Supplementary Materials

Table S1. Oligonucleotide sequences and PCR conditions used to amplify the indicated genes. TA = annealing temperature; gDNA = genomic DNA; cDNA = complementary DNA; c = concentration.

Gene	Oligo Sequence (5'>3')	T _A in °C	c [MgCl] in mM	PCR-Product Size
EF	tcccttcaacactcctttatagc	58	2.5	gDNA: 2354 bp
	aacagtctatgcgacacgtca	30		cDNA: 1500 bp
H2A	cggggaaaggtgctaaaggtt	55	2.5	gDNA: 690 bp
	aaatgcctcggcgagatacgt	33	2.3	cDNA: 282 bp
LTP1	ctccaaggtggtgtcattcc	57	2.5	gDNA: 450 bp
	cccatataacaagaacaccacaa	31		cDNA: 380 bp
LTP3	tgttatcacccaaaaagaagttca	57	2.5	gDNA: 703 bp
	ttattattacgttcgtatgcgttgg	57		cDNA: 610 bp
LTP4	tatcacccaaaagagaagagca	<i>5</i>	2.5	gDNA: 649 bp
	acacaagtatacaacataacaaagc	54		cDNA: 510 bp
AT1G12090	cccaattcactcacaacctagc	58	1.5	gDNA: 630 bp
	atcacccaatgaacaccag			cDNA: 630 bp
LEDO	gcaacaacaaagaaaccccta	58	2.5	gDNA: 885 bp
LTP8	ttaggacaagatggaccattga			cDNA: 448 bp
AT5G05960	tcatcactcaagaatggaaacc		2.5	gDNA: 390 bp
	acgtctattgctttctgtctgc	56		cDNA: 479 bp
AT4G22610	taaatccaagcctcacctc		2.5	gDNA: 501 bp
	cagcaacaactacgatcatgc	55		cDNA: 501 bp
AT3G22620	cacacttcaaacacaaaaccac		2.5	gDNA: 979 bp
	cttattccacaaagcaatgacc	67		cDNA: 790 bp
AT4G33550	cggaccaaatattcgcattc	70	2.5	gDNA: 490 bp
	ccggaatggtgtaacctataaca	58		cDNA: 490 bp
AT1G62510	tcccaattcacacataccaaag	27	1.5	gDNA: 605 bp
	ggtagaaatcatcttgtctgtcca	27		cDNA: 605 bp
Pb Actin	agctggcgtacgtggcgcag	- 1	2.5	gDNA: 342 bp
	ccttgacgcgcatcgacgac	64		cDNA: 342 bp
nos-terminator	tatagcggccgcggatcgttcaaacatttggcaata	55	2.5	272 hm
	gegegageteatetagtaacataga	55	2.5	273 bp

Table S2. Oligonucleotide sequences and PCR conditions used to verify the T-DNA integration. TA = annealing temperature; c = concentration.

T-DNA Insertion Line Acc. No/ Disrupted Gene	Oligos Used 5'>3'	Site of T-DNA Insertion	T _A in	c [MgCl] in mM	PCR-Product Size without T-DNA
N647582/	cgatcctgtttcgcgtagat	Promoter 60		2,5	961
AT1G12090	atcacccaatgaacaccag	Tiomotei	00	2,3	
N520925/	tgttgaaagagcttgtagtaatgg	3' utr	58	2,5	1535
LTP8	gcaacaacaaagaaaccccta	3 uu			
N648038/	tccgaatacgacctacaatga	Promoter/	58	2,5	1564
AT3G22620	cttattccacaaagcaatgacc	5' utr			
N527420/	ttgaggactggagaaaagga	Promoter	58	2,5	872
AT5G06960	acgtctattgctttctgtctgc	Promoter 38		۷,3	072
N500561/	aaggattcttgcagtttcat	Duamatan	50	2.5	2002
LTP4	acacaagtatacaacataacaaagc	Promoter 58		2,5	2093
N595248/	tgttatcacccaaaaagaagttca	Even	58	2,5	703
LTP3	ttattattacgttcgtatgcgttgg	Exon			

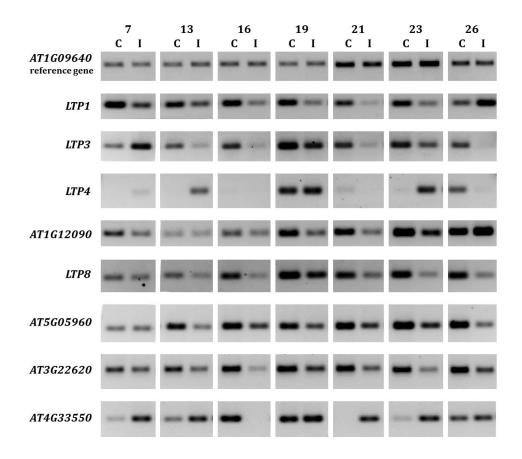


Figure S1. Expression of selected LTP genes in clubroot infected A. thaliana roots. The expression for some LTP genes during clubroot infection between 7 and 26 days after inoculation is shown. For the gene AT1G62510 the expression was not detectable. For infection a field isolate [44] was used. C = control; I = inoculated.

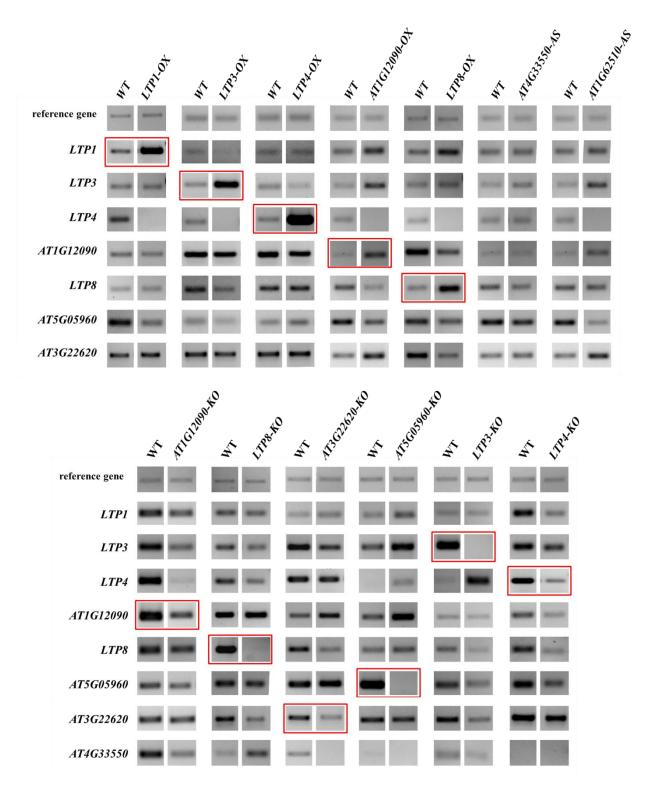


Figure S2. Expression of some *LTP* genes in roots of LTP mutants. The expression for the indicated *LTP* genes (left) in comparison to the wild type (WT) is shown. The elongation factor 1B gamma (*AT1G09640*) and histon H2A (*AT1G52740*) were used as reference genes. For the semi-quantitative expression analyses the RNA was extracted from 24-day-old *A. thaliana* roots. The red squares mark the expression of the overexpressed (OX) or down-regulated (KO) gene. For *AT1G62510* and *AT4G33550* the expression in the transgenic plants as well as in the wild type plants and for *AT1G62510* in the T-DNA insertion lines and their wild type control the gene expression was not detectable.

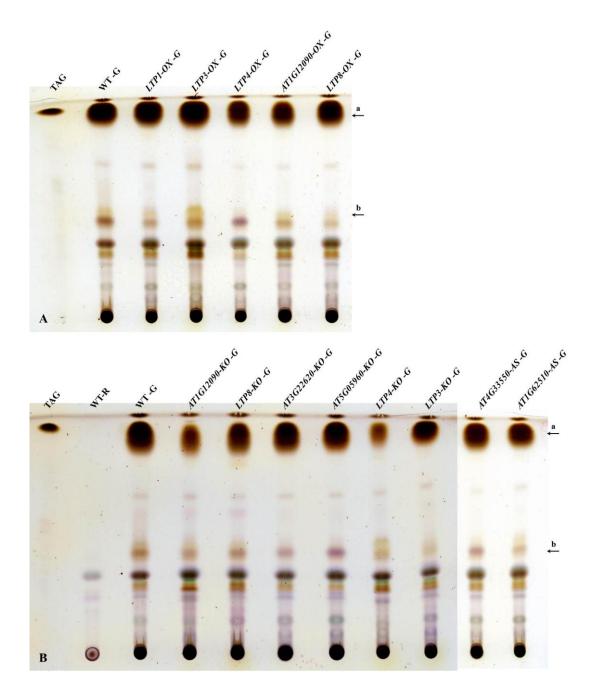


Figure S3. Lipid composition of uninfected roots and galls of *A. thaliana*. Results of thin layer chromatography from non-polar lipids isolated from equal amounts of infected roots (galls) 30 days after inoculation or of healthy root material of the same age. Plant material was from wild type (WT) and LTP mutants that (**A**) overexpress (OX) the genes *LTP1*, *LTP3*, *LTP4*, *AT1G12090* and *LTP8* and with reduced LTP gene expression (**B**) from T-DNA-insertion lines (KO) for the genes *AT1G12090*, *LTP8*, *AT3G22620*, *AT5G05960*, *LTP4*, *LTP3* and antisense lines (AS) for the genes *AT4G33550* and *AT1G62510*. Two biological replicates with approx. 25 plants each were analyzed. Triacylglycerol (TAG) was used as a standard. R = uninfected root; G = gall (infected root). For explanation of bands a and b see main text.

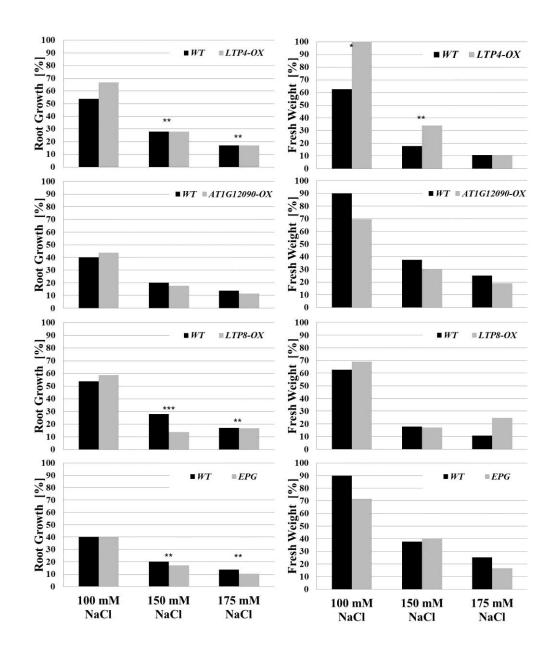


Figure S4. Growth reduction due to salt stress conditions. Root growth and whole plant fresh weight from wild type (WT), LTP mutants (LTP4-OX, AT1G12090-OX, LTP8-OX) and the empty vector control (EPG) in response to salt stress are shown. To calculate the root growth and fresh weight (in %) the root growth and fresh weight from unstressed plants (0 mM NaCl) were set to 100%. Therefore the graphs show the growth reduction due to salt stress. $n \ge 50$. Asterisks indicates a significant difference (for ** p < 0.01; *** p < 0.001). OX: overexpression of the indicated gene.

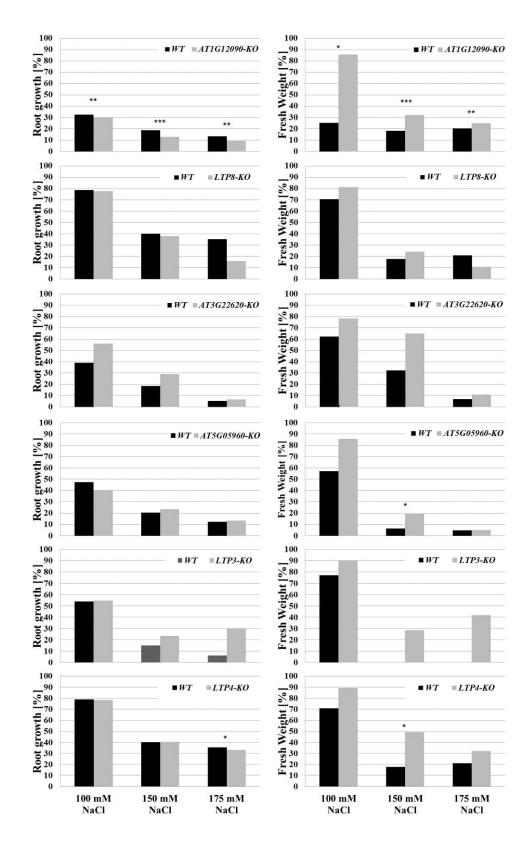


Figure S5. Growth reduction due to salt stress conditions. Root growth and whole plant fresh weight from wild type (WT) and LTP mutants (AT1G12090-KO, LTP8-KO, AT3G22620, AT5G05960-KO, LTP3-KO, LTP4-KO) in response to salt stress are shown. To calculate the root growth and fresh weight (in %) the root growth and fresh weight from unstressed plants (0 mM NaCl) were set to 100%. Therefore the graphs show the growth reduction due to salt stress. Since these graphs show a ratio based on the mean value the standard deviation could not be plotted on this graph. $n \ge 30$, KO: knockout or knockdown of the indicated gene. Asterisks indicates a significant difference (for * p < 0.05; *** p < 0.01; **** p < 0.001).

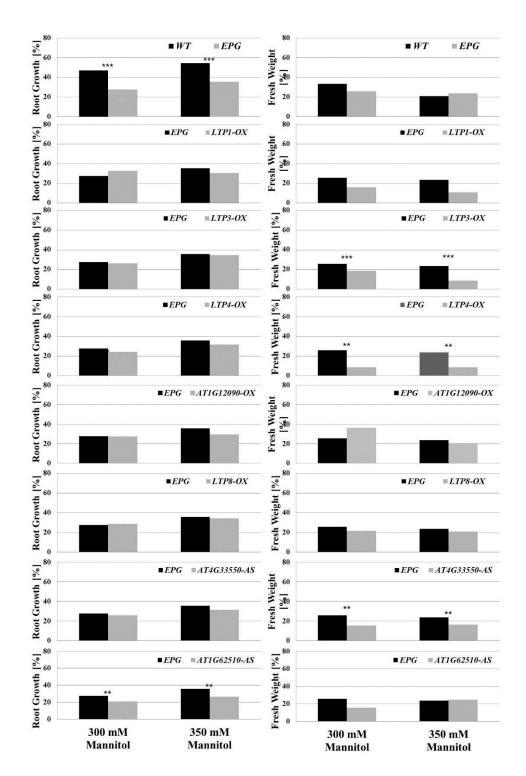


Figure S6. Growth reduction due to osmotic stress conditions. Root growth and whole plant fresh weight from wild type (WT) and LTP mutants (*LTP1-OX*, *LTP3-OX*, *LTP4-OX*, *AT1G12090-OX*, *LTP8-OX* and *AT4G33550-AS*, *AT1G62510-AS*) in response to mannitol treatment (300 mM, 350 mM) are shown. To calculate the root growth and fresh weight (in %) the root growth and fresh weight from unstressed plants (0 mM NaCl) were set to 100%. Because of the obvious differences between the vector control (EPG) and the wild type plants, the mutants were compared to the vector control plants (EPG). Since these graphs show a ratio based on the mean value the standard deviation could not be plottet on this graph. $n \ge 30$, OX: overexpression of the indicated gene; AS: silencing of the indicated gene using antisense technique. Asterisks indicates a significant difference (for ** p < 0.01; *** p < 0.001).

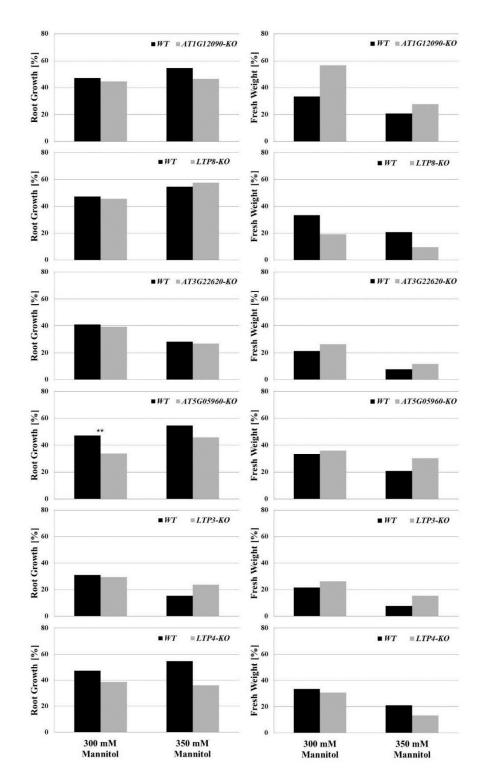


Figure S7. Growth reduction due to osmotic stress conditions. Root growth and whole plant fresh weight from wild type (WT) and LTP mutants (AT4G33550-AS, AT1G62510-AS, AT1G12090-KO, LTP8-KO, AT3G22620-KO, AT5G05960-KO, LTP3-KO, LTP4-KO) in response to mannitol treatment (300 mM, 350 mM) are shown. To calculate the root growth and fresh weight (in %) the root growth and fresh weight from unstressed plants (0 mM NaCl) were set to 100%. Since these graphs show a ratio based on the mean value the standard deviation could not be plotted on this graph. Because of the obvious differences between the vector control (EPG) and the wildtype plants (see S7). $n \ge 30$, KO: knockout or knockdown of the indicated gene, Asterisks indicates a significant difference (for ** p < 0.01).