

## Article

# A New Benzoxazole-Based Fluorescent Macrocyclic Chemosensor for Optical Detection of Zn<sup>2+</sup> and Cd<sup>2+</sup>

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**Abstract:** Background: Benzoxazole-containing ligands find many applications both in medicinal chemistry, catalysis and fluorescence chemosensing. Benzoxazole-containing macrocycles could be therefore a good strategy to achieve stable and selective fluorescent complexes with suitable metal ions. In this work, the synthesis, binding, and photochemical properties of a new fluorescent ligand (**L**) are reported. **L** is a cyclophane macrocycle containing the 1,3-bis(benzo[*d*]oxazol-2-yl)phenyl (BBzB) fluorophore and an aliphatic tetra-amine chain to form the macrocyclic skeleton. Methods: Spectrophotometric and spectrofluorimetric measurements, <sup>1</sup>H NMR analysis, and DFT calculations were performed. Results: **L** behaves as a PET-mediated chemosensor, being emissive at 390 nm at acidic pH and non-emissive at basic pH. The chemosensor is able to detect Zn<sup>2+</sup> and Cd<sup>2+</sup> in an aqueous medium (acetonitrile–water, 4:1 *v/v*) at neutral pH through a CHEF effect upon metal ion coordination. Paramagnetic metal ions (Cu<sup>2+</sup>) and heavy atoms (Pb<sup>2+</sup>, Hg<sup>2+</sup>) resulted in a quenching of fluorescence or very low emission. Conclusions: The new cyclophane macrocycle **L** was revealed to be a selective PET-regulated chemosensor for Zn<sup>2+</sup> and Cd<sup>2+</sup> in an aqueous medium, being able to bind up to two and one metal cations, respectively. The molecule showed a shifted emission towards the visible region compared to similar systems, suggesting a co-planar conformation of the aromatic fragment upon metal coordination. All these data are supported by both experimental measurements and theoretical calculations.

**Keywords:** fluorescent sensor; macrocycles; N-ligands; DFT calculations



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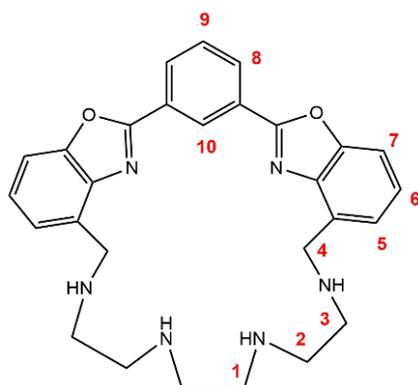
## 1. Introduction

Benzoxazoles (BZs) represent a common and versatile class of heterocycles. Numerous natural products showing biological activities contain BZ moieties (e.g., the fungal-derived antibiotic Caboxamycin and anticancer agent Nocarbenzoxazole G, the marine-derived anticancer agent Nakijinol B and antitubercular agent Pseudopteroxazole) [1]; indeed, many BZ fragments are often used as building blocks in medicinal chemistry for the synthesis of compounds with pharmacological activities (antimicrobial, antitumor, anti-inflammatory, antihistaminic, anti-helminthic, anti-tubercular, anticonvulsant, hypoglycaemic, HIV-1 reverse transcriptase inhibitor, just to name a few) [2,3]. More applications are derived from the excellent metal ion chelating properties of BZs. Transition metal complexes of BZ derivatives have been employed in the synthesis of metal catalysts [4], while other complexes have shown luminescence properties in the solid state [5,6].

Transition metal ions, such as Zn<sup>2+</sup> and Cd<sup>2+</sup> in particular, and optical methodologies devoted to their detection and quantification have long attracted attention from researchers [7–26]. Both metal ions feature a closed-shell d<sup>10</sup> configuration and, as a consequence, show similar chemical properties. Nevertheless, whilst Zn<sup>2+</sup> is essential to life

(playing crucial roles in many biological processes, such as regulation of apoptosis, signal transmission, gene expression, and enzyme function) [27,28],  $\text{Cd}^{2+}$  is toxic for animals and human beings and is classified as a human carcinogen [29–33]. Notably,  $\text{Zn}^{2+}$  could also become an issue for humans, since its deficiency results in an unbalanced metabolism, which could give rise to several neurological dysfunctions [34,35], while excessive amounts can cause cytotoxicity, arousing disorders such as skin disease, diabetes, prostatic adenocarcinoma, and pancreatic islets dysfunction [36].

In light of the desirable development of effective methods for monitoring  $\text{Cd}^{2+}$  and  $\text{Zn}^{2+}$  levels in biological and environmental matrices, and following our interest in fluorescent chemosensors for metal cations [16,17,37,38], we exploited the luminescence and coordination properties of BZs to develop a new BZ-based polyamine macrocycle able to bind and signal the presence of  $\text{Cd}^{2+}$  and  $\text{Zn}^{2+}$  in solution for possible analytical applications. A macrocycle topology has been exploited due to the greater selectivity they offer compared to open-chain ligands and the higher stability of their metal complexes [39–41]. Ligand **L** (Hexacyclo [28.3.1.1<sup>2,5</sup>.1<sup>26,29</sup>.0<sup>9,4</sup>.0<sup>22,27</sup>]-35,36-dioxo-3,11,14,17,20,28-hexaaza-2,4,6,8,22,24,26,28,30,32,1<sup>34</sup>-tetratricontaendecaene, Figure 1) has been synthesized by incorporating the 1,3-bis(benzo[*d*]oxazol-2-yl)phenyl (BBzB) fluorophore [42–46], containing two BZ units attached at positions 1 and 3 of a benzene ring, into a polyamine cyclophane macrocycle. The molecule contains two distinct compartments: 1,4,7,10-tetraazadecane polyamine as the binding unit, able to coordinate metal ions, and conjugated BBzB as the signaling unit, which could also participate in the coordination of guest species.



**Figure 1.** Structure of ligand **L** together with  $^1\text{H}$  NMR labelling scheme.

Herein, the synthesis and sensing properties in solution towards transition metal ions of the ligand **L** are reported.  $^1\text{H}$  NMR, UV-Vis and fluorescence spectroscopies, and DFT calculations were performed to assess the binding ability of the system towards  $\text{Cd}^{2+}$  and  $\text{Zn}^{2+}$ .

## 2. Materials and Methods

### 2.1. General Methods

All chemicals were purchased in the highest quality commercially available (Merck, Milan, Italy). The solvents were RP grade, unless otherwise indicated, and used without further purification. All reactions involving moisture-sensitive reagents were carried out under a nitrogen atmosphere using standard vacuum line techniques and glassware that was flame-dried before use. Elemental analyses were performed with a Thermo Finnigan Flash 1112 EA CHN analyzer (Milan, Italy). Mass spectra were performed with an Agilent 1200 Series HPLC system equipped with a binary pump and a C18 column (Phenomenex Synergi<sup>TM</sup> 4  $\mu\text{m}$  Fusion-RP 80  $\text{\AA}$ , LC Column 50  $\times$  2  $\text{mm}^2$ , Ea, Agilent Technologies, Milan, Italy), coupled to a SCIEX mod. API 4000 QqQ triple quadrupole mass spectrometer with ESI source (SCIEX, Milan, Italy). The High-Resolution Mass Spectrum of **L** was performed with an Orbitrap Exploris 240 Mass Spectrometer (Thermo Fisher Scientific, Milan, Italy) by

direct infusion at 5 microliters per minute and resolution set to 180K FWHM. AGC target was set to standard mode and Max Injection time was set to 100 ms.

## 2.2. Synthesis

### 2.2.1. 1,3-bis(4-Methyl-2-benzoxazolyl)phenyl (3)

2-Amino-m-cresol (**2**, 18.4 g, 150 mmol) and commercially available isophthalic acid (**1**, 12.4 g, 75 mmol) were suspended in polyphosphoric acid (PPA, 120.0 g) and heated overnight at 200 °C. The reaction mixture was allowed to cool down to room temperature (RT) and was poured into 3 dm<sup>3</sup> of water at 0 °C. After neutralization with Na<sub>2</sub>CO<sub>3</sub>, the mixture was filtered, and the residue was washed with methanol and dried under reduced pressure, obtaining product **3** as a white solid (18.6 g, yield 73%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25 °C) δ = 2.73 (s, 6H), 7.20 (d, *J* = 7.3 Hz, 2H), 7.30 (t, *J* = 7.8 Hz, 2H), 7.47 (d, *J* = 8.1 Hz, 2H), 7.70 (t, *J* = 7.8 Hz, 1H), 8.46 (dd, *J*<sub>1</sub> = 7.8 Hz, *J*<sub>2</sub> = 1.8 Hz, 2H), 9.14 (t, *J* = 1.5 Hz, 1H) ppm.

### 2.2.2. 1,3-bis(4-Bromomethyl-2-benzoxazolyl)phenyl (4)

A solution of **3** (2.0 g, 6.0 mmol), N-bromosuccinimide (NBS, 7.0 mmol), and 2,2'-azobis(2-methylpropionitrile) (AIBN, 1.20 mmol) in 50 mL of CCl<sub>4</sub> was refluxed for 24 h under a nitrogen atmosphere, and then the reaction mixture was allowed to cool down to RT. The obtained white precipitate was filtered off and the filtrate was concentrated under vacuum. The crude residue was crystallized from methanol, giving pure **4** as a pink solid (1.5 g, yield 50%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, 25 °C) δ = 5.00 (s, 4H), 7.74–7.41 (m, 7H), 8.50 (dd, *J* = 7.8, 1.8 Hz, 2H), 9.16 (t, *J* = 1.8 Hz, 1H) ppm (Figure S1). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 25 °C) δ = 27.6, 110.8, 124.7, 125.6, 126.9, 127.0, 128.0, 129.6, 130.7, 140.8, 150.9, 162.2 ppm (Figure S2). MS-ESI (*m/z*): calculated 498.95 (100%), 496.95 (51.4%), 500.95 (48.6%); found 497.3, 499.1, 501.3 for [M + H]<sup>+</sup> (Figure S3).

### 2.2.3. Hexacyclo[28.3.1.1<sup>2,5</sup>.1<sup>26,29</sup>.0<sup>9,4</sup>.0<sup>22,27</sup>]-35,36-dioxo-3,11,14,17,20,28-hexaaza-2,4,6,8,22,24,26,28,30,32,1<sup>34</sup>-tetratricontaendecaene tetrahydrobromide (L-4HBr)

A solution of 1,4,7,10-tetrakis(4-toluensulphonyl)-1,4,7,9-tetraazadecane [47] (**5**, 3.0 mmol) in 200 cm<sup>3</sup> of anhydrous dimethylformamide (DMF) was added to an excess of K<sub>2</sub>CO<sub>3</sub> (4 g, 30 mmol) and refluxed under a nitrogen atmosphere. To this mixture, a suspension of the brominated fluorophore **4** (1.5 g, 3 mmol) in 100 cm<sup>3</sup> of anhydrous DMF was added over a period of 8 h. After the addition, the mixture was refluxed for a further 12 h. Then, the reaction mixture was allowed to cool down to RT and the solid residue was filtered off. The filtrate was concentrated and put into cold water, obtaining a yellow precipitate that was collected by filtration and dried. The crude product was then used in the next step without further purification.

The yellow solid was suspended under a nitrogen atmosphere in 15 mL of 48% HBr/CH<sub>3</sub>COOH; next, phenol was added (2.5 g, 27.0 mmol), and then the mixture was heated at 90 °C for 24 h. The reaction was then allowed to cool down to RT. The obtained brown precipitate was collected by filtration and then washed with 2 mL mixture of ethanol and water (99:1), obtaining pure product L-4HBr as a white solid. Finally, the ligand was purified by re-crystallization from hot ethanol (1.48 g, yield 61%). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O, 25 °C) δ 3.27 (s, 4H), 3.40 (t, *J* = 7.0 Hz, 4H), 3.49 (t, *J* = 6.5 Hz, 4H), 4.63 (s, 4H), 7.34–7.44 (m, 4H), 7.63–7.74 (m, 3H), 8.23 (dd, *J* = 7.8, 1.9 Hz, 2H), 8.89 (s, 1H), ppm. <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O, 25 °C) δ = 43.2, 43.4, 43.9, 46.6, 112.9, 120.9, 126.4, 126.5, 126.8, 127.1, 130.4, 131.0, 140.3, 150.5, 163.1 ppm (Figure S4). HRMS: (positive mode, *m/z*) calculated 483.2508, found 483.2503 for [M + H]<sup>+</sup> (Figure S5). Anal. for C<sub>28</sub>H<sub>34</sub>Br<sub>4</sub>N<sub>6</sub>O<sub>2</sub> (806.23): calculated C 41.71, H 4.25, N 10.42; found C 41.5, H 4.4, N 10.3.

## 2.3. UV-Vis and Fluorescence Measurements

Spectrophotometric measurements were carried out at 298.1 K using a Varian Cary 100 spectrophotometer equipped with a temperature control unit. Uncorrected emission

spectra were obtained with a Varian Cary Eclipse fluorescence spectrophotometer. Luminescence quantum yields were determined using 2,2'-biphenol in acetonitrile ( $\Phi = 0.29$ ) [48].

UV-Vis and fluorescence emission screenings of **L** were performed in variable acetonitrile–aqueous HEPES (0.001 mol dm<sup>-3</sup>, pH 7.4) solvent mixtures (% aqueous HEPES: from 100 to 20% with respect to ACN) at 25 °C, with a ligand concentration of  $3.2 \times 10^{-6}$  mol dm<sup>-3</sup> and by exciting at  $\lambda_{\text{ex}} = 305$  nm.

UV-Vis and fluorescence emission titrations of **L** were performed in acetonitrile–aqueous medium 80:20 *v/v* at 25 °C, with a ligand concentration of  $3.2 \times 10^{-6}$  mol dm<sup>-3</sup> and exciting at  $\lambda_{\text{ex}} = 305$  nm. pH titrations of **L** were performed in acetonitrile/water in the 2–12 pH range; the pH was adjusted by using 0.01 mol dm<sup>-3</sup> HCl and NaOH solutions. Titrations with Zn(ClO<sub>4</sub>)<sub>2</sub> and Cd(ClO<sub>4</sub>)<sub>2</sub> were performed by adding to a solution of the ligand (10 cm<sup>3</sup>) in acetonitrile–aqueous HEPES (0.001 mol dm<sup>-3</sup>, pH 7.4) increasing volumes of aqueous solutions of the metal ions (0.1 equivalents at a time up to 5 equivalents).

Limit of detection (LOD) and limit of quantitation (LOQ) were calculated by linear regression of normalized emission intensity as a function of concentration of Zn<sup>2+</sup> and Cd<sup>2+</sup> [49] (for more details, see Supplementary Material).

#### 2.4. NMR Studies

<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on a Bruker Avance 400 instrument (Bruker Italia, Milan, Italy), operating at 400.13 and 100.61 MHz, respectively, and equipped with a variable temperature controller. The temperature of the NMR probe was calibrated using 1,2-ethanediol as a calibration sample. Chemical shifts ( $\delta$  scale) are presented in ppm and referenced by residual solvent peak. Coupling constants (*J* values) are given in hertz (Hz). <sup>1</sup>H-<sup>1</sup>H and <sup>1</sup>H-<sup>13</sup>C correlation experiments were performed to assign the signals. All measurements were performed at 298.1 K, in mixed acetonitrile-*d*<sub>3</sub>/D<sub>2</sub>O 60:40 *v/v* solution (to prevent solubility issues) for the pH titration, and in D<sub>2</sub>O for titrations with Zn(ClO<sub>4</sub>)<sub>2</sub> and Cd(ClO<sub>4</sub>)<sub>2</sub>, at a ligand concentration of  $5 \times 10^{-3}$  mol dm<sup>-3</sup>. The pH titration was performed in the pH range of 2–12, starting from pH 2 and raising the pH by using a KOD solution. Titrations with Zn(ClO<sub>4</sub>)<sub>2</sub> and Cd(ClO<sub>4</sub>)<sub>2</sub> were performed at pH 7.4 by adding the metal ions dissolved in D<sub>2</sub>O 0.1 equivalents at a time directly in the NMR tube.

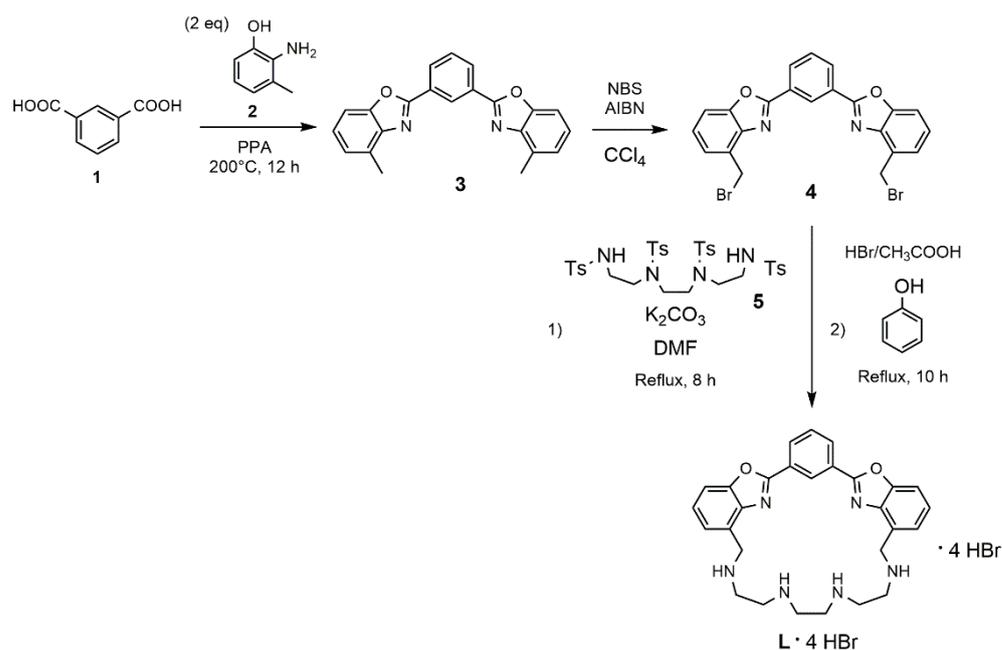
#### 2.5. Computational Analysis

The structures of **L** and of its complexes with Zn<sup>2+</sup> and Cd<sup>2+</sup> were optimized with Gaussian 09 software [50] and calculated for an isolated molecule using Density Functional Theory (DFT) [51–55] at the BP86/6-31G(d,p) and B3LYP/TZVP level of theory for the ligand and BP86/LanL2DZ and B3LYP/LanL2DZ [56] for the complexes. For more details, see the Supplementary Material.

### 3. Results and Discussion

#### 3.1. Synthesis

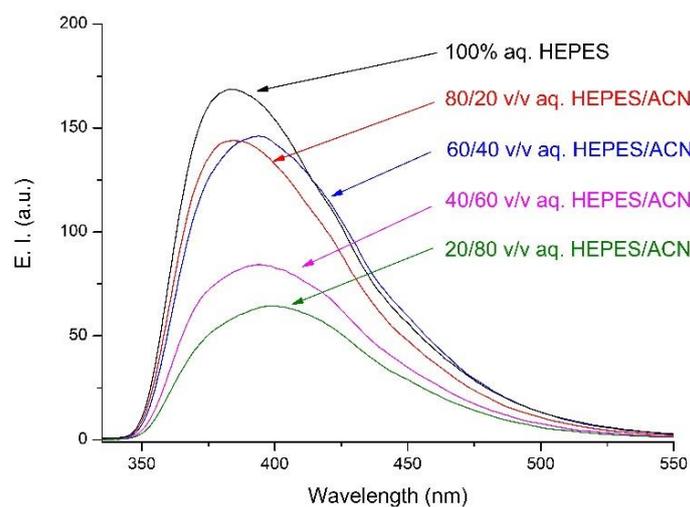
The synthetic route to obtain ligand **L** is depicted in Scheme 1. The BBzB fluorophore **3** was obtained by adapting a known procedure [42], via a double condensation of two equivalents of isophthalic acid (**1**) with one equivalent of 2-amino-*m*-cresol (**2**) in the presence of polyphosphoric acid. Compound **4**, obtained by benzylic bromination of **3**, was reacted with 1,4,7,10-tetrakis(4-toluensulphonyl)-1,4,7,9-tetraazadecane (**5**) [47] via Richmann–Atkins anellation [57,58]. In brief, this is a double nucleophilic substitution performed in high-dilution conditions to avoid polymer formation and favor the cyclization reaction. After the removal of the four protecting sulphonyl groups on the amine functions by treatment with HBr in acetic acid in the presence of phenol as a scavenger, the ligand was obtained as tetrahydrobromide salt (**L**·4HBr).



**Scheme 1.** Synthetic pathway of ligand **L**.

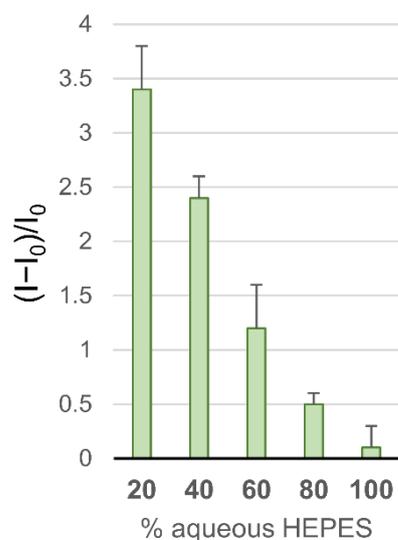
### 3.2. Preliminary Screening

To assess the better experimental conditions to investigate the photochemical properties of the ligand, solutions of **L** in different ratios of water (HEPES buffer, pH 7.4) and acetonitrile (ACN) were analyzed (Figure 2). The emission of the ligand increased along with the percentage of the aqueous solution: in a photoinduced intramolecular electron transfer (PET)-mediated sensor, this would suggest a higher protonation degree of the ligand, translating to a more efficient prevention of the PET effect and a higher fluorescence emission (vide infra).



**Figure 2.** Fluorescence emission of **L** in different aqueous HEPES/ACN mixtures.  $[L] = 3.2 \times 10^{-6}$  mol dm $^{-3}$ ;  $[HEPES] = 1.0 \times 10^{-3}$  mol dm $^{-3}$ ; pH 7.4;  $\lambda_{ex} = 305$  nm.

A preliminary screening towards the Cd $^{2+}$  ion was performed in the same mixed aqueous solutions (from 20 to 100% aqueous HEPES buffer with respect to ACN), revealing that working in ACN—aqueous HEPES buffer (pH 7.4) in a 4:1 *v/v* ratio returned the best performance of the system, with the emission increase upon the addition of the metal ion being the highest within the series (3.4-fold at 20% aqueous HEPES vs. 2.4-, 1.2-, 0.5-, and 0.1-fold at 40, 60, 80, and 100% aqueous HEPES, respectively; Figure 3).



**Figure 3.** Fluorescence emission increase upon addition of 1 equivalent of  $\text{Cd}^{2+}$  to a solution of **L** in different ACN—aqueous HEPES buffer *v/v* solvent mixtures.  $[\text{L}] = 3.2 \times 10^{-6} \text{ mol dm}^{-3}$ ;  $[\text{HEPES}] = 1.0 \times 10^{-3} \text{ mol dm}^{-3}$ ; pH 7.4;  $\lambda_{\text{ex}} = 305 \text{ nm}$ .  $\lambda_{\text{em}} = 390 \text{ nm}$  (100% aqueous HEPES), 394 nm (80% aqueous HEPES), 397 nm (60% aqueous HEPES), 398 nm (40% aqueous HEPES), and 398 nm (20% aqueous HEPES).

### 3.3. Acid–Base Behavior

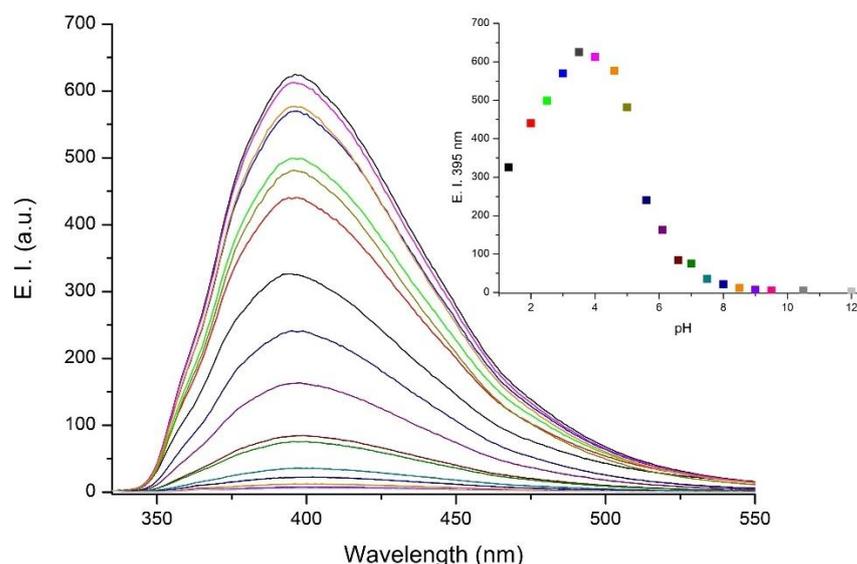
#### 3.3.1. UV-Vis Studies

After ascertaining the better performance of the system in ACN—water in a 4:1 *v/v* ratio, the acid–base behavior of the ligand was investigated under the same experimental conditions by spectrophotometric and spectrofluorimetric analysis. Spectra at different pH values were recorded to understand the role of the BBzB fluorophore in the acid–base behavior of the system.

Under the present experimental conditions, the free ligand shows two absorption bands at 234 nm ( $\epsilon = 18,440 \text{ cm}^{-1} \text{ mol}^{-1} \text{ dm}^3$ ) and 305 nm ( $\epsilon = 36,250 \text{ cm}^{-1} \text{ mol}^{-1} \text{ dm}^3$ ). While UV-Vis absorption measurements did not show any significant variation, the fluorescence emission is strongly pH dependent (Figure 4). By exciting at 305 nm at acidic pH (2), the ligand featured a strong emission with a maximum at  $\lambda_{\text{em}} = 402 \text{ nm}$ ; when increasing the pH, the emission also increased, doubling its value and reaching the maximum at pH 3.5. Then, the emission dropped down, with the system becoming totally quenched above pH 9.5. This behavior can be rationalized in terms of quenching of the excited state due to the PET effect from the HOMO of the polyamine nitrogen atoms to the excited fluorophore moiety [59]. In similar systems, it has been observed that the PET-mediated quenching of the fluorescence, which occurs when at least one amine function sufficiently close to the fluorophore is neither protonated nor involved in any hydrogen bond, is mainly due to the benzyl nitrogen atoms [60–65].

Considering these spectroscopic data, the pKa and the proton distribution in similar systems [62,63,65], and the key role of the benzylic nitrogen lone pairs in the quenching mechanism [61,62], it can be suggested that at pH 2, the fully protonated and fluorescent  $\text{H}_4\text{L}^{4+}$  species probably prevails in solution; the increase in the emission intensity between pH 2 and 3.5 suggests that a deprotonation process occurs in this pH range, forming the  $\text{H}_3\text{L}^{3+}$  species, and, more likely, a central amine function of the tetraamine chain, rather than an amine function in the benzylic position, is involved in the deprotonation step. In addition, the emission increase with the pH could be explained by a decrease of the molecular stiffness on moving from  $\text{H}_4\text{L}^{4+}$  to  $\text{H}_3\text{L}^{3+}$  species, as commonly observed in polyaza-macrocycles [62]. As a consequence, the conformation adopted by the BBzB system in the two species could be affected differently, resulting in  $\text{H}_4\text{L}^{4+}$  being less emissive than  $\text{H}_3\text{L}^{3+}$ . This behavior goes along with the hypothesis that in the  $\text{H}_4\text{L}^{4+}$  and  $\text{H}_3\text{L}^{3+}$  emissive

species, both benzyl nitrogen atoms are supposed to be protonated and the PET effect is prevented.



**Figure 4.** Fluorescence emission spectra of **L** registered in an ACN–water 4:1 *v/v* solvent mixture in the pH range of 2–12. Inset: trend of emission intensity at 395 nm at different pH values.  $[L] = 3.2 \times 10^{-6} \text{ mol dm}^{-3}$ ,  $\lambda_{\text{ex}} = 305 \text{ nm}$ . pH adjusted by adding HCl and NaOH aqueous solutions.

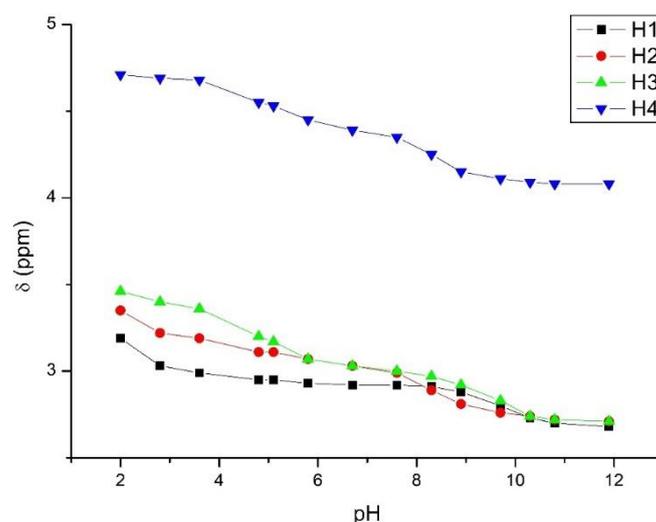
Above pH 3.5, the emission trend indicates further deprotonation processes at least up to pH 9.5, where the emission became totally quenched. In the 3.5–9.5 pH range, two deprotonation steps could be hypothesized to give the  $\text{H}_2\text{L}^{2+}$  and  $\text{HL}^+$  species. The observed emission drop suggests that the deprotonation involves the benzylic nitrogen atoms, partially restoring the PET effect at a high pH. The PET effect, however partial, is enough to make the emission drop by 87% upon moving from pH 3.5 to 7, with total quenching above pH 9.5. This reminds us that in similar systems, where the emission is PET regulated, a free lone pair on an amine function in benzylic position is enough to quench the emission; thus, it is presumable that above pH 9.5, at least one of the two nitrogen atoms of **L** in the benzylic position resulted in nonprotonation.

### 3.3.2. $^1\text{H}$ NMR Analysis

To better understand the proton distribution in the possible species of the ligand, fluorescence data were combined with  $^1\text{H}$  NMR measurements.  $^1\text{H}$  NMR spectra at various pH values were registered in acetonitrile- $d_3/\text{D}_2\text{O}$  60:40 over the same 2–12 pH range (Figure 5 and Figure S1): the initial pH of the ligand ( $\text{L}\cdot 4\text{HBr}$ , pH = 2) was raised directly in the NMR tube through the addition of KOD. 2D standard correlation experiments were performed to assign all the signals. A mixed aqueous medium was used to prevent solubility issues in  $\text{D}_2\text{O}$  at a high pH.

At pH = 12, the  $^1\text{H}$  NMR spectrum of **L** shows three aliphatic and four aromatic resonances ( $\delta = 2.70 \text{ ppm}$  ( $H_1$ , 4H), 2.72 ppm ( $H_2 + H_3$ , 8H), 4.08 ppm ( $H_4$ , 4H), 7.32 ppm ( $H_5$ , 2H), 7.40 ppm ( $H_6$ , 2H), 7.59 ppm ( $H_7$ , 2H), 7.77 ppm ( $H_9$ , 1H), 8.29 ppm ( $H_8$ , 2H), 9.16 ppm ( $H_{10}$ , 1H)) (see Figure S6 for the spectrum and Figure 1 for the assignments). The  $^{13}\text{C}$  spectrum exhibits a total of 15 signals (see experimental section): 4 for the aromatic and 11 for the aliphatic resonances. Along with the  $^1\text{H}$  spectrum, this indicates a  $\text{C}_{2v}$  symmetry of **L** mediated on the NMR timescale.

Starting from pH = 12, where the presence of the neutral **L** species can be hypothesized in solution, and moving towards acidic pH values, no significant changes were detected up to pH 10.5, indicating the presence in solution in this pH range of the neutral **L** species (Figure 5).



**Figure 5.** Trend of selected  $^1\text{H}$  NMR signals of aliphatic L protons in an acetonitrile- $d_3$ / $\text{D}_2\text{O}$  60:40 solution as a function of the pH.  $[\text{L}] = 5 \times 10^{-3} \text{ mol dm}^{-3}$ . See Figure 1 for the assignments.

In the pH range of 10.5–7.6, all the aliphatic resonances (mainly those belonging to H2, H3 and H4 protons) shifted downfield, suggesting that the first protonation step mainly involves an amine function in the benzylic position of BBzB to give the  $\text{HL}^+$  species. By lowering the pH to 5.2, another protonation process occurred (mainly involving H3 and H4 resonances), suggesting that this step is dependent again on a benzyl amine function to give the  $\text{H}_2\text{L}^{2+}$  species. However, these protons, although mainly located at those positions, are probably shared and stabilized via H-bonding with the closer amine functions, as suggested by both the downfield shift of all aliphatic resonances and the only slight switching ON of the fluorescence, due to partial prevention of the PET effect (vide supra), as already observed in similar systems containing the same aliphatic polyamine chain [62].

At lower pH values, the downfield shift of the aliphatic resonances suggests two further protonation steps, fixing the acidic protons on the aliphatic amine functions of the polyamine chain, mainly on the central ones; the greatest shifts refer to the H3 and H4 signals, in the pH range of 5.2–3.5, to give the  $\text{H}_3\text{L}^{3+}$  species, and to the H1 and H2 resonances, moving from pH 3.5 to 2, to give the fully protonated  $\text{H}_4\text{L}^{4+}$  species.

In general, all resonances move downfield when lowering the pH, with the aromatic resonances exhibiting smaller shifts than the aliphatic ones. This suggests that the aromatic fluorophore is not directly involved in any protonation step. The most evident shifts associated with the aromatic resonances occurred in the 3.5–5.2 and 7.6–10.5 pH ranges. H7 also showed a variation in the 5.2–7.6 pH range (Figure S7). These shifts are probably due to the presence or absence of hydrogen bonds involving the benzylic nitrogen atoms, which can affect the electron density on the aromatic rings and the conformation of the whole system. Regarding the aliphatic resonances, notwithstanding the shifts described above, it must be noted that they show a continuous perturbation throughout the whole pH range, suggesting, besides the protonation steps, the involvement of the amine functions in hydrogen bonding.

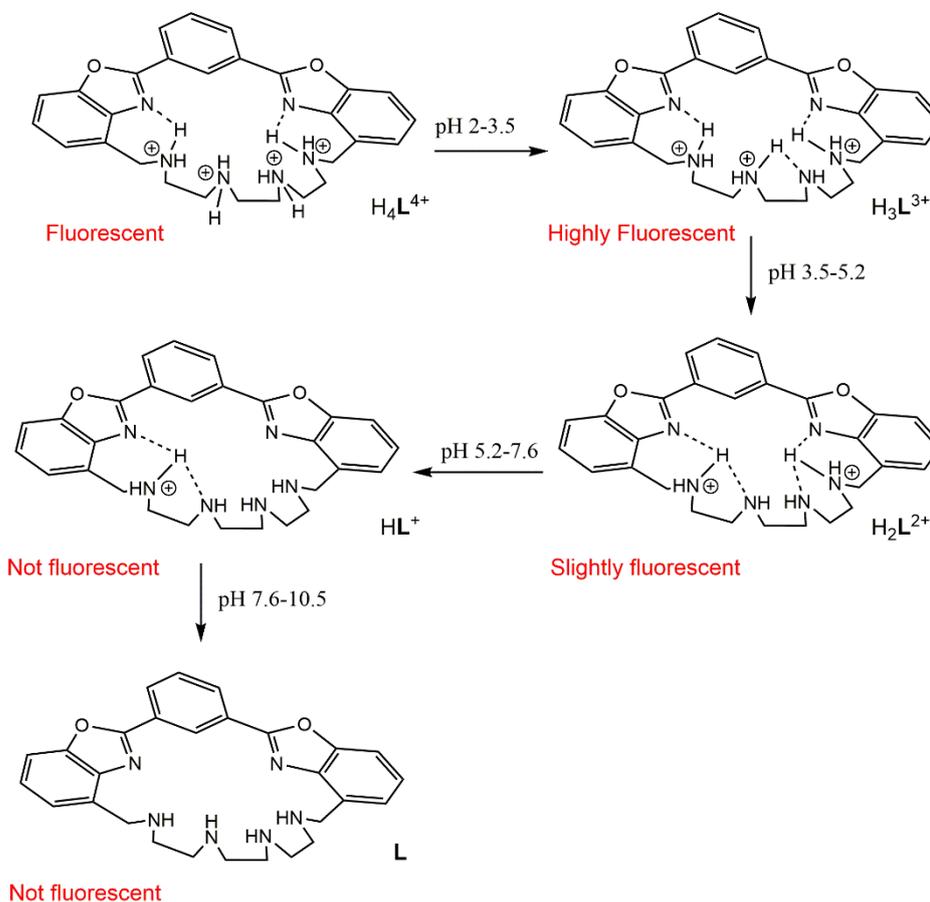
Taking into account both NMR and spectrofluorimetric data, the proton distribution scheme depicted in Figure 6 could be proposed.

### 3.4. Metal Ion Complexation

#### 3.4.1. UV-Vis Studies

Spectrophotometric and spectrofluorimetric measurements were performed to investigate the photochemistry of the BBzB fluorophore and the coordination behavior towards metal ions.

Since the ligand exhibits PET-mediated emission properties (*vide supra*), the coordination of a suitable metal ion within the macrocycle cavity should prevent the PET effect, enabling the emission of the complex.

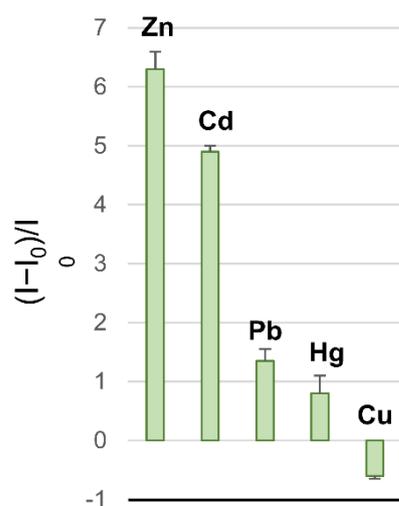


**Figure 6.** Proposed disposition of  $H^+$  in the protonated forms of the ligand.

A preliminary screening was performed to evaluate the response of the system to selected transition metal ions ( $Zn^{2+}$ ,  $Cd^{2+}$ ,  $Pb^{2+}$ ,  $Cu^{2+}$ ,  $Hg^{2+}$ ), chosen among heavy metals, paramagnetic metals, and generally responsive  $d^{10}$  metals. The screening was performed in the same experimental conditions found above (ACN—aqueous HEPES (pH 7.4) 4:1 *v/v*), by adding two equivalents of metal ion to the solution of **L** (Figure 7).

The addition of a solution of  $Cd^{2+}$  as perchlorate salt did not greatly affect the absorption spectrum, while the addition of  $Zn^{2+}$  as perchlorate salt provoked the variation of the band at a higher energy, suggesting a certain degree of involvement of the BBzB moiety in the coordination of the cation, especially regarding the second equivalent of  $Zn^{2+}$ . The addition of  $Pb^{2+}$ ,  $Hg^{2+}$ , and  $Cu^{2+}$  as perchlorate salts showed the growing of new bands at 251 nm, 261 nm, and 296 nm, respectively, which can be attributed either to the involvement of the BBzB moiety in the coordination or to a metal-induced distortion of the aromatic system (Figure S8).

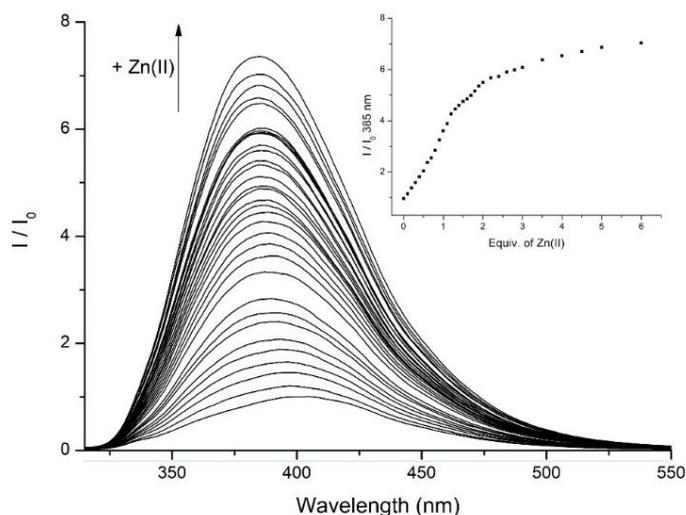
Among the tested cations, only  $Zn^{2+}$  and  $Cd^{2+}$  showed a pronounced increase of the emission intensity of the ligand under the chosen experimental conditions, while  $Pb^{2+}$  and  $Hg^{2+}$  only slightly affected the emission of the system, probably due to the heavy metal effect. As expected, the emission is quenched in the presence of  $Cu^{2+}$ , due to the paramagnetic effect and the resulting favored ISC from the  $S_1$  to the  $T_1$  state (Figure 7).



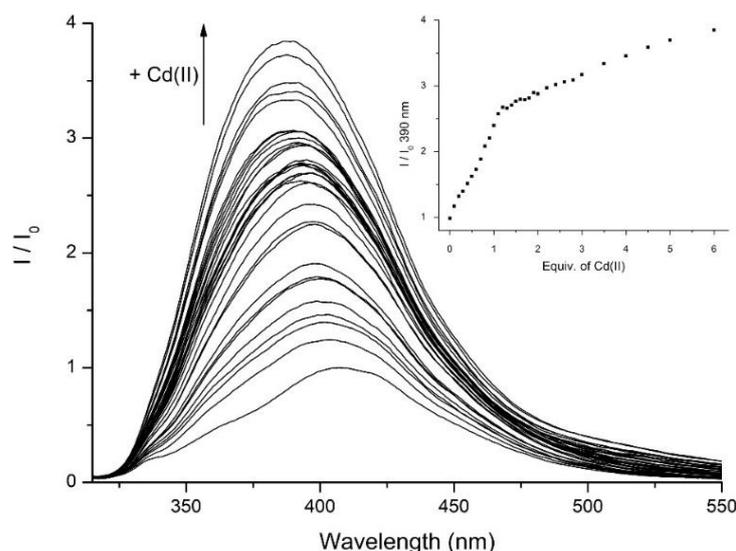
**Figure 7.** Bar-plot showing the emission of **L** upon addition of 2 equivalents of different metal cations ( $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Hg}^{2+}$ ).  $I$  and  $I_0$  are the emission intensities of the ligand in the presence and absence of the metal ions, respectively.  $[\text{L}] = 3.2 \times 10^{-6} \text{ mol dm}^{-3}$  (ACN–aqueous HEPES (pH 7.4) 4:1 *v/v* solvent mixture).  $\lambda_{\text{ex}} = 305 \text{ nm}$ .  $\lambda_{\text{em}} = 385 \text{ nm}$  ( $\text{Zn}^{2+}$ ),  $390 \text{ nm}$  ( $\text{Cd}^{2+}$ ),  $395 \text{ nm}$  ( $\text{Pb}^{2+}$ ),  $397 \text{ nm}$  ( $\text{Hg}^{2+}$ ), and  $410 \text{ nm}$  ( $\text{Cu}^{2+}$ ).

More metal ions were investigated ( $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Ag}^+$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ ), with none of them inducing a fluorescence response (Figure S9).

Following these preliminary results, spectrofluorimetric titrations of the ligand in ACN–aqueous HEPES pH 7.4, 4:1 *v/v* with  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  were performed to assess the behavior of the chemosensor in the presence of such cations. The measurements revealed great fluorescence emission enhancement upon the coordination of both  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  (chelation enhancement of fluorescence (CHEF) effect). Figures 8 and 9 report the emission spectra obtained by the titration of a buffered (pH 7.4) solution of **L** (ACN–aqueous HEPES 4:1 *v/v* solvent mixture) with  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  perchlorate, upon excitation at 305 nm. The free ligand exhibits an emission band with a maximum at 402 nm (emission quantum yield,  $\Phi_{\text{em}} = 0.006$ ; Stokes shift =  $7910 \text{ cm}^{-1}$ ); a blue shift was observed upon addition of the metal ion, reaching 385 and 390 nm in the presence of two equivalents of  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$ , respectively, with PET-mediated enhancement of the fluorescence compared to free **L**.



**Figure 8.** Variation of the fluorescence emission spectra of **L** by adding  $\text{Zn}(\text{ClO}_4)_2$  registered in an ACN–aqueous HEPES pH 7.4, 4:1 *v/v* solvent mixture. Inset: trend of emission intensity of **L** at 385 nm.  $[\text{L}] = 3.2 \times 10^{-6} \text{ mol dm}^{-3}$ .  $\lambda_{\text{ex}} = 305 \text{ nm}$ . Emission intensities are relative to the free ligand.



**Figure 9.** Variation of the fluorescence emission spectra of **L** by adding  $\text{Cd}(\text{ClO}_4)_2$  registered in an ACN–aqueous HEPES pH 7.4, 4:1 *v/v* solvent mixture. Inset: trend of emission intensity of **L** at 390 nm.  $[\text{L}] = 3.2 \times 10^{-6} \text{ mol dm}^{-3}$ .  $\lambda_{\text{ex}} = 305 \text{ nm}$ . Emission intensities are relative to the free ligand.

The two titrations seemed to suggest a different stoichiometry for the two complexes. The addition of the  $\text{Zn}^{2+}$  ion induced a fluorescence emission increase up to two equivalents, suggesting the formation of the dinuclear complex  $[\text{M}_2\text{L}]^{4+}$ , while the addition of the  $\text{Cd}^{2+}$  ion seemed to suggest the formation of the mononuclear complex  $[\text{ML}]^{2+}$ . The emission quantum yields of **L** in the presence of 2 equivalents of  $\text{Zn}^{2+}$  or 1 equivalent of  $\text{Cd}^{2+}$  are  $\Phi_{\text{em}} = 0.036$  and  $\Phi_{\text{em}} = 0.015$ , respectively, under these experimental conditions.

Statistical analysis of the trend furnished a limit of detection (LOD) of 0.02 and 0.03 ppm ( $3.1 \times 10^{-7}$  and  $2.9 \times 10^{-7} \text{ mol dm}^{-3}$ ) and a limit of quantitation (LOQ) of 0.06 and 0.09 ppm ( $9.6 \times 10^{-7}$  and  $8.9 \times 10^{-7} \text{ mol dm}^{-3}$ ) in  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$ , respectively ( $R > 0.99$ ), associated with this method (Figures S10 and S11) [49].

The emission wavelength of the newly synthesized BBzB fluorophore (402 nm) is redshifted by 40–50 nm with respect to the previously reported POXAPy (358 nm) [12] and PPD/PPyD (350 nm) [14,61,62,64] systems, featuring oxadiazole (ODA) rings and a total of five and three conjugated aromatic rings, respectively (Figure S12). The BBzB emission is therefore more shifted towards the visible region. The absorption wavelength of the fluorophore (305 nm) is the same as of that of POXAPy (306 nm) and is longer compared to those of PPD/PPyD (280 nm), with a greater Stokes shift compared to POXAPy ( $7910 \text{ vs. } 4750 \text{ cm}^{-1}$ ) (Table S1). This could be the consequence of the presence of five aromatic rings, four of which are fused in the two benzoxazole systems, allowing the whole system to be coplanar (*vide infra*), resulting in a  $\pi$ -system extension that redshifts the emission of the fluorophore. This was not the case for the POXAPy system: even if containing more aromatic rings compared to the PPD unit, it featured a non-coplanarity between the five aromatic rings, which prevented the  $\pi$ -system extension and the redshift [12].

### 3.4.2. NMR Studies

To gain more insight on the coordination behavior of **L** towards  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$ , NMR experiments were performed.  $^1\text{H}$  NMR titrations were carried out by adding the metal ion to a  $5 \times 10^{-3} \text{ mol dm}^{-3}$  solution of the ligand in  $\text{D}_2\text{O}$  at pH 7.4.

The spectra obtained during the titrations with  $6 \times 10^{-2} \text{ mol dm}^{-3}$  solutions of  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  as perchlorate salts to **L** are reported in Figures S13 and S14. The addition of the two metal ions induced similar perturbations in the spectrum of **L**, with all resonances, both aromatic and aliphatic, showing an upfield shift. This behavior suggests the coordination of the metal ion by the ligand, probably due to the aliphatic nitrogen atoms, accompanied

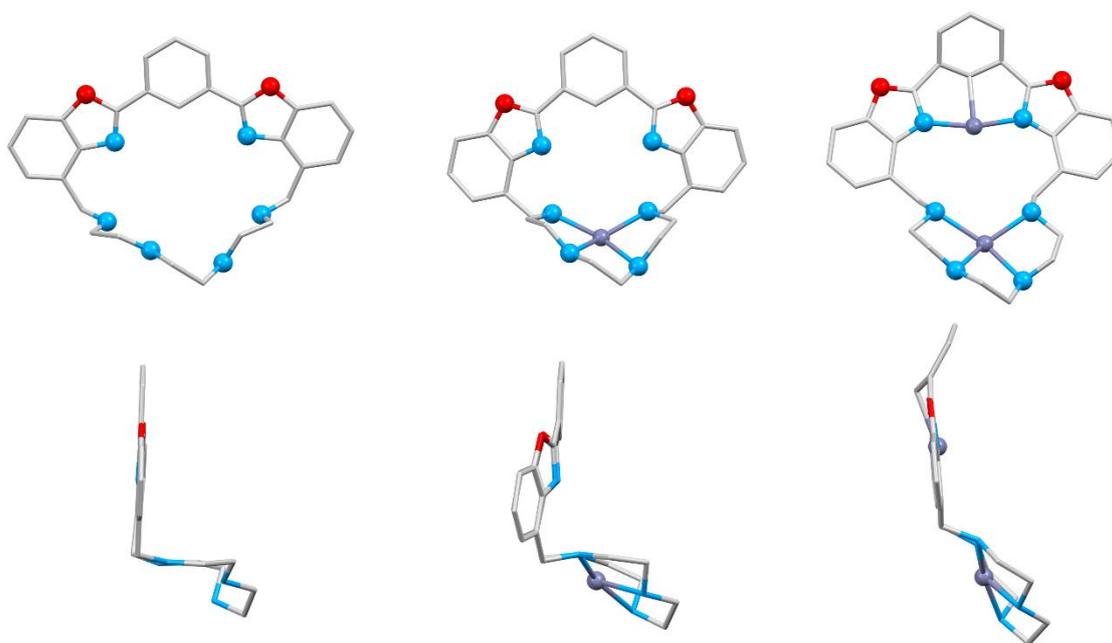
by the involvement of the benzoxazole nitrogen atoms and/or a distortion of the BBzB aromatic fragment.

The main difference between the titrations with the two metal ions is due to the invariance of the spectrum observed after the addition of one equivalent of  $\text{Cd}^{2+}$ , while further resonance shifts are induced by  $\text{Zn}^{2+}$  up to the addition of two equivalents of metal ion. This behavior suggests a different stoichiometry of the two complexes; in particular, a mononuclear  $[\text{ML}]^{2+}$  species is formed in the presence of  $\text{Cd}^{2+}$ , while a dinuclear  $[\text{M}_2\text{L}]^{4+}$  species is achieved in the presence of  $\text{Zn}^{2+}$ . These data are in agreement with those derived from fluorescence titrations, confirming the stoichiometry of the complexes with  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$ . Moreover, in both cases, the same  $\text{C}_{2v}$  symmetry mediated on the NMR timescale was preserved during the whole titration experiments, suggesting the symmetric involvement of the ligand in the coordination of the metal ion (Figures S13 and S14).

Finally, whereas the addition of  $\text{Cd}^{2+}$  to the preformed  $[\text{Zn}_2\text{L}]^{4+}$  species did not cause a significant variation of the  $^1\text{H}$  NMR spectrum of the complex, the addition of  $\text{Zn}^{2+}$  to the preformed  $[\text{CdL}]^{2+}$  species provoked an upfield shift of almost all resonances (Figures S15 and S16). This suggests a higher affinity of the ligand towards  $\text{Zn}^{2+}$  and a greater stability of the  $\text{Zn}^{2+}$  complex compared to the  $\text{Cd}^{2+}$  one.

### 3.4.3. Computational Studies

DFT calculations were used to study the relative stability of the  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  complexes with the ligand L. Results are presented in Figure 10 and Figure S17 and Table 1.



**Figure 10.** Front view (top) and side view (bottom) of the computed structures of (from left to right) L,  $[\text{ZnL}]^{2+}$  and  $[\text{Zn}_2\text{L}]^{4+}$ . Hydrogen atoms are omitted for the sake of clarity.

In detail, following the experimental solution studies (*vide supra*), the structures of both the mononuclear and dinuclear  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  complexes, i.e.,  $[\text{ZnL}]^{2+}$ ,  $[\text{Zn}_2\text{L}]^{4+}$ ,  $[\text{CdL}]^{2+}$ , and  $[\text{Cd}_2\text{L}]^{4+}$ , have been evaluated. The overall shapes of the two mononuclear and the two dinuclear metal complexes are almost identical (see Figures 10 and S17), as well as the coordination geometry around the metal cations, with the first cation coordinated by the four aliphatic nitrogen atoms of the polyamine chain, and the second one by the two nitrogen atoms of the benzoxazole rings and by the carbon atom of the central phenyl ring.

**Table 1.** Results of reactions 1 and 2 obtained by DFT calculations for Zn<sup>2+</sup> and Cd<sup>2+</sup> complexes of L.

Compound	$\Delta G$ (kcal/mol) <sup>1</sup>	$\Delta G$ (kcal/mol) <sup>2</sup>	DIST1 (Å)	DIST2 (Å)
[ZnL] <sup>2+</sup>	−331.8	−331.3	0.36 <sup>3</sup>	8.79 <sup>5</sup>
[Zn <sub>2</sub> L] <sup>4+</sup>	−367.5	−366.4	0.33 <sup>3</sup> 0.072 <sup>4</sup>	8.84 <sup>5</sup> 6.20 <sup>6</sup>
[CdL] <sup>2+</sup>	−289.8	−288.1	0.47 <sup>3</sup>	9.53 <sup>5</sup>
[Cd <sub>2</sub> L] <sup>4+</sup>	−277.1	−275.2	0.49 <sup>3</sup> 0.23 <sup>4</sup>	9.57 <sup>5</sup> 6.79 <sup>6</sup>

<sup>1</sup> Energies calculated by BP86 functional. <sup>2</sup> Energies calculated by B3LYP functional. <sup>3</sup> Distance of the metal ion from the mean plane containing the aliphatic nitrogen atoms (plane 1). <sup>4</sup> Distance of the metal ion from the mean plane containing the aromatic nitrogen and carbon atoms (plane 2). <sup>5</sup> Sum of M-N<sub>aliphatic</sub> distances. <sup>6</sup> Sum of M-N<sub>aromatic</sub> and M-C<sub>phenyl</sub> distances.

In the most stable mononuclear [ZnL]<sup>2+</sup> and [CdL]<sup>2+</sup> geometries (Figures 10 and S17), the metal ion interacts with the ligand L through the four nitrogen atoms of the polyamine chain without involving the BBzB fragment. In contrast, BBzB is involved in the coordination of the second metal ion in the dinuclear complexes [Zn<sub>2</sub>L]<sup>4+</sup> and [Cd<sub>2</sub>L]<sup>4+</sup> (Figures 10 and S17), through two nitrogen atoms of the benzoxazole rings and the central phenyl ring. This latter M⋯CH interaction could be described as an agostic interaction (i.e., 3-center–2-electron M–H–C bonds), where the coordinatively unsaturated zinc and cadmium ions can interact with the closer aryl C–H bond, with the transition metal cation drawing the ligand towards itself and forming an additional  $\sigma$ -bond [66,67].

Both free ligand and mononuclear complexes feature an overall “chair” conformation, while the dinuclear complexes adopt a more extended conformation (Figures 10 and S17).

The aromatic BBzB fragment, notwithstanding the slight distortion induced by the coordination of the metal ions, maintained a coplanar structure, which explains the redshifted emission due to the  $\pi$ -system extension observed for L (vide supra).

The Gibbs free energies for the formation of the complexes (i.e., the difference between the energy of the complex and the sum of the energy of the ligand and the metal ion placed at an infinite distance; Equations (1) and (2)) were calculated and compared (as shown in Table 1).



Based on these results, the Zn<sup>2+</sup> complexes are more stable than the Cd<sup>2+</sup> ones, and the dinuclear [Zn<sub>2</sub>L]<sup>4+</sup> is the most stable among all the studied species, in agreement with the experimental results. This is probably due to the different dimensions of the two metal cations (ionic radius of 0.6 vs. 0.78 Å for Zn<sup>2+</sup> and Cd<sup>2+</sup>, respectively): the larger the ionic radius of the metal cation, the less stable the complex. Interestingly, while in the case of the Zn<sup>2+</sup> complexes, the dinuclear species is more stable than the mononuclear one, in the case of Cd<sup>2+</sup> complexes, the situation reverses, with the mononuclear [CdL]<sup>2+</sup> species being the most stable. Again, these results are probably connected to the dimension of the metal ions: in the case of the smaller Zn<sup>2+</sup>, two ions can be lodged in the macrocycle cavity of L via the interaction with the nitrogen atoms of the BBzB and the polyamine chain fragments, without significant distortions of the macrocycle, gaining stability by moving from one to two Zn<sup>2+</sup> ions being coordinated. Conversely, in the case of the bigger Cd<sup>2+</sup>, the coordination of the second metal ion determines an energetically expensive distortion of the phenyl ring that is not adequately balanced by the favorable energetic contribution provided by the coordination of a second metal ion. As a consequence, in agreement with the experimental results, the [Cd<sub>2</sub>L]<sup>4+</sup> complex is the least stable complex within the series (Table 1). Indeed, the Zn<sup>2+</sup> ions are located closer to both mean planes containing the four nitrogen atoms of the aliphatic polyamine chain (plane 1) and the two nitrogen atoms and

the aryl CH of the BBzB moiety (plane 2) than the  $\text{Cd}^{2+}$  ions (DIST1, Table 1). Analogous energetic considerations can be drawn out for both BP86 and B3LYP functionals.

Finally, the sum of the coordination bond distances could give an insight of the macrocyclic hole size: the presence of different metal cations revealed the ability of the ligand to adapt to their size. Indeed, the bond sum is always lower for the  $\text{Zn}^{2+}$  complexes than for the  $\text{Cd}^{2+}$  complexes (DIST2, Table 1).

#### 4. Conclusions

The cyclophane macrocycle **L** containing the new 1,3-bis(benzo[*d*]oxazol-2-yl)phenyl (BBzB) fluorophore as sensing unit and an aliphatic polyamine chain was synthesized. **L** behaves as a PET-mediated chemosensor, being fluorescent only at acidic pH, with a maximum at 402 nm. **L** is able to signal the presence of  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  metal ions via a fluorescent response, in a mixed acetonitrile–aqueous medium at physiological pH 7.4, via a CHEF effect. Both experimental and theoretical studies suggested the formation of both mononuclear and dinuclear complexes in the case of  $\text{Zn}^{2+}$ , while only the mononuclear complex was detected in the case of  $\text{Cd}^{2+}$ . The redshifted emission wavelength together with the theoretical studies suggested that the whole aromatic system remained coplanar upon metal coordination.

Future developments may involve the variation of the central phenyl ring of the fluorophore moiety or of the polyamine chain, to switch the selectivity towards bigger or smaller metal cations or to use the complexes as metal receptors for anions of neutral species.

**Supplementary Materials:** The following supporting information can be downloaded at <https://www.mdpi.com/article/10.3390/chemosensors10050188/s1>. Figure S1:  $^1\text{H}$  NMR spectrum of **4** in  $\text{CDCl}_3$ , Figure S2:  $^{13}\text{C}$  NMR spectrum of compound **4** in  $\text{CDCl}_3$ , Figure S3: MS-ESI spectrum of compound **4**, Figure S4:  $^{13}\text{C}$  NMR spectrum of ligand **L** in  $\text{D}_2\text{O}$ .  $[\text{L}] = 7.7 \times 10^{-3} \text{ mol dm}^{-3}$ , Figure S5: HRMS spectrum of **L**, Figure S6:  $^1\text{H}$  NMR spectrum of **L** at pH 12 in  $\text{D}_2\text{O}$ .  $[\text{L}] = 5 \times 10^{-3} \text{ mol dm}^{-3}$ . See Figure 1 in the main text for proton assignment, Figure S7: Trend of selected  $^1\text{H}$  NMR signals of aromatic **L** protons in an acetonitrile– $d_3/\text{D}_2\text{O}$  60:40 solution as a function of the pH.  $[\text{L}] = 5 \times 10^{-3} \text{ mol dm}^{-3}$ , Figure S8: UV-Vis absorption spectra of **L** registered upon addition of 2 equivalents of different metal cations ( $\text{Zn(II)}$ ,  $\text{Cd(II)}$ ,  $\text{Pb(II)}$ ,  $\text{Hg(II)}$ ,  $\text{Cu(II)}$ ).  $[\text{L}] = 3.2 \times 10^{-6} \text{ mol dm}^{-3}$  (ACN–aqueous HEPES pH 7.4, 4:1 *v/v* solvent mixture), Figure S9: Bar-plot showing the emission of **L** upon addition of 2 equivalents of different metal cations ( $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Ag}^+$ ,  $\text{Mn}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Na}^+$ ,  $\text{K}^+$ ). The response to  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  has been reported as a reference.  $I$  and  $I_0$  are the emission intensities of the ligand in the presence and absence of the metal ions, respectively.  $[\text{L}] = 3.2 \times 10^{-6} \text{ mol dm}^{-3}$  (ACN–aqueous HEPES (pH 7.4) 4:1 *v/v* solvent mixture).  $\lambda_{\text{ex}} = 305 \text{ nm}$ .  $\lambda_{\text{em}} = 385 \text{ nm}$  ( $\text{Zn}^{2+}$ ),  $390 \text{ nm}$  ( $\text{Cd}^{2+}$ ),  $402 \text{ nm}$  for all other tested metal ions, Figure S10: Trend of the emission intensity at  $385 \text{ nm}$  ( $\lambda_{\text{ex}} = 305 \text{ nm}$ ) upon increasing amounts of  $\text{Zn}^{2+}$ , as described in the main text. Inset: linear regression from 0 to 0.4 ppm, Figure S11: Trend of the emission intensity at  $390 \text{ nm}$  ( $\lambda_{\text{ex}} = 305 \text{ nm}$ ) upon increasing amounts of  $\text{Cd}^{2+}$ , as described in the main text. Inset: linear regression from 0 to 0.45 ppm, Figure S12: Molecular structures of **L** (a) and the similar systems cited in the main text (b–d), Figure S13:  $^1\text{H}$  NMR spectra of **L** recorded in  $\text{D}_2\text{O}$  at pH 7.4 upon addition of  $\text{Zn}(\text{ClO}_4)_2$  up to 3 equivalents.  $[\text{L}] = 5 \times 10^{-3} \text{ mol dm}^{-3}$ , Figure S14:  $^1\text{H}$  NMR spectra of **L** recorded in  $\text{D}_2\text{O}$  at pH 7.4 upon addition of  $\text{Cd}(\text{ClO}_4)_2$  up to 2 equivalents.  $[\text{L}] = 5 \times 10^{-3} \text{ mol dm}^{-3}$ , Figure S15:  $^1\text{H}$  NMR spectra of **L** (bottom), **L** + 2 equivalents of  $\text{Zn}^{2+}$  (middle), and **L** + 2 equivalents of  $\text{Zn}^{2+}$  + 2 equivalents of  $\text{Cd}^{2+}$  (top) recorded in  $\text{D}_2\text{O}$  at pH 7.4.  $[\text{L}] = 5 \times 10^{-3} \text{ mol dm}^{-3}$ , Figure S16:  $^1\text{H}$  NMR spectra of **L** (bottom), **L** + 1 equivalents of  $\text{Cd}^{2+}$  (middle), and **L** + 1 equivalents of  $\text{Cd}^{2+}$  + 1 equivalents of  $\text{Zn}^{2+}$  (top) recorded in  $\text{D}_2\text{O}$  at pH 7.4.  $[\text{L}] = 5 \cdot 10^{-3} \text{ mol dm}^{-3}$ , Figure S17: Front view (top) and side view (bottom) of the computed structures of (from left to right)  $[\text{CdL}]^{2+}$  and  $[\text{Cd}_2\text{L}]^{4+}$ , Table S1: Experimental parameters of **L** and similar systems and their complexes with  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$ , where applicable. Supplementary Materials Refs. [12,14,49–56,61,62].

**Author Contributions:** Conceptualization, L.G.; investigation, D.P., E.M., M.F. and M.V.; writing—original draft preparation, E.M.; writing—review and editing, E.M., V.F., L.G., M.F. and L.C.; supervision, V.F.; funding acquisition, V.F. All authors have read and agreed to the published version of the manuscript.

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**Conflicts of Interest:** The authors declare no conflict of interest.

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