-exists with an increase in the level of TrkB, which is a BDNF receptor and occurs during the neural differentiation [22]. Another study provided evidence that *MAP2* in culture was upregulated by the neurotrophic factor NT3 [23] whose role in the central nervous system is similar to the role of BDNF.

Forming a network with neighboring cells is absolutely crucial for neuron-differentiated cells. These morphological and immunocytochemical features have been demonstrated. One of many genes that are involved in synaptogenesis is the *SYP* gene, which codes synaptophysin protein. We observed that the co-overexpression of combined *BCL2* + *BDNF* enhanced the *SYP* gene expression level. A similar effect in which the *SYP* expression level was also increased, along with increasing *MAP2*, was previously observed by other authors [24]. Moreover, an increased amount of *BDNF* in the brain environment is directly connected with an increased level of tyrosine kinase receptor B (trkB)—a receptor for *BDNF*. Other authors investigated whether signaling through tyrosine kinase receptor B influences neuronal survival, differentiation and synaptogenesis [25]. The overexpression of the *BDNF* gene may explain the increase in the expression of not only the *SYP* and *MAP2* genes but also the increase in the expression exhibited by the genes characterizing certain types of mature neurons, which was demonstrated in previous experiments using the same type of genetic modification. *MAP2*-positive cells could express different mature-neuronal markers such as *CHATA* marker of cholinergic neurons; *TH*, a marker of dopaminergic neurons; or *TPH1*, a marker of serotonergic neurons. The confirmed, increased expression of *MAP2* and *SYP* provided the basis for investigating whether the co-overexpression of the *BCL* and *BDNF* affects the *GAD67* gene and its protein expression level, which is a marker of GABA-ergic neurons. We observed that the co-overexpression of *BCL* and *BDNF* increased the protein expression level. In one recent study, the authors administrated dental pulp stem cells intravenously in an HD rat model. They demonstrated the stem cells homed in the striatum, cortex and subventricular zone using specific markers for human cells. Thirty days after the administration, an immunohistochemistry quantity analysis revealed a significant increase in the amount of *BDNF*, which proves its important role in GABA-ergic differentiation [26]. Other authors identified the GABA-ergic synapse pathway as being one of the major targets of the MSC-activated genes [27]. The enhancement of the GABA synapses was impaired by treatment with a Trk/neurotrophin receptors blocker and by TrkB receptors, thereby suggesting the involvement of *BDNF* as a mediator of such effect [28].

When investigating a model of differentiation, the co-overexpression of *BCL2* and *BDNF* seems to be especially important because of the correlation of the GAD-positive cells with the release of gamma-aminobutyric acid as a result of depolarization. Another author investigated this phenomenon in several central nervous system areas and demonstrated that *BDNF* can modulate the GABA-ergic synapses by regulating transmitter release [29]. A facilitatory action of *BDNF* on GABA release was previously reported by Pezet et al. [30], who demonstrated that an acute application of *BDNF* caused an increase in the KCl-elicited GABA release in a supernatant that had been collected from the isolated spinal cord in adult rats.

Since we have established that MSCs differentiate into GABA-ergic neurons, the evaluated cell therapy might be a great chance for patients suffering from HD to receive a promising treatment. However, the other types of neurons that various researchers obtained remain an auspicious treatment strategy in other neurodegenerative disorders as well. What has been proved by Venkatesh and Sen [31] is that MSCs may also differentiate into dopaminergic neurons, which could be potentially used as a cell therapy in Parkinson's disease. Moreover, Ranjbaran et al. [32] conducted research in which it was stated that the MSCs may be a useful measure in treating anxiety regarding septic patients with encephalopathy by reducing the inflammation process and changing serotonergic neurotransmission. Furthermore, Shwartz et al. [33] proposed a unique stem cell depression treatment by differentiating MSCs into cells exhibiting increased levels of functional glutamate transporters. In this case, it was proved that this type of MSC induces glutamatergic neurotransmission in the hippocampus, which leads to a meaningful improvement in the therapy used for the mentioned disorder.

Our study identified a novel model of genetically engineered MSCs as a tool for delivering antiapoptotic proteins (*BCL*) and neurotrophic factor (*BDNF*) directly into the

brain microenvironment. Additionally, in the investigated model, the genetically modified MSCs could easily differentiate into functional GABA-ergic neurons and, therefore, due to the secreted *BCL* and *BDNF*, promote endogenous neuronal growth, encourage the formation of synaptic connections in damaged neurons, decrease apoptosis and, according to the literature, reduce the levels of free radicals [34] and regulate inflammation [24].

5. Conclusions

Based on the results of this study, it can be concluded that the co-overexpression of both *BCL* and *BDNF* can increase the ability of MSC to differentiate into GAD-positive cells and that the co-overexpression of *BCL2* and *BDNF* seems to be especially important as it correlates with the ability of the modified cells to release gamma-aminobutyric acid as a result of depolarization.

Author Contributions: Conceptualization, P.B. and J.K.; Methodology, P.B.; Formal analysis, J.M.; Investigation, P.B. and A.Z.; Data curation, P.B. and J.M.; Writing—original draft, P.B.; Writing—review & editing, J.M.; Funding acquisition, J.K. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by a grant from the Medical University of Silesia, Katowice, Poland (PCN-1-194/N/2/B).

Institutional Review Board Statement: The study was approved by the Bioethical Committee of the Medical University of Silesia in Katowice (Resolution No. KNW/0022/KB/195/14).

Informed Consent Statement: The participants (mothers) were informed of the procedure and gave their written consent to use the umbilical cords.

Data Availability Statement: The datasets that were used and/or analyzed during the presented study are available from the corresponding author upon reasonable request.

Conflicts of Interest: The authors declare that they have no conflict of interest.

References

1. Staff, N.P.; Jones, D.T.; Singer, W. Mesenchymal Stromal Cell Therapies for Neurodegenerative Diseases. *Mayo Clin. Proc.* **2019**, *5*, 892–905. [[CrossRef](#)]
2. Kim, J.; Lee, Y.; Lee, S.; Kim, K.; Song, M.; Lee, J. Mesenchymal Stem Cell Therapy and Alzheimer’s Disease: Current Status and Future Perspectives. *J. Alzheimers Dis.* **2020**, *1*, 1–14. [[CrossRef](#)] [[PubMed](#)]
3. Heris, R.M.; Shirvaliloo, M.; Abbaspour-Aghdam, S.; Hazrati, A.; Shariati, A.; Youshanlouei, H.R.; Niaragh, F.J.; Valizadeh, H.; Ahmadi, M. The potential use of mesenchymal stem cells and their exosomes in Parkinson’s disease treatment. *Stem. Cell Res. Ther.* **2022**, *1*, 371. [[CrossRef](#)] [[PubMed](#)]
4. Pittenger, M.F.; Mackay, A.M.; Beck, S.C.; Jaiswal, R.K.; Douglas, R.; Mosca, J.D.; Moorman, M.A.; Simonetti, D.W.; Craig, S.; Marshak, D.R. Multilineage potential of adult human mesenchymal stem cells. *Science* **1999**, *2*, 143–147. [[CrossRef](#)]
5. Bianco, P.; Riminucci, M.; Gronthos, S.; Robey, P.G. Bone marrow stromal stem cells: Nature, biology, and potential applications. *Stem. Cells* **2001**, *19*, 180–192. [[CrossRef](#)]
6. Jacobs, S.A.; Roobrouck, V.D.; Verfaillie, C.M.; Van Gool, S.W. Immunological characteristics of human mesenchymal stem cells and multipotent adult progenitor cells. *Immunol. Cell Biol.* **2013**, *91*, 32–39. [[CrossRef](#)]
7. Yasuhara, T.; Matsukawa, N.; Hara, K.; Maki, M.; Ali, M.M.; Yu, S.J.; Bae, E.; Yu, G.; Xu, L.; McGrogan, M.; et al. Notch-induced rat and human bone marrow stromal cell grafts reduce ischemic cell loss and ameliorate behavioral deficits in chronic stroke animals. *Stem. Cells Dev.* **2009**, *18*, 1501–1514. [[CrossRef](#)] [[PubMed](#)]
8. Shen, L.H.; Li, Y.; Chen, J.; Zacharek, A.; Gao, Q.; Kapke, A.; Lu, M.; Raginski, K.; Vanguri, P.; Smith, A.; et al. Therapeutic benefit of bone marrow stromal cells administered 1 month after stroke. *J. Cereb. Blood Flow Metab.* **2007**, *27*, 6–13. [[CrossRef](#)]
9. Zhao, L.R.; Duan, W.M.; Reyes, M.; Keene, C.D.; Verfaillie, C.M.; Low, W.C. Human bone marrow stem cells exhibit neural phenotypes and ameliorate neurological deficits after grafting into the ischemic brain of rats. *Exp. Neurol.* **2002**, *174*, 11–20. [[CrossRef](#)]
10. Shen, L.H.; Ye, M.; Ding, X.S.; Han, Q.; Zhang, C.; Liu, X.F.; Huang, H.; Wu, E.B.; Huang, H.F.; Gu, X.S. Protective effects of MCI-186 on transplantation of bone marrow stromal cells in rat ischemic stroke model. *Neuroscience* **2012**, *223*, 315–324. [[CrossRef](#)]
11. Li, Y.; Chopp, M.; Chen, J.; Wang, L.; Gautam, S.C.; Xu, Y.X.; Zhang, Z. Intrastratal transplantation of bone marrow non-hematopoietic cells improves functional recovery after stroke in adult mice. *J. Cereb. Blood Flow Metab.* **2000**, *20*, 1311–1319. [[CrossRef](#)]

12. Shichinohe, H.; Kuroda, S.; Lee, J.B.; Nishimura, G.; Yano, S.; Seki, T.; Ikeda, J.; Tamura, M.; Iwasaki, Y. In vivo tracking of bone marrow stromal cells transplanted into mice cerebral infarct by fluorescence optical imaging. *Brain Res. Brain Res. Protoc.* **2004**, *13*, 166–175. [[CrossRef](#)]
13. Borkowska, P.; Zielińska, A.; Paul-Samojedny, M.; Stojko, R.; Kowalski, J. Synergistic Effect of the Long-Term Overexpression of Bcl-2 and BDNF Lentiviral in Cell Protecting against Death and Generating TH Positive and CHAT Positive Cells from MSC. *Int. J. Mol. Sci.* **2021**, *13*, 7086. [[CrossRef](#)] [[PubMed](#)]
14. Borkowska, P.; Morys, J.; Zielińska, A.; Sadlocha, M.; Kowalski, J. Survival and Neurogenesis-Promoting Effects of the Co-Overexpression of BCLXL and BDNF Genes on Wharton’s Jelly-Derived Mesenchymal Stem Cells. *Life* **2022**, *9*, 1406. [[CrossRef](#)] [[PubMed](#)]
15. Hayase, M.; Kitada, M.; Wakao, S.; Itokazu, Y.; Nozaki, K.; Hashimoto, N.; Takagi, Y.; Dezawa, M. Committed Neural Progenitor Cells Derived from Genetically Modified Bone Marrow Stromal Cells Ameliorate Deficits in a Rat Model of Stroke. *J. Cereb. Blood Flow Metab.* **2009**, *8*, 1409–1420. [[CrossRef](#)] [[PubMed](#)]
16. Borkowska, P.; Zielińska, A.; Paul-Samojedny, M.; Stojko, R.; Kowalski, J. Evaluation of reference genes for quantitative real-time PCR in Wharton’s Jelly-derived mesenchymal stem cells after lentiviral transduction and differentiation. *Mol. Biol. Rep.* **2020**, *47*, 1107–1115. [[CrossRef](#)] [[PubMed](#)]
17. Weber, K.; Bartsch, U.; Stocking, C.; Fehse, B. A multicolor panel of novel lentiviral “gene ontology” (LeGO) vectors for functional gene analysis. *Mol. Ther.* **2008**, *16*, 698–706. [[CrossRef](#)]
18. Reichert, C.O.; de Freitas, F.A.; Sampaio-Silva, J.; Rokita-Rosa, L.; De Lima Barros, P.; Levy, D.; Bydlowski, S.P. Ferroptosis Mechanisms Involved in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2020**, *21*, 8765. [[CrossRef](#)] [[PubMed](#)]
19. Browne, S.E.; Beal, M.F. Oxidative Damage in Huntington’s Disease Pathogenesis. *Antioxid. Redox. Signal.* **2006**, *8*, 11–12. [[CrossRef](#)] [[PubMed](#)]
20. Godin, J.D.; Poizat, G.; Hickey, M.A.; Maschat, F. Mutant huntingtin-impaired degradation of β -catenin causes neurotoxicity in Huntington’s disease. *EMBO J.* **2010**, *29*, 2433–2445. [[CrossRef](#)]
21. Navone, S.E.; Marfia, G.; Canzi, L.; Ciusani, E.; Canazza, A.; Visintini, S.; Campanella, R.; Parati, E.A. Expression of neural and neurotrophic markers in nucleus pulposus cells isolated from degenerated intervertebral disc. *J. Orthop. Res.* **2012**, *9*, 1470–1477. [[CrossRef](#)]
22. San José, I.; Vázquez, E.; García-Atares, N.; Huerta, J.J.; Vega, J.A.; Represa, J. Differential expression of microtubule associated protein MAP-2 in developing cochleovestibular neurons and its modulation by neurotrophin. *Int. J. Dev. Biol.* **1997**, *3*, 509–519.
23. Valeri, A.; Chiricosta, L.; Gugliandolo, A.; Pollastro, F.; Salamone, S.; Zingale, V.D.; Silvestro, S.; Mazzon, E. Cannabinol and NSC-34 Transcriptomic Analysis: Is the Dose Who Makes Neuronal Differentiation? *Int. J. Mol. Sci.* **2022**, *7*, 7541. [[CrossRef](#)] [[PubMed](#)]
24. Romanczyk, T.B.; Weickert, C.S.; Webster, M.J.; Herman, M.M.; Akil, M.; Kleinman, E.J. Alterations in trkB mRNA in the human prefrontal cortex throughout the lifespan. *Eur. J. Neurosci.* **2002**, *2*, 269–280. [[CrossRef](#)]
25. Wenceslau, C.V.; de Souza, D.M.; Mambelli-Lisboa, N.C.; Ynoue, L.H.; Araldi, R.P.; da Silva, J.M.; Pagani, E.; Haddad, M.S.; Kerkis, I. Restoration of BDNF, DARPP32, and D2R Expression Following Intravenous Infusion of Human Immature Dental Pulp Stem Cells in Huntington’s Disease 3-NP Rat Model. *Cells* **2022**, *10*, 1664. [[CrossRef](#)] [[PubMed](#)]
26. Cao, T.; Chen, H.; Huang, W.; Xu, S.; Liu, P.; Zou, W.; Pang, M.; Xu, Y.; Bai, X.; Liu, B.; et al. hUC-MSC-mediated recovery of subacute spinal cord injury through enhancing the pivotal subunits $\beta 3$ and $\gamma 2$ of the GABA A receptor. *Theranostics* **2022**, *7*, 3057–3078. [[CrossRef](#)]
27. Mauri, M.; Lentini, D.; Gravati, M.; Foudah, D.; Biella, G.; Costa, B.; Toselli, M.; Parenti, M.; Coco, S. Mesenchymal stem cells enhance GABAergic transmission in co-cultured hippocampal neurons. *Mol. Cell Neurosci.* **2012**, *4*, 395–405. [[CrossRef](#)]
28. Bardoni, R.; Ghirri, A.; Salio, C.; Prandini, M.; Merighi, A. BDNF-mediated modulation of GABA and glycine release in dorsal horn lamina II from postnatal rats. *Dev. Neurobiol.* **2007**, *7*, 960–975. [[CrossRef](#)] [[PubMed](#)]
29. Pezet, S.; Cunningham, J.; Patel, J.; Grist, J.; Gavazzi, I.; Lever, I.J.; Malcangio, M. BDNF modulates sensory neuronsynaptic activity by a facilitation of GABA transmission in the dorsal horn. *Mol. Cell Neurosci.* **2002**, *21*, 51–62. [[CrossRef](#)] [[PubMed](#)]
30. Meyerrose, T.; Olson, S.; Pontow, S.; Kalomoiris, S.; Jung, Y.; Annett, G.; Bauer, G.; Nolta, J.A. Mesenchymal stem cells for the sustained in vivo delivery of bioactive factors. *Adv. Drug Deliv. Rev.* **2010**, *62*, 1167–1174. [[CrossRef](#)]
31. Venkatesh, K.; Sen, D. Mesenchymal Stem Cells as a Source of Dopaminergic Neurons: A Potential Cell Based Therapy for Parkinson’s Disease. *Curr. Stem. Cell Res. Ther.* **2017**, *12*, 326–347. [[CrossRef](#)] [[PubMed](#)]
32. Ranjbaran, M.; Kianian, F.; Kadkhodaei, M.; Seifi, B.; Ashabi, G.; Akhondzadeh, F.; Adelipour, M.; Izad, M.; Abdolmohammadi, K. Mesenchymal stem cells and their conditioned medium as potential therapeutic strategies in managing comorbid anxiety in rat sepsis induced by cecal ligation and puncture. *Iran J. Basic Med. Sci.* **2022**, *25*, 690–697. [[PubMed](#)]

33. Shwartz, A.; Betzer, O.; Kronfeld, N.; Kazimirsky, G.; Cazacu, S.; Finniss, S.; Lee, H.K.; Motiei, M.; Dagan, S.Y.; Popovtzer, R.; et al. Therapeutic Effect of Astroglia-like Mesenchymal Stem Cells Expressing Glutamate Transporter in a Genetic Rat Model of Depression. *Theranostics* **2017**, *7*, 2690–2703. [[CrossRef](#)] [[PubMed](#)]
34. Urrutia, M.; Fernández, S.; González, M.; Vilches, R.; Rojas, P.; Vásquez, M.; Kurte, M.; Vega-Letter, A.M.; Carrión, F.; Figueroa, F.; et al. Overexpression of Glutamate Decarboxylase in Mesenchymal Stem Cells Enhances Their Immunosuppressive Properties and Increases GABA and Nitric Oxide Levels. *PLoS ONE* **2016**, *11*, e0163735. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.