

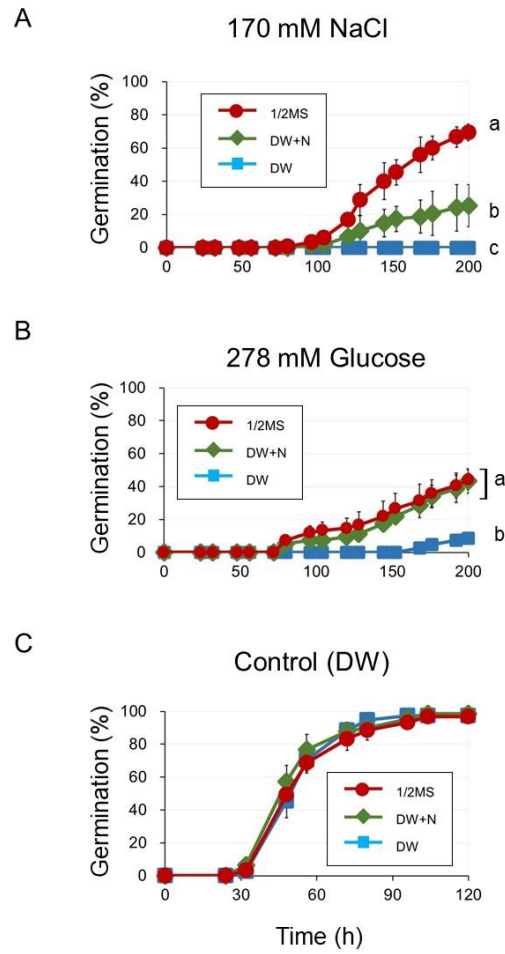


Figure S1. Seed germination on 1/2 MS, nitrogen components of 1/2 MS (DW+N), and DW in the presence of (A) 170 mM NaCl, (B) 278 mM glucose, and (C) no additional compounds (control). Freshly harvested seeds were incubated at 23 °C without stratification. Data are means $\pm$ SD for three replicates. Each replicate containing 50 seeds. Different letters indicate significant differences in germination at the final time point. (ANOVA and Tukey's test, $p < 0.05$).

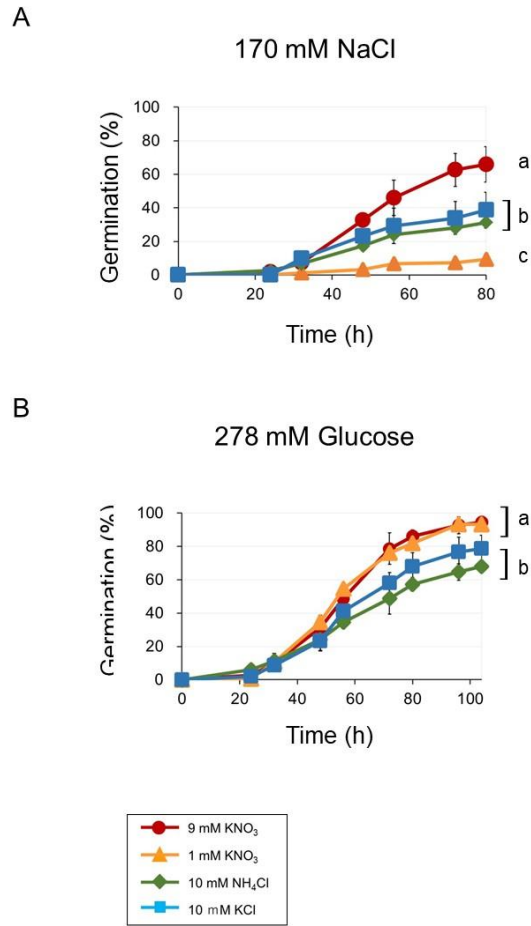


Figure S2. The effect of KCl and NH₄Cl on seed germination in the presence of (A) 170 mM NaCl and (B) 278 mM Glucose. Seeds were stratified for three days before incubation at 23 °C. Germination was scored after transferred to 23 °C. Data are means $\pm$ SD for three replicates. Each replicate containing 50 seeds. Different letters indicate significant differences in germination at the final time point. (ANOVA and Tukey's test, $p < 0.05$).

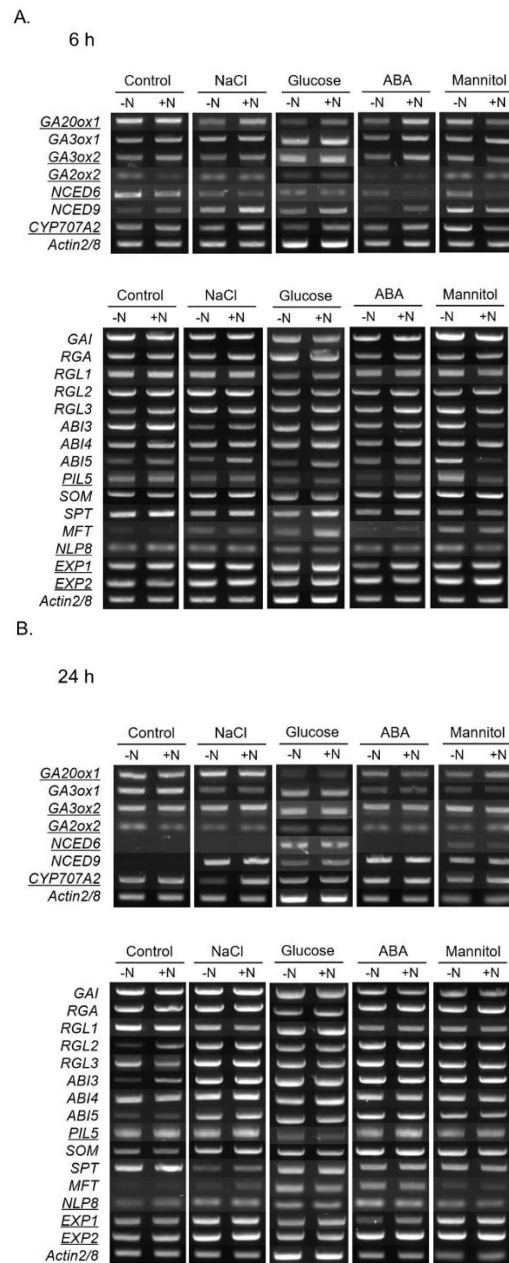


Figure S3. Expression of genes involved in the regulation of seed germination in the presence in the presence (+N) or absence (-N) of 10 mM KNO₃. Seeds were imbibed in the presence of 170 mM NaCl, 278 mM Glucose, 5 mM ABA, and 500 mM Mannitol for (A) 6 and (B) 24 h. The gene expression was analyzed by semi-quantitative RT-PCR. The Actin 2/8 gene was used as control. The gene amplification reactions were carried out for 30 PCR cycles or 35 PCR cycles (underlined genes).

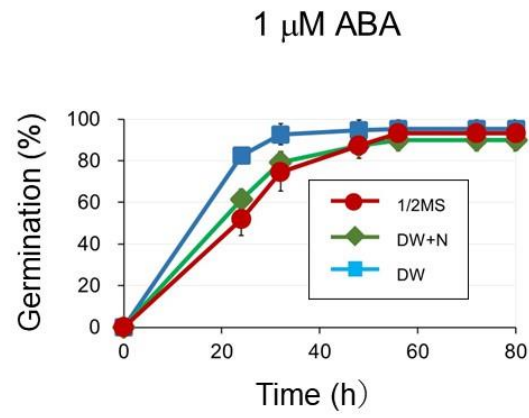


Figure S4. Seed germination on 1/2 MS, nitrogen components of 1/2 MS (DW + N), and DW in the presence of 1 μ M ABA. Seeds were stratified for two days before incubation at 23 °C. Data are means $\pm$ SD for three replicates. Each replicate containing 50 seeds.

Table S1. List of genes used in this study.

Gene Name	Protein	Key role	References
<i>GA20ox1 (At4g25420)</i> <i>GA20-OXIDASE 1</i>	gibberellin 20-oxidase	GA biosynthesis	[47,48]
<i>GA3ox1 (At1g15550)</i> <i>GA3-OXIDASE 1</i>	Gibberellin 3-b-dioxygenase	GA biosynthesis	[49] [50]
<i>GA3ox2 (AT1G80340)</i> <i>GA3-OXIDASE 2</i>	Gibberellin 3-b-dioxygenase	GA biosynthesis	[49]
<i>GA2ox2 (AT1G30040)</i> <i>GA2-OXIDASE 2</i>	Gibberellin 2-b-dioxygenase	GA catabolism	[51,52]
<i>NCED6 (AT3G24220)</i> <i>NINE-CIS- EPOXYCAROTENOID DIOXYGENASE 6</i>	9-cis-epoxycarotenoid dioxygenase	ABA biosynthesis	[53]
<i>NCED9 (AT1G78390)</i> <i>NINE-CIS- EPOXYCAROTENOID DIOXYGENASE 9</i>	9-cis-epoxycarotenoid dioxygenase	ABA biosynthesis	[53]
<i>CYP707A2 (AT2G29090)</i> <i>ABA 8'-HYDROXYLASE CYTOCHROME P450-2</i>	Absciscic acid 8'-hydroxylase	ABA degradation	[54,55]
<i>GAI (AT1G14920)</i> <i>GA INSENSITIVE</i>	DELLA transcription factor	GA signaling	[56]
<i>RGA (AT2G01570)</i> <i>REPRESSOR OF GAI-3</i>	DELLA transcription factor	GA signaling	[57]
<i>RGL1 (AT1G66350)</i> <i>RGA-LIKE 1</i>	DELLA transcription factor	GA signaling	[58]
<i>RGL2 (AT3G03450)</i> <i>RGA-LIKE 2</i>	DELLA transcription factor	GA signaling	[59]
<i>RGL3 (AT5G17490)</i> <i>RGA-LIKE 3</i>	DELLA transcription factor	GA signaling	[60]
<i>ABI3 (AT3G24650)</i> <i>ABA INSENSITIVE 3</i>	b-ZIP transcription factor	ABA signaling	[60]
<i>ABI4 (AT2G40220)</i> <i>ABA INSENSITIVE 4</i>	AP2 transcription factor	ABA signaling	[61]
<i>ABI5 (AT2G36270)</i> <i>ABA INSENSITIVE 5</i>	b-ZIP transcription factor	ABA signaling	[60]
<i>PIL5 (AT2G20180)</i> <i>PHYTOCHROME INTERACTING FACTOR 3-LIKE 5</i>	PIF1, bHLH transcription factor	light signaling	[62,63]
<i>SOM (AT1G03790)</i> <i>SOMNUS</i>	zinc finger CCCH-type transcription factor	seed germination	[12]
<i>SPT (AT4G36930)</i> <i>SPATULA</i>	bHLH transcription factor	floral organogenesis.seed germination	[64]
<i>MFT (AT1G18100)</i> <i>MOTHER OF FT AND TFL1</i>	phosphatidylethanolamine-binding protein	seed dormancy, germination	[65]
<i>NLP8 (AT2G43500)</i> <i>NIN-LIKE PROTEIN 8</i>	NIN-like transcription factor	nitrate signaling	[19]
<i>EXP1 (AT1G69530)</i> <i>EXPANSIN 1</i>	expansin	loosening and extension of plant cell walls	[66]
<i>EXP2 (AT5G05290)</i> <i>EXPANSIN 2</i>	expansin	loosening and extension of plant cell walls	[67]

Table S2. Primers used in this study.

Semi-quantitative RT-PCR (GA/ABA metabolism)	
GA20ox1 F	ATGGCCGTAAGTTTCGTAAC
GA20ox1 R	TTAGATGGGTTTGGTGAGCC
GA3ox1 F	ATGCCTGCTATGTTAACAG
GA3ox1 R	TCATTCTTCTCTGTGATTTC
GA3ox2 F	ATGAGTTCAACGTTGAGCG
GA3ox2 R	TTAATTTCTAATAATGGAAAG
GA2ox2 F	ATGGTGGTTTGGCCACAGCC
GA2ox2 R new	TCATACAAGGGTTTATGATTG
NCED6 F	ATGCAACACTCTCTTCGTTC
NCED6 R	TCAGAAAACCTGTTCCTTCAAC
NCED9 3rd ATG- F	ATGGCTTCTACTACCTTACT
NCED9 R	CTAACACAAAGCTTGCTTCG
CYP707A2 F	ATGCAAATCTCATCTTCATC
CYP707A2 R	TTAAATCGGGGTTACTCTTATTG
Semi-quantitative RT-PCR (GA/ABA signaling)	
Y GAI EcoRI F	GGAATTCATGAAGAGAGATCATC
Y GAI BamHI R	GGGATCCCTAATTGGTGGAGAGTTTCC
RGA1 XbaI F	GTCTAGAATGAAGAGAGATCATCACCATTCCAAG
RGA1 BamHI R	GGGATCCTCAGTACGCCGCCGTCGAGAGTTTCCAAGC
Y RGL1 BamHI F	GGGATCCCGATGAAGAGAGAGCACAACCAC
Y RGL1 SalI R	GGTCGACTTATTCCACACGATTGATTTC
Y RGL2 SalI F	GGTCGACCGATGAAGAGAGGATACGGAGAAAC
Y RGL2 PstI R	GCTGCAGTCAGGCGAGTTTCCACGCCG
Y RGL3 SalI F	GGTCGACGATGAAACGAAGCCATCAAG
Y RGL3 PstI R	GCTGCAGCTACCGCCGCAACTCCGCCG
ABI3 XbaI F	GTCTAGAATGAAAAGCTTGCATGTGGC
ABI3 BamHI R	GGGATCCTCATTTAACAGTTTGAGAAG
ABI4 XbaI F for effector	GTCTAGAATGGACCCCTTTAGCTTCCC
ABI4 KpnI R for effector	GGGTACCTTAATAGAATTCCCCCAAG
ABI5 BamHI F	GGGATCCATGGTAAGTACGAGAAACGAAG
ABI5 KpnI R	GGGTACCTTAGAGTGGACAACCTCGGGTTC
SPATULA KpnI F	CGGTACCATGATATCACAGAGAGAAG
SPATULA EcoRI R	GGAATTCTCAAGTAATTCGATCTTTTAG
Y MFT EcoRI F	GGAATTCATGGCGGCTTCTGTGTGATCC
Y MFT SalI R	GGTCGACCTAGCGTCTGCGTGAAGCAG
PIL5 XbaI F	GTCTAGAATGCATCATTTTGTCCCTG
PIL5 KpnI R	GGGTACCTTAACCTGTTGTGTGGTTTC
SOMNUS BamHI F	GGGATCCATGGATGTCGTTTGTACGG
SOMNUS KpnI R	GGGTACCTCAAGTCAAGAGATCATTG
EXP1 F	ATGGCTCTTGTACCTTCTTG
EXP1 R new	TCAAGCACTCGAAGCACCAC
EXP2 F	ATGAATCTTACAGAATATTC
EXP2 R	CTAAAATTGTCCGCCTTC
NLP8_1500bp_F	TTTGAGAACTGTTTCAGAAG
NLP8_2000bp_R	TCGCCAGTTGCTGAGTCGAAC
Actin2/8 F	CTGGTGATGGTGTGTCTCAC
Actin2/8 R	GACCTTAATCTTCATGCTGC
Quantitative RT-PCR	
Real-time GA20ox1(Roche) F	CATGGGTTTCAGCCATTG
Real-time GA20ox1(Roche) R	CTCTAAAGTAGTCCCGTTTACGC
Real-time GA2ox2(Roche) F	CTCCTCCTCAATGCTAATCCTC
Real-time GA2ox2(Roche) R	CTCCTCCACCGACTCACG
Real-time NCED6(Roche) F	TAGCAATAGCCGATCCTTGG
Real-time NCED6(Roche) R	GCCGTTTGTATATCTACCTTCG
Real-time NCED9(Roche) F	AAAGTATATTTACGGAGAGGGGAAAT
Real-time NCED9(Roche) R	CGTCACCGGAAGGTAGAAAC
Real-time CYP707A2(Roche) F	GCTTCTCAAGTACTTACACGACCA
Real-time CYP707A2(Roche) R	TGTCGAATGCTGAATTGCTC
Real-time PIL5 (Roche) F	AGCGAGGAAACAAAACAAGC

Real-time PIL5 (Roche) R	TCATTGATCCTATCTCTCCGTTT
Real-time SOM (Roche) F	GAGAACGTTTCCGGTGCTC
Real-time SOM (Roche) R	CAAGCAACATGGATTCTTCG
Real-time SPT (Roche) F	GCAGAGCTGCTGAAGTTCATAAT
Real-time SPT (Roche) R	TTTGAATTAGGGATGAGACTTTGTAA
Real-time MFT F (Xi et al. 2010)	CGAGCCGAACATGAGAGAAT
Real-time MFT R (Xi et al. 2010)	AAGTATCTCTTTTCCTCTTGAGGG
Real-time ABI3 (Roche) F	GCTGGCTCAGCTTCTGCTAT
Real-time ABI3 (Roche) R	AAATTCTTTCTGGTTCCATCC
Real-time ABI4 (Roche) F	CCGCTTCTTCTCCTTCCAC
Real-time ABI4 (Roche) R	GAGGGAGGAGAGGTCTTAGGG
Real-time ABI5 (Roche) F	TGGAGAGAAGACAGAGGAGGA
Real-time ABI5 (Roche) R	GTTCACTTCCAATTCCACTG
Real-time RGL2 (Roche) F	CTTCTGCGTTTCCAAAGGA
Real-time RGL2 (Roche) R	TCGGATCCTCTTGCTGCTA
Real-time NLP8 (Roche) F	AGGCACTTTGGAGCAGGA
Real-time NLP8 (Roche) R	GCTCACATTTTCTCTGTAGTGCT
Real-time EF1aA4 (Roche) F	CTTGGTGCAAGCAGATGATT
Real-time EF1aA4 (Roche) R	CGTACCTAGCCTTGGAGTATTG

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