

Review



Drought Stress Effects and Olive Tree Acclimation under a Changing Climate

Cátia Brito, Lia-Tânia Dinis, José Moutinho-Pereira and Carlos M. Correia *

CITAB—Centre for the Research and Technology of Agro-Environmental and Biological Sciences, University of Trás-os-Montes and Alto Douro, 5000-801 Vila Real, Portugal

* Correspondence: ccorreia@utad.pt

Received: 6 June 2019; Accepted: 15 July 2019; Published: 17 July 2019

Abstract: Increasing consciousness regarding the nutritional value of olive oil has enhanced the demand for this product and, consequently, the expansion of olive tree cultivation. Although it is considered a highly resilient and tolerant crop to several abiotic stresses, olive growing areas are usually affected by adverse environmental factors, namely, water scarcity, heat and high irradiance, and are especially vulnerable to climate change. In this context, it is imperative to improve agronomic strategies to offset the loss of productivity and possible changes in fruit and oil quality. To develop more efficient and precise measures, it is important to look for new insights concerning response mechanisms to drought stress. In this review, we provided an overview of the global status of olive tree ecology and relevance, as well the influence of environmental abiotic stresses in olive cultivation. Finally, we explored and analysed the deleterious effects caused by drought (e.g., water status and photosynthetic performance impairment, oxidative stress and imbalance in plant nutrition), the most critical stressor to agricultural crops in the Mediterranean region, and the main olive tree responses to withstand this stressor.

Keywords: climate change; drought; Olea europaea; recovery; tolerance mechanisms

1. Introduction

The olive tree (Olea europaea L.) is one of the emblematic crops of the Mediterranean region, where most of the world's olive oil is produced [1]. Olive oil is widely known as the main source of fat in the so-called Mediterranean diet, being related to several beneficial effects on human health, due to its balanced fatty acid composition and antioxidant properties. Therefore, the consumption and demand for olive oil is increasing all over the world [1]. The Mediterranean region is characterized by severe summer conditions, including low rainfall, excessive heat load and high daily irradiance. Among the constituents of summer stress, drought is usually the most critical, although it is highly exacerbated by the others. Moreover, the Mediterranean region is a particularly susceptible area to climate change, a major challenge for agriculture [2]. According to the Intergovernmental Panel on Climate Change (IPCC) [2], climate change scenarios predict that the temperature will rise and precipitation patterns will shift, leading to higher evaporative demand and decreased soil water availability. Moreover, night-time temperatures will increase to a greater extent than daytime temperatures and the frequency and severity of drought and heat wave spells are likely to increase as well. These environmental factors cause adverse pleiotropic effects on plants' growth and development. Specifically, a water deficit has negative repercussions on water relations, nutrient uptake, carbon assimilation, canopy dimension, oxidative pathways, phenology and reproduction processes [1–9] and, thus, affect crop yield and quality [10–12]. Meanwhile, some of these plant responses to adverse conditions are connected with defence adaptation strategies. Although the olive tree is a well-adapted species against drought [4,5,13–16], considerable energy resources will be used in these protective processes, compromising plant growth and productivity [16].

In the predicted scenarios of climate change, the risks for the olive sector will increase, particularly under rainfed conditions, which may jeopardize its economic viability. This may lead to the abandonment of traditional groves, with devastating socioeconomic (e.g., income and employment reduction in marginal regions) and environmental (e.g., soil erosion, increased risk of wildfires, changes in wildlife communities) consequences. On the other hand, the carbon sequestered by olive tree orchards (biomass + soil) could surpass the emissions created from farming operations and oil manufacturing, with the carbon footprint value (calculated for the production and sale of olive oil) being very low or even negative [17]. This outcome claims worldwide importance in that olive tree plantations may have significant CO₂ sinks and mitigate greenhouse gas emissions caused by farming activities [17]. Thus, it is important to act to make this crop more sustainable, productive and resilient under severe adverse conditions, which are likely to be exacerbated in the Mediterranean region. Understanding how olive trees respond to drought stress is the first step to improving its profitability, allowing the selection of more resistant cultivars and identification of tolerant characteristics useful in breeding programs and in genetic engineering, as well as the development of accurate adaptation strategies according to necessities. In this context, the implications of climate change projections and an overview of the optimum conditions for olive cultivation are provided. In the following, the impact of drought on the morphological, physiological and biochemical traits, as well the acclimatisation responses of the olive tree to this stressor are critically discussed.

2. Olive Tree Growth Conditions and Distribution

The olive tree, belonging to the botanical family Oleaceae and genus *Olea* [18], is one of the oldest cultivated plants native of the Mediterranean Basin [19]. Ever since, it has contributed to the economy, health, nutrition, culture and sustainability of this region. Although the Mediterranean region remains the main area of cultivation, nowadays, this area extends to southern Africa, South and North America, Australia, Japan and China [1]. Olive cultivation worldwide is limited by edaphoclimatic factors of Mediterranean isoclimatic zones lying between the 30th and 45th parallels on the northern and southern hemispheres [20]. The Mediterranean climate is typically mild and wet during the winter and hot and dry during the summer [21], being the Mediterranean area, it is usually also exposed to high daily irradiances, including UV radiation. Temperature is the most significant environmental factor that limits olive growing areas, while water availability is the most significant factor that limits olive yield.

Proper olive cultivation areas have a mean annual temperature of 15–20 °C, with a minimum of 4 °C and a maximum of 40 °C [18]. Usually, the optimum temperature for olive vegetative growth ranges between 10 °C and 30 °C, while carbohydrate synthesis occurs at higher rates at temperatures ranging from 20 °C to 30 °C [22]. Olive trees require a period of low temperatures (0–7 °C) for flowering bud differentiation [18]. On the other hand, temperatures constantly above 16 °C prevent bud differentiation [22]. However, the minimum temperature should not drop below –7 °C, which can seriously damage trees, and if the temperature reaches –12 °C, can kill them. High altitudes (>800 m) are not appropriate for olive cultivation, due to the incidences of frost and the short vegetative period in those locals [18].

Despite being able to grow well even in poor, dry, calcareous and gravelly soils, the best conditions for olive tree annual bearing are deep, sandy-loam adequately supplied with nitrogen, phosphorus, potassium and water [18], while the optimal pH values range between 5.5 and 8.5 [23]. Although in some cases, olive trees can grow with a rainfall of 200 mm year⁻¹ [24], it should be above 400 mm year⁻¹, and values of 600 mm year⁻¹, 800 mm year⁻¹ and 1000 mm year⁻¹ are considered sufficient, moderate and good, respectively [22]. Still, 500 mm year⁻¹ is the lower limit for commercial olive yields under rainfed conditions [25].

Under low levels of photosynthetic photon flux density (PPFD), the percentage of flower bud induction and differentiation falls, and the same occurs with net photosynthetic rate (A). For the

majority of the olive leaves, the ideal PPFD, depending on the genotype, must be in the range between 600 and 1000 μ mol m⁻² s⁻¹, the light saturation point. On the other hand, PPFD of olive leaves must be above 20–30 μ mol m⁻² s⁻¹, the light compensation point, to obtain higher assimilation rates than respiration rates [25].

The growing awareness of the nutritional value of olive oil has led to the expansion of olive tree cultivation [1]. The total estimated global land-use area for cultivating olive trees was over 10.6 and 10.8 million ha in 2016 and 2017, respectively. In both years, Spain was the country with a higher total harvesting area, followed by Tunisia, Italy, Morocco and Greece [26].

Globally, olive production was 20,344,597 and 20,872,788 tons in 2016 and 2017, respectively. Spain was 1st place, followed by Greece, Italy, Turkey and Morocco [26]. About 90% of the world's olives production is for oil extraction and the remaining 10% for table olives [27]. Almost 92% of the world's olive oil production comes from the Mediterranean region, with European Union countries (i.e., Spain, Italy, France, Greece and Portugal) responsible for 67% of global production [1]. To increase production, large areas were irrigated and fertilized, trees were adjusted to mechanical pruning and harvesting, and new orchards were planted in high and super-high-density plots [19].

3. Implication of the Change in Environmental Conditions for the Olive Tree

The impacts of recent extreme climate-related events, such as heat waves, droughts, floods, and wildfires, have revealed the significant vulnerability and exposure of some ecosystems to current climate variability [2]. However, while the above records are concerned, the forecasted scenarios may be worse, accounting with global temperatures rising, with special prominence at night-time, and increase in extreme events intensity and frequency [2]. Due to the uniqueness of its geographic location—in a transition zone between the arid climate of North Africa and the temperate and rainy climate of central Europe—the Mediterranean Basin is particularly vulnerable to present and future climate variability and climate change [23]. According to the Fifth Assessment Report (AR5) of the Intergovernmental Panel on Climate Change [2], the projections for the Mediterranean region also show warming in all seasons, especially in summer. Precipitation is not projected to change or will be moderately reduced in the winter half year (October to March), while it will be markedly reduced in the summer half year (April to September). The length, frequency and/or intensity of warm spells or heat waves are very likely to increase throughout the region.

The effects of weather and climate in agriculture can be felt at different levels, as changes in CO₂ atmospheric concentration, temperature and water resource availability, among other factors, affect plants' development and productivity, the possibility to execute agricultural operations and the geographical distribution of crops.

It is assumed that the CO₂ assimilation rate and olive yield will decrease substantially in the context of climate change [28]. Regarding the increase in atmospheric CO₂ concentration, a crop model projected an increase in the potential assimilation rate, and thus, in the overall productivity, even if reduced water availability controls and limits this tendency [29]. In fact, it is known that stomatal conductance (g_s) decreases with increasing CO₂ levels, while photosynthesis increases, leading to greater water use efficiency of several Mediterranean species, including the olive tree [30]. On the other hand, greater CO₂ concentrations will promote weed growth, and therefore, competition with olive crops, although the nature of the damaging effects depends on the weed species [18]. An increase in other atmospheric pollutants is also expected, such as tropospheric ozone [16], whose increase has already been shown to cause reductions of g_s and A in olive trees [31]. In addition, the co-occurrence of other stressing factors (temperature increase and consequent increase in evapotranspiration and water demand; the decrease in water availability and the increase in saline water use for irrigation) will overcome the influence of increased atmospheric CO₂ on A [30].

The expected increases in spring and autumn temperatures will prolong the growing season [32]. If there is adequate water availability during these periods, a general anticipation of a flowering date, of 1 to 2 weeks, could be expected [33]. On the other hand, very high temperatures could be catastrophic to flowering quality and, therefore, they could contribute to a lower olive yield [18]. In fact, it has been observed that trees exposed to insufficient chilling temperatures and high

temperature events can flower, but the flowers are of low quality and have a low set percentage. This phenomenon has been documented in olive growing areas at low latitudes, where some olive varieties produce deformed floral buds and fruit [34]. Higher temperatures and evapotranspiration also accelerate fruit ripening, claiming the necessity to harvest early, probably at a lower maturity index than used today [35].

Warmer conditions will determine a possible north range expansion of cropping activities into regions where lower temperature was a limitation in the past [2]. Conversely, a reduction in the southerly cropping areas and crop yields will be expected. Indeed, potentially cultivable areas for olive cultivation are expected to extend northward and to higher altitudes [36,37], increasing by 25% in 50 years [37]. These shifts are expected due to the enlargement of the growing season in winter and because some southern areas will not satisfy the minimum chilling requirements [36]. The low temperature role in releasing dormancy of potentially reproductive olive tree buds was demonstrated in several studies [38–40], highlighting that 7.2 °C was sufficient to complete chilling requirements, while 12.5 °C provided both chilling requirement fulfilment and adequate temperature for subsequent floral bud growth and differentiation. Moreover, areas along the Atlantic shores may become viable very quickly, due to the increasingly milder winters [41]. For all these reasons, olive trees may be considered as one of the best bioindicators of climate evolution in the Mediterranean Basin [41].

Warming will also boost pest and disease generations [18]. In particular, warming will affect olive fly infestation levels across the Mediterranean Basin, altering olive production and decreasing the profitability of small olive farms in many marginal areas of Europe and elsewhere in the basin [42].

4. Drought Effects in Plants Morphological, Physiological and Biochemical Mechanisms

Drought is considered the most limiting factor for agricultural productivity worldwide [43]. In plants, water deficits occur when there is not enough water to absorb in order to replace water losses by evapotranspiration, or when having difficulty absorbing water (e.g., saline soils, low temperature, flooding). Consequently, plants then have a lower amount of water than it contains when in a state of maximum hydration, triggering a variety of physiological and biochemical responses at the cellular and organism levels. Thus, this section discusses in detail how water deficits affect plants, including the olive tree, their morphological characteristics and physiological and biochemical mechanisms, in order to further understand the specific olive tree mechanisms to deal with drought imposition.

4.1. Influence on Water Status, Growth and Plant Morphology

With drought imposition, as plant water content decreases, the cells shrink, and the cell wall relaxes, resulting in loss of turgor [44], causing a reduction in leaf water potential (Ψ) and in cell division and expansion [8]. If a water deficit is imposed early in the development, the inhibition of cell expansion results in a reduced leaf area, while if it is imposed after a substantial leaf area has developed, leaves will senesce and can fall off [44]. The number of leaves can also be affected, associated with a decrease in the number of branches and growth rate [43,44]. These responses limit the photosynthetic area, and thus, contribute to the decline in whole-canopy photosynthesis [43].

During soil drying and/or when evaporative demand is high, leaf transpiration often exceeds water transport capacity. Thus, xylem water potential decreases, increasing the susceptibility to cavitation—the aspiration of air into the transpiration stream. Cavitation events can cause embolism when the air fills the entire conduit, blocking water transport, reducing the number of functional conduits and increasing hydraulic resistance [16].

4.2. Influence on Stomatal and Mesophyll Conductance, Photosynthesis, Respiration and Water Use Efficiency

One of the primary drought consequences is the regulation of the stomatal aperture to restrict water losses [45]. The olive tree presents a tight control of stomatal behaviour to maintain Ψ within

an adequate level, avoiding critical values and keeping them in a safe range to avoid embolism [15,46]. Stomatal regulation is influenced by both hydraulic and hormonal signals (abscisic acid, ABA), but may also vary under increasing drought and recovery conditions [46]. The actual role of each component in the stomatal control mechanism is not fully understood [45]. Hernandez-Santana et al. [45] found that in olive trees, the variable most strongly related to g_s response to water deficit is the leaf hydraulic conductance (K_{leaf}), which decreases exponentially with leaf water potential. Moreover, K_{leaf} starts to decline immediately with dehydration, while the drop in g_s began only after a substantial K_{leaf} loss, suggesting a protective role of the stomata for K_{leaf} maintenance. Torres-Ruiz et al. [46] also recognized that g_s decline in olive trees during water deficit was more related to the loss of hydraulic functioning at the most distal organs of the plant (i.e., roots and leaves) than to the increase of ABA levels on leaves, stems and roots. Still, ABA's effect on stomatal regulation should not be put aside, as it might promote stomatal closure in a dual way, by a direct biochemical action on guard cells and by an indirect hydraulic action through a decrease in leaf water permeability within leaf vascular tissues [47].

As stomatal aperture decreases, the entering of CO_2 into the mesophyll also decreases, with negative consequences on A [6,48]. Moreover, the mesophyll compactness increases under drought conditions to restrict water diffusion, what also restricts the supply of CO₂ to the carboxylation sites [49]. Thus, both diffusional limitations, i.e., stomatal (g_s) and mesophyll (g_m) conductance, contribute to A decline, showing a close relationship with each other [50,51]. Nonetheless, diffusional limitations to photosynthesis were not exclusively associated with leaf anatomical traits. In fact, especially in harsh environmental conditions, changes in leaf biomechanical and biochemical traits can lead to a reduction in mesophyll conductance to CO₂ [16,50], e.g., carbonic anhydrase (CA) and aquaporins (AQPs) [51]. Carbonic anhydrase is a zinc metalloenzyme that catalyses the reversible hydration of CO₂ [52], being involved in the maintenance of the equilibrium between CO₂ and HCO₃- [53]. Thus, CA may facilitate the diffusion of CO₂ through the chloroplast membrane and, therefore, the CO₂ conductance within the chloroplast [49,50]. The relationship between the increase of water deficit severity and the reduction in CA expression and g_m in olive leaves [51] suggests that g_m can be enzymatically regulated by the CA. Furthermore, Gillon and Yakir [54] demonstrated that CAmediated diffusion takes special importance in sclerophyllous species, such as the olive tree, which offers more resistance to CO₂ in their cell walls, being necessary to offset this limitation by optimizing the chloroplast conductance. There is also other evidence supporting the relationship between gm and AQPs, since some of them are involved in CO₂ transport [51,55]. Additionally, AQPs can also be involved in gs regulation [51,55] via their effects on water transport [56].

In response to moderately stressful conditions, g_s and g_m are the main limitations to A [16,57], but at severe stress levels the biochemical component of photosynthesis can also be inhibited [14,48,58]. Meanwhile, limitations in net CO₂ assimilation may lead to an overexcitation and subsequent photo-inhibitory damage of photosystem II (PSII), as demonstrated in several studies by the reduction of relevant photochemical traits, as the photosynthetic electron transport rate (ETR), effective quantum efficiency of photosystem II (Φ PSII), maximal quantum efficiency of photosystem II (F_v/F_m), capture efficiency of excitation energy by open PSII reaction centres (F'_v/F'_m) and photochemical quenching (qP) and by the increase of non-photochemical quenching (NPQ) [4,9,48,59,60]. In line with the detrimental effect of drought on the photochemical processes of olive leaves, a reduced abundance of several proteins related to the photosynthetic Calvin–Benson cycle was also observed (mainly Rubisco downregulation) [61].

Under conditions of drought and high light stress, the decrease in CO₂ assimilation is usually also associated with the increase in the photorespiration rate, owing to the nature of ribulose-1,5bisphosphate carboxylase/oxygenase (Rubisco) [43]. Nonetheless, under severe drought, photorespiration is involved in energy dissipation, reducing photoinhibition [62]. Severe drought can also cause the impairment of Rubisco and other photosynthetic enzymes [62,63], as well as the degradation of photosynthetic pigments [3,48,57]. A summary of the possible mechanisms that lead to a decrease in photosynthesis under drought is shown in Figure 1.

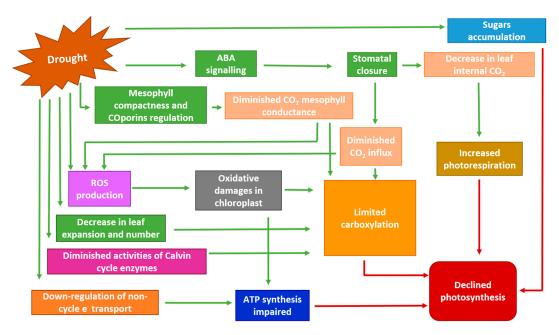


Figure 1. Possible mechanisms that lead to a decrease in photosynthesis under drought. ABA: abscisic acid; ROS: reactive oxygen species; ATP: adenosine triphosphate. Adapted from Farooq et al. [43].

Although unexpected as there is no opportunity for carbon gain [64], a substantial night-time stomatal conductance (g_{night}) and leaf transpiration (E_{night}) were observed in a wide range of species from different functional groups and ecosystems [65–67], including the olive tree [7]. This generalized response suggests potential benefits related to the continued water loss during the night. For instance, it was proposed that it may improve nutrient uptake [7,65], reduce foliar temperature, limit the consumption of storage products through respiration (R) [7], and maintain the sap flux for O₂ delivery for internal sapwood respiration and/or stem corticular photosynthesis [68]. Although the implication of overnight water loss in physiological processes remains unclear [66,67], the occurrence of this phenomenon affects the plant water balance and water-use efficiency [67], which may be detrimental under limited water conditions.

Respiration and A are strongly coupled and intrinsically interdependent because A provides photosynthetic substrates to R, and R supplies adenosine triphosphate (ATP) and carbon skeletons to sustain the requirements of plant energy processes [69]. However, in response to drought, the R trend is still not clear. Studies demonstrated that R varies, generally from inhibition, with low to moderate stress (due to the decrease of energy demand for plant growth and the impairment of some enzymes involved in R), to stimulation, with severe stress (due to the changes in metabolism to extra repair costs to offset serious damage) [70,71]. On the other hand, it was reported that olive trees are able to maintain reduced R rates during moderate to severe drought stress (relative water content of 64.7%) [7]. By reducing the metabolism, this species is able to conserve photosynthates, important for plant regrowth. Additionally, because phloem transport depends on turgor, the decrease in water potential in the phloem under severe drought may inhibit the amount of assimilates exported [44], and sugars accumulation in leaves may control photosynthesis by feedback processes [72].

Derived from changes in A and g_s, the intrinsic water-use efficiency (WUE_i), defined as A/g_s, will eventually be affected by water availability. Under mild to moderate drought, WUE_i typically increases [6], while it may decrease under severe drought conditions, as illustrated in olive trees by Bacelar et al. [4]. However, improving WUE_i may not necessarily result in improving whole plant water-use efficiency (WUE_{WP}) [73] and the whole plant carbon and biomass acquisition per amount of transpired water [74]. The difference in time-scale of both processes (from seconds to months) and non-accounted energy expenses in growth and maintenance in long-term water use can justify such differences [75]. Effectively, Bacelar et al. [5] reported an absence of a significant association between WUE_i and WUE_{WP} in different cultivars of olive trees under drought conditions.

4.3. Influence on Minerals Uptake and Allocation

Drought stress affects uptake, transport, and subsequent distribution of minerals within the plant [8], causing an imbalance in plant nutrition. This disequilibrium may cause important perturbations on physiological functions and on biomass accumulation [6,7], since minerals serve numerous functions in plants, as structural components in macromolecules, co-factors in enzymatic reactions, osmotic solutes and in the maintenance of charge balance in cellular compartments [76].

It is well accepted that reduced water availability results in limited nutrient uptake [77] since (i) drought reduces the whole-plant transpiration rate, due to the inferior stomatal conductance and total leaf area [7]; (ii) drought declines the soil water potential, slowing the diffusion rate of nutrients between the soil matrix and the root surface [8]; (iii) drought reduces the nutrient supply through mineralization [78]; (iv) drought impairs the activity of enzymes involved in nutrient assimilation, disturbing nutrient acquisition [40] and changing membrane permeability [77]; and (v) drought decreases the concentration of root nutrient-uptake proteins [79]. Although a lower tissue concentration of minerals is a regular response [8,77], this is not always necessarily true, as the concentration effect due to the lower production of biomass can be traduced in higher concentrations of nutrients. However, as reported previously with young olive trees, drought stress did not affect the uptake, transport, allocation and physiological use efficiency of all minerals to the same extent [80], suggesting an adaptive mechanism for growth under water limitation. For instance, drought increased both the concentration and the allocation of Mg and B to stems and roots, respectively, and increased the allocation of N to roots, at the expenses of leaves, and of P to leaves, at the expenses of stems. Phosphorus use efficiency was also improved by drought, suggesting a better distribution of phosphorus resources among the different metabolic processes involved in biomass production.

4.4. Influence on Redox Status

Drought increases the generation of reactive oxygen species (ROS) due to the accumulation of excess energy, which increases the photo-oxidative effect [6,61]. Usually, under mild and moderate water deficits, drought tolerant plants enhance the concentration of enzymatic and non-enzymatic antioxidants [4,63]. On the other hand, under severe drought stress, an imbalance between ROS production and the antioxidant defence system may occur [43], damaging lipids, proteins, carbohydrates, pigments and deoxyribonucleic acid (DNA) [3,4,8,9,59], which also results in increased cellular membrane damage and electrolyte leakage [7,61].

4.5. Influence on Hormonal Dynamics

Stress conditions often stimulate changes in the production, distribution or signal transduction of phytohormones. In fact, through the action of these molecules, plants respond to the adverse conditions modifying their physiology and biochemistry [81]. Usually considered the main stress hormone, ABA biosynthesis and accumulation is stimulated by drought, in association to a key role in g_s regulation, but a water deficit also influences other stress signalling pathways [47]. Thus, ABA is unequally distributed within and between the different plant organs [6,7,82,83]. Under drought stress, ethylene (ET), another important stress hormone, might inhibit plant growth [84], is involved in leaf abscission, and thus, in the reduction of water loss [85], induces remobilization of minerals from the leaves [84]; recent evidence also suggests that it might increase the accumulation of compatible solutes and reduce oxidative stress damage [86]. Moreover, it was suggested that under stress, ABA and ET act antagonistically [87]. Ethylene prevents ABA accumulation and inhibits ABA-induced stomatal closure [88] and, on the other hand, higher concentrations of ABA prevent excess ET production, which allows for shoot and root growth under drought conditions [89].

Compared to the ABA and ET responses to drought stress, no consensus has yet been achieved concerning the responses of the growth hormones auxins (Aux) and gibberellins (GA). Nevertheless, there is increasing evidence that a reduction in the levels of GA or signalling contribute to plant growth restriction, once GA inhibits the accumulation of growth suppressors, namely, DELLA (aspartic acid–glutamic acid–leucine–leucine–alanine) proteins [81]. On the other hand, the Aux

responses to drought are usually contradictory, varying from decreases [90,91] to increases [92,93]. Meanwhile, Wang et al. [94] reported a marked decrease in Aux with a transient increase during the initial stage of drought adaptation. Furthermore, some studies have shown that drought influences the local Aux concentration and distribution by changes in Aux transport, allowing to maintain a balance between vegetative growth and survival [95,96]. In fact, it was well demonstrated that the Aux was unequally distributed within and among the different plant organs [6,82,83]. In addition, it was suggested that Aux could act as a stress hormone, directly or indirectly, once it was observed that several auxin-responsive genes operate during stress signalling [97]. However, hormone action cannot be considered in isolation, as the cross-talk among the different plant hormones results in synergistic or antagonistic interactions that play crucial roles in the response of plants to abiotic stress [98].

5. Drought Influence on Olive Crop Yield and Quality

In general, a water deficit has a negative effect on yield, fruit dry mass and oil accumulation [10-12,99,100], while it accelerates fruit maturation [100,101]. Regarding fruit and olive oil phenolics composition, dissimilar responses may occur. However, the general trend is the parallel increase with drought intensity [10,11,99–103], but oils are occasionally characterized as excessively bitter [10]. Nevertheless, from a certain threshold, the increase in quality, conferred by phenolic compounds, is not compensated by the losses in quantity, with deficit irrigation being the better option [10,103]. In line, a deficit irrigation strategy has been shown to be effective in producing quality Extra Virgin Olive Oil, by increasing the content of total phenols and sensory quality [104,105]. Meanwhile, the water deficit influence on qualitative olive oil indexes and fatty acid composition are inconsistent in the literature. Generally, the minor effects of irrigation were felt in peroxide value, free acidity and specific absorption coefficients (K_{232} , K_{270} , ΔK) [10,99,103]. Regarding the fatty acids profile, while in some studies no significant influence was detected [11,99], in other works, the growing season highly determined the responses [10,103], making it difficult to establish a pattern. Although there is not a direct effect from a water deficit, the severity and frequency of frosts may be higher with reduced soil and air moisture. This may be a critical event for olive quality, as harvests usually occur when early frosts start coming. Some studies have already reported that frost events damage olive fruits and, consequently, the quality of the extracted olive oil, including a decrease in pigments and phenolic compound concentrations [106–109].

6. Olive Tree Strategies to Withstand Drought

Plants respond to adapt and survive under limited moisture supply by inducing different strategies, which can be divided into three distinct mechanisms: drought escape, which involves a shortened life cycle or growing season, allowing plants to reproduce before the environment becomes dry; drought avoidance, which involves the adoption of mechanisms that reduce water loss from plants; and drought tolerance, which is defined as the ability to grow, flower and display economic yield under sub-optimal water supply [8].

The capacity of olive to grow under harsh conditions is due to the development of certain morphological, anatomical, physiological and biochemical responses [16], benefiting from the memory effects caused by stress pre-exposure [60]. However, these mechanisms are activated at considerable expenses to the plant in terms of energy, which causes a decrease in current-season production and compromises vegetative development, impairing next year's production. Olive trees can slow the onset of stress (avoidance) with the ability to extract water from the soil and restricting water losses to the atmosphere. Moreover, tolerance is the ultimate drought strategy, displayed by the ability to sustain a large internal water deficit and maintain enough metabolic activity for survival [19]. However, as argued by Chen et al. [110], drought adaptability integrates much more than the drought resistance concept (i.e., drought escape, drought avoidance and drought tolerance), recovery capacity also plays a fundamental role in plants' growth and survival. This takes special importance in Mediterranean-type ecosystems, where plants are continuously exposed to repeated cycles of drought re-watering during their life. Nevertheless, compared to development during drought, the

study of recovery has been neglected. Although drought is considered the primary stressor, others such as heat and high irradiance, especially in association with each other, also impair plant functions and, therefore, different adaptive mechanisms are adopted by plants.

Olive leaves are small, with high mesophyll compactness, grouped along sclereids in spongy parenchyma and two/three palisade layers associated with the upper epidermis [13,111], being the lignin accumulation [61], the thickness and density especially marked under adverse conditions [48,56,111,112]. This particular structure reduces the internal conductance to water vapor transport [113] and provides a greater resistance to physical damage driven by desiccation [114]. Olive leaves also present a thick cuticle that prevents water diffusion through the cuticular layer. In fact, cuticular conductance is negligible when compared with g_s, meaning that most of the transpiration is associated to the stomata [19]. Moreover, leaf surface, especially the abaxial surface, is covered with a waxy layer and stellar peltate trichomes hiding the small and abundant stomata [13], which usually increases under drought conditions [7]. These structures increase water-use efficiency, by increasing leaf boundary-layer resistance, and allowing leaves to take advantage of light rain or water condensation [115].

The stomata of olive leaves are small and present only on the abaxial surface (hypostomatous), being even smaller and denser in water shortage situations, allowing better control of water loss by transpiration [48,111]. Moreover, an efficient control of the stomatal aperture helps to maintain xylem water potential values above the safety threshold for loss of hydraulic conductance [15,16]. Although strong evidence shows that g_s decreases as plant Ψ becomes more negative [48,56,112], under severe conditions, stomatal control over transpiration may be not enough to prevent the loss of hydraulic conductance [112]. For some plant species, the permanent wilting point is reached when $\Psi = -1.5$ MPa [116], but since olive tissues can withstand very negative values of Ψ [117], the wilting point for olive ranges approximately between -2.5 MPa [118] and -3.5 MPa [119] or even has a huge capacity to sustain values below -8 MPa [120]. In fact, Moriana et al. [120] reported that rainfed olive trees with Ψ around -8 MPa extracted more 40 mm of water below the conventional wilting point (-1.5 MPa). To rainfed orchards in arid regions, this amount has significant importance since it represents around 10–15% of annual transpiration [121]. During recovery, olive trees typically show conservative behaviour, rapidly restoring water status, but exhibiting a slow recovery of g_s [6,15,51]. Torres-Ruiz et al. [46] found that neither hydraulic nor non-hydraulic factors were able to explain the delay in the full recovery of gs. These authors proposed two explanations, one involving the restoration of certain aquaporins activities, not affecting leaf hydraulic conductance directly, but the balance of osmolytes in the cells; and the other involving the occurrence of a metabolic limitation, as the increase in ABA in guard cells under drought induces the expression of hexokinases, which accelerates the stomatal closure. On the other hand, the hexokinases are also involved in sugar sensing and stimulation of the osmolytes balance that should be restarted after the recovery of water status. In addition, Brito et al. [6] showed that in line with a delay in gs restoration, the intense ABA signal in droughted olive leaves after stress relief was stronger closer to the upper epidermis, suggesting its relocalisation after rehydration and a "memory" effect, which might enable a rapid response under drought restoration. Olive trees pre-exposed to drought also recover A faster than gs after stress relief [6,60].

Olive trees show a high resistance to drought-induced embolism, essentially due to the small diameter of the xylem vessels and high density, leading to low xylem hydraulic conductivity that limit transpiration [5,15,122]. Furthermore, the olive root system grows quite parallel to the soil and the highest root density is found close to the trunk surface, being more suitable to absorb the light and intermittent rainfall, typical of its habitat, than water from deep layers [23]. Nevertheless, olive root growth and distribution depend largely on the soil conditions [19]. As rainfed olive trees need to explore larger soil volumes than irrigated trees to collect similar amounts of water, the total root system is greater in dry than in wet conditions [123]. In addition, under low water potential, olive trees also slow or even stop canopy growth, but still present some net photosynthesis, allowing the production of photo-assimilates that are particularly accumulated in the root system [58,61]. As a consequence, an increase in the root/canopy ratio is usual [6,58,61] in order to adjust the demand for

transpiration and soil water uptake. Olive trees also benefit from hydraulic redistribution — the ability of deep roots to uptake water in moist soil layers to maintain transpiration during the hot dry season and to redistribute soil water through different root types, reducing the intensive drying of the upper soil layers [124].

To ensure the hydraulic conductance and the maintenance of water flow from roots to leaves, olive trees decrease the water potential of their tissues, establishing a particularly high gradient between leaves and roots [58,125]. Under drought conditions, the olive tree displays a strong capacity to osmotic adjustment (OA)-the accumulation of solutes-both in the leaves and roots [58,60,61,118,119]. This mechanism decreases the osmotic potential, creating a soil-plant water gradient, which enables the extraction of water from the soil at a water potential below the wilting point [119]. Osmotic adjustment is linked with passive and active osmotic regulation mechanisms, an increase in solute concentration resulting from symplastic water loss [119] and an accumulation or de novo synthesis of solutes within cells [126], respectively. Two major classes of solutes can lower the osmotic potential of tissues: inorganic cations and anions and organic compatible solutes, such as sugars, sugar alcohols, amino acids (notably proline), and quaternary ammonium compounds (notably glycine betaine) [126,127]. Some of the organic solutes can also protect cellular proteins, enzymes and cellular membranes and allow the metabolic machinery to continue functioning [14,126]. On the other hand, changes in cell wall elasticity can also contribute to drought adaptability, as demonstrated in different olive genotypes [14] and water regimes [3]. In these studies, it was interesting to notice that both increases and decreases in cell wall elasticity may aid survival under low water availability. In fact, more elastic cell walls can shrink more easily when subjected to stress, helping the maintenance of higher turgor pressure and protecting cell walls from rupturing [128], while more rigid cells may help to maintain lower water potential at any given volume than elastic ones, resulting in a higher gradient of water potential between the soil and the plant, thereby promoting more effective water uptake from drying soils [127].

Although the AQPs' relevance in olive tree drought tolerance is still poorly explored, their involvement in precise water movement regulation underpin this [129]. The change in AQPs' activity may serve to ensure that during stress, water moves to where it is required or is retained and where it is most critical [130]. Additionally, AQPs may be important in whole-plant rehydration during the recovery period, displaying also an important role in xylem conduit refilling after drought-induced embolism [131,132]. A downregulation of AQP genes, both under moderate or severe droughts, followed by an upregulation upon re-watering and then a return to normal levels were identified in olive trees [131,132]. Furthermore, AQPs' responses can be correlated with the isohydric and anisohydric behaviour of plants, which can eventually switch from one to another [56] in response to changing environmental conditions, as reported for grapevines [133] or to fruit load, as stated for the olive tree [134].

Finally, the regulation of the antioxidant system is one of the most relevant mechanisms against oxidative stress caused by ROS. Reactive oxygen species play a double role in plant physiology, but whether ROS would act as signalling molecules or might cause oxidative stress to the tissues depend on the refined balance between its production and scavenging [135]. The increase in carotenoids and the carotenoids/chlorophylls ratio is considered one of the mechanisms developed by the olive tree to protect the photosynthetic apparatus against photooxidation [60]. Moreover, the increment in some antioxidant enzymes activities, such as ascorbate peroxidase, catalase, superoxide dismutase, glutathione reductase and/or in non-enzymatic antioxidant mechanisms, such as the accumulation of phenolic compounds, tocopherols, carotenoids, ascorbate and glutathione, were commonly described in olive trees under drought conditions [3,4,9,58-60]. On the other hand, in a study conducted by Abdallah et al. [60], it was demonstrated that upon re-watering, olive trees still exhibited higher levels of hydrogen peroxide (H_2O_2), a known signalling ROS, possibly to keep the antioxidative system on alert. Moreover, olive trees that were drought-primed showed an alleviation in oxidative stress in relation to plants exposed to drought for the first time [60]. A summary of the strategies adopted by the olive tree to improve drought adaptability (i.e., drought avoidance, tolerance and recovery capacity) is shown in Figure 2.

Drought intensity			Rehydration
Avoidance		Tolerance	Recovery capacity
Ability to extract water from soil: High root density close to trunk surface; Large root system; Small and dense xylem vessels Hydraulic redistribution; Nightime stomatal conductance; High root/canopy ratio; Capacity to decrease water potential; Osmotic adjustment.	Restriction water losses: Paraheliotropism; Small and sclerophyllous leaves; Leaf mesophyll compactness; Mypostomatous leaves; Small and dense stomata; Dense trichome layer; High root/canopy ratio; Reduced stomatal conductance; Aquaporins regulation.	Ability to sustain large internal water deficit and metabolic activity; Aquaporins regulation; Osmotic adjustment; High carotenoids and carotenoids/chlorophyll ratios; Efficient enzymatic and non-enzymatic antioxidant responses. 	 Conservative behaviour: ✓ Slow stomatal conductance restauration; ✓ Rapid water status restauration; ✓ Aquaporins regulation; ✓ ABA persistence in leaves; ✓ H₂O₂ persistence to keep antioxidant system in alert.

Olive tree strategies to improve drought adaptability

Figure 2. Strategies adopted by the olive tree to improve drought adaptability (i.e., drought avoidance, tolerance and recovery capacity).

7. Cultivars' Response to Drought

Considerable genotypic differences are present among different cultivars, which employ different mechanisms to cope with drought [3,5,9,13,14,136,137]. In general, olive cultivars native to dry regions have more capability to acclimate to drought conditions than cultivars which originated in regions with a more temperate climate [14]); still, the identification of the traits of the more resistant cultivars is ambiguous, as it depends on the cultivars compared in the studies available.

Bacelar and colleagues [3,5,13,14] studied Portuguese and Spanish cultivars, including Cobrançosa, Manzanilla, Negrinha, Blanqueta, Arbequina, Madural and Verdeal Transmontana. Cobrançosa exhibited good protection against water loss through high-density foliar tissue and by thick cuticle and trichome layers, while Manzanilla and Negrinha enhanced their sclerophyll by building parenchyma tissues and increasing protective structures like the upper cuticle and both the upper and lower epidermis [13]. Among Cobrançosa, Madural and Verdeal Transmontana cultivars, Cobrançosa had a more efficient water transport through the xylem, a more enhanced water-use efficiency of biomass production [5] and a high capability to osmotic adjustment and protection against oxidative stress [3]. Madural also had a thick upper epidermis, a thick palisade parenchyma, a high stomatal density, high capability for osmotic adjustment and increased tissue rigidity, but less effective mechanisms against oxidative stress [3]. Meanwhile, Cobrançosa, Manzanilla and Negrinha employ a prodigal water-use strategy and high tissue elasticity [14]. Moreover, Manzanilla accumulated high levels of proline [14]. Arbequina had a thinner trichome layer, implying that the leaves were less protected against water loss, but the development of smaller leaves may reduce water loss at the whole-plant level [13]. Conversely, present high levels of soluble proteins, which may represent an increased activity of oxidative stress defence enzymes [14]. Verdeal Transmontana did not exhibit osmotic adjustment capacity but was able to increase tissue elasticity and total soluble protein concentration [3]. Blanqueta had larger leaves and some anatomical traits that may lead to high water loss, especially from the adaxial surface [13]. Arbequina and Blanqueta had high tissue rigidity, employed a conservative water-use strategy, had lower photosynthetic rates and a high midday depression in photosynthesis [14].

Among Greek cultivars, *Gaidourelia* showed higher phenolic concentration and antioxidant activity and lower lipid peroxidation and photochemical damage than *Kalamon, Koroneiki* and *Megaritiki* [9], while in a study comparing *Konservolia*, *Zard* and *Amigdalolia* cultivars, *Konservolia* had higher chlorophyll and total carbohydrates concentrations and higher antioxidant enzymes activities, whereas the highest total phenol and proline levels were recorded in the *Zard* cultivar [136].

In another study, the effect of drought on the inhibition of growth was higher in *Fishomi* than in *Dezful, Amigdalolia* and *Conservolia* cultivars. *Dezful, Amigdalolia* and *Conservolia* were able to preserve higher levels of relative leaf water content and membrane stability index under drought stress, due to the higher concentrations of soluble carbohydrates, proline, potassium and calcium in their leaves [137].

Future climatic conditions in the Mediterranean region will certainly cause harmful consequences to olive tree physiology and yield, particularly under rainfed conditions. Although the olive tree has several mechanisms that allow for good acclimation to drought, they are activated at the expense of carbon reserves and may be detrimental with the increased duration and intensity of the stress. This consequence may lead to the abandonment of traditional olive groves and desertification in extensive areas, with devastating economic, social and environmental costs. Moreover, young plantations require some irrigation for their establishment due to their poorly developed root system and reduced capacity for storing water, that will be traduced in a great amount of water consumption. Thus, understanding the main effects and the response mechanisms adopted by the olive tree to cope with drought is crucial to improving crop management strategies and to designing more sustainable and productive crop systems and saving water resources. This review consolidated knowledge on how the olive tree responds to drought, but also demonstrated that much more effort is needed to fully understand these responses and their implication from the plant to the ecosystem level in a changing world. For instance, night-time stomatal conductance and sap flux, different classes of aquaporins activities, leaf hydraulic conductance, mesophyll conductance, the role of phytohormones cross-talk and other signalling molecules, such as H2O2, and recovery processes are topics less explored in olive tree response to environmental stressors. Moreover, higher investment in integrated approaches with proteome and the corresponding physiological responses can profoundly change our understanding of olive tree adaptability.

Abbreviations and Symbols:

A: net photosynthetic rate A/gs: intrinsic water use efficiency ABA: abscisic acid **AQPs:** aquaporins ATP: adenosine triphosphate Aux: auxins CA: carbonic anhydrase CO2: carbon dioxide DNA: deoxyribonucleic acid Enight: night-time transpiration ET: ethylene ETR: photosynthetic electron transport rate F'_v/F'_m : capture efficiency of excitation energy by open PSII reaction centres Fv/Fm: maximal quantum efficiency of photosystem II GA: gibberellins gm: mesophyll conductance gnight: night-time stomatal conductance gs: stomatal conductance H2O2: hydrogen peroxide HCO3⁻: bicarbonate anion K232: excitation coefficient at 232 nm K270: excitation coefficient at 270 nm Kleaf: leaf hydraulic conductance NPQ: non-photochemical quenching O2: oxygen OA: osmotic adjustment PPFD: photosynthetic photon flux density PSII: photosystem II a qP: photochemical quenching R: respiration

ROS: reactive oxygen species
WUE_i: intrinsic water use efficiency
WUE_{WP}: whole plant water use efficiency
ΔK: variation of the specific extinction
ΦPSII: effective quantum efficiency of photosystem II
Ψ: leaf water potential

Acknowledgements: This work was financed by project Interact: Integrative Research in Environment, Agro-Chain, and Technology, operation NORTE-01-0145-FEDER-000017, research line ISAC, co-funded by European Regional Development Fund (FEDER) through NORTE 2020 (Programa Operacional Regional do Norte 2014/2020), and supported by the project "Novas práticas em olivais de sequeiro: estratégias de mitigação e adaptação às alterações climáticas", PDR2020-101-032119, financed by the European Agricultural Fund for Rural Development (EAFRD) and the Portuguese State under Ação 1.1 "Grupos Operacionais", integrada na Medida 1. "Inovação" do PDR 2020: Programa de Desenvolvimento Rural do Continente. Institution CITAB, for its financial support through National Funds by FCT: Portuguese Foundation for Science and Technology, under the project UID/AGR/04033/2019.

Author Contributions: Writing—original draft preparation and editing, C.B.; writing—review and editing, C.M.C.; critical corrections, L.-T.D. and J.M.-P.

Funding: Project Interact: Integrative Research in Environment, Agro-Chain, and Technology, operation NORTE-01-0145-FEDER-000017, research line ISAC, co-funded by European Regional Development Fund (FEDER) through NORTE 2020 (Programa Operacional Regional do Norte 2014/2020).

Conflicts of Interest: Conflicts of interest: The authors declare no conflict of interest.

References

- 1. International Olive Concil (IOC). Available online: http://www.internationaloliveoil.org (acessed on 5 June 2019).
- Stocker, T.F.; Qin, D.; Plattner, G.-K.; Tignor, M.; Allen, S. K.; Boschung, J.; Nauels, A.; Xia, Y.; Bex, V.; Midgley, P. M. Climate Change 2013: The Physical Science Basis: Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change 2013; Cambridge University Press: New York, NY, USA, 2013.
- Bacelar, E.A.; Santos, D.L.; Moutinho-Pereira, J.M.; Gonçalves, B.C.; Ferreira, H.F.; Correia, C.M. Immediate responses and adaptative strategies of three olive cultivars under contrasting water availability regimes: Changes on structure and chemical composition of foliage and oxidative damage. *Plant Sci.* 2006, *170*, 596– 605, doi:10.1016/j.plantsci.2005.10.014.
- Bacelar, E.A.; Santos, D.L.; Moutinho-Pereira, J.M.; Lopes, J.I.; Gonçalves, B.C.; Ferreira, T.C.; Correia, C.M. Physiological behaviour, oxidative damage and antioxidative protection of olive trees grown under different irrigation regimes. *Plant Soil* 2007, 292, 1–12, doi:10.1007/s11104-006-9088-1.
- Bacelar, E.A.; Moutinho-Pereira, J.M.; Gonçalves, B.C.; Ferreira, H.F.; Correia, C.M. Changes in growth, gas exchange, xylem hydraulic properties and water use efficiency of three olive cultivars under contrasting water availability regimes. *Environ. Exp. Bot.* 2007, *60*, 183–192, doi:10.1016/j.envexpbot.2006.10.003.
- Brito, C.; Dinis, L.-T.; Meijón, M.; Ferreira, H.; Pinto, G.; Moutinho-Pereira, J.; Correia, C. Salicylic acid modulates olive tree physiological and growth responses to drought and rewatering events in a dose dependent manner. J. Plant Physiol. 2018, 230, 21–32, doi:10.1016/j.jplph.2018.08.004.
- Brito, C.; Dinis, L.-T.; Ferreira, H.; Moutinho-Pereira, J.; Correia, C. The role of nighttime water balance on Olea europaea plants subjected to contrasting water regimes. J. Plant Physiol. 2018, 226, 56–63, doi:10.1016/j.jplph.2018.04.004.
- 8. Farooq, M.; Wahid, A.; Kobayashi, N.; Fujita, D.; Basra, S.M.A. Plant drought stress: Effects, mechanisms and management. *Agron. Sustain. Dev.* **2009**, *29*, 185–212, doi:10.1051/agro:2008021.
- Petridis, A.; Therios, I.; Samouris, G.; Koundouras, S.; Giannakoula, A. Effect of water deficit on leaf phenolic composition, gas exchange, oxidative damage and antioxidant activity of four Greek olive (*Olea europaea* L.) cultivars. *Plant Physiol. Biochem.* 2012, 60, 1–11, doi:10.1016/j.plaphy.2012.07.014.
- Servili, M.; Esposto, S.; Fabiani, R.; Urbani, S.; Taticchi, A.; Mariucci, F.; Selvaggini, R.; Montedoro, G.F. Phenolic compounds in olive oil: Antioxidant, health and organoleptic activities according to their chemical structure. *Inflammopharmacology* 2009, *17*, 76–84, doi:10.1007/s10787-008-8014-y.

- 11. Greven, M.; Neal, S.; Green, S.; Dichio, B.; Clothier, B. The effects of drought on the water use, fruit development and oil yield from young olive trees. *Agric. Water Manag.* 2009, *96*, 1525–1531, doi:10.1016/j.agwat.2009.06.002.
- Fernandes-Silva, A.A.; Ferreira, T.C.; Correia, C.M.; Malheiro, A.; Villalobos, F.J. Influence of different irrigation regimes on crop yield and water use efficiency of olive. *Plant Soil* 2010, 333, 35–47, doi:10.1007/s11104-010-0294-5.
- Bacelar, E.A.; Correia, C.M.; Moutinho-Pereira, J.M.; Goncalves, B.C.; Lopes, J.I.; Torres-Pereira, J.M. Sclerophylly and leaf anatomical traits of five field-grown olive cultivars growing under drought conditions. *Tree Physiol.* 2004, 24, 233–239, doi:10.1093/treephys/24.2.233.
- Bacelar, E.A.; Moutinho-Pereira, J.M.; Gonçalves, B.C.; Lopes, J.I.; Correia, C.M. Physiological responses of different olive genotypes to drought conditions. *Acta Physiol. Plant.* 2009, *31*, 611–621, doi:10.1007/s11738-009-0272-9.
- Torres-Ruiz, J.M.; Diaz-Espejo, A.; Morales-Sillero, A.; Martín-Palomo, M.J.; Mayr, S.; Beikircher, B.; Fernández, J.E. Shoot hydraulic characteristics, plant water status and stomatal response in olive trees under different soil water conditions. *Plant Soil* 2013, 373, 77–87, doi:10.1007/s11104-013-1774-1.
- 16. Fernández, J.-E. Understanding olive adaptation to abiotic stresses as a tool to increase crop performance. *Environ. Exp. Bot.* **2014**, *103*, 158–179, doi:10.1016/j.envexpbot.2013.12.003.
- Lopez-Bellido, P.J.; Lopez-Bellido, L.; Fernandez-Garcia, P.; Muñoz-Romero, V.; Lopez-Bellido, F.J. Assessment of carbon sequestration and the carbon footprint in olive groves in Southern Spain. *Carbon Manag.* 2016, 7, 161–170, doi:10.1080/17583004.2016.1213126.
- 18. Therios, I. Olives: Crop Production Science in Horticulture 18; CABI Publishing: Wallingford, UK, 2009.
- Connor, D.J.; Fereres, E. The physiology of adaptation and yield expression in olive. *Hortic. Rev.* 2005, 31, 155–193, doi:10.1002/9780470650882.ch4.
- Böhm, J., Antunes, M.T. A evolução da espécie olea europaea. In O Grande Livro da Oliveira e do Azeite Portugal Oleícola; Böhm, J., Ed.; Dinalivro Editora: Lisboa, Portugal, 2013; pp. 34–49.
- Giorgi, F.; Lionello, P. Climate change projections for the Mediterranean region. *Glob. Planet Chang.* 2008, 63, 90–104, doi:10.1016/j.gloplacha.2007.09.005.
- Tombesi, A.; Tombesi, S. Orchard planning and planting. In *Production Techniques in Olive Growing*; Sbitri, M.O., Serafini, F., Eds.; International Olive Council: Madrid, Spain, 2007; pp. 17–40.
- 23. Fernández, J.E.; Moreno, F. Water use by the olive tree. J. Crop Prod. 1999, 2, 101–162, doi:10.1300/J144v02n02_05.
- 24. Guerrero, A. Nueva Olivicultura, 5th ed.; Ediciones Mundi-Prensa: Madrid, Spain, 2002.
- Ponti, L.; Gutierrez, A.P.; Basso, B.; Neteler, M.; Ruti, P.M.; Dell'Aquila, A.; Iannetta, M. Olive agroecosystems in the Mediterranean Basin: Multitrophic analysis of climate effects with process-based representation of soil water balance. *Procedia Environ. Sci.* 2013, 19, 122–131, doi:10.1016/j.proenv.2013.06.014.
- 26. Food and Agriculture Organization of the United Nations (FAOSTAT). Available online: http://www.fao.org/faostat/en/#home (acessed on 4 March 2019).
- Gucci, R.; Fereres, E. Crop Yield Response to Water, FAO Irrigation and Drainage Paper 66; Steduto, P., Hsiao, T.C., Fereres, E.D.R., Eds.; Food and Agriculture Organization of the United Nations (FAO): Rome, Italy, 2012; pp. 300–315.
- Viola, F.; Caracciolo, D.; Pumo, D.; Noto, L.V. Olive yield and future climate forcings. *Procedia Environ. Sci.* 2013, 19, 132–138, doi:10.1016/j.proenv.2013.06.015.
- 29. Viola, F.; Caracciolo, D.; Pumo, D.; Noto, L.; Loggia, G. Future climate forcings and olive yield in a Mediterranean orchard. *Water* **2014**, *6*, 1562–1580, doi:10.3390/w6061562.
- Chartzoulakis, K.; Psarras, G. Global change effects on crop photosynthesis and production in Mediterranean: The case of Crete, Greece. *Agric. Ecosyst. Environ.* 2005, 106, 147–157, doi:10.1016/j.agee.2004.10.004.
- Minnocci, A.; Panicucci, A.; Sebastiani, L.; Lorenzini, G.; Vitagliano, C. Physiological and morphological responses of olive plants to ozone exposure during a growing season. *Tree Physiol.* 1999, 19, 391–397, doi:10.1093/treephys/19.6.391.
- Pérez-López, D.; Ribas, F.; Moriana, A.; Rapoport, H.F.; De Juan, A. Influence of temperature on the growth and development of olive (*Olea europaea* L.) trees. *J. Hortic. Sci. Biotechnol.* 2008, *83*, 171–176, doi:10.1080/14620316.2008.11512366.

- Aguilera, F.; Fornaciari, M.; Ruiz-Valenzuela, L.; Galán, C.; Msallem, M.; Ben Dhiab, A.; Díaz-de la Guardia, C.; Trigo, M.; Bonofiglio, T.; Orlandi, F. Phenological models to predict the main flowering phases of olive (*Olea europaea* L.) along a latitudinal and longitudinal gradient across the Mediterranean region. *Int. J. Biometeorol.* 2015, *59*, 629–641, doi:10.1007/s00484-014-0876-7.
- Torres, M.; Pierantozzi, P.; Searles, P.; Rousseaux, M.C.; García-Inza, G.; Miserere, A.; Bodoira, R.; Contreras, C.; Maestri, D. Olive cultivation in the southern hemisphere: Flowering, water requirements and oil quality responses to new crop environments. *Front. Plant Sci.* 2017, *8*, 1830, doi:10.3389/fpls.2017.01830.
- 35. Dag, A.; Harlev, G.; Lavee, S.; Zipori, I.; Kerem, Z. Optimizing olive harvest time under hot climatic conditions of Jordan Valley, Israel. *Eur. J. Lipid Sci. Technol.* **2014**, *116*, 169–176, doi:10.1002/ejlt.201300211.
- Orlandi, F.; Garcia-Mozo, H.; Dhiab, A.B.; Galán, C.; Msallem, M.; Romano, B.; Abichou, M.; Dominguez-Vilches, E.; Fornaciari, M. Climatic indices in the interpretation of the phenological phases of the olive in mediterranean areas during its biological cycle. *Clim. Chang.* 2013, *116*, 263–284, doi:10.1007/s10584-012-0474-9.
- Tanasijevic, L.; Todorovic, M.; Pereira, L.S.; Pizzigalli, C.; Lionello, P. Impacts of climate change on olive crop evapotranspiration and irrigation requirements in the Mediterranean region. *Agric. Water Manag.* 2014, 144, 54–68, doi:10.1016/j.agwat.2014.05.019.
- Rallo, L.; Martin, G.C. The role of chilling in releasing olive floral buds from dormancy. J. Am. Soc. Hortic. Sci. 1991, 116, 1058–1062, doi:10.21273/JASHS.116.6.1058.
- Fabbri, A.; Benelli, C. Flower bud induction and differentiation in olive. J. Hortic. Sci. Biotec. 2000, 75, 131– 141, doi:10.1080/14620316.2000.11511212.
- Ramos, A.; Rapoport, H.F.; Cabello, D.; Rallo, L. Chilling accumulation, dormancy release temperature, and the role of leaves in olive reproductive budburst: Evaluation using shoot explants. *Sci. Hortic.* 2018, 231, 241–252, doi:10.1016/j.scienta.2017.11.003.
- Moriondo, M.; Trombi, G.; Ferrise, R.; Brandani, G.; Dibari, C.; Ammann, C.M.; Lippi, M.M.; Bindi, M. Olive trees as bio-indicators of climate evolution in the Mediterranean Basin. *Glob. Ecol. Biogeogr.* 2013, 22, 818– 833, doi:10.1111/geb.12061.
- Ponti, L.; Gutierrez, A.P.; Ruti, P.M.; Dell'Aquila, A., Fine-scale ecological and economic assessment of climate change on olive in the Mediterranean Basin reveals winners and losers. *Proc. Natl. Acad. Sci. USA* 2014, *111*, 5598–5603, doi:10.1073/pnas.1314437111.
- 43. Taiz, L.; Zeiger, E. Plant Physiology, 4th ed.; Sinauer Associates, Inc.: Sunderland, MA, USA, 2006.
- Farooq, M.; Hussain, M.; Wahid, A.; Siddique, K.H.M. Drought stress in plants: An overview. In *Plant Responses to Drought Stress—From Morphological to Molecular Features*; Aroca, R., Ed.; Springer: Heidelberg, Germany, 2012; pp. 1–33.
- Hernandez-Santana, V.; Rodriguez-Dominguez, C.M.; Fernandez, J.E.; Diaz-Espejo, A. Role of leaf hydraulic conductance in the regulation of stomatal conductance in almond and olive in response to water stress. *Tree Physiol.* 2016, *36*, 725–735, doi:10.1093/treephys/tpv146.
- Torres-Ruiz, J.M.; Diaz-Espejo, A.; Perez-Martin, A.; Hernandez-Santana, V. Role of hydraulic and chemical signals in leaves, stems and roots in the stomatal behaviour of olive trees under water stress and recovery conditions. *Tree Physiol.* 2015, 35, 415–424, doi:10.1093/treephys/tpu055.
- 47. Pantin, F.; Monnet, F.; Jannaud, D.; Costa, J.M.; Renaud, J.; Muller, B.; Simonneau, T.; Genty, B. 2013. The dual effect of abscisic acid on stomata. *New Phytol.* **2013**, *197*, 65–72, doi:10.1111/nph.12013.
- 48. Boughalleb, F.; Hajlaoui, H. Physiological and anatomical changes induced by drought in two olive cultivars (cv Zalmati and Chemlali). *Acta Physiol. Plant.* **2011**, *33*, 53–65, doi:10.1007/s11738-010-0516-8.
- Tomás, M.; Flexas, J.; Copolovici, L.; Galmes, J.; Hallik, L.; Medrano, H.; Ribas-Carbo, M.; Tosens, T.; Vislap, V.; Niinemets, U. Importance of leaf anatomy in determining mesophyll diffusion conductance to CO₂ across species: Quantitative limitations and scaling up by models. *J. Exp. Bot.* 2013, *64*, 2269–2281, doi:10.1093/jxb/ert086.
- Perez-Martin, A.; Flexas, J.; Ribas-Carbo, M.; Bota, J.; Tomás, M.; Infante, J.M.; Diaz-Espejo, A. Interactive effects of soil water deficit and air vapour pressure deficit on mesophyll conductance to CO₂ in *Vitis vinifera* and *Olea europaea. J. Exp. Bot.* 2009, *60*, 2391–2405, doi:10.1093/jxb/erp145.
- Perez-Martin, A.; Michelazzo, C.; Torres-Ruiz, J.M.; Flexas, J.; Fernández, J.E.; Sebastiani, L.; Diaz-Espejo, A. Regulation of photosynthesis and stomatal and mesophyll conductance under water stress and recovery in olive trees: Correlation with gene expression of carbonic anhydrase and aquaporins. *J. Exp. Bot.* 2014, 65, 3143–3156, doi:10.1093/jxb/eru160.

- 52. Ferreira, F.J.; Guo, C.; Coleman, J.R. Reduction of plastid-localized carbonic anhydrase activity results in reduced arabidopsis seedling survivorship. *Plant Physiol.* **2008**, *147*, 585–594, doi:10.1104/pp.108.118661.
- Tholen, D.; Zhu, X.-G. The mechanistic basis of internal conductance: A theoretical analysis of mesophyll cell photosynthesis and CO₂ diffusion. *Plant Physiol.* 2011, *156*, 90–105, doi:10.1104/pp.111.172346.
- Gillon, J.S.; Yakir, D. Internal Conductance to CO₂ diffusion and C18OO discrimination in C3 Leaves. *Plant Physiol.* 2000, 123, 201–214.
- Heckwolf, M.; Pater, D.; Hanson, D.T.; Kaldenhoff, R. The Arabidopsis thaliana aquaporin AtPIP1;2 is a physiologically relevant CO₂ transport facilitator. *Plant J.* 2011, 67, 795–804, doi:10.1111/j.1365-313X.2011.04634.x.
- Moshelion, M.; Halperin, O.; Wallach, R.; Oren, R.; Way, D.A. Role of aquaporins in determining transpiration and photosynthesis in water-stressed plants: Crop water-use efficiency, growth and yield. *Plant Cell Environ.* 2015, *38*, 1785–1793, doi:10.1111/pce.12410.
- Guerfel, M.; Baccouri, O.; Boujnah, D.; Chaïbi, W.; Zarrouk, M. Impacts of water stress on gas exchange, water relations, chlorophyll content and leaf structure in the two main Tunisian olive (*Olea europaea* L.) cultivars. *Sci. Hortic.* 2009, *119*, 257–263, doi:10.1016/j.scienta.2008.08.006.
- Sofo, A.; Manfreda, S.; Fiorentino, M.; Dichio, B.; Xiloyannis, C. The olive tree: A paradigm for drought tolerance in Mediterranean climates. *Hydrol. Earth Syst. Sci. Discuss.* 2008, 12, 293–301.
- Guerfel, M.; Ouni, Y.; Boujnah, D.; Zarrouk, M. Photosynthesis parameters and activities of enzymes of oxidative stress in two young 'Chemlali' and 'Chetoui' olive trees under water deficit. *Photosynthetica* 2009, 47, 340, doi:10.1007/s11099-009-0054-z.
- Abdallah, B.M.; Methenni, K.; Nouairi, I.; Zarrouk, M.; Youssef, N.B. Drought priming improves subsequent more severe drought in a drought-sensitive cultivar of olive cv. *Chétoui. Sci. Hortic.* 2017, 221, 43–52, doi:10.1016/j.scienta.2017.04.021.
- Abdallah, B.M.; Trupiano, D.; Polzella, A.; De Zio, E.; Sassi, M.; Scaloni, A.; Zarrouk, M.; Youssef, N.B.; Scippa, G. S. Unraveling physiological, biochemical and molecular mechanisms involved in olive (*Olea europaea* L. cv. Chétoui) tolerance to drought and salt stresses. *J. Plant Physiol.* 2018, 220, 83–95, doi:10.1016/j.jplph.2017.10.009.
- Flexas, J.; Medrano, H. Energy dissipation in C3 plants under drought. Funct. Plant Biol. 2002, 29, 1209– 1215.
- Flexas, J.; Gallé, A.; Galmés, J.; Ribas-Carbo, M.; Medrano, H. The response of photosynthesis to soil water stress. In *Plant Responses to Drought Stress—From Morphological to Molecular Features*; Aroca, R., Ed.; Springer: Heidelberg, Germany, 2012; pp. 129–144.
- 64. Caird, M.A.; Richards, J.H.; Donovan, L.A. Nighttime stomatal conductance and transpiration in C3 and C4 plants. *Plant Physiol.* **2007**, *143*, 4–10, doi:10.1104/pp.106.092940.
- Scholz, F.G.; Bucci, S.J.; Goldstein, G.; Meinzer, F.C.; Franco, A.C.; Miralles-Wilhelm, F. Removal of nutrient limitations by long-term fertilization decreases nocturnal water loss in savanna trees. *Tree Physiol.* 2007, 27, 551–559, doi:10.1093/treephys/27.4.551.
- 66. Ogle, K.; Lucas, R.W.; Bentley, L.P.; Cable, J.M.; Barron-Gafford, G.A.; Griffith, A.; Ignace, D.; Jenerette, G.D.; Tyler, A.; Huxman, T.E.; et al. Differential daytime and night-time stomatal behavior in plants from North American deserts. *New Phytol.* 2012, 194, 464–476, doi:10.1111/j.1469-8137.2012.04068.x.
- Escalona, J.M.; Fuentes, S.; Tomás, M.; Martorell, S.; Flexas, J.; Medrano, H. Responses of leaf night transpiration to drought stress in *Vitis vinifera* L. *Agric. Water Manag.* 2013, 118, 50–58, doi:10.1016/j.agwat.2012.11.018.
- Chen, X.; Gao, J.; Zhao, P.; McCarthy, H.R.; Zhu, L.; Ni, G.; Ouyang, L., Tree species with photosynthetic stems have greater nightime sap flux. *Front. Plant Sci.* 2018, *9*, 30, doi:10.3389/fpls.2018.00030.
- 69. Cannell, M.G.R.; Thornley, J.H.M. Modelling the components of plant respiration: Some guiding principles. *Ann. Bot.* **2000**, *85*, 45–54, doi:10.1006/anbo.1999.0996.
- Flexas, J.; Galmes, J.; Ribas-Carbo, M.; Medrano, H. The effects of water stress on plant respiration. In *Plant Respiration: From Cell to Ecosystem*; Lambers, H., Ribas-Carbo, M., Eds.; Springer: Berlin, Germany, 2005; pp. 85–94.
- Varone, L.; Gratani, L. Leaf respiration responsiveness to induced water stress in Mediterranean species. *Environ. Exp. Bot.* 2015, 109, 141–150, doi:10.1016/j.envexpbot.2014.07.018.
- Paul, M.J.; Pellny, T.K. Carbon metabolite feedback regulation of leaf photosynthesis and development. J. Exp. Bot. 2003, 54, 539–547, doi:10.1093/jxb/erg052.

- 73. Flexas, J.; Galmés, J.; Gallé, A.; GulíAs, J.; Pou, A.; Ribas-Carbo, M.; Tomásw, M.; Medrano, H. Improving water use efficiency in grapevines: Potential physiological targets for biotechnological improvement. *Aust. J. Grape Wine Res.* 2010, *16*, 106–121, doi:10.1111/j.1755-0238.2009.00057.x.
- Bacelar, E.; Moutinho-Pereira, J.M.; Gonçalves, B.; Brito, C.; Gomes-Laranjo, J.; Ferreira, H.; Correia, C.M. Water use strategies of plants under drought conditions. In *Plant Responses to Drought Stress—From Morphological to Molecular Features*; Aroca, R., Ed.; Springer: New York, NY, USA, 2012; pp. 145–170.
- Maroco, J.P.; Pereira, J.S.; Manuela Chaves, M. Growth, photosynthesis and water-use efficiency of two C4 sahelian grasses subjected to water deficits. J. Arid Environ. 2000, 45, 119–137, doi:10.1006/jare.2000.0638.
- 76. Grusak, M.A. Plant macro- and micronutrient minerals. *Encyclopedia Life Sci.* 2001, 1–5, doi:10.1002/9780470015902.a0001306.pub2.
- 77. Silva, E.; Nogueira, R.; Silva, M.; Albuquerque, M. Drought stress and plant nutrition. *Plant Stress* **2011**, *5*, 32–41.
- Sanaullah, M.; Rumpel, C.; Charrier, X.; Chabbi, A. How does drought stress influence the decomposition of plant litter with contrasting quality in a grassland ecosystem? *Plant Soil* 2012, 352, 277–288, doi:10.1007/s11104-011-0995-4.
- Bista, D.R.; Heckathorn, S.A.; Jayawardena, D.M.; Mishra, S.; Boldt, J.K. Effects of drought on nutrient uptake and the levels of nutrient-uptake proteins in roots of drought-sensitive and -tolerant grasses. *Plants* 2018, 7, 28, doi:10.3390/plants7020028.
- Brito, C.; Dinis, L.-T.; Ferreira, H.; Coutinho, J.; Moutinho-Pereira, J.; Correia, C. Salicylic acid increases drought adaptability of young olive trees by changes on redox status and ionome. *Plant Physiol. Biochem.* 2019,141, 315-324, doi:10.1016/j.plaphy.2019.06.011.
- Colebrook, E.H.; Thomas, S.G.; Phillips, A.L.; Hedden, P. The role of gibberellin signalling in plant responses to abiotic stress. J. Exp. Biol. 2014, 217, 67–75, doi:10.1242/jeb.089938.
- De Diego, N.; Rodriguez, J.L.; Dodd, I.C.; Perez-Alfocea, F.; Moncalean, P.; Lacuesta, M. Immunolocalization of IAA and ABA in roots and needles of radiata pine (*Pinus radiata*) during drought and rewatering. *Tree Physiol.* 2013, 33, 537–549, doi:10.1093/treephys/tpt033.
- Brito, C.; Dinis, L.-T.; Luzio, A.; Silva, E.; Gonçalves, A.; Meijón, M.; Escandón, M.; Arrobas, M.; Rodrigues, M.A.; Moutinho-Pereira, J.; et al. Kaolin and salicylic acid alleviate summer stress in rainfed olive orchards by modulation of distinct physiological and biochemical responses. *Sci. Hortic.* 2019, 246, 201–211, doi:10.1016/j.scienta.2018.10.059.
- Xiong, M. Aabscisic acid in plant response and adaptation to drought and salt stress. In Advances in Molecular Breeding toward Drought and Salt Tolerant Crops; Jenks, M., Hasegawa, P., Jain, S., Eds.; Springer Science & Business Media: New York, NY, USA, 2007; pp. 193–221.
- Gómez-Cadenas, A.; Tadeo, F.; Talon, M.; Primo-Millo, E. Leaf abscission Induced by ethylene in waterstressed Intact seedlings of cleopatra mandarin requires previous abscisic acid accumulation in roots. *Plant Physiol.* **1996**, *112*, 401–408, doi:10.1104/pp.112.1.401.
- Cui, M.; Lin, Y.; Zu, Y.; Efferth, T.; Li, D.; Tang, Z. Ethylene increases accumulation of compatible solutes and decreases oxidative stress to improve plant tolerance to water stress in Arabidopsis. *J. Plant Biol.* 2015, 58, 193–201, doi:10.1007/s12374-014-0302-z.
- Wilkinson, S.; Kudoyarova, G.R.; Veselov, D.S.; Arkhipova, T.N.; Davies, W.J. Plant hormone interactions: Innovative targets for crop breeding and management. *J. Exp. Bot.* 2012, 63, 3499–3509, doi:10.1093/jxb/ers148.
- Tanaka, Y.; Sano, T.; Tamaoki, M.; Nakajima, N.; Kondo, N.; Hasezawa, S. Ethylene inhibits abscisic acidinduced stomatal closure in Arabidopsis. *Plant Physiol.* 2005, *138*, 2337–2343, doi:10.1104/pp.105.063503.
- Sharp, R.E.; LeNoble, M.E. ABA, ethylene and the control of shoot and root growth under water stress. J. Exp. Bot. 2002, 53, 33–37, doi:10.1093/jexbot/53.366.33.
- 90. Man, D.; Bao, Y.-X.; Han, L.-B.; Zhang, X. Drought tolerance associated with proline and hormone metabolism in two tall fescue cultivars. *HortScience* **2011**, *46*, 1027–1032, doi:10.21273/HORTSCI.
- Du, H.; Liu, H.; Xiong, L. Endogenous auxin and jasmonic acid levels are differentially modulated by abiotic stresses in rice. *Front. Plant Sci.* 2013, 4, 1–10, doi:10.3389/fpls.2013.00397.
- De Diego, N.; Perez-Alfocea, F.; Cantero, E.; Lacuesta, M.; Moncalean, P. Physiological response to drought in radiata pine: Phytohormone implication at leaf level. *Tree Physiol.* 2012, 32, 435–449, doi:10.1093/treephys/tps029.

- Pustovoitova, T.N.; Zhdanova, N.E.; Zholkevich, V.N. Changes in the levels of IAA and ABA in cucumber leaves under progressive soil drought. Russ. *J. Plant Physiol.* 2004, *51*, 513–517, doi:10.1023/B:RUPP.0000035745.82509.bc.
- Wang, C.; Yang, A.; Yin, H.; Zhang, J. Influence of water stress on endogenous hormone contents and cell damage of maize seedlings. *J. Integr. Plant Biol.* 2008, 50, 427–434, doi:10.1111/j.1774-7909.2008.00638.x.
- 95. Shen, C.; Bai, Y.; Wang, S.; Zhang, S.; Wu, Y.; Chen, M.; Jiang, D.; Qi, Y. Expression profile of PIN, AUX/LAX and PGP auxin transporter gene families in Sorghum bicolor under phytohormone and abiotic stress. *FEBS J.* 2010, 277, 2954–2969, doi:10.1111/j.1742-4658.2010.07706.x.
- Shojaie, B.; Mostajeran, A.; Esmaeili, A. Different drought conditions could modulate growth responses of Arabidopsis thaliana through regulation of mRNA expression of genes encoding plasma membrane PIN proteins. *Int. J. Adv. Res. Biol. Sci.* 2015, 2, 241–254.
- Sharma, E.; Sharma, R.; Borah, P.; Jain, M.; Khurana, J.P. Emerging Roles of Auxin in Abiotic Stress Responses. In *Elucidation of Abiotic Stress Signaling in Plants: Functional Genomics Perspectives*; Pandey, G.K., Ed.; Springer: New York, NY, USA, 2015; pp. 299–328.
- Peleg, Z.M.; Blumwald, E. Hormone balance and abiotic stress tolerance in crop plants. *Curr. Opin. Plant Biol.* 2011, 14, 290–295, doi:10.1016/j.pbi.2011.02.001.
- Patumi, M.; d'Andria, R.; Marsilio, V.; Fontanazza, G.; Morelli, G.; Lanzac, B. Olive and olive oil quality after intensive monocone olive growing (*Olea europaea* L., cv. Kalamata) in different irrigation regimes. *Food Chem.* 2002, 77, 27–34, doi:10.1016/S0308-8146(01)00317-X.
- Bartolini, S.; Leccese, A.; Andreini, L. Influence of canopy fruit location on morphological, histochemical and biochemical changes in two oil olive cultivars. *Plant Biosyst.* 2014, 148, 1221–1230, doi:10.1080/11263504.2014.980360.
- 101. Machado, M.; Felizardo, C.; Fernandes-Silva, A.A.; Nunes, F.M.; Barros, A. Polyphenolic compounds, antioxidant activity and l-phenylalanine ammonia-lyase activity during ripening of olive cv. "Cobrançosa" under different irrigation regimes. *Food. Res. Int.* 2013, *51*, 412–421, doi:10.1016/j.foodres.2012.12.056.
- 102. Bucelli, P.; Costantini, E.A.C.; Barbetti, R.; Franchini, E. Soil water availability in rainfed cultivations affects more than cultivar some nutraceutical components and the sensory profile of virgin olive oil. *J. Agric. Food. Chem.* 2011, 59, 8304–8313, doi:10.1021/jf201278r.
- 103. Caruso, G.; Guccia, R.; Urbani, S.; Esposto, S.; Taticchi, A.; Di Maio, I.; Selvaggini, R.; Maurizio Servili, M. Effect of different irrigation volumes during fruit development onquality of virgin olive oil of cv. *Frantoio. Agric. Water Manag.* 2014, 134, 94–103, doi:10.1016/j.agwat.2013.12.003.
- Gómez del Campo, M.; García, J.M. Summer deficit-irrigation strategies in a hedgerow olive cv. Arbequina orchard: Effect on oil quality. J. Agric. Food Chem. 2013, 61, 8899–8905, doi:10.1021/jf402107t.
- 105. García, J.M.; Morales-Sillero, A.; Pérez-Rubio, A.G.; Diaz-Espejo, A.; Montero, A.; Fernández, J.E. Virgin olive oil quality of hedgerow 'Arbequina' olive trees under deficit irrigation. J. Sci. Food Agric. 2017, 97, 1018–1026, doi:10.1002/jsfa.7828.
- Houliston, A.; Vanhanen, L.; Savage, G. Frost protection of olives using glycine betaine. *Sci. Thecnol.* 2007, 108, 31–40.
- Morelló, J.-R.; Motilva, M.-J.; Ramo, T.; Romero, M.-P. Effect of freeze injuries in olive fruit on virgin olive oil composition. *Food Chem.* 2003, *81*, 547–553, doi:10.1016/S0308-8146(02)00488-0.
- 108. Morelló, J.R.; Romero, M.P.; Motilva, M.J. Influence of seasonal conditions on the composition and quality parameters of monovarietal virgin olive oils. J. Am. Oil Chem. Soc. 2006, 83, 683–690, doi:10.1007/s11746-006-5024-z.
- 109. Brito, C.; Dinis, L.-T.; Silva, E.; Gonçalves, A.; Matos, C.; Rodrigues, M.A.; Moutinho-Pereira, J.; Barros, A.; Correia, C. Kaolin and salicylic acid foliar application modulate yield, quality and phytochemical composition of olive pulp and oil from rainfed trees. *Sci. Hortic.* 2018, 237, 176–183, doi:10.1016/j.scienta.2018.04.019.
- 110. Chen, D.; Wang, S.; Cao, B.; Cao, D.; Leng, G.; Li, H.; Yin, L.; Shan, L.; Deng, X. Genotypic variation in growth and physiological response to drought stress and re-watering reveals the critical role of recovery in drought adaptation in maize seedlings. *Front. Plant Sci.* 2016, *6*, 1–15, doi:10.3389/fpls.2015.01241.
- Ennajeh, M.; Vadel, A.M.; Cochard, H.; Khemira, H. Comparative impacts of water stress on the leaf anatomy of a drought-resistant and a drought-sensitive olive cultivar. *J. Hortic. Sci. Biotechnol.* 2010, 85, 289– 294, doi:10.1080/14620316.2010.11512670.

- 113. Chartzoulakis, K.; Patakas, A.; Bosabalidis, A.M. Changes in water relations, photosynthesis and leaf anatomy induced by intermittent drought in two olive cultivars. *Environ. Exp. Bot.* **1999**, *42*, 113–120, doi:10.1016/S0098-8472(99)00024-6.
- Mediavilla, S.; Escudero, A.; Heilmeier, H. Internal leaf anatomy and photosynthetic resource-use efficiency: Interspecific and intraspecific comparisons. *Tree Physiol.* 2001, 21, 251–259, doi:10.1093/treephys/21.4.251.
- 115. Savé, R.; Biel, C.; de Herralde, F. Leaf pubescence, water relations and chlorophyll fluorescence in two subspecies of *Lotus Creticus* L. *Biol. Plant.* 2000, 43, 239–244, doi:10.1023/A:1002704327076.
- Veihmeyer, F.J.; Hendrickson, A.H. Soil moisture at permanent wilting of plants. *Plant Physiol.* 1928, 3, 355–357.
- Xiloyannis, C.; Dichio, B. Irrigazione. In Olea: Trattato di Olivicoltura; Fiorino, P., Ed.; Il Sole 24 ORE Edagricole Srl: Bologna, Italy, 2003; pp. 365–389.
- Dichio, B.; Xiloyannis, C.; Angelopoulos, K.; Nuzzo, V.; Bufo, S.A.; Celano, G. Drought-induced variations of water relations parameters in *Olea europaea*. *Plant Soil* 2003, 257, 381–389, doi:10.1023/A:1027392831483.
- Dichio, B.; Xiloyannis, C.; Sofo, A.; Montanaro, G. Osmotic regulation in leaves and roots of olive trees during a water deficit and rewatering. *Tree Physiol.* 2006, 26, 179–185, doi:10.1093/treephys/26.2.179.
- Moriana, A.; Orgaz, F.; Pastor, M.; Fereres, E. Yield responses of a mature olive orchard to water deficits. J. Am. Soc. Hortic. Sci. 2003, 128, 425–431, doi:10.21273/JASHS.12.
- Orgaz, F.; Fereres, E.; Barranco, D.; Fernández-Escobar, R.; Rallo, L. *El Cultivo del Olivo*, 6th ed.; Ediciones Mundi-Prensa y Junta de Andalucía: Madrid, Spain, 2008; pp. 337–362.
- 122. Dichio, B.; Montanaro, G.; Sofo, A.; Xiloyannis, C. Stem and whole-plant hydraulics in olive (*Olea europaea*) and kiwifruit (*Actinidia deliciosa*). *Trees* **2013**, *27*, 183–191, doi:10.1007/s00468-012-0787-3.
- Fernández Luque, J.E.; Aranda, M.; Moreno Lucas, F.; Rapoport, H.F. Anatomical response of olive roots to dry and irrigated soils. *Adv. Hortic. Sci.* 1994, 8, 141–144.
- 124. Nadezhdina, N.; Ferreira, M.I.; Conceição, N.; Pacheco, C.A.; Häusler, M.; David, T.S. Water uptake and hydraulic redistribution under a seasonal climate: Long-term study in a rainfed olive orchard. *Ecohydrology* 2015, *8*, 387–397, doi:10.1002/eco.1545.
- 125. Dichio, B.; Margiotta, G.; Xiloyannis, C.; Bufo, S.A.; Sofo, A.; Cataldi, T.R.I. Changes in water status and osmolyte contents in leaves and roots of olive plants (*Olea europaea* L.) subjected to water deficit. *Trees* 2009, 23, 247–256, doi:10.1007/s00468-008-0272-1.
- Sanders, G.J.; Arndt, S.K. Osmotic adjustment under drought conditions. In *Plant Responses to Drought Stress—From Morphological to Molecular Features*; Aroca, R., Ed.; Springer: New York, NY, USA, 2012; pp. 199–230.
- 127. Patakas, A.; Nikolaou, N.; Zioziou, E.; Radoglou, K.; Noitsakis, B. The role of organic solute and ion accumulation in osmotic adjustment in drought-stressed grapevines. *Plant Sci.* 2002, 163, 361–367, doi:10.1016/S0168-9452(02)00140-1.
- Joly, R.J.; Zaerr, J.B. Alteration of cell-wall water content and elasticity in Douglas-fir during periods of water deficit. *Plant Physiol.* 1987, 83, 418–422, doi:c10.1104/pp.83.2.418.
- 129. Zargar, S.M.; Nagar, P.; Deshmukh, R.; Nazir, M.; Wani, A.A.; Masoodi, K.Z.; Agrawal, G.K.; Rakwal, R. Aquaporins as potential drought tolerance inducing proteins: Towards instigating stress tolerance. J. Proteom. 2017, 169, 233–238, doi:10.1016/j.jprot.2017.04.010.
- 130. Šurbanovski, N.; Grant, O.M. The emerging role of aquaporins in plant tolerance of abiotic stress, emerging technologies and management of crop stress tolerance. In *Emerging Technologies and Management of Crop Stress Tolerance, Volume 2: A Sustainable Approach*; Ahmad, P.; Rasool, S., Eds.; Academic Press: San Diego, CA, USA, 2014; pp. 431–447.
- Secchi, F.; Lovisolo, C.; Schubert, A. Expression of OePIP2.1 aquaporin gene and water relations of *Olea* europaea twigs during drought stress and recovery. *Ann. Appl. Biol.* 2007, 150, 163–167, doi:10.1111/j.1744-7348.2007.00118.x.
- 132. Secchi, F.; Lovisolo, C.; Uehlein, N.; Kaldenhoff, R.; Schubert, A. Isolation and functional characterization of three aquaporins from olive (*Olea europaea* L.). *Planta* **2007**, 225, 381–392, doi:10.1007/s00425-006-0365-2.

- Rogiers, S.Y.; Greer, D.H.; Hatfield, J.M.; Hutton, R.J.; Clarke, S.J.; Hutchinson, P.A.; Somers, A. Stomatal response of an anisohydric grapevine cultivar to evaporative demand, available soil moisture and abscisic acid. *Tree Physiol.* 2012, 32, 249–261, doi:10.1093/treephys/tpr131.
- 134. Naor, A.; Schneider, D.; Ben-Gal, A.; Zipori, I.; Dag, A.; Kerem, Z.; Birger, R.; Peres, M.; Gal, Y. The effects of crop load and irrigation rate in the oil accumulation stage on oil yield and water relations of 'Koroneiki' olives. *Irrig. Sci.* 2013, 31, 781–791, doi:10.1007/s00271-012-0363-z.
- Mattos, L.; Moretti, C. Oxidative stress in plants under drought conditions and the role of different enzymes. *Enzyme Eng.* 2015, 5, 1–6, doi:10.4172/2329-6674.1000136.
- Ahmadipour, S.; Arji, I.; Ebadi, A.; Abdossi, V. Physiological and biochemical responses of some olive cultivars (*Olea europaea* L.) to water stress. *Cell Mol. Biol.* 2018, 64, 20–29.
- 137. Karimi, S.; Rahemi, M.; Rostami, A.A.; Sedaghat, S. Drought effects on growth, water content and osmoprotectants in four olive cultivars with different drought tolerance. *Int. J. Fruit Sci.* 2018, 18, 254–267, doi:10.1080/15538362.2018.1438328.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).