

Figure S1. Representative confocal image of neurons in WT mouse hippocampus stained with DiIC₁₈; scale bar 25 μ m.

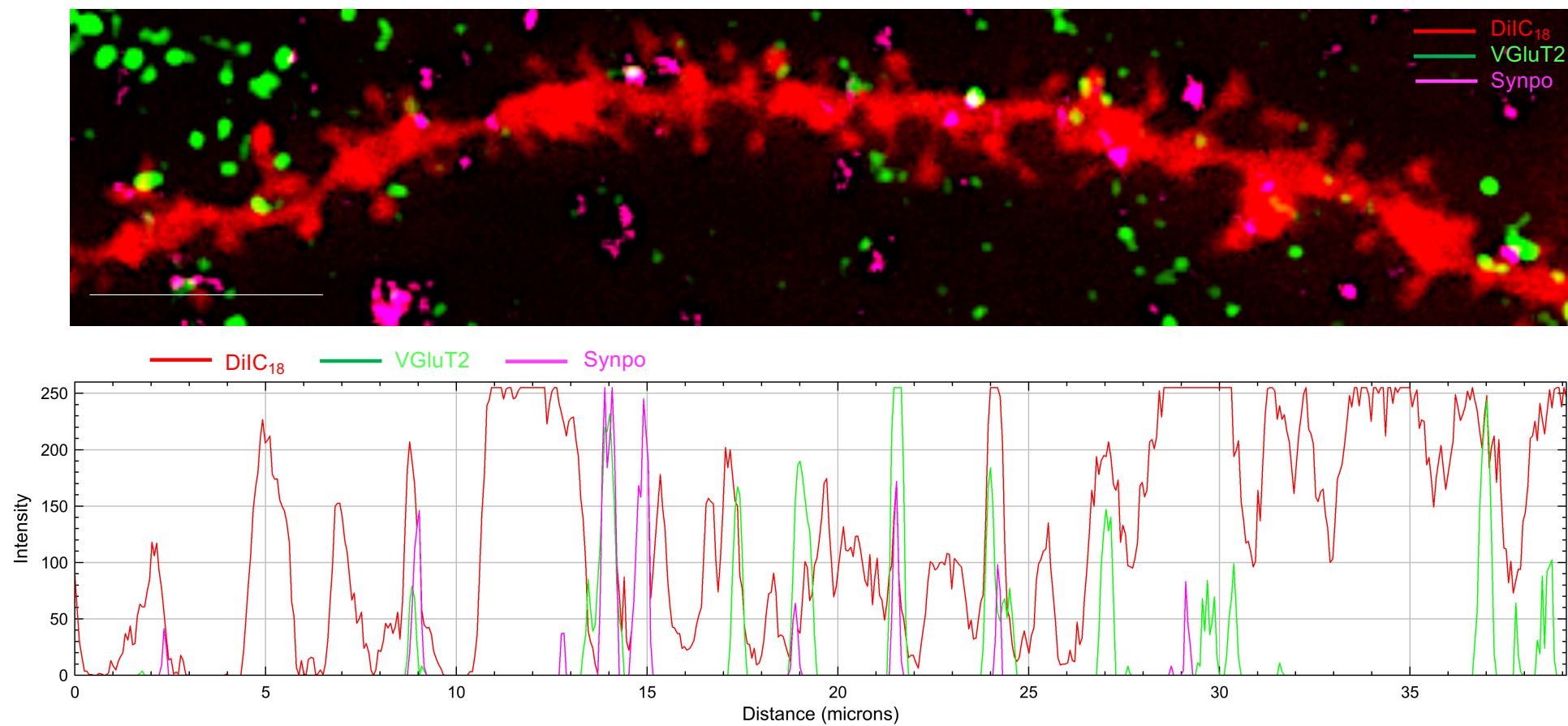


Figure S2. Representative line scan illustrating signal intensity for DiIC₁₈ (red), VGluT2 (green), and Synpo (Magenta) in function of distance. Scale bar, 5 μ m.

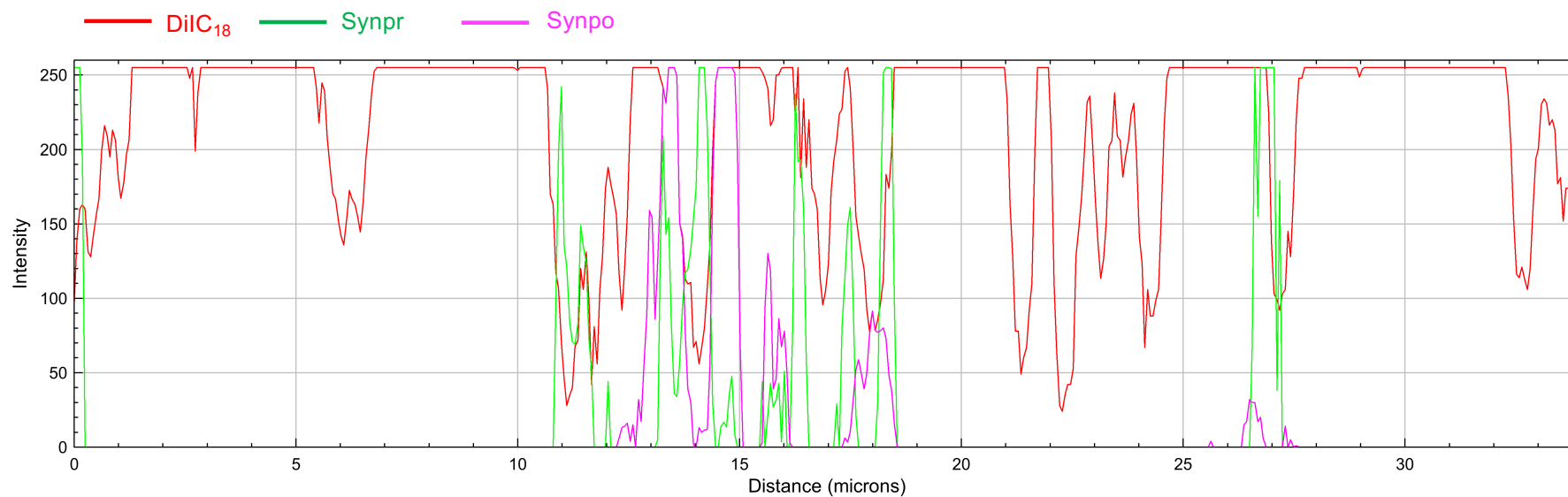
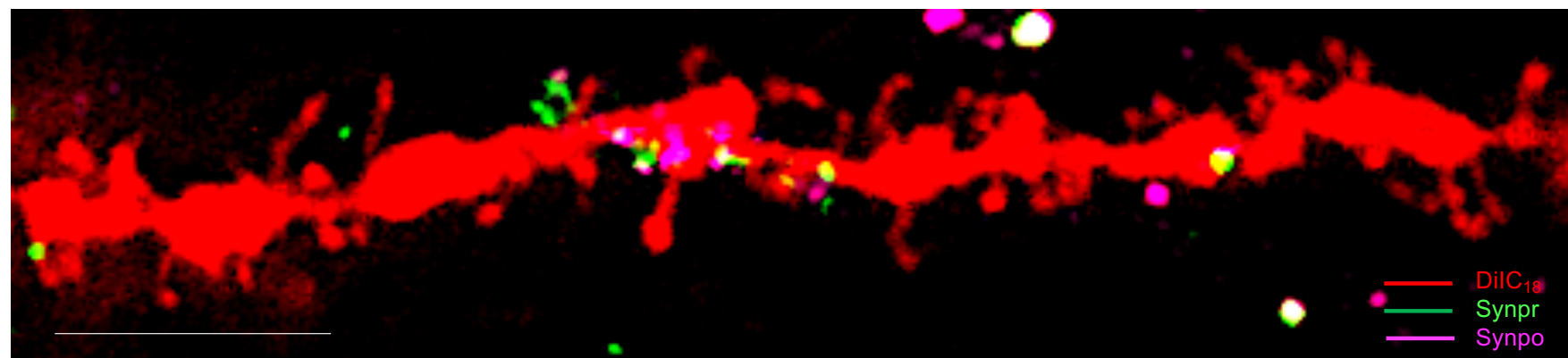


Figure S3. Representative line scan illustrating signal intensity for DiIC₁₈ (red), Synpr (green), and Synpo (Magenta) in function of distance. Scale bar, 5 μ m.

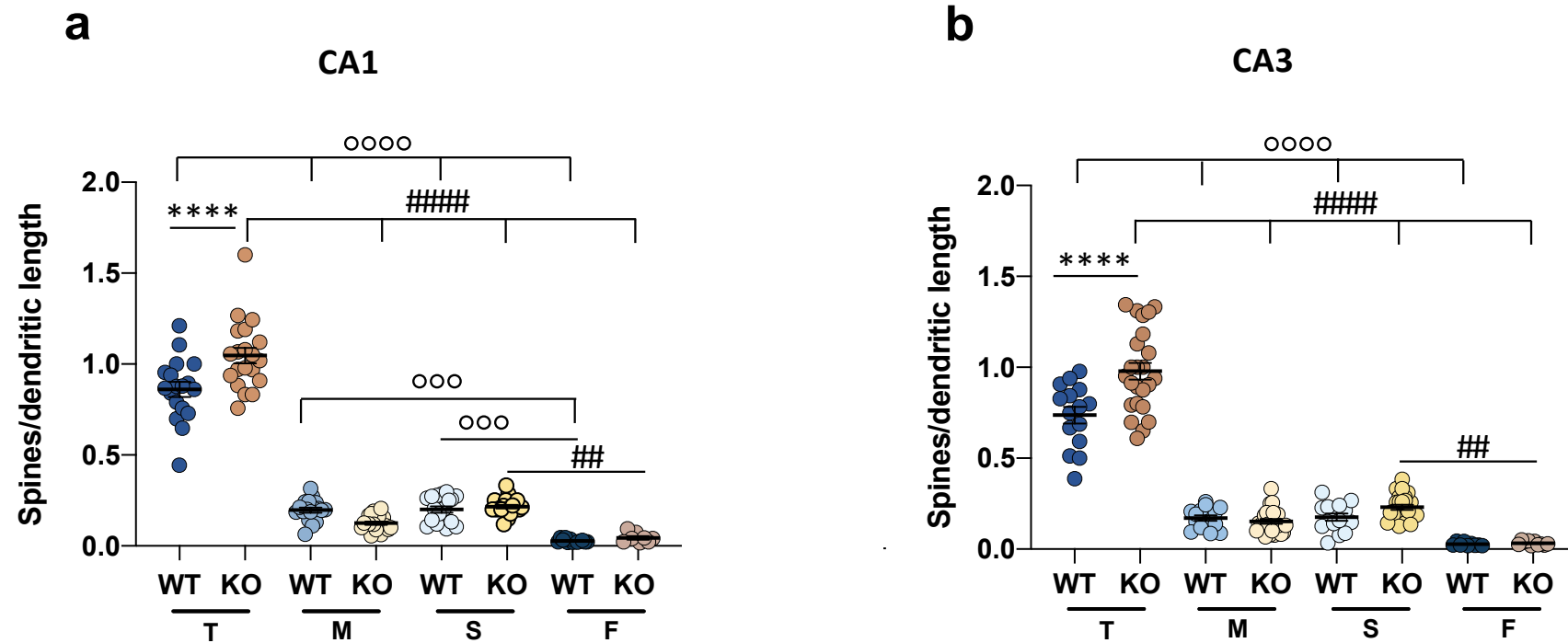


Figure S4. DiIC₁₈ staining reveals dysgenesis of hippocampal spines in juvenile Fmr1 KO mice. (a) Quantification of thin (T), mushroom (M), stubby (S), and filopodia (F) spines per dendritic length (μm) in CA1 of WT and Fmr1 KO mice. Differences were evaluated by two-way ANOVA followed by Tukey's post-hoc multiple comparisons test **** $p < 0.0001$ KO vs WT, oooo $p < 0.0001$ WT vs WT, ooo $p \leq 0.0007$ WT vs WT, ### $p < 0.0001$ KO vs KO, ## $p = 0.0044$ KO vs KO. A two-factor ANOVA demonstrated a significant effect of spine type ($p < 0.0001$) but a non-significant effect of genotype ($p = 0.057$), and the interaction of genotype by spine's type is significant ($p < 0.0001$). (b) Quantification of spines per dendritic length (μm) in CA3 of WT and Fmr1 KO mice. Differences were evaluated by two-way ANOVA followed by Tukey's post-hoc multiple comparisons test **** $p < 0.0001$, oooo $p < 0.0001$ WT vs WT, ## $p < 0.0011$ KO vs KO. A two-factor ANOVA demonstrated a significant effect of genotype ($p = 0.0022$) and spine's type ($p < 0.0001$), and the interaction of genotype by spine's type is significant ($p < 0.0001$).