

Article

Silicon Supply Improves Nodulation and Dinitrogen Fixation and Promotes Growth in *Trifolium incarnatum* Subjected to a Long-Term Sulfur Deprivation

Raphaël Coquerel ¹, Mustapha Arkoun ², Quentin Dupas ¹, Fanny Leroy ³, Philippe Lainé ^{1,†} and Philippe Etienne ^{1,*,†}

¹ Unicaen, INRAE, UMR 950 EVA, SF Normandie Végétal (FED4277), Normandie Université, 14000 Caen, France; raphael.coquerel@unicaen.fr (R.C.); quentin.dupas@unicaen.fr (Q.D.); philippe.laine@unicaen.fr (P.L.)

² Laboratoire de Nutrition Végétale, Agro Innovation International-TIMAC AGRO, 35400 Saint-Malo, France; mustapha.arkoun@roullier.com

³ Plateau Technique d'Isotopie de Normandie (PLATIN'), Unité de Services EMERODE, Normandie Université, 14000 Caen, France; fanny.leroy@unicaen.fr

* Correspondence: philippe.etienne@unicaen.fr

† These authors contributed equally to this work.

Abstract: In many crops species, sulfur (S) deprivation negatively affects growth, seed yield quality and plant health. Furthermore, silicon (Si) is known to alleviate many nutritional stresses but the effects of Si supply on plants subjected to S deficiency remain unclear and poorly documented. The objective of this study was to evaluate whether Si supply would alleviate the negative effects of S deprivation on root nodulation and atmospheric dinitrogen (N₂) fixation capacity in *Trifolium incarnatum* subjected (or not) to long-term S deficiency. For this, plants were grown for 63 days in hydroponic conditions with (500 μM) or without S and supplied (1.7 mM) or not with Si. The effects of Si on growth, root nodulation and N₂ fixation and nitrogenase abundance in nodules have been measured. The most important beneficial effect of Si was observed after 63 days. Indeed, at this harvest time, a Si supply increased growth, the nitrogenase abundance in nodules and N₂ fixation in S-fed and S-deprived plants while a beneficial effect on the number and total biomass of nodules was only observed in S-deprived plants. This study shows clearly for the first time that a Si supply alleviates negative effects of S deprivation in *Trifolium incarnatum*.

Keywords: N₂ fixation; nodules; S deficiency; silicon; ¹⁵N labeling; *Trifolium incarnatum*



Citation: Coquerel, R.; Arkoun, M.; Dupas, Q.; Leroy, F.; Lainé, P.; Etienne, P. Silicon Supply Improves Nodulation and Dinitrogen Fixation and Promotes Growth in *Trifolium incarnatum* Subjected to a Long-Term Sulfur Deprivation. *Plants* **2023**, *12*, 2248. <https://doi.org/10.3390/plants12122248>

Academic Editor: Berca Mihai

Received: 24 May 2023

Revised: 5 June 2023

Accepted: 6 June 2023

Published: 8 June 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

At the end of the 20th century, various policies were implemented to drastically decrease SO₂ emissions into the atmosphere [1,2] in order to improve air quality and reduce the negative effects of these emissions on human health [3]. Consequently, this legislation has led to a strong decrease in sulfur (S) deposits on agricultural soils and has increased the occurrence of S deficiency [4], especially in plants requiring high S such as brassicas [5–7] and legumes [8–11]. A sulfur deficiency can lead to many metabolic perturbations [12,13] such as reduced atmospheric dinitrogen (N₂) fixation [8,14] and nitrogen (N) uptake from soil [15] and it usually leads to sharp drops in the yield and quality of the harvested product [9,16]. Finally, due to the decrease in seed quality, sulfur deficiency has indirect negative impacts on the health of human and animal consumers [17]. Currently, use of S fertilizers at a dose of about 30 kg S ha⁻¹ [18] is the most widely used strategy to support S deficient crops. However, in the global agroecological context of reducing inputs, it is urgent that other strategies be developed to alleviate the stress of plants experiencing S deficiency. Among them is consideration of supplementation with silicon (Si), whose beneficial effects on plants subjected to biotic [19,20] and abiotic [21] stresses has been widely described in

the literature. For example, a recent review reported that Si mitigates the negative effect of abiotic stresses as early as seed germination [22] and several studies have reported that Si supply enables alleviation of plants experiencing drought [23] or heat stress [24]. In the same way, a Si supply enables better tolerance to salt stress by sustaining various biological process such as photosynthesis or by stimulating detoxification of reactive oxygen species (ROS) [25]. Moreover, it has been shown that Si supplementation is beneficial in plants subjected to nutrient deficiency, such as deficits in N, P and K, and to a lesser extent S [26–28]. In legumes, the beneficial effects of silicon have also been demonstrated in plants subject to a large panel of abiotic stresses. For example, in several legume species exposed to salt stress, Si supply leads to maintenance of atmospheric N₂ fixation by increasing the number of nodules and their leghemoglobin concentration and nitrogenase activity [29,30]. Although Si has several beneficial effects under a variety of adverse conditions, there are currently no studies that have focused on the effects of Si application to legumes subjected to S-deficiency. Indeed, currently only three studies demonstrate beneficial effects of Si application in barley [31], rice [32] and rapeseed [28] subjected to S-deficiency. In this study, we focused on *Trifolium incarnatum* because previous studies have shown that this legume species is of agroecological interest especially when it cultivated in mixed crops. For example, in *Trifolium incarnatum*-*Brassica napus* intercrop, several works recently report shown that the transfer of nitrogenous compounds from clover to rapeseed improved the agronomic performance of this field crop [18,33,34].

The objectives of this study were to assess the effect of Si supply in *Trifolium incarnatum* subjected (or not) to a long-term S deficiency (63 days). For this, the effect of Si supply on the growth, nodulation (number, biomass and nitrogenase abundance of nodules) and atmospheric N₂ fixation capacity was performed in S-deprived or S-fed plants and those supplied or not with Si (1.7 mM) over 21, 42 and 63 days. Finally, the beneficial effects of Si supply on growth, nodulation and N₂ fixation observed in S-fed and mainly S-deprived plants have been discussed.

2. Results

2.1. Effect of Si Supply on Growth of *Trifolium incarnatum* Subjected or Not to S-Deprivation

At D21, whole plant biomasses were similar and reached about 5 g plant⁻¹, regardless of the S and Si supplies. At day 42, irrespective of the Si supply (-Si or +Si), S-deprivation had no impact on whole plant biomasses (Figure 1). However, it is notable that the Si supply led to a significant increase in shoot biomass (around 1.5-fold) in both S-fed and S-deprived plants. In comparison to S-fed plants, at D63 a S-deprivation led to a significant decrease in total biomass when plants were cultivated without Si, which was as expected. In contrast, whichever S treatment was considered (+S or -S), Si supplementation resulted in an increase in total plant biomass (by 1.2- and 2-fold in +S+Si and -S+Si plants, respectively; Figures 1 and S1). In S-fed plants (+S+Si), this effect is explained by an increase in shoot biomass alone. In S-deprived plants, this was partly the result of an increase in root biomass, but mainly due to a strong increase in shoot biomass. Interestingly, it could be highlighted that a Si supply offset the negative effect of S deprivation on plant growth because the total biomass of -S+Si plants was increased by around 13 g plant⁻¹ compared to -S-Si plants, and reached a biomass similar to S-fed plants (24.75 g plant⁻¹ ± 2.5 g) (Figures 1 and S1).

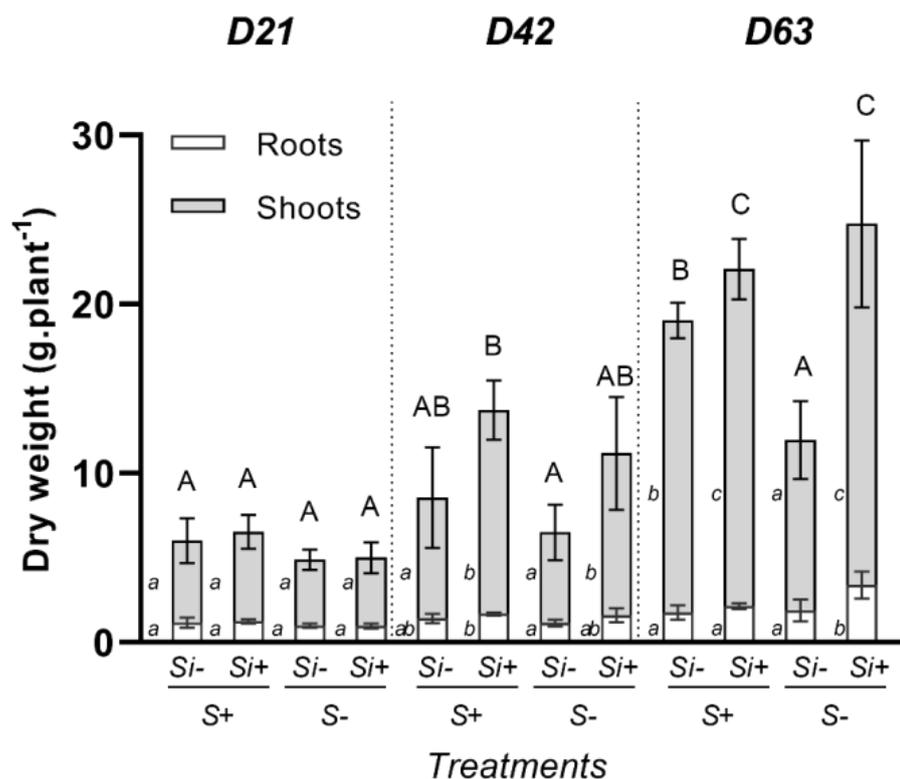


Figure 1. Root and shoot dry weights of *Trifolium incarnatum* plants cultivated in hydroponic conditions for 21 (D21), 42 (D42) and 63 days (D63) with (+S; 500 μ M) or without (-S) sulfur and supplied with (+Si; 1.7 mM) or without (-Si) Si. Data are means \pm SE ($n = 5$). Different lowercase and uppercase letters indicate that the mean dry weight between organs or whole plants harvested at the same time (D21, D42 or D63) are significantly different ($p < 0.05$), respectively.

2.2. Determination of Total Sulfur (S) and Silicon (Si) Concentrations in Plants

As expected, the S concentration in roots and shoots in S-deprived plants were always lower than in those of S-fed plants (Figure 2A,B), whatever the harvest date (D21, D42 or D63). In general, Si supply had no effect on S concentrations in plants compartments except at D21 where the S concentrations in roots and shoots in -S+Si plants were slightly lower than those of -S-Si plants (Figure 2A,B). In the same manner, whichever S treatment was considered (-S or +S), the Si concentrations in roots and shoots of plants cultivated without Si (+S-Si or -S-Si) were always lower than those of plants supplied with Si (-S+Si or +S+Si). Moreover, in plants supplied with Si, it was evident that the Si concentrations in roots increased throughout the treatment (from 1.8 to 3.5 mg g⁻¹ DW on average), while the Si concentration in shoots was always lower and remained stable (around 1.1 mg g⁻¹ DW), regardless of the harvest date (Figure 2C,D). These data show that, as previously demonstrated in *Brassica napus* by [26], *Trifolium incarnatum* preferentially accumulates Si in roots. Finally, it was observed that Si concentrations were not affected by the presence or the lack of S in the nutrient solution (Figure 2C,D).

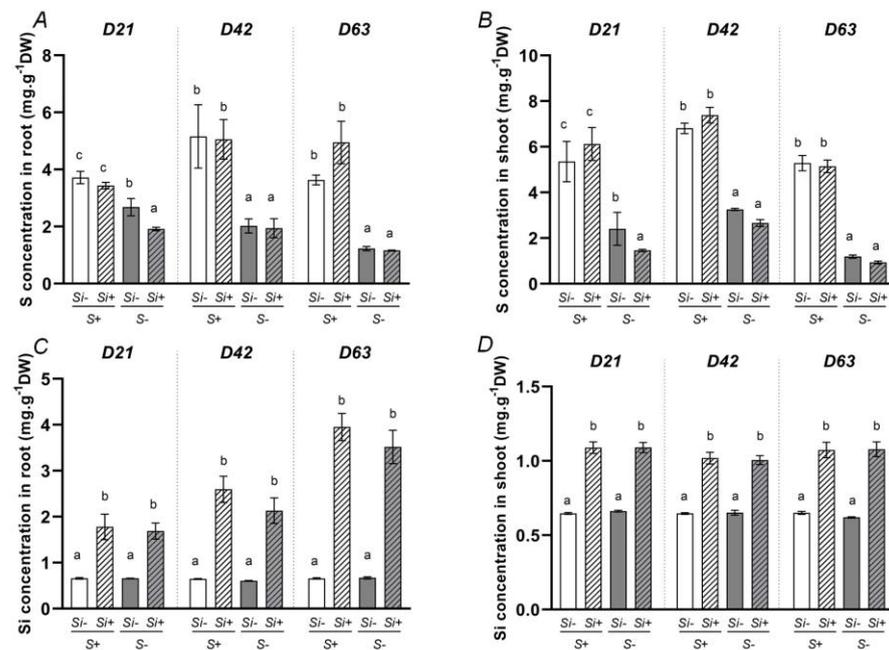


Figure 2. Sulfur (A,B) and silicon (C,D) concentrations in roots and shoots of *Trifolium incarnatum* plants cultivated in hydroponic conditions for 21 (D21), 42 (D42) and 63 days (D63) with (+S; 500 μ M) or without (-S) sulfur and supplied with (+Si; 1.7 mM) or without (-Si) Si. Data are means \pm SE ($n = 5$). Different lowercase letters indicate that the means between plants harvested at the same time (D21, D42 or D63) are significantly different ($p < 0.05$).

2.3. Effect of Silicon Supply on Root Nodulation

In S-fed plants, regardless of the harvest date (D21, D42 or D63) or whether plants were treated with or without Si, the number of nodules per plant (approximately 7000 nodules plant⁻¹; Figure 3A), the weight of a single nodule (Figure 3B), and the weight of all nodules per plant (Figure 3C) remained similar. Compared to +S-Si plants, in -S-Si plants, the number of nodules (around 4000 nodules plant⁻¹), the weight of a single of nodule (Figure 3B) and the weight of all nodules by plant (Figure 3C) decreased for all treatment durations. These results showing a negative effect of S-deprivation on nodulation agree with previously reported work performed in *Trifolium repens*, another clover species [8]. In contrast, surprisingly, the provision of a Si supply to -S+Si plants alleviated the negative effect of S deprivation on the number of nodules per plant, which again was similar to the observations in S-fed plants at D21 and D42 (between 6000 and 7000 nodules plant⁻¹) and even to a high increase at 63 days (15,000 vs. 7000 nodules plant⁻¹ in -S+Si and +S-Si, respectively) (Figure 3A). In addition, while the addition of Si in these -S+Si plants did not affect the weights of individual nodules or even decreased their individual weights at D63, it nevertheless led to an increase in the total weight of the nodules per plant at D21 (around 2-fold), D42 (around 1.5-fold) and more especially at D63 (around 4-fold) compared to the -S-Si plants (Figure 3C).

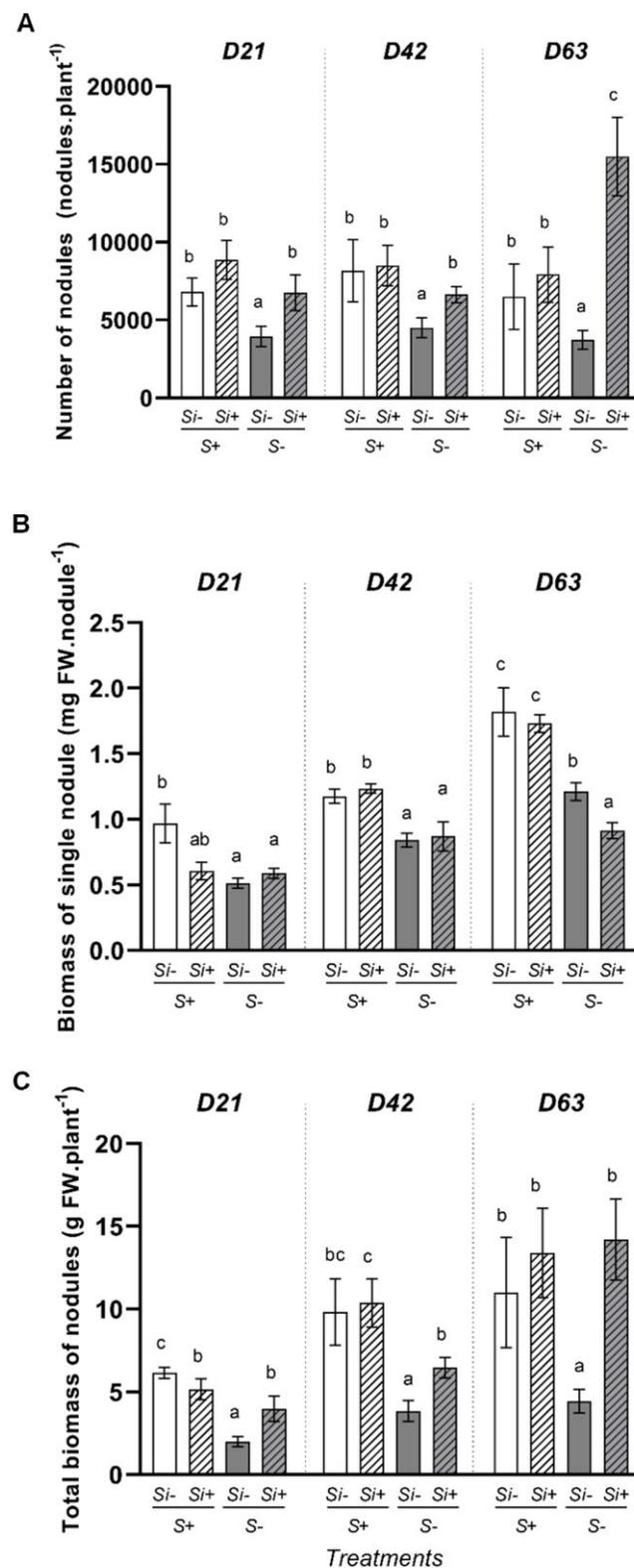


Figure 3. Number of nodules (A), fresh weight of individual nodules (B) and total fresh weight of all the nodules (C) of *Trifolium incarnatum* plants cultivated in hydroponic conditions for 21 (D21), 42 (D42) and 63 days (D63) with (+S; 500 μ M) or without (-S) sulfur and with (+Si; 1.7 mM) or without (-Si) Si supply. Data are means \pm SE ($n = 5$). Different lowercase letters indicate that the means between plants harvested at the same time (D21, D42 or D63) are significantly different ($p < 0.05$).

2.4. Effect of Silicon Supply on the Total Plant N Derived from Uptake and N₂ Fixation

Whatever the S treatment (+S or -S), at D21 the Si supply had no effect on either the amount of N taken up from nutrient solution or the amount of N derived from atmospheric N₂ fixation (Figure 4A). In contrast, at D42, a Si supply led only to an increase in the N amount derived from atmospheric N₂ fixation in plants cultivated with or without S. For example, at D42, in S-fed plants, the Si supply led to an increase in the amount of N₂ derived from the atmosphere by around 1.8-fold and contributed to an increase in the total N amount from 296 ± 28 to 434 ± 23 mg plant⁻¹ compared to +S-Si plants (Figure 4A). In addition, at D63, it can be pointed out that in plants grown without Si, S deprivation significantly reduced (by 1.5-fold) the total amount of nitrogen in plants from 644 ± 45 to 400 ± 41 mg plant⁻¹. This decrease is explained by a decrease in the amount of nitrogen taken up from the nutrient solution and especially by a decrease in N₂ fixation capacity, which dropped from 45 to 30% (Figure 4B). At this harvest time, irrespective of the S treatment (+S or -S), a Si supply increased the N₂ fixation capacity, which resulted in an increase in the total N amount in plants. However, this Si effect was exacerbated in S-deprived plants. Indeed, in a spectacular way, supplying Si to S-deprived plants allowed a strong increase in both the amount of N fixed (around 2.7-fold) and the total N (around 1.8-fold), which reached values comparable to those observed in S-fed plants. Thus, by improving the N₂ fixation of S-deprived plants (by 30–55%; Figure 4B), a Si supply mitigates the negative effect of S deprivation on N nutrition and thus maintains the total amount of N in -S plants at a level similar to that observed in +S plants (Figure 4A).

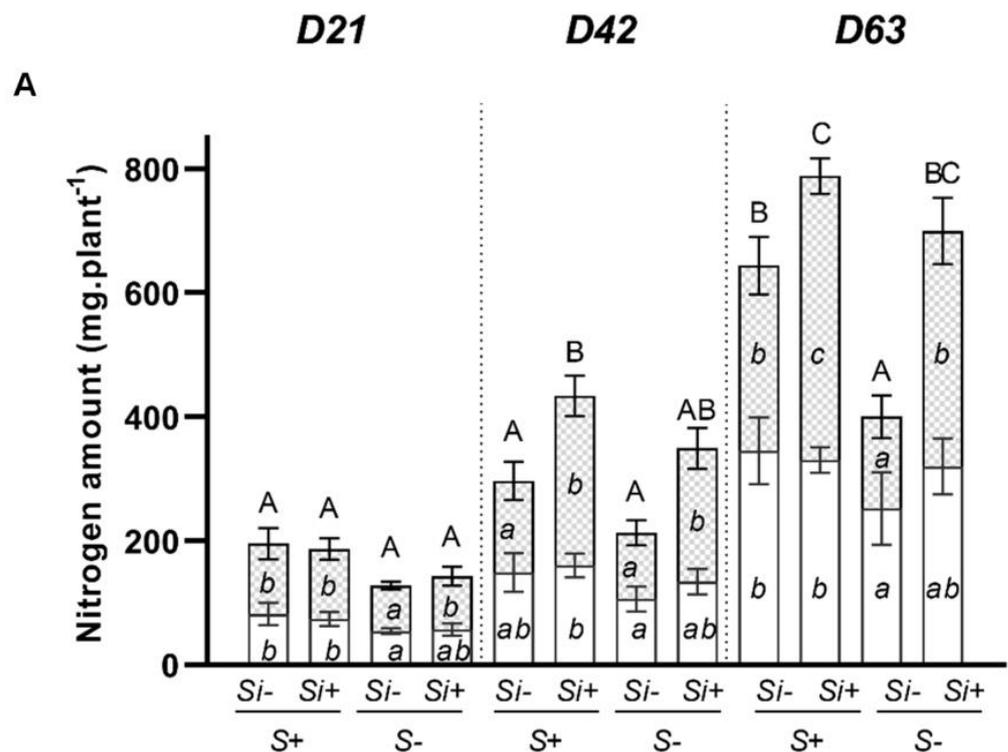


Figure 4. Cont.

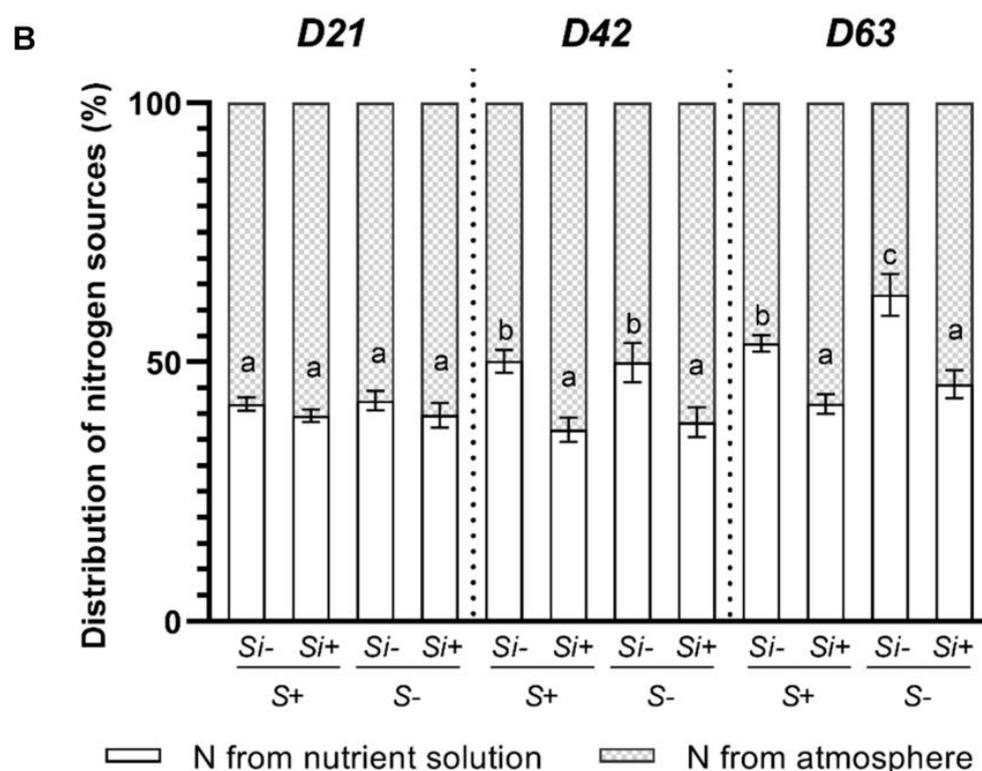


Figure 4. Root and shoot total nitrogen amounts (A) and distribution of nitrogen sources (B) in *Trifolium incarnatum* plants cultivated in hydroponic conditions for 21 (D21), 42 (D42) and 63 days (D63) with (+S; 500 μ M) or without (-S) sulfur and with (+Si; 1.7 mM) or without (-Si) Si. Data are means \pm SE ($n = 5$). Different lowercase italic letters to the left of the histogram and different uppercase letters above or inside of the histogram indicate that the means between plants harvested at the same time (D21, D42 or D63) are significantly different ($p < 0.05$).

2.5. Effect of Silicon Supply on Abundance of Nitrogenase in Nodules

At each harvest date and for each treatment, the concentration of nitrogenase in nodules (U.A. mg^{-1} FW) was determined using Western blot analysis (S2). From these data, at each harvest date, the total abundance of nitrogenase in all nodules of one plant was assessed (see Materials and Method for details) and expressed relative to +S-Si plants (considered as controls) to study the effects of S and Si treatments on this parameter (Figure 5). At D21, a negative effect of S-deprivation on nitrogenase relative abundance was observed and Si supply had no effect on this parameter, irrespective of the S treatment. At 42 and D63, the negative effect of S-deprivation on nitrogenase relative abundance was observed only in plants cultivated without Si. Indeed, compared to -S-Si plants, the Si supply increased the nitrogenase relative abundance by 5-fold and 6-fold at D42 and D63, respectively. This increase is a consequence of the increase in nitrogen concentration in nodules (S2) and of the increase in the total nodule biomass (Figure 3C) in -S+Si plants compared to -S-Si plants. Interestingly, the nitrogenase relative abundance in -S+Si plants at D63 again became similar to the values observed in +S+Si plants, suggesting a new time period in which supplying Si alleviated the negative effect of S-deprivation. In S-fed plants, even though a Si supply also resulted in an increase (around 3-fold) in the nitrogenase relative abundance at D42 and D63 (Figure 5), this increase was only due to an increase in the nitrogenase concentration in nodules of +S+Si plants (S2) because the total biomass of nodules was similar in +S-Si and +S+Si plants (Figure 3C).

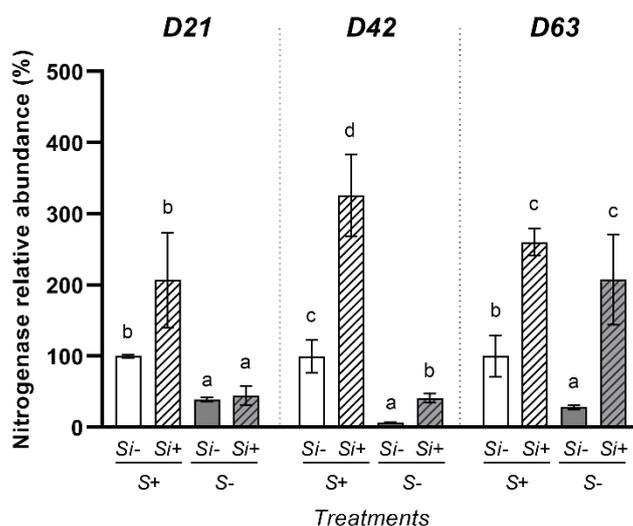


Figure 5. Change in the total amount of nitrogenase in nodules of *Trifolium incarnatum* cultivated in hydroponic conditions for 21 (D21), 42 (D42) and 63 days (D63) with (+S; 500 μ M) or without (–S) sulfur and with (+Si; 1.7 mM) or without (–Si) Si supply. For each time (D21, D42, or D63) and each treatment the total amount of nitrogenase was expressed relative to the value obtained for the +S-Si plants. Data are means \pm SE ($n = 3$). Different lowercase letters indicate that the mean between plants harvested at the same time (D21, D42 or D63) are significantly different ($p < 0.05$).

3. Discussion

Silicon is largely known to alleviate negative effects of abiotic stress in plants [26,28,35,36] and to improve plant health [30]. However, the effect of Si supply on S-deprived plants is currently poorly documented, as reported by [27,37]. The main objective of this study was to evaluate the effect of Si supply on the growth, nodulation and atmospheric N_2 fixation capacity of *Trifolium incarnatum* subjected (or not) to long-term sulfur deprivation (63 days). This study demonstrated that the growth of *Trifolium incarnatum* was not affected by S deficiency during the 21 first days of treatment (Figure 1). This result agrees with previous work performed in *Brassica napus* cultivated in hydroponic conditions and subjected to the same duration of S-deprivation [28]. In contrast, when the S-deprivation was prolonged (D63), this study showed that a strong decrease in both shoot and total biomasses occurred in –S-Si plants compared to +S-Si plants (Figure 1). This result agrees with previous studies reporting that long-term S deficiency limits growth in several leguminous plant species [8,9]. A supply of Si to these S-deprived plants led to an increase in shoot biomass at D42, and more interestingly to an increase in root and shoot biomass at D63, which allowed –S+Si plants to reach a total biomass similar to and even higher than that of S-fed plants (Figure 1). These beneficial effects of Si on the biomass of S-deprived plants agree with earlier studies showing that Si alleviates the negative effects on growth of plants experiencing macronutrient deficiencies (for review see [27]) such as N [26], K [38] or P [39,40]. However, with the beneficial effects of Si being poorly documented in plants subjected to S-deficiency [27,37], this study is the first to demonstrate enhancement of growth of S-deprived clover plants following application of Si. In addition, in our study, it was surprising to find that Si supply also promoted the growth of S-fed plants by enhancing shoot biomass from D42 to D63 (Figure 1). Indeed, in the literature it is usually stated that the benefits of Si for growth are mainly observed in plants under biotic and abiotic stresses [41].

To explain these beneficial effects of Si in *Trifolium incarnatum*, our study has focused on N nutrition, which is one of the major macronutrients and considered the most important component for supporting plant growth [42,43]. Our results show that at D63, compared to S-fed plants, the S-deprivation in –S-Si plants resulted in a significant decrease in the total amount of N, and this was mainly explained by a significant decrease in N_2 atmospheric fixation while the uptake of N from the nutrient solution was not affected (Figure 4). This decrease in N_2 fixation is associated with a decrease in the number of nodules, the biomass

of individual nodules and the total biomass of nodules per plant (Figure 3). Further, it was also associated with a significant decrease in total nitrogenase abundance (Figure 5) in roots in -S-Si plants, which occurred after 21 days of S-deficiency. These negative effects of S-deficiency on N₂ fixation, the number and biomass of nodules the nitrogenase abundance all agree with previous studies [44,45]. In particular, there are close parallels with work performed on *Trifolium repens* by [8], who reported that S-deficiency reduced cysteine and methionine contents, both of which are key amino acids for nitrogenase synthesis. As previously reported, these responses to S-deprivation could be explained by an increase in ROS [46], which will directly impact the structures and the activity of nodules [47]. Compared to -S-Si plants, supplying Si to S-deprived plants (-S+Si) led to an increase in the amount of total N, mainly due to the strong increase in N₂ fixation from D42 onwards (Figure 4B). This improvement in N₂ fixation was associated with an increase in number and total biomass of nodules from D21 (Figure 3) and later (from D42) with a higher nitrogenase abundance in the nodules (Figures 5 and S2) of -S+Si plants relative to -S-Si plants. This beneficial effect of Si agrees with numerous studies that have reported exogenous Si supply as being one of the most effective strategies to enhance nodulation and N₂ fixation in legumes under stressed conditions, including metal toxicity, salinity, alkalinity and pathogen attack [29,30,48]. Since nitrogen is a macronutrient required for plant growth, the increase in nitrogen fixation caused by Si leads to an increase in the amount of total nitrogen -S+Si plants (Figure 4) and improves their growth (Figure 1). However, our study is currently the only one to show these beneficial effects of Si in response to S-deficiency. Given the central role of S in numerous biological processes (such as N₂ fixation, nitrogenase and leghemoglobin synthesis and ROS detoxification), it is therefore difficult to explain how a Si supply is able to alleviate the negative impacts of S-deprivation, and particularly regarding nodulation and N₂ fixation. In addition, in S-fed plants, supplying Si also led to an enhancement of the total N and N₂ fixation (Figure 4) without any effect on the number and biomass of nodules (Figure 3). In this case, the improvement in N₂ fixation is essentially due to the strong increase in the nitrogenase concentration in nodules (S2). This result agrees with a previous study showing that a Si application on *Medicago truncatula* increased the nitrogenase concentration in nodules and improved their N₂ fixation capacity [30].

The current study has shown clearly that supplying Si alleviates the negative effects of S-deprivation in *Trifolium incarnatum*, especially by promoting root nodulation and N₂ fixation. Nevertheless, it is very difficult to explain the mechanisms related to these Si beneficial effects without being speculative. Indeed, regardless of the explanations advanced by earlier studies on other abiotic stresses where beneficial effects of Si on nodulation and N₂ fixation in legumes have been reported, the underlying mechanisms remain unclear and only some assumptions can be formulated [29]. For example, [30] reported that Si supplementation might affect key symbiotic signals such as flavonoid compounds and would promote the attraction of rhizobia and increase the degree of nodulation. Another assumption supported by the same authors concerned the silicification of nodules, which would enhance their N₂ fixation efficiency. This hypothesis is compatible with the preferential accumulation of Si in the roots of *Trifolium incarnatum* (Figure 2C) observed in this study. Finally, it is well known that S-deprivation leads to ROS production, which might be significantly reduced by Si supply as shown recently by [28]. As suggested by some authors, the decrease in oxidative stress in -S+Si plants can lead to better nitrogenase activity, with this enzyme known to be inhibited in the presence of ROS [30,35]. Even if our pioneering study has demonstrated for the first time that Si supply alleviates the negative effect of S-deficiency in *Trifolium incarnatum*, further investigation is required to verify all or part of these hypotheses and to decipher the fine mechanisms that remain unclear or misunderstood.

In conclusion, the results of this study suggest that Si fertilization could be considered as part of emerging agroecological and agricultural practices to optimize fertilizer inputs. For example, to reduce the quantity of N inputs applied during *Brassica napus* crop growth,

several recent studies have explored mixed cropping of this plant with legume species [49], and in particular with *Trifolium incarnatum* [18,33]. Indeed, the high sulfur requirements of *Brassica napus* for growth can decrease the availability of S in the soil and lead to decreases in the N₂ fixation capacity of the companion legume. Thus, as suggested by this study, Si fertilization could alleviate this by increasing the atmospheric N₂ fixation capacity of clover and eventually lead to an enhancement of the agronomic performance of this mixed cropping by improving the transfer of exuded N compounds from *Trifolium incarnatum* to *Brassica napus* [34].

4. Materials and Methods

4.1. Plant Growth Conditions and Experimental Design

The experimental design is summarized in Figure 6. In a greenhouse, seeds of *Trifolium incarnatum* L. were germinated on perlite over deionized water for four days in the dark [18]. When the first leaf emerged, seedlings were transferred to natural light conditions and supplied with nutrient solution for two weeks containing: KNO₃ (1 mM), KH₂PO₄ (0.25 mM), KCl (1 mM), CaCl₂ (3 mM), MgSO₄ (0.5 mM), EDTA-2NaFe (0.2 mM), H₃BO₃ (14 μM), MnSO₄ (5 μM), ZnSO₄ (3 μM), CuSO₄ (0.7 μM), (NH₄)₆Mo₇O₂₄ (0.7 μM), CoCl₂ (0.1 μM). After two weeks, the seedlings were transplanted in hydroponics tanks (20 L, nine plants per tank) containing the nutrient solution described above and inoculated with *Rhizobium leguminosarum* bv *trifolii* (T354) [33] previously grown in YEM liquid medium [50] for 72 h. After four weeks (D0), plants were separated into four sets: the first two sets corresponded to plants maintained on S (+S; 500 μM) and supplied (+S+Si) or not (+S-Si) with 1.7 mM silicon for 63 days (D63). The nutrient solution described above with the addition of 1.7 mM silicon (Si as sodium metasilicate: Na₂SiO₃) or NaCl (3.4 mM to compensate the sodium supplied by the Si treatment) to the +S+Si and +S-Si plants, respectively. The other two sets corresponded to plants deficient in S (-S) and supplied (-S+Si) or not (-S-Si) with silicon for 63 days. For this, the nutrient solution was modified as follows to remove S: KH₂PO₄ (0.25 mM), KCl (1 mM), CaCl₂ (3 mM), MgCl₂ (0.5 mM), EDTA-2NaFe (0.2 mM), H₃BO₃ (14 μM), MnCl₂ (5 μM), ZnCl₂ (3 μM), CuCl₂ (0.7 μM), (NH₄)₆Mo₇O₂₄ (0.7 μM) and CoCl₂ (0.1 μM). In addition, plants were supplied with K¹⁵NO₃ (1 mM, 0.5% isotopic excess) in order to discriminate between the N taken up from nutrient solution and the N derived from atmospheric fixation in the plants. Throughout the culture duration, natural light was supplied by high pressure sodium lamps (Philips, MASTER GreenPower T400W) with photosynthetically active radiation of 450 μmol photons·m⁻²·s⁻¹ at canopy height. Nutrient solutions, continuously aerated with a compressed air bubbling system, were replaced every three days. The pH of the nutrient solution was monitored each day and adjusted if necessary to 5.8 ± 0.2. At each solution renewal, 10 mL of medium containing *Rhizobium leguminosarum* bv *trifolii* (T354) was added to each tank. After 21, 42 and 63 days of cultivation, plants were harvested (Figure 6). At each harvest, the nodules were counted, separated from roots, weighed and frozen at -80%. In addition, shoots and roots were separated and weighed. Then, an aliquot of each plant compartment was placed in liquid nitrogen and stored at -80 °C and the rest was dried at 60 °C. After dry weight determination, samples were ground to perform the elemental analyses.

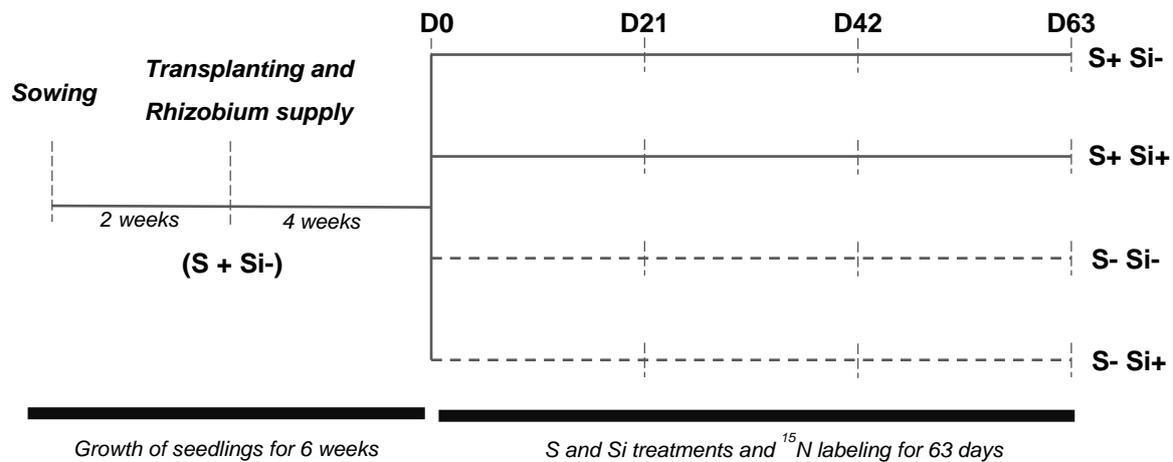


Figure 6. Experimental design used to study the effect of silicon supply on *Trifolium incarnatum* L. grown with (+S) or without (-S) sulfur. After six weeks of cultivation with sulfur (+S; 500 μ M) (D0), plants were separated into four batches: the first two batches (full line) corresponded to plants that continued to be fed with S (+S; 500 μ M) and supplied (+Si; 1.7 mM) or not (-Si) with Si for 63 days (D63). The other two batches (dotted lines) corresponded to plants deficient in S (-S) and supplied (+Si; 1.7 mM) or not (-Si) with silicon. At D0, all plants were fed with $K^{15}NO_3$ (1 mM, 0.5% isotopic excess).

4.2. Determination of Total, S, Si, N and ^{15}N Concentrations and Calculation of the Distribution of Nitrogen Sources in Plants

For S and Si concentration determinations, approximately 1 g of the dried powder was analyzed with an X-ray-fluorescence spectrometer (XEPOS, Ametek, Berwyn, PA, USA) using calibration curves obtained from international standards.

To determine the total N and ^{15}N concentrations, 1.5 mg of each dried powder was precisely weighed and placed into tin capsules before analysis with a continuous flow isotope ratio mass spectrometer (IRMS, Horizon, NU Instruments, Wrexham, UK) linked to a C/N/S analyzer (EA3000, Euro Vector, Milan, Italy). The total N amount (N_{tot}) in each organ or in the whole plant was calculated as:

$$N_{tot} = \frac{\%N \times W}{100}$$

The amount of ^{15}N (^{15}N) was determined by the following:

$$^{15}N = \frac{\%^{15}N \times N_{tot}}{100}$$

As the nutrient solution was labeled with a 0.5% isotopic excess, it was possible to calculate the total amount of nitrogen taken up from the nutrient solution (N_{ns}):

$$N_{ns} = \frac{^{15}N_{total} \times 100}{0.5}$$

The amount of nitrogen from dinitrogen (N_2) atmospheric fixation (N_{atm}) was then obtained by subtracting the amount of nitrogen taken up from the nutrient solution from the total nitrogen in the plant. Then, the proportion of each N source (% of N_{ns} or % N_{atm}) was calculated as follows:

$$\%N_{ns} \text{ (or } \%N_{atm}) = \frac{N_{ns} \text{ (or } N_{atm}) \times 100}{N_{tot} \text{ (whole plant)}}$$

4.3. Extraction and Quantification of Proteins from Nodules

Total proteins were extracted from nodules using the protocol described by [8]. Fresh nodules were ground in liquid nitrogen in a mortar and approximately 30 mg of the resulting powder was mixed in 2 mL of cold acetone containing 10% trichloroacetic acid (*w/v*). This mixture was centrifuged at $16,000\times g$ (4 min at 4 °C), the supernatant was then removed the pellet was resuspended with 1.75 mL of ammonium acetate (0.1 M)/methanol (80%) buffer to precipitate the proteins. After centrifugation at $16,000\times g$ (3 min at 4 °C), the supernatant was removed, and the pellet was washed with 1 mL of acetone (80%) and centrifuged again. The resulting pellet was resuspended with 800 μL of phenol pH 8 and 800 μL of sodium dodecyl sulfate (SDS) buffer [30% saccharose (*w/v*), 2% SDS (*w/v*), 0.1 M TRIS-HCl (*w/v*), 0.5% β -mercaptoethanol (*v/v*), pH 8] then centrifuged at $16,000\times g$ (10 min at 4 °C). The upper phenolic phase was recovered and 800 μL of ammonium acetate (0.1 M)/methanol (80%) buffer was added. After storing overnight at $-20\text{ }^{\circ}\text{C}$, the mixture was centrifuged at $16,000\times g$ (10 min at 4 °C). Then the pellet was washed with 1 mL of methanol (100%) and after centrifugation at $16,000\times g$ (10 min at 4 °C) it was washed again with 1 mL of acetone (80%). The pellet was resuspended with a buffer containing dithiothreitol (DTT, 11.11 mM), thiourea (2.22 M), urea (6.66 M) and Tris buffer 2 M (33.33 mM) and used for the determination of the protein concentration using the Bradford method [51].

4.4. Immunodetection and Quantification of Nitrogenase in Root Nodules

The abundance of nitrogenase in nodules was determined after Western blotting of 20 μg of total proteins previously separated by SDS-PAGE (12% polyacrylamide gels at a constant current: 250 V, 75 mA, 1 h). Western blots were carried out on a polyvinylidene fluoride (PVDF) membrane (Immobilon-PVDF, Millipore Bedford, USA) by semi-dry electroblotting (Pierce™ G2 Fast Blotter, Thermo Fisher Scientific, Waltham, USA). Conditions for protein transfer and immunoblotting were carried out as previously described [8]. Briefly, after Western blotting, PVDF membrane was incubated overnight with chicken polyclonal anti-nitrogenase (NifH) from IgG antibodies (Agriser, SE 911, Vanadas, Sweden; dilution 1:2000). Subsequently the PVDF membrane was washed four times with TBST (Tris-base 10 mM, NaCl 150 mM, pH 8, 0.1% Tween 20) before incubation with secondary antibody (rabbit anti chicken immunoglobulin Y coupled to phosphatase alkaline; dilution 1:6000) for 1 h. The nitrogenase–antibody complex was revealed using the Bio-Rad alkaline phosphatase kit as a single signal at a molecular weight between 25 and 37 kDa (S2). Independently performed gels and Western blots were scanned and analyzed using the Millipore Bioimage computerized image analysis system to determine the intensity of signal (U.A.) that was proportional to the nitrogenase abundance in each nodule sample. The nodule nitrogenase concentration (U.A. g^{-1} FW) was calculated by dividing the intensity value by the fresh nodule biomass (20 μg) used for Western blot analysis. Then the total abundance of nitrogenase in all the nodules of one plant was estimated by multiplying the nitrogenase concentration by total nodule biomass of this plant. Finally, at each harvest time, the results were expressed relative to the abundance of nitrogenase in the total nodules from control plants (+S-Si).

4.5. Statistical Analysis

The experiment was performed with five independent replicates. All data are indicated as the mean \pm S.E ($n = 5$). Statistical analyses were performed using R software (version 4.2.0: R Core Team, 2022). At each harvest date (D21, D42 and D63), significant differences between treatments were determined using Student's *t*-test ($p < 0.05$).

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants12122248/s1>, Figure S1: Representative pictures of roots and shoots of *Trifolium incarnatum* plants cultivated in hydroponic conditions at 63 days (D63) with (+S; 500 μ M) or without (-S) sulphur and with (+Si; 1.7 mM) or without (-Si) Si supply; Figure S2: Change in nitrogenase abundance in nodules of *Trifolium incarnatum* plants cultivated in hydroponic conditions during 21 (D21), 42 (D42) and 63 days (D63) with (+S; 500 μ M) or without (-S) sulphur and with (+Si; 1.7 mM) or without (-Si) Si supply.

Author Contributions: Conceptualization, R.C., P.L. and P.E.; realization and methodology, R.C., P.L., Q.D., F.L. and P.E.; data analysis, R.C., P.L., Q.D., F.L. and P.E.; writing—original draft preparation, R.C., P.L. and P.E.; writing—reviewing and editing, R.C., P.L., M.A., Q.D., F.L. and P.E.; supervision, P.L. and P.E. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

International Review Board Statement: Not applicable.

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: Not applicable.

Acknowledgments: The authors thank the team of the PLATIN' (Plateau d'Isotopie de Normandie) core facility for the element analysis. The authors would like to thank the "Region Normandie" for its support for research in Normandy and especially for the RIN grant to R.C.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Hoesly, R.M.; Smith, S.J.; Feng, L.; Klimont, Z.; Janssens-Maenhout, G.; Pitkanen, T.; Seibert, J.J.; Vu, L.; Andres, R.J.; Bolt, R.M.; et al. Historical (1750–2014) Anthropogenic Emissions of Reactive Gases and Aerosols from the Community Emissions Data System (CEDS). *Geosci. Model Dev.* **2018**, *11*, 369–408. [\[CrossRef\]](#)
- Aas, W.; Mortier, A.; Bowersox, V.; Cherian, R.; Faluvegi, G.; Fagerli, H.; Hand, J.; Klimont, Z.; Galy-Lacaux, C.; Lehmann, C.M.B.; et al. Global and Regional Trends of Atmospheric Sulfur. *Sci. Rep.* **2019**, *9*, 953. [\[CrossRef\]](#) [\[PubMed\]](#)
- Orellano, P.; Reynoso, J.; Quaranta, N. Short-Term Exposure to Sulphur Dioxide (SO₂) and All-Cause and Respiratory Mortality: A Systematic Review and Meta-Analysis. *Environ. Int.* **2021**, *150*, 106434. [\[CrossRef\]](#)
- Feinberg, A.; Stenke, A.; Peter, T.; Hinckley, E.-L.S.; Driscoll, C.T.; Winkel, L.H.E. Reductions in the Deposition of Sulfur and Selenium to Agricultural Soils Pose Risk of Future Nutrient Deficiencies. *Commun. Earth Env.* **2021**, *2*, 101. [\[CrossRef\]](#)
- D'Hooghe, P.; Escamez, S.; Trouverie, J.; Avicé, J.-C. Sulphur Limitation Provokes Physiological and Leaf Proteome Changes in Oilseed Rape That Lead to Perturbation of Sulphur, Carbon and Oxidative Metabolisms. *BMC Plant Biol.* **2013**, *13*, 23. [\[CrossRef\]](#) [\[PubMed\]](#)
- Sorin, E.; Etienne, P.; Maillard, A.; Zamarreño, A.-M.; Garcia-Mina, J.-M.; Arkoun, M.; Jamois, F.; Cruz, F.; Yvin, J.-C.; Ourry, A. Effect of Sulphur Deprivation on Osmotic Potential Components and Nitrogen Metabolism in Oilseed Rape Leaves: Identification of a New Early Indicator. *J. Exp. Bot.* **2015**, *66*, 6175–6189. [\[CrossRef\]](#)
- Etienne, P.; Sorin, E.; Maillard, A.; Gallardo, K.; Arkoun, M.; Guerrand, J.; Cruz, F.; Yvin, J.-C.; Ourry, A. Assessment of Sulfur Deficiency under Field Conditions by Single Measurements of Sulfur, Chloride and Phosphorus in Mature Leaves. *Plants* **2018**, *7*, 37. [\[CrossRef\]](#)
- Varin, S.; Cliquet, J.-B.; Personeni, E.; Avicé, J.-C.; Lemauiel-Lavenant, S. How Does Sulphur Availability Modify N Acquisition of White Clover (*Trifolium repens* L.)? *J. Exp. Bot.* **2010**, *61*, 225–234. [\[CrossRef\]](#)
- Zuber, H.; Poignavent, G.; Le Signor, C.; Aimé, D.; Vieren, E.; Tadla, C.; Lugan, R.; Belghazi, M.; Labas, V.; Santoni, A.-L.; et al. Legume Adaptation to Sulfur Deficiency Revealed by Comparing Nutrient Allocation and Seed Traits in *Medicago truncatula*. *Plant J.* **2013**, *76*, 982–996. [\[CrossRef\]](#)
- Gao, Y.; Li, X.; Tian, Q.-Y.; Wang, B.-L.; Zhang, W.-H. Sulfur Deficiency Had Different Effects on *Medicago truncatula* Ecotypes A17 and R108 in Terms of Growth, Root Morphology and Nutrient Contents. *J. Plant Nutr.* **2016**, *39*, 301–314. [\[CrossRef\]](#)
- Becana, M.; Wienkoop, S.; Matamoros, M.A. Sulfur Transport and Metabolism in Legume Root Nodules. *Front. Plant Sci.* **2018**, *9*, 1434. [\[CrossRef\]](#) [\[PubMed\]](#)
- Hasanuzzaman, M.; Bhuyan, M.H.M.B.; Parvin, K.; Bhuiyan, T.F.; Anee, T.I.; Nahar, K.; Hossen, M.S.; Zulfiqar, F.; Alam, M.M.; Fujita, M. Regulation of ROS Metabolism in Plants under Environmental Stress: A Review of Recent Experimental Evidence. *Int. J. Mol. Sci.* **2020**, *21*, 8695. [\[CrossRef\]](#) [\[PubMed\]](#)
- Ristova, D.; Kopriva, S. Sulfur Signaling and Starvation Response in Arabidopsis. *iScience* **2022**, *25*, 104242. [\[CrossRef\]](#) [\[PubMed\]](#)

14. Scherer, H.W.; Pacyna, S.; Spoth, K.R.; Schulz, M. Low Levels of Ferredoxin, ATP and Leghemoglobin Contribute to Limited N₂ Fixation of Peas (*Pisum sativum* L.) and Alfalfa (*Medicago sativa* L.) under S Deficiency Conditions. *Biol. Fertil. Soils* **2008**, *44*, 909–916. [[CrossRef](#)]
15. Karmoker, J.; Clarkson, D.T.; Saker, L.R.; Rooney, J.M.; Purves, J.V. Sulphate Deprivation Depresses the Transport of Nitrogen to the Xylem and the Hydraulic Conductivity of Barley (*Hordeum vulgare* L.) Roots. *Planta* **1991**, *185*, 269–278. [[CrossRef](#)] [[PubMed](#)]
16. Girondé, A.; Etienne, P.; Trouverie, J.; Bouchereau, A.; Le Cahérec, F.; Lepout, L.; Orsel, M.; Niogret, M.-F.; Nési, N.; Carole, D.; et al. The Contrasting N Management of Two Oilseed Rape Genotypes Reveals the Mechanisms of Proteolysis Associated with Leaf N Remobilization and the Respective Contributions of Leaves and Stems to N Storage and Remobilization during Seed Filling. *BMC Plant Biol.* **2015**, *15*, 59. [[CrossRef](#)] [[PubMed](#)]
17. Yu, Z.; She, M.; Zheng, T.; Diepeveen, D.; Islam, S.; Zhao, Y.; Zhang, Y.; Tang, G.; Zhang, Y.; Zhang, J.; et al. Impact and Mechanism of Sulphur-Deficiency on Modern Wheat Farming Nitrogen-Related Sustainability and Gliadin Content. *Commun. Biol.* **2021**, *4*, 945. [[CrossRef](#)]
18. Génard, T.; Laîné, P.; Diquélou, S.; Nési, N.; Yvin, J.-C.; Etienne, P. Impact of Sulfur Applications on the Agronomic Performance of Rapeseed–Clover Mixtures. *J. Plant Nutr. Soil Sci.* **2017**, *180*, 676–682. [[CrossRef](#)]
19. Verma, K.K.; Song, X.-P.; Tian, D.-D.; Guo, D.-J.; Chen, Z.-L.; Zhong, C.-S.; Nikpay, A.; Singh, M.; Rajput, V.D.; Singh, R.K.; et al. Influence of Silicon on Biocontrol Strategies to Manage Biotic Stress for Crop Protection, Performance, and Improvement. *Plants* **2021**, *10*, 2163. [[CrossRef](#)]
20. Bakhat, H.F.; Bibi, N.; Zia, Z.; Abbas, S.; Hammad, H.M.; Fahad, S.; Ashraf, M.R.; Shah, G.M.; Rabbani, F.; Saeed, S. Silicon Mitigates Biotic Stresses in Crop Plants: A Review. *Crop Prot.* **2018**, *104*, 21–34. [[CrossRef](#)]
21. Moraes, S.P.D.O.; Rodrigues, J.F.M.; Escobar, M.E.O.; Araújo, F.S.; Oliveira, T.S.D. Beneficial Effects of Silicon in Plants under Abiotic Stress Conditions: A New Approach. *RCA* **2022**, *53*, e2021821. [[CrossRef](#)]
22. El Moukhtari, A.; Ksaa, M.; Zorrig, W.; Cabassa, C.; Abdelly, C.; Farissi, M.; Savoure, A. How Silicon Alleviates the Effect of Abiotic Stresses During Seed Germination: A Review. *J. Plant Growth Regul.* **2022**, *42*, 3323–3341. [[CrossRef](#)]
23. Wang, D.; Hou, L.; Zhang, L.; Liu, P. The Mechanisms of Silicon on Maintaining Water Balance under Water Deficit Stress. *Physiol. Plant.* **2021**, *173*, 1253–1262. [[CrossRef](#)] [[PubMed](#)]
24. Mustafa, T.; Sattar, A.; Sher, A.; Ul-Allah, S.; Ijaz, M.; Irfan, M.; Butt, M.; Cheema, M. Exogenous Application of Silicon Improves the Performance of Wheat under Terminal Heat Stress by Triggering Physio-Biochemical Mechanisms. *Sci. Rep.* **2021**, *11*, 23170. [[CrossRef](#)]
25. El Moukhtari, A.; Cabassa-Hourton, C.; Crilat, E.; Carol, P.; Lamsaadi, N.; Hidri, R.; Farissi, M.; Savouré, A. Salt Stress Is Alleviated by Either Proline or Silicon But Not by Their Combination in Alfalfa (*Medicago sativa* L.) Inoculated with a Salt-Tolerant Ensifer Meliloti Strain. *J. Plant Growth Regul.* **2022**. [[CrossRef](#)]
26. Haddad, C.; Arkoun, M.; Jamois, F.; Schwarzenberg, A.; Yvin, J.-C.; Etienne, P.; Laîné, P. Silicon Promotes Growth of *Brassica napus* L. and Delays Leaf Senescence Induced by Nitrogen Starvation. *Front. Plant Sci.* **2018**, *9*, 516. [[CrossRef](#)]
27. Pavlovic, J.; Kostic, L.; Bosnic, P.; Kirkby, E.A.; Nikolic, M. Interactions of Silicon With Essential and Beneficial Elements in Plants. *Front. Plant Sci.* **2021**, *12*, 697592. [[CrossRef](#)]
28. Laîné, P.; Coquerel, R.; Arkoun, M.; Trouverie, J.; Etienne, P. Assessing the Effect of Silicon Supply on Root Sulfur Uptake in S-Fed and S-Deprived *Brassica napus* L. *Plants* **2022**, *11*, 1606. [[CrossRef](#)]
29. El Moukhtari, A.; Lamsaadi, N.; Oubenali, A.; Mouradi, M.; Savoure, A.; Farissi, M. Exogenous Silicon Application Promotes Tolerance of Legumes and Their N₂ Fixing Symbiosis to Salt Stress. *Silicon* **2022**, *14*, 6517–6534. [[CrossRef](#)]
30. Putra, R.; Vandeger, R.K.; Karan, S.; Powell, J.R.; Hartley, S.E.; Johnson, S.N. Silicon Enrichment Alters Functional Traits in Legumes Depending on Plant Genotype and Symbiosis with Nitrogen-fixing Bacteria. *Funct. Ecol.* **2021**, *35*, 2856–2869. [[CrossRef](#)]
31. Maillard, A.; Ali, N.; Schwarzenberg, A.; Jamois, F.; Yvin, J.-C.; Hosseini, S.A. Silicon Transcriptionally Regulates Sulfur and ABA Metabolism and Delays Leaf Senescence in Barley under Combined Sulfur Deficiency and Osmotic Stress. *Environ. Exp. Bot.* **2018**, *155*, 394–410. [[CrossRef](#)]
32. Réthoré, E.; Ali, N.; Yvin, J.-C.; Hosseini, S.A. Silicon Regulates Source to Sink Metabolic Homeostasis and Promotes Growth of Rice Plants under Sulfur Deficiency. *Int. J. Mol. Sci.* **2020**, *21*, 3677. [[CrossRef](#)] [[PubMed](#)]
33. Génard, T.; Etienne, P.; Diquélou, S.; Yvin, J.-C.; Revellin, C.; Laîné, P. Rapeseed-Legume Intercrops: Plant Growth and Nitrogen Balance in Early Stages of Growth and Development. *Heliyon* **2017**, *3*, e00261. [[CrossRef](#)] [[PubMed](#)]
34. Génard, T.; Etienne, P.; Laîné, P.; Yvin, J.-C.; Diquélou, S. Nitrogen Transfer from *Lupinus albus* L., *Trifolium incarnatum* L. and *Vicia sativa* L. Contribute Differently to Rapeseed (*Brassica napus* L.) Nitrogen Nutrition. *Heliyon* **2016**, *2*, e00150. [[CrossRef](#)] [[PubMed](#)]
35. Mostofa, M.G.; Rahman, M.M.; Ansary, M.M.U.; Keya, S.S.; Abdelrahman, M.; Miah, M.G.; Phan Tran, L.-S. Silicon in Mitigation of Abiotic Stress-Induced Oxidative Damage in Plants. *Crit. Rev. Biotechnol.* **2021**, *41*, 918–934. [[CrossRef](#)]
36. Laîné, P.; Haddad, C.; Arkoun, M.; Yvin, J.-C.; Etienne, P. Silicon Promotes Agronomic Performance in Brassica Napus Cultivated under Field Conditions with Two Nitrogen Fertilizer Inputs. *Plants* **2019**, *8*, 137. [[CrossRef](#)]
37. Ali, N.; Réthoré, E.; Yvin, J.-C.; Hosseini, S.A. The Regulatory Role of Silicon in Mitigating Plant Nutritional Stresses. *Plants* **2020**, *9*, 1779. [[CrossRef](#)]
38. Dos Santos Sarah, M.M.; De Mello Prado, R.; De Souza Júnior, J.P.; Teixeira, G.C.M.; Dos Santos Duarte, J.C.; De Medeiros, R.L.S. Silicon Supplied via Foliar Application and Root to Attenuate Potassium Deficiency in Common Bean Plants. *Sci. Rep.* **2021**, *11*, 19690. [[CrossRef](#)]

39. Hu, A.Y.; Xu, S.N.; Qin, D.N.; Li, W.; Zhao, X.Q. Role of Silicon in Mediating Phosphorus Imbalance in Plants. *Plants* **2020**, *10*, 51. [[CrossRef](#)]
40. Beier, S.; Marella, N.C.; Yvin, J.-C.; Hosseini, S.A.; Von Wirén, N. Silicon Mitigates Potassium Deficiency by Enhanced Remobilization and Modulated Potassium Transporter Regulation. *Environ. Exp. Bot.* **2022**, *198*, 104849. [[CrossRef](#)]
41. Epstein, E. Silicon. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **1999**, *50*, 641–664. [[CrossRef](#)] [[PubMed](#)]
42. Engels, C.; Marschner, H. Plant Uptake and Utilization of Nitrogen. In *Nitrogen Fertilization in the Environment*; Marcel Dekker, Inc.: New York, NY, USA, 1995; pp. 41–81.
43. Leghari, S.J.; Wahocho, N.A.; Laghari, G.M.; HafeezLaghari, A.; MustafaBhabhan, G.; HussainTalpur, K.; Lashari, A.A. Role of Nitrogen for Plant Growth and Development: A Review. *Adv. Environ. Biol.* **2016**, *10*, 209–219.
44. Jamal, A.; Moon, Y.-S.; Abdin, M. Sulphur—A General Overview and Interaction with Nitrogen. *Aust. J. Crop Sci.* **2010**, *4*, 523–529.
45. Zenda, T.; Liu, S.; Dong, A.; Duan, H. Revisiting Sulphur—The Once Neglected Nutrient: It’s Roles in Plant Growth, Metabolism, Stress Tolerance and Crop Production. *Agriculture* **2021**, *11*, 626. [[CrossRef](#)]
46. Chandra, N.; Pandey, N. Influence of Sulfur Induced Stress on Oxidative Status and Antioxidative Machinery in Leaves of *Allium Cepa* L. *Int. Sch. Res. Not.* **2014**, *2014*, 568081. [[CrossRef](#)]
47. Mhadhbi, H.; Chihaoui, S.; Mhamdi, R.; Mnasri, B.; Jebara, M.; Mhamdi, R. A Highly Osmotolerant Rhizobial Strain Confers a Better Tolerance of Nitrogen Fixation and Enhances Protective Activities to Nodules of *Phaseolus vulgaris* under Drought Stress. *Afr. J. Biotechnol.* **2011**, *10*, 4555–4563. [[CrossRef](#)]
48. Putra, R.; Waterman, J.M.; Mathesius, U.; Wojtalewicz, D.; Powell, J.R.; Hartley, S.E.; Johnson, S.N. Benefits of Silicon-Enhanced Root Nodulation in a Model Legume Are Contingent upon Rhizobial Efficacy. *Plant Soil* **2022**, *477*, 201–217. [[CrossRef](#)]
49. Jamont, M.; Piva, G.; Fustec, J. Sharing N Resources in the Early Growth of Rapeseed Intercropped with Faba Bean: Does N Transfer Matter? *Plant Soil* **2013**, *371*, 641–653. [[CrossRef](#)]
50. Gulati, S.L. New Nonsynthetic Medium ForRhizobium Culture Production from Wastes. *Biotechnol. Bioeng.* **1979**, *21*, 1507–1515. [[CrossRef](#)]
51. Bradford, M.M. A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Anal. Biochem.* **1976**, *72*, 248–254. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.