



Article

Hormesis Responses of Growth and Photosynthetic Characteristics in *Lonicera japonica* Thunb. to Cadmium Stress: Whether Electric Field Can Improve or Not?

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Abstract: “Hormesis” is considered a dose–response phenomenon mainly observed at hyperaccumulator plants under heavy metals stress. In this study, the effects of electric fields on hormesis responses in *Lonicera japonica* Thunb. under cadmium (Cd) treatments were investigated by assessing the plant growth and photosynthetic characteristics. Under Cd treatments without electric fields, the parameters of plant growth and photosynthetic characteristics increased significantly when exposed to 5 mg L^{−1} Cd, and decreased slightly when exposed to 25 mg L^{−1} Cd, showing an inverted U-shaped trend, which confirmed that low concentration Cd has a hormesis effect on *L. japonica*. Under electric fields, different voltages significantly promoted the inverted U-shaped trend of the hormesis effect on the plant, especially by 2 V cm^{−1} voltage. Under 2 V cm^{−1} voltage, the dry weight of the root and leaf biomass exposed to 5 mg L^{−1} Cd increased significantly by 38.38% and 42.14%, and the photosynthetic pigment contents and photosynthetic parameters were also increased significantly relative to the control, indicating that a suitable electric field provides better improvements for the hormesis responses of the plant under Cd treatments. The synergistic benefits of the 5 mg L^{−1} Cd and 2 V cm^{−1} electric field in terms of the enhanced hormesis responses of growth and photosynthetic characteristics could contribute to the promoted application of electro-phytotechnology.

Keywords: hormesis; electric fields; heavy metal; *Lonicera japonica* Thunb.; phytoremediation



Citation: Liu, Z.; Tian, L.; Chen, M.; Zhang, L.; Lu, Q.; Wei, J.; Duan, X. Hormesis Responses of Growth and Photosynthetic Characteristics in *Lonicera japonica* Thunb. to Cadmium Stress: Whether Electric Field Can Improve or Not? *Plants* **2023**, *12*, 933. <https://doi.org/10.3390/plants12040933>

Academic Editor: Werther Guidi Nissim

Received: 30 December 2022

Revised: 16 February 2023

Accepted: 16 February 2023

Published: 18 February 2023



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1. Introduction

With rapid population growth and intensive human activities, large quantities of chemical contaminants, especially heavy metals released into the environment, have recently attracted global attention [1,2]. Heavy metals in soils are mainly derived from metalliferous mining and waste water irrigation, overuse of agricultural fertilizers and pesticides, warfare and military training, and over recent decades, heavy metals have become ubiquitous environmental contaminants all over the world [3,4]. Increasing emissions of heavy metals pose a significant threat to human health, because they may be accumulated in plants, animals or microorganisms and enter into the food chain [5–8]. Among heavy metals, cadmium (Cd) is one of the most hazardous pollutants and can cause leaf chlorosis, nutritional imbalance, and growth and photosynthesis inhibition [9–15]. Current methods for remediating Cd-contaminated soils rely primarily on physical and chemical techniques, which have the disadvantages of high operation cost, limited site scope, and complicated operation and may easily bring secondary contaminations or negative environmental effects [16,17].

In contrast, the phytoremediation technology of heavy metal-contaminated soil is widely considered as a promising and sustainable remediation strategy because of its superior characteristics such as being green, low cost and causing less secondary contamination. The key role of phytoremediation technology is hyperaccumulator uptake or the extraction of heavy metals from contaminated soils [18]. It is considered that the concentrations in hyperaccumulators for accumulating heavy metal elements in contaminated soils can reach more than 100 times of those found in non-accumulators [19,20]. Recently, several studies have indicated that electric fields could improve heavy metal accumulation and stimulate seed germination, growth and development of different plants responding to various environmental stresses [21–25]. The combination of hyperaccumulators and electric fields has also been proposed as a new method to promote remediation efficiency [26–30]. However, limited information is available on the effect of electric fields on the growth and photosynthetic characteristics of Cd-hyperaccumulators.

It is known that numerous studies focus on the toxicity of high-dose environmental contaminants [31–34]; however, some low-dose environmental contaminants may have beneficial effects on organisms [35–37]. The beneficial effects of low-dose environmental contaminants is widely recognized in the field of toxicology and medicine, where it is defined as “hormesis”, characterized by a biphasic adaptive response [38–40]. It is also found that hormesis can improve the adaptation of plants to some adverse environments, such as the low doses of nitrogen, lanthanum, ozone, ultraviolet radiation and herbicides [41–48]. Some researchers observed that hormesis can protect plants against environmental stress and enhance plant biomass productivity and functional components [49–51]. Nevertheless, few studies focus on the relationship of hormesis and hyperaccumulation. Thus, it is very necessary to investigate the hormesis response strategy of hyperaccumulators, especially the electric field-assisted effects on the hormesis responses of growth and photosynthetic characteristics in a hyperaccumulator.

Lonicera japonica Thunb. (Japanese honeysuckle) is a popular ornamental plant and has become established in temperate and tropical regions worldwide in the past 150 years [52]. It is commonly cultivated as a highly valued garden plant in urban greening because of its high biomass and easy cultivation, and its deep roots and shoots could reach as long as 150 cm. It also possesses the characteristics of extensive competitive ability, wide geographic distribution, and strong resistance to environmental stresses such as bacterial, viral and oxidative interference [53]. Our previous studies showed that *L. japonica* has a strong tolerance and good accumulation capability for Cd in plant tissues (the stem and shoot Cd accumulated concentrations in *L. japonica* can reach 344.49 ± 0.71 and $286.12 \pm 9.38 \mu\text{g g}^{-1}$ DW, respectively), and it is recognized as a new woody ornamental Cd-hyperaccumulator [9,54,55]. Moreover, we also found that the growth, photosynthetic pigment and relative water contents in *L. japonica* were stimulated by hormesis under low concentrations of Cd stress [9,54–57]. In the present study, we selected *L. japonica* as a model plant to investigate the effect of different electric fields on the hormesis responses of the growth, photosynthetic pigment composition and photosynthesis of the plant. The specific objectives are to explore whether an electric field can improve the hormesis responses of the plant under different concentrations of Cd stress. It will contribute to a better understanding of the hormesis response strategy of hyperaccumulators and promote the application of electro-phytotechnology.

2. Materials and Methods

2.1. Plant Materials and Experimental Treatments

The experiment was carried out in a greenhouse of Shenyang Agricultural University ($41^{\circ}44' \text{ N}$ and $123^{\circ}27' \text{ E}$, 44.7 m a.s.l.). Seedlings of *L. japonica* were collected from a non-contaminated experimental field and cultivated in sterilized sand by culture medium. The culture medium was Hoagland solution containing the following composition (mmol L^{-1}): $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ 5.00, $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ 2.00, KNO_3 5.00, KH_2PO_4 1.00, H_3BO_3 0.05, $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ 0.80×10^{-3} , $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$ 9.00×10^{-3} , $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ 0.30×10^{-3} ,

$(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot 4\text{H}_2\text{O}$ 0.02×10^{-3} , Fe-EDTA 0.10 [54,58]. The pH was measured by a pH meter and the pH value was 5.8 ± 0.1 .

After the plants were cultivated for 8 weeks, $\text{CdCl}_2 \cdot 2.5\text{H}_2\text{O}$ (Kermel Chemical Reagent Co., Ltd., Tianjin, China, >99%) was added into the culture medium and the Cd treatments were 0, 5 and 25 mg L^{-1} . Subsequently, an electric field (EF), which contains a pair of graphite electrodes (10.0 cm in length) associated with a DC power supply (220 V, 50 Hz), was daily 6 h and referred to 0, 1, 2 and 3 V cm^{-1} according to Liu et al. (2022) [59]. The EF-Cd treatments are shown in Table 1. The experiment consisted of three independent replicates. After one week, the plants were harvested for analysis.

Table 1. The treatments in the study.

Different Treatments	EF-Cd Treatment	Electric Field (V cm^{-1})	Cd Treatment (mg L^{-1})
T ₁	0-0	0 (V0)	0 (Cd0)
T ₂	0-5	0 (V0)	5 (Cd5)
T ₃	0-25	0 (V0)	25 (Cd25)
T ₄	1-0	1 (V1)	0 (Cd0)
T ₅	1-5	1 (V1)	5 (Cd5)
T ₆	1-25	1 (V1)	25 (Cd25)
T ₇	2-0	2 (V2)	0 (Cd0)
T ₈	2-5	2 (V2)	5 (Cd5)
T ₉	2-25	2 (V2)	25 (Cd25)
T ₁₀	3-0	3 (V3)	0 (Cd0)
T ₁₁	3-5	3 (V3)	5 (Cd5)
T ₁₂	3-25	3 (V3)	25 (Cd25)

2.2. Measurements of Plant Biomass and Cd Content

The harvested plants were washed with tap water, and the roots of the plants were immersed in 20 mM $\text{Na}_2\text{-EDTA}$ for 15 min and then washed with deionized water to remove Cd adhering to the root surface [9]. The plants were separated into leaves and roots. These plant tissues were dried at 105 °C for 20 min, then at 70 °C until the weight was constant. Subsequently, the dry weight (g) of the root and leaf biomass was obtained.

Dried plant materials were ground to fine powder by a grinder. The powders were digested with a concentrated acid mixture of $\text{HNO}_3/\text{HClO}_4$ (3:1, v/v). The plant Cd concentrations in *L. japonica* were determined with a flame atomic absorption spectrophotometer (Perkin-Elmer, Waltham, MA, USA) after microwave digestion.

2.3. Determination of Photosynthetic Pigment Contents

The photosynthetic pigments were measured by the uniform and similar leaf samples. The leaf samples (0.2 g) were cut into small pieces, and then soaked in 25 mL 95% (v/v) ethanol at 4 °C in darkness until the tissues became white. The extracting solution absorbance at 649, 665 and 470 nm was measured. The contents of chlorophyll a (Chla), chlorophyll b (Chlb), total chlorophyll (Total Chl) and carotenoid (Car) were calculated by a modified method according to Lichtenthaler and Wellburn (1983) and Lichtenthaler (1987) [60,61].

2.4. Assays of Photosynthetic Parameters

The photosynthetic parameters were determined by a portable photosynthesis system (LI-6400, Li-Cor Inc. Lincoln, NE, USA) under different treatments. The photosynthetic parameters consisted of net photosynthetic rate (Pn), stomatal conductance (Gs), transpiration rate (Tr) and intercellular CO_2 concentration (Ci). During different treatments, the parameters inside the leaf chamber (light level, CO_2 concentration and leaf temperature) were maintained constant at 1000 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD, 25 ± 0.3 °C and 380 ± 5 $\mu\text{mol CO}_2 \text{mol}^{-1}$. The upper second fully expanded leaves were used for the determination according to the method of Pandey et al. (2003) [62].

2.5. Statistical Analyses

All measurements in the study were replicated three times. The data analyses were performed as the means \pm SD. The statistical analysis of variance was carried out with SPSS 22.0. The significant difference was presented at the $p < 0.05$ level. The least significant difference (LSD) test was used to determine the multiple comparison between treatments.

3. Results and Discussion

3.1. Effects of Different EF-Cd Treatments on Plant Cd Concentrations

The Cd accumulations in *L. japonica* under different treatments were shown in Figure 1. With the increase of Cd concentrations in the medium without an electric field (T_1 – T_3 , Table 1), the plant Cd concentrations in *L. japonica* had an increased trend, which ranged from 11.11, 137.43 to 419.05 mg kg^{−1}. Under the T_4 – T_6 treatments (V1-Cd0, V1-Cd5 and V1-Cd25), a slight increase of plant Cd concentrations in *L. japonica* under different concentrations by Cd stress was observed by 1 V cm^{−1} voltages, which ranged from 12.31, 198.92 to 654.65 mg kg^{−1}. Under the T_7 – T_9 treatments (V2-Cd0, V2-Cd5 and V2-Cd25), the plant Cd concentrations in *L. japonica* under different concentrations by Cd stress had a significant increased trend compared with the T_1 – T_3 treatments (under Cd treatments without electric field), which ranged from 13.23, 358.30 to 1440.00 mg kg^{−1}. Under the T_{10} – T_{13} treatments (V3-Cd0, V3-Cd5 and V3-Cd25), the plant Cd concentrations in *L. japonica* under different concentrations by Cd stress had a more significant increase compared with the T_1 – T_3 treatments (under Cd treatments without electric field), which ranged from 13.99, 414.58 to 1630.84 mg kg^{−1}. It was demonstrated that the concentrations of several heavy metals (Cd, Cu, Zn and Pb) in plants were promoted because of the application of electric fields [27,63]. In the present study, the electric fields significantly enhanced the plant Cd concentrations in *L. japonica* exposed to different concentrations Cd compared with T_1 – T_3 treatments (under Cd treatments without electric fields). The significant increase of plant Cd concentrations under the electric field were observed when the plants were exposed to different concentrations of Cd, especially exposed to high concentrations (25 mg L^{−1}) Cd. Under different concentrations of Cd treatments, the plant Cd concentrations in *L. japonica* were increased significantly by 2 V cm^{−1} voltage and 3 V cm^{−1} voltage, which reached 1440.00 (T_9 , V2-Cd25) and 1630.84 mg kg^{−1} (T_{12} , V3-Cd25), which were 3.44 and 3.89 times of the T_3 treatment (V0-Cd25), respectively. The positive effect of the electric field may be correlated with the variety of metal ions polarity and cell membrane properties in plants [64,65]. The similar results have been reported by Klink et al. (2019) and Yuan et al. (2021), which mainly resulted from the electric field-induced increase of the membrane polarization rate, cell metabolism and activated ion channels [25,66].

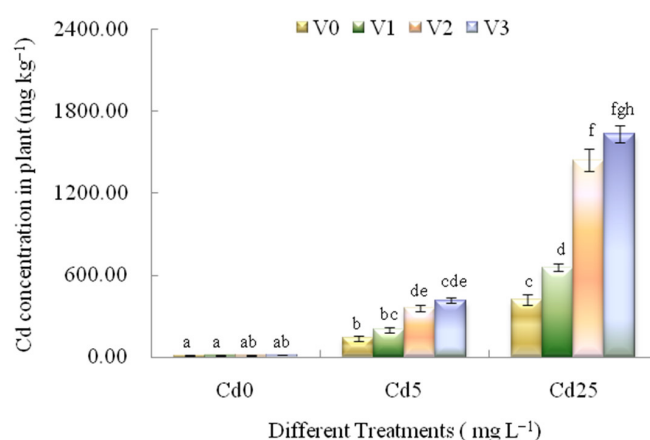


Figure 1. Effects of different EF-Cd treatments on plant Cd concentrations in *L. japonica*. V0, V1, V2 and V3 showed 0, 1, 2 and 3 V cm^{−1} electric fields. Cd0, Cd5, and Cd25 showed 0, 5 and 25 mg L^{−1} Cd treatments. Different letters indicate significant differences at the $p < 0.05$ level. Values represent mean \pm SD.

3.2. Effect of Different EF-Cd Treatments on Plant Growth

It is well known that the biomass of plants is an important highly sensitive indicator responding to heavy metal or other abiotic stresses [32,33]. The growth responses in the form of dry weight of the root and leaf biomass in *L. japonica* under different treatments is displayed in Figure 2. Under the T₁–T₃ treatments (under Cd treatments without electric fields), the dry weight of the root biomass exposed to 5 mg L^{−1} Cd increased significantly by 10.12% relative to the T₁ treatment (V0-Cd0), and had a slight decrease when exposed to 25 mg L^{−1} Cd, indicating an inverted U-shaped curve, which confirmed that low concentration Cd has the hormesis effect on the root growth of *L. japonica*. The results correspond to our previous studies, which showed that the growth characteristics, photosynthetic pigments contents, relative water contents and other physiological parameters of Cd treatments all significantly indicated an inverted U-shaped dose–response curve, confirming that the hormesis effect of low concentration Cd occurred in *L. japonica* [9,54–57]. Under the T₄–T₁₂ treatments (under electric field), the dry weight of root biomass had an increased trend compared with the T₁–T₃ treatments (under Cd treatments without electric fields), which showed that different voltages significantly promoted the inverted U-shaped trend of dry weight of root biomass in *L. japonica*, especially by 1 V cm^{−1} voltage and 2 V cm^{−1} voltage. Under 1 V cm^{−1} voltage and 2 V cm^{−1} voltage, the dry weight of root biomass exposed to 5 mg L^{−1} Cd increased significantly by 20.54% and 38.38% relative to the T₂ treatment (V0-Cd5), which investigated that the medium voltage (2 V cm^{−1}) has more improvements to the hormesis effect of low concentration Cd on the plant growth of *L. japonica*. He et al. (2017) have also reported that the dry weight of root biomass in maize under a drought environment was enhanced by a pulsed electric field, which could be derived from the improvement of the respiration metabolism and substance transformation through the pulsed electric field [67]. Under different voltages, the dry weight of root biomass exposed to 25 mg L^{−1} Cd increased by 15.12%, 34.30% and 10.47% relative to the T₃ treatment (V0-Cd25), which confirmed our previous study, indicating the electric field-enhanced tolerance of *L. japonica* responded to high concentrations of Cd.

In contrast, under the T₁–T₃ treatments (under Cd treatments without electric fields), the dry weight of leaf biomass exposed to 5 mg L^{−1} Cd increased significantly by 17.78% relative to the T₁ treatment (V0-Cd0), and decreased slightly when exposed to 25 mg L^{−1} Cd, showing a similar inverted U-shaped trend of the hormesis effect with the dry weight of root biomass. Wiewiórka (2013) observed that a high-intensity electric field had limited impacts on the growth of tomatoes in a hydroponic culture [64]. However, in the present study, under the T₄–T₁₂ treatments (under electric field), the dry weight of leaf biomass had an increased trend compared with the T₁–T₃ treatments (under Cd treatments without electric field), which indicated that different voltages significantly promoted the inverted U-shaped trend of the dry weight of leaf biomass in *L. japonica*, especially by 1 V cm^{−1} voltage and 2 V cm^{−1} voltage. Under 1 V cm^{−1} voltage and 2 V cm^{−1} voltage, the dry weight of leaf biomass exposed to 5 mg L^{−1} Cd increased significantly by 15.09% and 42.14% relative to the T₂ treatment (V0-Cd5). Compared with the results of the dry weight of root biomass above, it indicated that the medium voltage (2 V cm^{−1}) more significantly enhanced the hormesis effect of low concentration Cd on the leaf biomass than the root biomass in *L. japonica*, which could be the reason that plant organs in *L. japonica* have different sensitivity and tolerance mechanisms when responding to environmental stress. In summary, a medium strength electric field (2 V cm^{−1}) could improve the hormesis responses of plant growth in *L. japonica* under different treatments. This is in accordance with those earlier studies that reported that the electric field stimulated the plant growth and productivity through regulating the different levels of plant growth hormones [22,64,66].

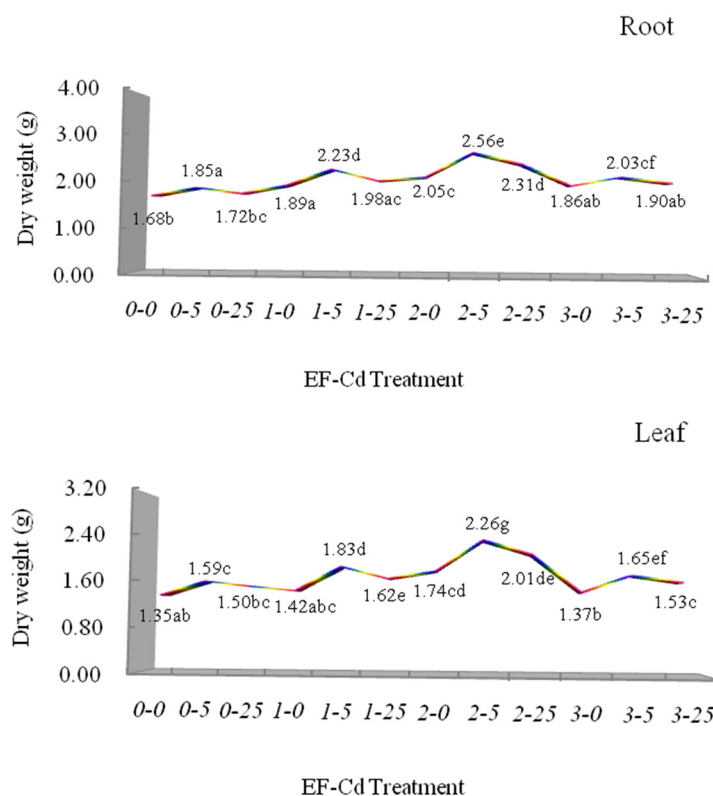


Figure 2. Effects of different EF-Cd treatments on dry weight of root and leaf biomass in *L. japonica*. Different colors showed the different responses in *L. japonica* under EF-Cd treatments. Different letters indicate significant differences at the $p < 0.05$ level. Values represent mean \pm SD.

3.3. Effect of Different EF-Cd Treatments on Photosynthetic Pigment Composition

The measured results of photosynthetic pigment composition including chlorophyll a (Chla), chlorophyll b (Chlb), total chlorophyll (Total Chl) and carotenoid (Car) in leaves of *L. japonica* are presented in Figure 3. Under the T_1 – T_3 treatments (under Cd treatments without electric fields), the contents of Chla, Chlb, Total Chl and Car exposed to 5 mg L^{-1} Cd increased significantly by 5.99%, 7.56%, 6.55% and 7.41% relative to T_1 treatment (V_0 -Cd0), and had a decrease exposed to 25 mg L^{-1} Cd, which showed an inverted U-shaped curve, indicating low concentration Cd has the hormesis effect on the photosynthetic pigment composition of *L. japonica*. The results confirmed that low concentration Cd could have a stimulatory effect on plant growth, the reasons of which may be the promoted dry matter accumulation and seedling biomass through the increased photosynthetic pigment contents [68–70]. When *L. japonica* was exposed to 25 mg L^{-1} Cd without electric fields (T_1 – T_3 treatments), the contents of Chla, Chlb, Total Chl and Car in the plant showed the decreased trend, which could have resulted from the substitution of chlorophyll Mg^{2+} in photosynthetic pigment composition by Cd^{2+} [71]. With the increase of Cd concentrations in the medium, more chlorophyll Mg^{2+} in the photosynthetic pigment composition are replaced spontaneously by Cd^{2+} and cause the degradation of photosynthetic pigments and even the inhibition of photosynthesis. Under the T_4 – T_{12} treatments (under electric fields), the contents of Chla, Chlb, Total Chl and Car had an increased trend compared with the T_1 – T_3 treatments (under Cd treatments without electric fields), which showed the electric field could improve the Cd-induced degradation of photosynthetic pigments and stimulate the protective mechanism in *L. japonica*. It was observed that the different voltages significantly promoted the inverted U-shaped trend of the contents of Chla, Chlb, Total Chl and Cars, especially by 1 V cm^{-1} voltage and 2 V cm^{-1} voltage. Moreover, different photosynthetic pigments have different sensibilities to environmental stress [72]. Under 1 V cm^{-1} voltage, the contents of Chla, Chlb, Total Chl and Cars exposed to 5 mg L^{-1} Cd increased significantly by 13.48%, 6.25%, 10.89% and 13.79% relative to the T_2 treatment

(V0-Cd5); by comparison, under 2 V cm^{-1} voltage, the contents of Chla, Chlb, Total Chl and Cars exposed to 5 mg L^{-1} Cd increased significantly by 24.78%, 23.44%, 24.30% and 22.41% relative to the T₂ treatment (V0-Cd5), which indicated that a medium voltage (2 V cm^{-1}) better promotes the hormesis effect of low concentration Cd on the photosynthetic pigment composition of *L. japonica*. The phenomenon is in agreement with the hormesis responses of plant growth in *L. japonica* under different treatments, which mainly result from electric field-induced uptake increase of Fe, Mg or other trace elements [32,73]. When the increased voltage reached 3 V cm^{-1} under the electric field, the contents of Chla, Chlb, Total Chl and Cars exposed to 5 mg L^{-1} Cd had no significant increases relative to the T₂ treatment (V0-Cd5), the contents of which were $2.45 \text{ mg g}^{-1}\text{FW}$, $1.29 \text{ mg g}^{-1}\text{FW}$, $3.74 \text{ mg g}^{-1}\text{FW}$ and $1.86 \text{ mg g}^{-1}\text{FW}$, respectively. The results indicated that a suitable electric field could have better improvement for the hormesis responses of photosynthetic pigment composition in *L. japonica* to Cd stress.

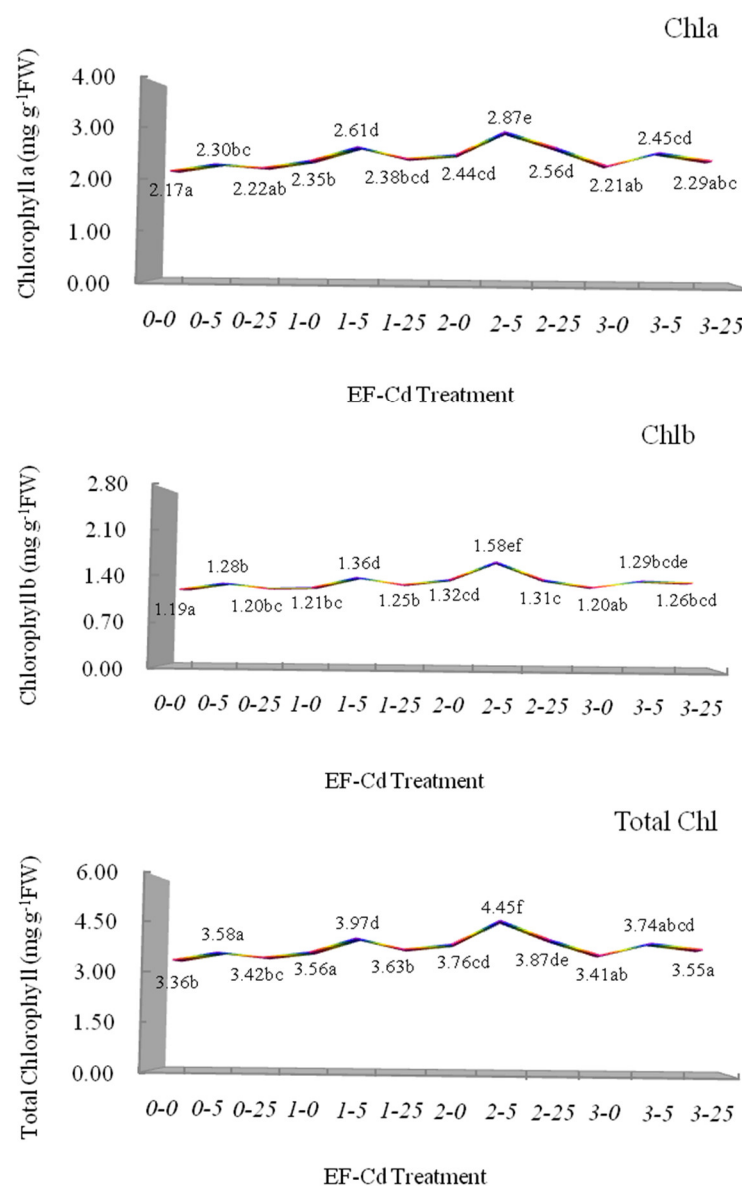


Figure 3. Cont.

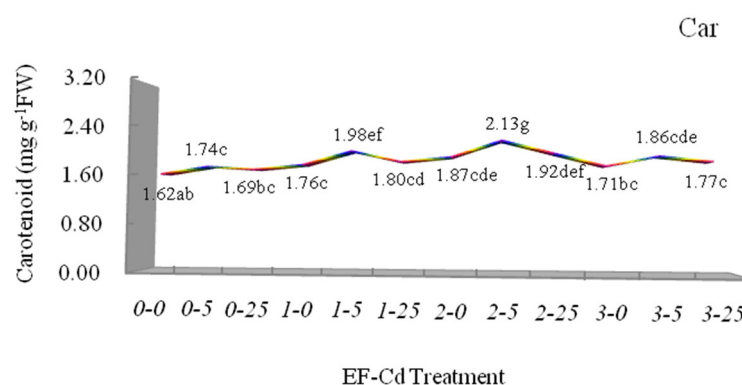


Figure 3. Effects of different EF-Cd treatments on the contents of Chlorophyll a (Chla), Chlorophyll b (Chlb), Total Chlorophyll (Chl) and Carotenoid (Car) in leaves of *L. japonica*. Different colors showed the different responses in *L. japonica* under EF-Cd treatments. Different letters indicate significant differences at the $p < 0.05$ level. Values represent mean \pm SD.

3.4. Effect of Different EF-Cd Treatments on Photosynthetic Parameters

Photosynthesis, as the basis of all plant growth and crop yield, is undoubtedly the most important biological process and is very susceptible to environments contaminated by Cd [74]. The photosynthesis responses in terms of the net photosynthetic rate (Pn), stomatal conductance (Gs), transpiration rate (Tr) and intercellular CO₂ concentration (Ci) in *L. japonica* under different treatments are evaluated in Table 2. Under the T₁–T₃ treatments (under Cd treatments without electric fields), when the plants were exposed to low concentration (5 mg L⁻¹) Cd, the contents of Pn, Gs, Tr and Ci in *L. japonica* had a significant increase compared with the T₁ treatment (V0-Cd0), which were $15.85 \pm 0.91 \mu\text{mol m}^{-2} \text{s}^{-1}$, $0.38 \pm 0.01 \text{mol m}^{-2} \text{s}^{-1}$, $2.52 \pm 0.05 \text{mmol m}^{-2} \text{s}^{-1}$ and $383.25 \pm 10.78 \mu\text{L L}^{-1}$, respectively. It is shown that the significant hormesis effect on Pn promoted the gas exchange and transpiration in *L. japonica* in the form of the increased Gs, Tr and Ci, the reasons of which may result from the stimulating impact of low concentration Cd on the Rubisco contents [74]. Under the T₄–T₁₂ treatments (under electric fields), when the plants were exposed to low concentration (5 mg L⁻¹) Cd, the contents of Pn, Gs, Tr and Ci in *L. japonica* were all increased significantly by 1 V cm⁻¹ voltage (T₅, V1-Cd5), 2 V cm⁻¹ voltage (T₈, V2-Cd5) and 3 V cm⁻¹ voltage (T₁₁, V3-Cd5), respectively. Under the T₄–T₁₂ treatments (under electric fields), different voltages significantly promoted the inverted U-shaped trend of the contents of Pn, Gs, Tr and Ci, especially by 2 V cm⁻¹ voltage. Under different EF-Cd treatments, the maximum value of Pn, Gs and Tr reached $22.95 \pm 0.98 \mu\text{mol m}^{-2} \text{s}^{-1}$, $1.19 \pm 0.05 \text{mol m}^{-2} \text{s}^{-1}$ and $3.33 \pm 0.08 \text{mmol m}^{-2} \text{s}^{-1}$, and under low concentration (5 mg L⁻¹) Cd treatment, the contents of Pn, Gs, Tr and Ci were all increased significantly by 2 V cm⁻¹ voltage (T₈, V2-Cd5). This is in agreement with the dry weight of root and leaf biomass, which showed that the combination of low concentration (5 mg L⁻¹) Cd and medium voltage (2 V cm⁻¹) was useful to improve the photosynthesis capacity and plant growth. The photosynthesis responses, in terms of Pn, Gs, Tr and Ci in *L. japonica* under different treatments, also have a good correlation with the change trend of the photosynthetic pigment composition. Several researchers observed that Cd stress had a negative impact on plant photosynthesis, which is probably traceable in the decreased chlorophyll biosynthesis and thylakoids or the inhibited plant growth [68,75–77]. However, in the present study, under high concentration (25 mg L⁻¹) Cd treatment, the contents of Pn, Gs, Tr and Ci in *L. japonica* were promoted significantly by electric fields relative to the T₃ treatment (V0-Cd25), which is probably associated with the adaptive mechanisms of hyperaccumulators responding to external stress [78–80].

Table 2. Effect of different EF-Cd treatments on photosynthetic parameters in *L. japonica*.

Different Treatments	Pn ($\mu\text{mol m}^{-2} \text{s}^{-1}$)	Gs ($\text{mol m}^{-2} \text{s}^{-1}$)	Tr ($\text{mmol m}^{-2} \text{s}^{-1}$)	Ci ($\mu\text{L L}^{-1}$)
T ₁	13.61 ± 0.45 a	0.16 ± 0.02 ab	1.77 ± 0.10 a	357.23 ± 8.15 a
T ₂	15.85 ± 0.91 b	0.38 ± 0.01 d	2.52 ± 0.05 bc	383.25 ± 10.78 b
T ₃	14.26 ± 0.36 c	0.22 ± 0.02 bc	2.38 ± 0.03 d	320.92 ± 16.51 cd
T ₄	15.65 ± 0.83 ab	0.36 ± 0.02 d	1.97 ± 0.07 ab	337.51 ± 11.02 c
T ₅	19.72 ± 0.42 d	0.77 ± 0.04 e	2.91 ± 0.04 e	342.97 ± 18.65 abc
T ₆	16.33 ± 0.85 abc	0.43 ± 0.02 bcd	2.59 ± 0.12 cd	341.86 ± 8.97 ab
T ₇	17.62 ± 0.51 bc	0.56 ± 0.03 cde	2.17 ± 0.06 d	316.45 ± 13.52 cd
T ₈	22.95 ± 0.98 ef	1.19 ± 0.05 fg	3.33 ± 0.08 gh	375.08 ± 19.33 b
T ₉	18.74 ± 0.72 cd	0.67 ± 0.03 ef	2.83 ± 0.11 def	365.63 ± 9.82 bc
T ₁₀	14.13 ± 0.69 a	0.21 ± 0.02 b	1.82 ± 0.09 ab	281.36 ± 17.20 ef
T ₁₁	17.45 ± 0.57 cde	0.54 ± 0.01 de	2.68 ± 0.06 def	320.27 ± 11.26 cd
T ₁₂	15.56 ± 0.62 b	0.35 ± 0.03 abc	2.51 ± 0.05 c	333.29 ± 14.91 bcd

Data are means ± SD. Different letters indicate significant differences at the $p < 0.05$ level. Pn: net photosynthetic rate; Gs: stomatal conductance; Tr: transpiration rate; Ci: intercellular CO₂ concentration.

4. Conclusions

Based on the previous study, it is shown that *L. japonica* is a good model plant to investigate the effect of different electric fields on the hormesis responses of the growth, photosynthetic pigment composition and photosynthesis of plants. In the study, under the T₁–T₃ treatments (under Cd treatments without electric fields), the parameters of plant growth (dry weight of root and leaf biomass), photosynthetic pigment composition (Chla, Chlb, Total Chl and Cars) and photosynthesis (Pn, Gs, Tr and Ci) increased significantly when exposed to 5 mg L^{−1} Cd ($p < 0.05$), and had a slight decrease when exposed to 25 mg L^{−1} Cd, showing an inverted U-shaped trend, which confirmed that low concentration Cd has a hormesis effect on *L. japonica*. Under the T₄–T₁₂ treatments (under electric field), different voltages significantly promoted the inverted U-shaped trend of the hormesis effect, especially by 2 V cm^{−1} voltage, which indicated that a suitable electric field better improves the hormesis responses of growth photosynthetic pigment composition and photosynthesis in *L. japonica* to Cd stress. The present results will be useful to explore the underlying mechanisms of the hormesis effect of Cd stress on hyperaccumulators for electric field-assisted phytoremediation.

Author Contributions: Conceptualization, Z.L., Q.L. and L.T.; data curation, Z.L. and M.C.; formal analysis, Z.L. and L.Z.; funding acquisition, L.T.; methodology, Z.L. and J.W.; software, Z.L. and X.D.; writing—original draft, Z.L.; writing—review and editing, Z.L., Q.L. and L.T. All authors have read and agreed to the published version of the manuscript.

Funding: This study is financially supported by the funding project of Northeast Geological S&T Innovation Center of China Geological Survey (No. QCJJ2022-44), Science and Technology Project for Black Soil Granary (XDA28080200, XDA28020400), the National Natural Science Foundation of China (42007043, 41920104008, 41771200, 41171399), Special Foundation for Basic Research Program in Wild China of CAS (XDA23070501), Key Laboratory Foundation of Mollisols Agroecology (2020ZKHT-02), National Key Research and Development Program of China (No. 2022YFD1500201) and Natural Science Foundation of Jilin Province (YDZJ202201ZYTS472).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in the study are available on request from the corresponding author. The data are not publicly available due to the restriction policy of the coauthors' affiliations.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Haso, H.W.; Dubale, A.A.; Chimdesa, M.A.; Atlabachew, M. High performance copper based metal organic framework for removal of heavy metals from wastewater. *Front. Mater.* **2022**, *9*, 840806. [\[CrossRef\]](#)
- Mukherjee, A.G.; Wanjari, U.R.; Renu, K.; Vellingiri, B.; Gopalakrishnan, A.V. Heavy metal and metalloid-induced reproductive toxicity. *Environ. Toxicol. Phar.* **2022**, *92*, 103859. [\[CrossRef\]](#) [\[PubMed\]](#)
- Pandey, A.K.; Zoric, L.; Su, T.; Karanovic, D.; Fang, P.P.; Borisev, M.; Wu, X.Y.; Lukovic, J.; Xu, P. The anatomical basis of heavy metal responses in legumes and their impact on plant-rhizosphere interactions. *Plants* **2022**, *11*, 2554. [\[CrossRef\]](#)
- Punia, P.; Bharti, M.K.; Dhar, R.; Thakur, P.; Thakur, A. Recent advances in detection and removal of heavy metals from contaminated water. *ChemBioEng. Rev.* **2022**, *9*, 351–369. [\[CrossRef\]](#)
- Zulkafflee, N.S.; Redzuan, N.A.M.; Nematbakhsh, S.; Selamat, J.; Ismail, M.R.; Praveena, S.M.; Lee, S.Y.; Razis, A.F.A. Heavy metal contamination in *Oryza sativa* L. at the eastern region of Malaysia and its risk assessment. *Int. J. Env. Res. Pub. He.* **2022**, *19*, 739. [\[CrossRef\]](#)
- Yang, L.; Wang, J.B.; Yang, Y.S.; Li, S.; Wang, T.X.; Oleksak, P.; Chrienova, Z.; Wu, Q.H.; Nepovimova, E.; Zhang, X.J.; et al. Phytoremediation of heavy metal pollution: Hotspots and future prospects. *Ecotox. Environ. Safe.* **2022**, *234*, 113403. [\[CrossRef\]](#)
- Shi, L.; Li, J.; Palansooriya, K.N.; Chen, Y.H.; Hou, D.Y.; Meers, E.; Tsang, D.C.W.; Wang, X.N.; Ok, Y.S. Modeling phytoremediation of heavy metal contaminated soils through machine learning. *J. Hazard. Mater.* **2023**, *441*, 129904. [\[CrossRef\]](#)
- Pande, V.; Pandey, S.C.; Sati, D.; Bhatt, P.; Samant, M. Microbial interventions in bioremediation of heavy metal contaminants in agroecosystem. *Front. Microbiol.* **2022**, *13*, 824084. [\[CrossRef\]](#)
- Liu, Z.L.; Chen, W.; He, X.Y. Influence of Cd²⁺ on growth and chlorophyll fluorescence in a hyperaccumulator—*Lonicera japonica* Thunb. *J. Plant Growth Regul.* **2015**, *34*, 672–676. [\[CrossRef\]](#)
- Andresen, E.; Kappel, S.; Stark, H.J.; Riegger, U.; Borovec, J.; Mattusch, J.; Heinz, A.; Schmelzer, C.E.H.; Matouskova, S.; Dickinson, B. Cadmium toxicity investigated at the physiological and biophysical levels under environmentally relevant conditions using the aquatic model plant *Ceratophyllum demersum*. *New Phytol.* **2016**, *210*, 1244–1258. [\[CrossRef\]](#)
- Borges, K.L.R.; Salvato, F.; Alcântara, B.K.; Nalin, R.S.; Piotto, F.Â.; Azevedo, R.A. Temporal dynamic responses of roots in contrasting tomato genotypes to cadmium tolerance. *Ecotoxicology* **2018**, *27*, 245–258. [\[CrossRef\]](#) [\[PubMed\]](#)
- He, J.L.; Zhuang, X.L.; Zhou, J.T.; Sun, L.Y.; Wan, H.X.; Li, H.F.; Lyu, D.G. Exogenous melatonin alleviates cadmium uptake and toxicity in apple rootstocks. *Tree Physiol.* **2020**, *40*, 746–761. [\[CrossRef\]](#) [\[PubMed\]](#)
- Zhang, Z.W.; Dong, Y.Y.; Feng, L.Y.; Deng, Z.L.; Xu, Q.; Tao, Q.; Wang, C.Q.; Chen, Y.E.; Yuan, M.; Yuan, S. Selenium enhances cadmium accumulation capability in two mustard family species—*Brassica napus* and *B. juncea*. *Plants* **2020**, *9*, 904. [\[CrossRef\]](#) [\[PubMed\]](#)
- Irshad, M.A.; Rehman, M.Z.U.; Anwar-ul-Haq, M.; Rizwan, M.; Nawaz, R.; Shakoar, M.B.; Wijaya, L.; Alyemeni, M.N.; Ahmad, P.; Ali, S. Effect of green and chemically synthesized titanium dioxide nanoparticles on cadmium accumulation in wheat grains and potential dietary health risk: A field investigation. *J. Hazard. Mater.* **2021**, *415*, 125585. [\[CrossRef\]](#)
- Liu, Z.L.; Chen, M.D.; Lin, M.S.; Chen, Q.L.; Lu, Q.X.; Yao, J.; He, X.Y. Cadmium uptake and growth responses of seven urban flowering plants: Hyperaccumulator or bioindicator? *Sustainability* **2022**, *14*, 619. [\[CrossRef\]](#)
- Liu, Z.L.; Chen, W.; He, X.Y. Evaluation of hyperaccumulation potentials to cadmium (Cd) in six ornamental species (compositae). *Int. J. Phytoremediat.* **2018**, *20*, 1464–1469. [\[CrossRef\]](#)
- Aboelkassem, A.; Alzamel, N.M.; Alzain, M.N.; Loutfy, N. Effect of Pb-contaminated water on *Ludwigia stolonifera* (Guill. & Perr.) P.H. Raven physiology and phytoremediation performance. *Plants* **2022**, *11*, 636.
- Durante-Yáñez, E.V.; Martínez-Macea, M.A.; Enamorado-Montes, G.; Combatt Caballero, E.; Marrugo-Negrete, J. Phytoremediation of soils contaminated with heavy metals from gold mining activities using *Clidemia sericea* D. Don. *Plants* **2022**, *11*, 597. [\[CrossRef\]](#)
- Pouresmaeli, M.; Ataei, M.; Forouzandeh, P.; Azizollahi, P.; Mahmoudifard, M. Recent progress on sustainable phytoremediation of heavy metals from soil. *J. Environ. Chem. Eng.* **2022**, *10*, 108482. [\[CrossRef\]](#)
- Skuza, L.; Szucko-Kociuba, I.; Filip, E.; Bozek, I. Natural molecular mechanisms of plant hyperaccumulation and hypertolerance towards heavy metals. *Int. J. Mol. Sci.* **2022**, *23*, 9335. [\[CrossRef\]](#)
- Harikumar, P.S.P.; Megha, T. Treatment of heavy metals from water by electro-phytoremediation technique. *J. Ecol. Eng.* **2017**, *18*, 18–26.
- Schmiedchen, K.; Petr, A.K.; Driessen, S.; Bailey, W.H. Systematic review of biological effects of exposure to static electric fields. Part II: Invertebrates and plants. *Environ. Res.* **2018**, *160*, 60–76. [\[CrossRef\]](#)
- Chang, J.H.; Dong, C.D.; Shen, S.Y. The lead contaminated land treated by the circulation-enhanced electrokinetics and phytoremediation in field scale. *J. Hazard. Mater.* **2019**, *368*, 894–898. [\[CrossRef\]](#)
- Cao, Y.B.; Wang, X.; Zhang, X.Y.; Misselbrook, T.; Bai, Z.H.; Ma, L. An electric field immobilizes heavy metals through promoting combination with humic substances during composting. *Bioresour. Technol.* **2021**, *330*, 124996. [\[CrossRef\]](#) [\[PubMed\]](#)
- Yuan, L.Z.; Guo, P.H.; Guo, S.H.; Wang, J.N.; Huang, Y.J. Influence of electrical fields enhanced phytoremediation of multi-metal contaminated soil on soil parameters and plants uptake in different soil sections. *Environ. Res.* **2021**, *198*, 111290. [\[CrossRef\]](#)
- Chirakkara, R.A.; Reddy, K.R.; Cameselle, C. Electrokinetic amendment in phytoremediation of mixed contaminated soil. *Electrochim. Acta.* **2015**, *181*, 179–191. [\[CrossRef\]](#)

27. Luo, J.; Cai, L.; Qi, S.; Wu, J.; Gu, X.S. Influence of direct and alternating current electric fields on efficiency promotion and leaching risk alleviation of chelator assisted phytoremediation. *Ecotoxicol. Environ. Saf.* **2018**, *149*, 241–247. [\[CrossRef\]](#) [\[PubMed\]](#)
28. Cameselle, C.; Gouveia, S. Phytoremediation of mixed contaminated soil enhanced with electric current. *J. Hazard. Mater.* **2019**, *361*, 95–102. [\[CrossRef\]](#)
29. Cameselle, C.; Gouveia, S.; Urrejola, S. Benefits of phytoremediation amended with DC electric field. Application to soils contaminated with heavy metals. *Chemosphere* **2019**, *229*, 481–488. [\[CrossRef\]](#)
30. Taamalli, M.; Ghabriche, R.; Amari, T.; Mnasri, M.; Zolla, L.; Lutts, S.; Abdely, C.; Ghnaya, T. Comparative study of Cd tolerance and accumulation potential between *Cakile maritima* L. (halophyte) and *Brassica juncea* L. *Ecol. Eng.* **2014**, *71*, 623–627. [\[CrossRef\]](#)
31. Zeng, P.; Guo, G.; Xiao, X.; Peng, C.; Feng, W. Phytoextraction potential of *Pteris vittata* L. co-planted with woody species for As, Cd, Pb and Zn in contaminated soil. *Sci. Total Environ.* **2019**, *650*, 594–603. [\[CrossRef\]](#)
32. Szopinski, M.; Sitko, K.; Rusinowski, S.; Zieleznik-Rusinowska, P.; Corso, M.; Rostanski, A.; Rojek-Jelonek, M.; Verbruggen, N.; Malkowski, E. Different strategies of Cd tolerance and accumulation in *Arabidopsis halleri* and *Arabidopsis arenosa*. *Plant Cell Environ.* **2020**, *43*, 3002–3019. [\[CrossRef\]](#)
33. Chen, L.; Hu, W.F.; Long, C.; Wang, D. Exogenous plant growth regulator alleviate the adverse effects of U and Cd stress in sunflower (*Helianthus annuus* L.) and improve the efficacy of U and Cd remediation. *Chemosphere* **2021**, *262*, 127809. [\[CrossRef\]](#)
34. Huang, Y.F.; Chen, J.H.; Sun, Y.M.; Wang, H.X.; Zhan, J.Y.; Huang, Y.N.; Zou, J.W.; Wang, L.; Su, N.N.; Cui, J. Mechanisms of calcium sulfate in alleviating cadmium toxicity and accumulation in pak choi seedlings. *Sci. Total Environ.* **2022**, *805*, 150115. [\[CrossRef\]](#)
35. Nascentes, R.F.; Carbonari, C.A.; Simoes, P.S.; Brunelli, M.C.; Velini, E.D.; Duke, S.O. Low doses of glyphosate enhance growth, CO₂ assimilation, stomatal conductance and transpiration in sugarcane and eucalyptus. *Pest Manag. Sci.* **2018**, *74*, 1197–1205. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Anunciato, V.M.; Bianchi, L.; Gomes, G.L.; Velini, E.D.; Duke, S.O.; Carbonari, C.A. Effect of low glyphosate doses on flowering and seed germination of glyphosate-resistant and -susceptible *Digitaria insularis*. *Pest Manag. Sci.* **2022**, *78*, 1227–1239. [\[CrossRef\]](#)
37. Yang, Y.F.; Xie, L.L.; Peng, Y.H.; Yan, H.P.; Huang, J.T.; Xiao, Z.H.; Lu, X.L. Single-cell transcriptional profiling reveals low-level tragus stimulation improves sepsis-induced myocardial dysfunction by promoting M₂ macrophage polarization. *Oxidative Med. Cell. Longev.* **2022**, *2022*, 3327583. [\[CrossRef\]](#) [\[PubMed\]](#)
38. Calabrese, E.J.; Baldwin, I.A. Toxicology rethinks its central belief: Hormesis demands a reappraisal of the way risks are assessed. *Nature.* **2003**, *421*, 691–692. [\[CrossRef\]](#) [\[PubMed\]](#)
39. Calabrese, E.J.; Blain, R.B. Hormesis and plant biology. *Environ. Pollut.* **2009**, *157*, 42–48. [\[CrossRef\]](#)
40. Agathokleous, E.; Calabrese, E.J. Hormesis: The dose response for the 21st century: The future has arrived. *Toxicology* **2019**, *425*, 152249. [\[CrossRef\]](#)
41. Sicard, P.; Anav, A.; De Marco, A.; Paoletti, E. Projected global ground-level ozone impacts on vegetation under different emission and climate scenarios. *Atmos. Chem. Phys.* **2017**, *17*, 12177–12196. [\[CrossRef\]](#)
42. Agathokleous, E.; Kitao, M.; Calabrese, E.J. The rare earth element (REE) lanthanum (La) induces hormesis in plants. *Environ. Pollut.* **2018**, *238*, 1044–1047. [\[CrossRef\]](#)
43. Belz, R.G. Herbicide hormesis can act as a driver of resistance evolution in weeds—PSII-target site resistance in *Chenopodium album* L. as a case study. *Pest Manag. Sci.* **2018**, *74*, 2874–2883. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Agathokleous, E.; Kitao, M.; Calabrese, E.J. Hormetic dose responses induced by lanthanum in plants. *Environ. Pollut.* **2019**, *244*, 332–341. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Heine, K.B.; Powers, M.J.; Kallenberg, C.; Tucker, V.L.; Hood, W.R. Ultraviolet irradiation increases size of the first clutch but decreases longevity in a marine copepod. *Ecol. Evolut.* **2019**, *9*, 9759–9767. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Ji, K.; Wang, Y.; Du, L.; Xu, C.; Liu, Y.; He, N.; Wang, J.; Liu, Q. Research progress on the biological effects of low-dose radiation in China. *Dose-Response* **2019**, *17*, 1559325819833488. [\[CrossRef\]](#) [\[PubMed\]](#)
47. Volkova, P.Y.; Clement, G.; Makarenko, E.S.; Kazakova, E.A.; Bitarishvili, S.V.; Lychenkova, M.A. Metabolic profiling of gamma-irradiated barley plants identifies reallocation of nitrogen metabolism and metabolic stress response. *Dose-Response* **2020**, *18*, 1559325820914186. [\[CrossRef\]](#)
48. Pishenin, I.; Gorbatova, I.; Kazakova, E.; Podobed, M.; Mitsenyk, A.; Shesterikova, E.; Dontsova, A.; Dontsov, D.; Volkova, P. Free amino acids and methylglyoxal as players in the radiation hormesis effect after low-dose gamma-irradiation of barley seeds. *Agriculture* **2021**, *11*, 918. [\[CrossRef\]](#)
49. Abbas, T.; Nadeem, M.A.; Tanveer, A.; Chauhan, B.S. Can hormesis of plant- released phytotoxins be used to boost and sustain crop production? *Crop Protect.* **2017**, *93*, 69–76. [\[CrossRef\]](#)
50. Agathokleous, E.; Calabrese, E.J. Hormesis can enhance agricultural sustainability in a changing world. *Glob. Food Sec.* **2019**, *20*, 150–155. [\[CrossRef\]](#)
51. Choudhary, R.C.; Kumaraswamy, R.V.; Kumari, S.; Sharma, S.S.; Pal, A.; Raliya, R.; Biswas, P.; Saharan, V. Zinc encapsulated chitosan nanoparticle to promote maize crop yield. *Int. J. Biol. Macromol.* **2019**, *127*, 126–135. [\[CrossRef\]](#)
52. Larson, B.M.H.; Catling, P.M.; Waldron, G.E. The biology of Canadian weeds. 135. *Lonicera japonica* Thunb. *Can. J. Plant Sci.* **2007**, *87*, 423–438. [\[CrossRef\]](#)

53. Thanabhorn, S.; Jaijoy, K.; Thamaree, S.; Ingkaninan, K.; Panthong, A. Acute and subacute toxicity study of the ethanol extract from *Lonicera japonica* Thunb. *J. Ethnopharmacol.* **2006**, *107*, 370–373. [\[CrossRef\]](#)
54. Liu, Z.L.; He, X.Y.; Chen, W.; Yuan, F.H.; Yan, K.; Tao, D.L. Accumulation and tolerance characteristics of cadmium in a potential hyperaccumulator—*Lonicera japonica* Thunb. *J. Hazard. Mater.* **2009**, *169*, 170–175. [\[CrossRef\]](#)
55. Liu, Z.L.; He, X.Y.; Chen, W. Effects of cadmium hyperaccumulation on the concentrations of four trace elements in *Lonicera japonica* Thunb. *Ecotoxicology* **2011**, *20*, 698–705. [\[CrossRef\]](#) [\[PubMed\]](#)
56. Liu, Z.L.; Chen, W.; He, X.Y.; Jia, L.; Huang, Y.Q.; Zhang, Y.; Yu, S. Cadmium-induced physiological response in *Lonicera japonica* Thunb. *Clean-Soil Air Water* **2013**, *41*, 478–484. [\[CrossRef\]](#)
57. Liu, Z.L.; Chen, W.; He, X.Y.; Jia, L.; Yu, S.; Zhao, M.Z. Hormetic responses of *Lonicera Japonica* Thunb. to cadmium stress. *Dose-Response* **2015**, *13*, 1–10. [\[CrossRef\]](#)
58. Hoagland, D.R.; Arnon, D.I. The water-culture method for growing plants without soil. *Calif. Agric. Exp. Stn. Circ.* **1950**, *347*, 32.
59. Liu, Z.L.; Chen, Q.L.; Lin, M.S.; Chen, M.D.; Zhao, C.; Lu, Q.X.; Meng, X.Y. Electric field-enhanced cadmium accumulation and photosynthesis in a woody ornamental hyperaccumulator—*Lonicera japonica* Thunb. *Plants* **2022**, *11*, 1040. [\[CrossRef\]](#) [\[PubMed\]](#)
60. Lichtenthaler, H.K.; Wellburn, A.R. Determinations of total carotenoids and chlorophylls a and b of leaf extracts in different solvents. *Biochem. Soc. Trans.* **1983**, *11*, 591–592. [\[CrossRef\]](#)
61. Lichtenthaler, H.K. Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. *Method. Enzymol.* **1987**, *148*, 350–382.
62. Pandey, S.; Kumar, S.; Nagar, P.K. Photosynthetic performance of *Ginkgo biloba* L. grown under high and low irradiance. *Photosynthetica* **2003**, *41*, 505–511. [\[CrossRef\]](#)
63. Bi, R.; Schlaak, M.; Siefert, E.; Lord, R.; Connolly, H. Influence of electrical fields (AC and DC) on phytoremediation of metal polluted soils with rapeseed (*Brassica napus*) and tobacco (*Nicotiana tabacum*). *Chemosphere* **2011**, *83*, 318–326. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Wiewiórka, Z. The effects of electromagnetic and electrostatic fields on the development and yield of greenhouse tomato plants. *Acta. Agrobot.* **2013**, *43*, 25–36. [\[CrossRef\]](#)
65. Sánchez, V.; Francisco, J.; López-Bellido, J.; Rodrigo, M.A.; Rodríguez, L. Enhancing the removal of atrazine from soils by electrokinetic-assisted phytoremediation using ryegrass (*Lolium perenne* L.). *Chemosphere* **2019**, *232*, 204–212. [\[CrossRef\]](#)
66. Klink, A.; Polechonska, L.; Dambiec, M.; Bienkowski, P.; Klink, J.; Salamacha, Z. The influence of an electric field on growth and trace metal content in aquatic plants. *Int. J. Phytoremediat.* **2019**, *21*, 246–250. [\[CrossRef\]](#)
67. He, R.R.; Xi, G.; Liu, K. Alleviating effect of extremely low frequency pulsed electric field on drought damage of maize seedling roots. *J. Lumin.* **2017**, *188*, 441–447. [\[CrossRef\]](#)
68. Mobin, M.; Khan, N.A. Photosynthetic activity, pigment composition and antioxidative response of two mustard (*Brassica juncea*) cultivars differing in photosynthetic capacity subjected to cadmium stress. *J. Plant Physiol.* **2007**, *164*, 601–610. [\[CrossRef\]](#)
69. González-Casado, S.; Martín-Belloso, O.; Elez-Martínez, P.; Soliva-Fortuny, R. Enhancing the carotenoid content of tomato fruit with pulsed electric field treatments: Effects on respiratory activity and quality attributes. *Postharvest Biol. Tec.* **2018**, *137*, 113–118. [\[CrossRef\]](#)
70. Agathokleous, E.; Feng, Z.; Peñuelas, J. Chlorophyll hormesis: Are chlorophylls major components of stress biology in higher plants? *Sci. Total Environ.* **2020**, *1*, 38637. [\[CrossRef\]](#)
71. Grajek, H.; Rydzynski, D.; Piotrowicz-Cieślak, A.; Herman, A.; Maciejczyk, M.; Wieczorek, Z. Cadmium ion-chlorophyll interaction e Examination of spectral properties and structure of the cadmium-chlorophyll complex and their relevance to photosynthesis inhibition. *Chemosphere* **2020**, *261*, 127434. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Kummerova, M.; Zezulka, S.; Kral'ova, K.; Masarovicova, E. Effect of zinc and cadmium on physiological and production characteristics in *Matricaria recutita*. *Biol. Plantarum.* **2010**, *54*, 308–314. [\[CrossRef\]](#)
73. Haque, A.M.; Tasnim, J.; El-Shehawi, A.M.; Rahman, M.A.; Parvez, M.S.; Ahmed, M.B.; Kabir, A.H. The Cd-induced morphological and photosynthetic disruption is related to the reduced Fe status and increased oxidative injuries in sugar beet. *Plant Physiol. Bioch.* **2021**, *166*, 448–458. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Ying, R.R.; Qiu, R.L.; Tang, Y.T.; Hu, P.J.; Qiu, H.; Chen, H.R.; Shi, T.H.; Morel, J.L. Cadmium tolerance of carbon assimilation enzymes and chloroplast in Zn/Cd hyperaccumulator *Picris divaricata*. *J. Plant Physiol.* **2010**, *167*, 81–87. [\[CrossRef\]](#) [\[PubMed\]](#)
75. Dai, H.P.; Wei, Y.; Zhang, Y.Z.; Gao, P.X.; Chen, J.W.; Jia, G.L.; Yang, T.X.; Feng, S.J.; Wang, C.F.; Wang, Y.; et al. Influence of photosynthesis and chlorophyll synthesis on Cd accumulation in *Populusxcanescens*. *J. Food Agric. Environ.* **2012**, *10*, 1020–1023.
76. Singh, S.; Prasad, S. IAA alleviates Cd toxicity on growth, photosynthesis and oxidative damages in eggplant seedlings. *Acta Physiol. Plant.* **2015**, *77*, 87–98. [\[CrossRef\]](#)
77. Dobrikova, A.G.; Apostolova, E.L.; Hanc, A.; Yotsova, E.; Borisova, P.; Sperdouli, I.; Adamakis, I.D.S.; Moustakas, M. Cadmium toxicity in *Salvia sclarea* L.: An integrative response of element uptake, oxidative stress markers, leaf structure and photosynthesis. *Ecotox. Environ. Safe.* **2021**, *209*, 111851. [\[CrossRef\]](#)
78. Calabrese, E.J.; Bachmann, K.A.; Bailer, A.J.; Bolger, P.M.; Borak, J.; Cai, L.; Cedergreen, N.; Cherian, M.G.; Chiueh, C.C.; Clarkson, T.W.; et al. Biological stress response terminology: Integrating the concepts of adaptive response and preconditioning stress within a hormetic dose-response framework. *Toxicol. Appl. Pharmacol.* **2007**, *222*, 122–128. [\[CrossRef\]](#)

-
79. Kim, S.A.; Lee, Y.M.; Choi, J.Y.; Jacobs, D.R., Jr.; Lee, D.H. Evolutionarily adapted hormesis-inducing stressors can be a practical solution to mitigate harmful effects of chronic exposure to low dose chemical mixtures. *Environ. Pollut.* **2018**, *233*, 725–734. [[CrossRef](#)]
 80. Erofeeva, E.A. Environmental hormesis of non-specific and specific adaptive mechanisms in plants. *Sci. Total Environ.* **2022**, *804*, 150059. [[CrossRef](#)]

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