

# Supporting Information

## Anticholinesterase Activity of Budmunchiamine Alkaloids Revealed by Comparative Chemical Profiling of Two *Albizia* spp., Molecular Docking and Dynamic Studies

Mai E. Hussein <sup>1,\*†</sup>, Osama G. Mohamed <sup>1,2,†</sup>, Ahlam M. El-Fishawy <sup>1</sup>, Hesham I. El-Askary <sup>1</sup>, Ahmed A. Hamed <sup>3</sup>, Marwa M. Abdel-Aziz <sup>4</sup>, Radwan Alnajjar <sup>5,6,7</sup>, Amany Belal <sup>8</sup>, Ahmed M. Naglah <sup>9,10</sup>, Abdulrahman A. Almehezia <sup>9</sup>, Ahmed A. Al-Karmalawy <sup>11</sup>, Ashootosh Tripathi <sup>2,12</sup> and Amira S. El Senousy <sup>1</sup>

<sup>1</sup> Pharmacognosy Department, Faculty of Pharmacy, Cairo University, Kasr el Aini St., Cairo 11562, Egypt

<sup>2</sup> Natural Products Discovery Core, Life Sciences Institute, University of Michigan, Ann Arbor, MI 48109, USA

<sup>3</sup> Microbial Chemistry Department, National Research Centre, 33 El-Buhouth Street, Dokki, Giza 12622, Egypt

<sup>4</sup> Regional Center for Mycology and Biotechnology (RCMB), Al-Azhar University, Cairo 11651, Egypt

<sup>5</sup> Department of Chemistry, Faculty of Science, University of Benghazi, Benghazi, Libya

<sup>6</sup> PharmD, Faculty of Pharmacy, Libyan International Medical University, Benghazi, Libya

<sup>7</sup> Department of Chemistry, University of Cape Town, Rondebosch 7701, South Africa

<sup>8</sup> Medicinal Chemistry Department, Faculty of Pharmacy, Beni-Suef University, Beni-Suef 62514, Egypt

<sup>9</sup> Drug Exploration and Development Chair (DEDC), Department of Pharmaceutical Chemistry, College of Pharmacy, King Saud University, Riyadh 11451, Saudi Arabia

<sup>10</sup> Peptide Chemistry Department, National Research Centre, Dokki, Cairo 12622, Egypt

<sup>11</sup> Pharmaceutical Chemistry Department, Faculty of Pharmacy, Ahram Canadian University, 6th of October City, Giza 12566, Egypt

<sup>12</sup> Department of Medicinal Chemistry, College of Pharmacy, University of Michigan, Ann Arbor, MI 48109, USA

\* Correspondence: mai.husein@pharma.cu.edu.eg

† These authors contributed equally to this work.

### List of Supporting Information

**Table S1:** Identified metabolites in the negative ion mode of ethanolic extracts of *A. lucidior* and *A. procera* leaves using UHPLC-MS/MS ..... 3-7

## List of Figures

<b>Figure S1.</b> GNPS molecular network of two <i>Albizia</i> spp. ethanolic extracts in the positive ion mode.....	8
<b>Figure S2.</b> GNPS molecular network of two <i>Albizia</i> spp. ethanolic extracts in the negative ion mode.....	9
<b>Figure S3.</b> Superimposition of the redocked MF2 700 inhibitor (green) over its native one (red).....	10

**Table S1:** Identified metabolites in the negative ion mode of ethanolic extracts of *A. lucidior* and *A. procera* leaves using UHPLC-MS/MS

No.	R <sub>t</sub> (min)	Name	Ion m/z ppm	Molecular Formula	MS/MS fragmentation product ions	Ref	Al	Ap
1	0.41	Dicaffeoylhexaric acid	533.1718	C <sub>23</sub> H <sub>33</sub> O <sub>14</sub> <sup>-</sup>	209.0653; 191.0561 85.0300	[1]	√	-
2	0.42	Quinic acid	191.0563	C <sub>7</sub> H <sub>11</sub> O <sub>6</sub> <sup>-</sup>	173.0447; 127.0396 93.0344; 85.0293	[2]	√	√
3	0.43	Xylonic acid	165.0403	C <sub>5</sub> H <sub>9</sub> O <sub>6</sub> <sup>-</sup>	147.0275; 135.0269 105.0185; 87.0084 75.0086	[2]	√	√
4	0.49	Gallic acid glucoside	331.0686	C <sub>13</sub> H <sub>15</sub> O <sub>10</sub> <sup>-</sup>	211.0237; 169.0147 151.0026; 125.0241	[1]	-	√
5	0.55	Gallic acid	169.0143	C <sub>7</sub> H <sub>5</sub> O <sub>5</sub> <sup>-</sup>	125.0258; 107.0153 97.0309; 79.0207	[3]	-	√
6	0.76	Protocatechuic acid	153.0190	C <sub>7</sub> H <sub>5</sub> O <sub>4</sub> <sup>-</sup>	109.0293	[4]	√	√
7	0.77	Vanillic acid-O-glucoside	329.0885	C <sub>14</sub> H <sub>17</sub> O <sub>9</sub> <sup>-</sup>	209.0452; 167.0345 125.0246; 123.0449	[1]	-	√
8	0.80	Catechol	109.0287	C <sub>6</sub> H <sub>5</sub> O <sub>2</sub> <sup>-</sup>	108.0217; 91.0191 81.0335; 66.0361	[5]	√	-
9	0.88	Gentisic acid-5-O-xyloside	285.0619	C <sub>12</sub> H <sub>13</sub> O <sub>8</sub> <sup>-</sup>	152.0114; 108.0216	[6]	√	-
10	0.91	Digalloyl-glucose	483.0779	C <sub>20</sub> H <sub>19</sub> O <sub>14</sub> <sup>-</sup>	313.0553; 271.0455 169.0137	[7]	-	√
11	0.99	Caffeic acid glucuronide	355.0685	C <sub>15</sub> H <sub>15</sub> O <sub>10</sub> <sup>-</sup>	209.0272; 191.0545 161.0240; 85.0292	[2]	√	-
12	1.05	Methyl gallate	183.0301	C <sub>8</sub> H <sub>7</sub> O <sub>5</sub> <sup>-</sup>	168.0046; 124.0164	[3]	-	√
13	1.06	<i>p</i> -coumaric acid-4-O-glucoside	325.0936	C <sub>15</sub> H <sub>17</sub> O <sub>8</sub> <sup>-</sup>	163.0395; 119.0499	[8]	√	√
14	1.07	Aesculin	339.0731	C <sub>15</sub> H <sub>15</sub> O <sub>9</sub> <sup>-</sup>	177.0195	[8]	√	-
15	1.22	Ferulic acid glucoside	355.1033	C <sub>16</sub> H <sub>19</sub> O <sub>9</sub> <sup>-</sup>	309.0453; 193.0499 178.0266; 134.0374	[9]	-	√
16	1.34	Shikimic acid	173.0452	C <sub>7</sub> H <sub>9</sub> O <sub>5</sub> <sup>-</sup>	136.9332; 111.0452 93.0360; 85.0303 83.0133	[10]	√	√
17	1.36	Caffeoylquinic acid	353.0880	C <sub>16</sub> H <sub>17</sub> O <sub>9</sub> <sup>-</sup>	191.0556; 173.0445 135.0446; 85.0297	[8]	√	√
18	1.36	Umbelliferone	161.0241	C <sub>9</sub> H <sub>5</sub> O <sub>3</sub> <sup>-</sup>	133.0288	[11]	√	-
19	1.37	Caffeic acid	179.0352	C <sub>9</sub> H <sub>7</sub> O <sub>4</sub> <sup>-</sup>	135.0459; 89.0028	[1]	√	√
20	1.46	Esculetin	177.0193	C <sub>9</sub> H <sub>5</sub> O <sub>4</sub> <sup>-</sup>	149.0253; 133.0296 105.0347; 89.0399	[3]	√	√

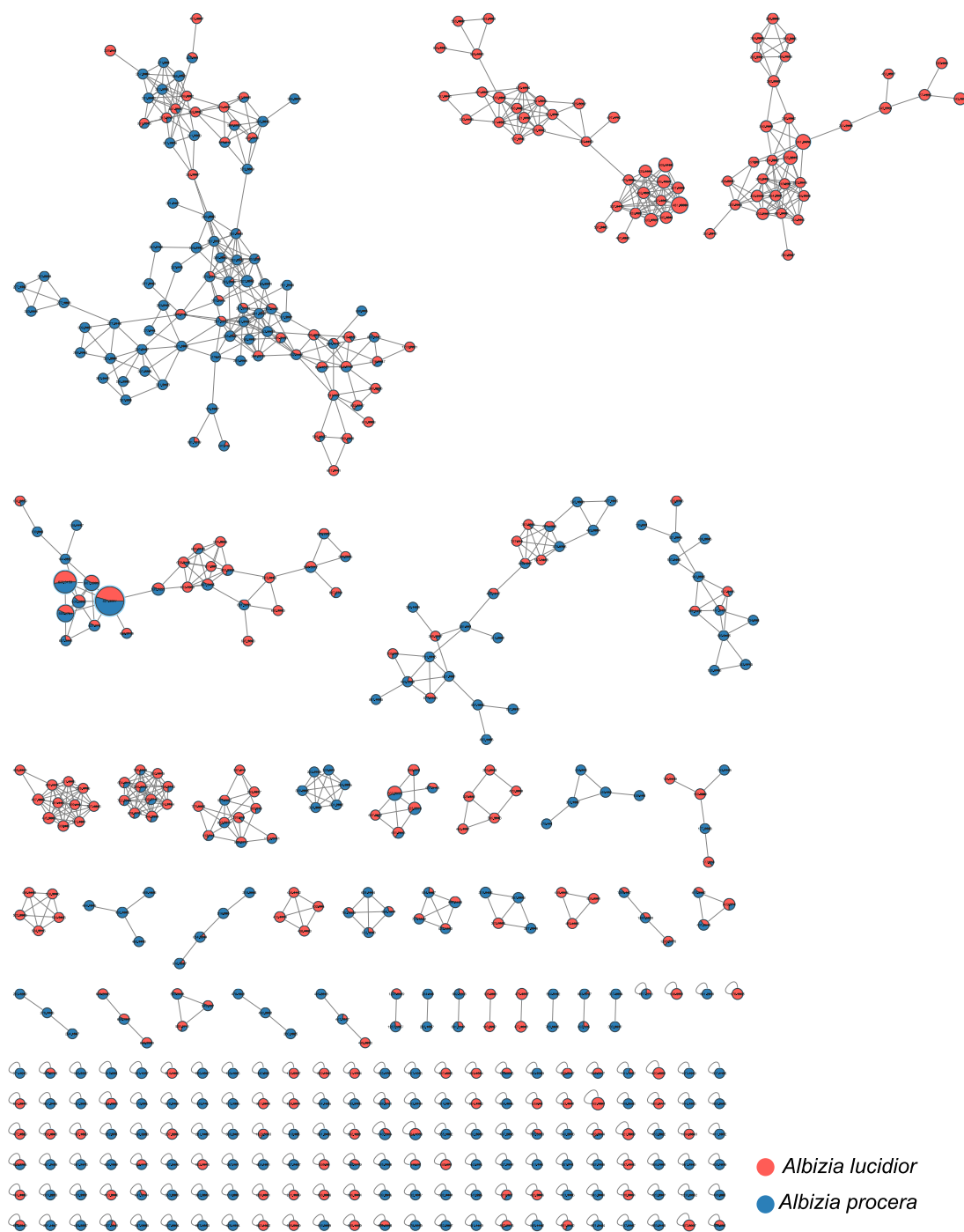
No.	R <sub>t</sub> (min)	Name	Ion m/z ppm	Molecular Formula	MS/MS fragmentation product ions	Ref	Al	Ap
21	1.92	Rosmarinic acid- <i>O</i> -glucoside	521.2047	C <sub>24</sub> H <sub>25</sub> O <sub>13</sub> <sup>-</sup>	359.1511; 344.1271	[12]	-	√
22	2.03	Syringic acid	197.0457	C <sub>9</sub> H <sub>9</sub> O <sub>5</sub> <sup>-</sup>	169.0142; 124.0168	[7]	-	√
23	2.09	Feruloyl quinic acid	367.1042	C <sub>17</sub> H <sub>19</sub> O <sub>9</sub> <sup>-</sup>	193.0521; 173.0454	[13]	-	√
24	2.10	<i>p</i> -coumaroyl quinic acid	337.0934	C <sub>16</sub> H <sub>17</sub> O <sub>8</sub> <sup>-</sup>	191.0556; 163.0394 119.0491; 93.0342	[12]	√	√
25	2.35	<i>p</i> -coumaric acid	163.0401	C <sub>9</sub> H <sub>7</sub> O <sub>3</sub> <sup>-</sup>	119.0510; 93.0350 91.0568	[1]	√	√
26	2.45	Myricetin-3- <i>O</i> -rutinoside	625.1415	C <sub>27</sub> H <sub>29</sub> O <sub>17</sub> <sup>-</sup>	317.0293; 316.0234 287.0917; 271.0252 178.9991; 151.0043	[14]	-	√
27	2.45	Coumaroyl- <i>O</i> -galloyl glucose	477.1040	C <sub>22</sub> H <sub>21</sub> O <sub>12</sub> <sup>-</sup>	313.0558; 169.0151	[7]	-	√
28	2.48	Hexosylrutin	771.1992	C <sub>33</sub> H <sub>39</sub> O <sub>21</sub> <sup>-</sup>	609.1453; 463.0886 301.0346; 300.0273	[9]	-	√
29	2.49	Methyl caffeoyl quinate	367.1033	C <sub>17</sub> H <sub>19</sub> O <sub>9</sub> <sup>-</sup>	191.0564; 179.0349 161.0249; 135.0452	[13]	√	√
30	2.51	Quercetin rhamnosyl- rutinoside	755.2053	C <sub>33</sub> H <sub>39</sub> O <sub>20</sub> <sup>-</sup>	609.1423; 301.0342 300.0280	[9]	√	-
31	2.51	Myricetin-3- <i>O</i> -glucoside	479.0837	C <sub>21</sub> H <sub>19</sub> O <sub>13</sub> <sup>-</sup>	317.0287; 316.0281 271.0235; 178.9972 151.0023	[1]	-	√
32	2.51	Myricetin-3- <i>O</i> -glucuronide	493.0630	C <sub>21</sub> H <sub>17</sub> O <sub>14</sub> <sup>-</sup>	317.0297; 299.0184 271.0262; 178.9974 151.0025	[15]	-	√
33	2.54	Naringenin-7- <i>O</i> -glucoside	433.1151	C <sub>21</sub> H <sub>21</sub> O <sub>10</sub> <sup>-</sup>	313.0707; 271.0618 193.0381; 119.0493	[10]	-	√
34	2.62	Myricetin-3- <i>O</i> -arabinoside	449.0760	C <sub>20</sub> H <sub>17</sub> O <sub>12</sub> <sup>-</sup>	317.0326; 316.0229 287.0182; 271.0238 178.9977	[14]	-	√
35	2.62	Kaempferol-rhamnosyl- galactoside-rhamnoside	739.2108	C <sub>33</sub> H <sub>39</sub> O <sub>19</sub> <sup>-</sup>	593.1507; 285.0403 284.0334; 255.0299	[9]	√	√
36	2.65	Quercetin-3- <i>O</i> -sambubioside	595.1318	C <sub>26</sub> H <sub>27</sub> O <sub>16</sub> <sup>-</sup>	301.0348; 300.0287 271.0252; 255.0304 178.9990; 151.0041	[16]	√	√
37	2.69	Kaempferol-3-(2 <i>G</i> - xylosylrutinoside)	725.1944	C <sub>32</sub> H <sub>37</sub> O <sub>19</sub> <sup>-</sup>	575.1360; 285.0391 284.0327; 255.0304 227.0334; 151.0034	[9]	√	√
38	2.69	Rutin	609.1468	C <sub>27</sub> H <sub>29</sub> O <sub>16</sub> <sup>-</sup>	445.0696; 343.0456 301.0351; 300.0275 271.0242; 255.0297 178.9975; 151.0029	[2]	√	√

No.	R <sub>t</sub> (min)	Name	Ion m/z ppm	Molecular Formula	MS/MS fragmentation product ions	Ref	Al	Ap
39	2.71	Quercetin- <i>O</i> -galloyl-glucoside	615.0990	C <sub>28</sub> H <sub>23</sub> O <sub>16</sub> <sup>-</sup>	463.0890; 313.0557 301.0354; 300.0281 271.0246; 255.0286 169.0143	[2]	√	-
40	2.76	Myricetin-3- <i>O</i> -rhamnoside (myricitrin)	463.0894	C <sub>21</sub> H <sub>19</sub> O <sub>12</sub> <sup>-</sup>	317.0298; 316.0232 287.0202; 271.0248 178.9984	[3]	√	√
41	2.79	Quercetin-3- <i>O</i> -glucuronide	477.0684	C <sub>21</sub> H <sub>17</sub> O <sub>13</sub> <sup>-</sup>	301.0354; 255.0295 178.9984; 151.0033	[15]	-	√
42	2.80	Kaempferol-3- <i>O</i> -rutinoside	593.1522	C <sub>27</sub> H <sub>29</sub> O <sub>15</sub> <sup>-</sup>	285.0403; 284.0323 255.0299; 227.0345 151.0033	[2]	√	√
43	2.85	Quercetin pentose deoxy- hexose	579.1375	C <sub>26</sub> H <sub>27</sub> O <sub>15</sub> <sup>-</sup>	447.0832; 301.0326 300.0288; 255.0314	[9]	√	-
44	2.85	<i>p</i> -Hydroxybenzoic acid	137.0245	C <sub>7</sub> H <sub>5</sub> O <sub>3</sub> <sup>-</sup>	93.0354; 65.0397	[13]	√	-
45	2.85	Azelaic acid	187.0975	C <sub>9</sub> H <sub>15</sub> O <sub>4</sub> <sup>-</sup>	169.0863; 125.0980 123.0825; 97.0669	[17]	√	-
46	2.93	Kaempferol-3- <i>O</i> -sambubioside	579.2086	C <sub>26</sub> H <sub>27</sub> O <sub>15</sub> <sup>-</sup>	417.1552; 285.0408	[18]	√	-
47	2.94	Quercetin-3- <i>O</i> -arabinoside (Avicularin)	433.0789	C <sub>20</sub> H <sub>17</sub> O <sub>11</sub> <sup>-</sup>	301.0344; 300.0275 271.0241; 255.0301 151.0043	[9]	√	√
48	2.96	Myrciacitrin	477.1030	C <sub>23</sub> H <sub>25</sub> O <sub>11</sub> <sup>-</sup>	317.0247; 316.0231 287.0188; 178.9986 151.0023	[9]	-	√
49	2.98	Kaempferol-3- <i>O</i> -glucuronide	461.0736	C <sub>21</sub> H <sub>17</sub> O <sub>12</sub> <sup>-</sup>	285.0406; 257.0453 229.0506	[14]	-	√
50	3.00	Quercetin 3- <i>O</i> -rhamnoside (Quercitrin)	447.0950	C <sub>21</sub> H <sub>19</sub> O <sub>11</sub> <sup>-</sup>	301.0356; 300.0281 271.0249; 255.0301 178.9987; 151.0043	[3]	√	√
51	3.06	Ferulic acid	193.0508	C <sub>10</sub> H <sub>9</sub> O <sub>4</sub> <sup>-</sup>	161.0247; 134.0374 133.0298	[1]	√	√
52	3.10	Phloridzin	435.1304	C <sub>21</sub> H <sub>23</sub> O <sub>10</sub> <sup>-</sup>	273.0775; 167.0353	[3]	-	√
53	3.18	Quercetin benzoyl-glucoside	583.4803	C <sub>28</sub> H <sub>23</sub> O <sub>14</sub> <sup>-</sup>	463.0891; 301.0322 300.0269; 271.0229 255.0282; 152.0106	[7]	√	-

No.	R <sub>t</sub> (min)	Name	Ion m/z ppm	Molecular Formula	MS/MS fragmentation product ions	Ref	Al	Ap
54	3.18	Kaempferol-3-O-rhamnoside	431.0988	C <sub>21</sub> H <sub>19</sub> O <sub>10</sub> <sup>-</sup>	285.0416; 284.0340 255.0310; 227.0360 151.0036	[2]	√	√
55	3.19	Myricetin	317.0309	C <sub>15</sub> H <sub>9</sub> O <sub>8</sub> <sup>-</sup>	178.9998; 151.0448 137.0250	[9]	-	√
56	3.21	Quercetin-3-O-glucosyl-6''-acetate	505.1002	C <sub>23</sub> H <sub>21</sub> O <sub>13</sub> <sup>-</sup>	329.0664; 301.0343 300.0280; 271.0241 255.0290; 151.0023	[10]	-	√
57	3.21	Myricetin-O-(O-galloyl)-3-rhamnoside	615.1002	C <sub>28</sub> H <sub>23</sub> O <sub>16</sub> <sup>-</sup>	463.0895; 317.0315 178.9998; 169.0148 151.0047	[3]	-	√
58	3.21	Quercetin-di-O-glucoside	625.1202	C <sub>27</sub> H <sub>29</sub> O <sub>17</sub> <sup>-</sup>	463.0891; 301.0359 300.0284; 255.0315	[1]	√	-
59	3.21	Chrysoeriol-O-glucoside	461.1084	C <sub>22</sub> H <sub>21</sub> O <sub>11</sub> <sup>-</sup>	299.0200	[19]	-	√
60	3.41	Quercetin galloyl rhamnoside	599.1051	C <sub>28</sub> H <sub>23</sub> O <sub>15</sub> <sup>-</sup>	447.0920; 383.1042 301.0368; 273.0402 178.9991; 151.0041	[2]	-	√
61	3.41	Kaempferol acetyl glycoside	489.1044	C <sub>23</sub> H <sub>21</sub> O <sub>12</sub> <sup>-</sup>	285.0409; 284.0335 255.0306; 227.0359	[1]	-	√
62	3.43	Quercetin coumaroyl glucoside	609.1263	C <sub>30</sub> H <sub>25</sub> O <sub>14</sub> <sup>-</sup>	463.0872; 301.0352 300.0276; 271.0238 178.9977; 151.0031	[1]	√	-
63	3.46	Quercetin feruloyl glucoside	639.1359	C <sub>31</sub> H <sub>27</sub> O <sub>15</sub> <sup>-</sup>	463.0881; 301.0350 300.0276; 271.0248 255.0291; 151.0025	[7]	√	-
64	3.48	Ethyl caffeate	207.0665	C <sub>11</sub> H <sub>11</sub> O <sub>4</sub> <sup>-</sup>	179.0352; 161.0250 135.0453; 134.0376	[13]	√	√
65	3.52	<i>p</i> -methoxycinnamic acid	177.0563	C <sub>10</sub> H <sub>9</sub> O <sub>3</sub> <sup>-</sup>	162.0317; 145.0296 133.0061; 118.0427	[20]	-	√
66	3.55	Quercetin	301.0345	C <sub>15</sub> H <sub>9</sub> O <sub>7</sub> <sup>-</sup>	178.9993; 151.0045 121.0296; 107.0147	[1]	√	√
67	3.55	Kaempferol	285.0396	C <sub>15</sub> H <sub>9</sub> O <sub>6</sub> <sup>-</sup>	243.0301; 217.0505 199.0403; 175.0397	[1]	√	√
68	3.60	Trihydroxyoctadienoic acid	327.2182	C <sub>18</sub> H <sub>31</sub> O <sub>5</sub> <sup>-</sup>	291.1969; 229.1449 211.1349; 171.1033	[2]	√	√
69	3.77	Trihydroxyoctadecenoic acid	329.2336	C <sub>18</sub> H <sub>33</sub> O <sub>5</sub> <sup>-</sup>	229.1452; 211.1348 171.1031	[2]	√	√
70	3.85	Naringenin	271.0623	C <sub>15</sub> H <sub>11</sub> O <sub>5</sub> <sup>-</sup>	151.0039; 119.0509 65.0045	[1]	-	√
71	3.85	Dihydroxy palmitic acid	287.2233	C <sub>16</sub> H <sub>31</sub> O <sub>4</sub> <sup>-</sup>	269.2144; 241.2189 223.2067	[2]	√	√

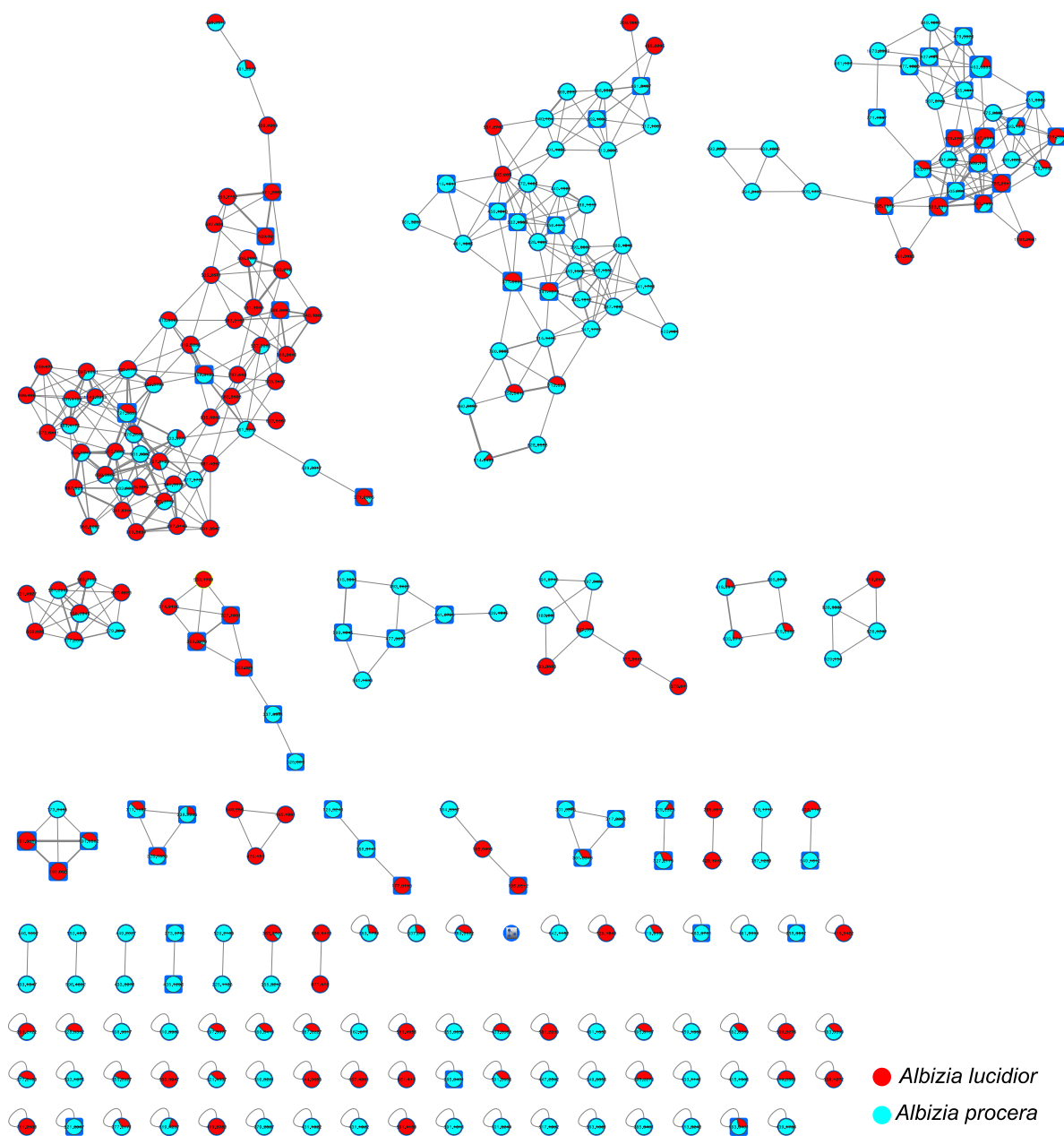
No.	R <sub>t</sub> (min)	Name	Ion m/z ppm	Molecular Formula	MS/MS fragmentation product ions	Ref	Al	Ap
72	4.42	Dihydroxyoctadecadienoic acid	311.2223	C <sub>18</sub> H <sub>31</sub> O <sub>4</sub> <sup>-</sup>	311.2208; 293.2096 275.2005; 223.1705	[19]	√	√
73	4.51	Epoxy-hydroxyoctadeca- dienoic acid	309.2073	C <sub>18</sub> H <sub>29</sub> O <sub>4</sub> <sup>-</sup>	291.1959; 273.1980 221.1541	[19]	√	√
74	4.56	Pinocembrin	255.0665	C <sub>15</sub> H <sub>11</sub> O <sub>4</sub> <sup>-</sup>	213.0556; 171.0443 151.0031; 107.0134 83.0136; 65.0037	[1]	-	√
75	4.58	Dihydroxyoctadecenoic acid	313.2386	C <sub>18</sub> H <sub>33</sub> O <sub>4</sub> <sup>-</sup>	295.2300; 277.2174 183.1392	[19]	-	√
76	4.98	Sphingolipid conjugate II	564.3315	C <sub>27</sub> H <sub>51</sub> O <sub>9</sub> NP <sup>-</sup>	279.2329	[19]	√	-
77	5.00	13-keto-octadeca- 9,11-dienoic acid	293.2126	C <sub>18</sub> H <sub>29</sub> O <sub>3</sub> <sup>-</sup>	275.2027; 223.1346 195.1396; 179.1463	[5]	√	√
78	5.13	Hydroxyoctadecenoic acid	297.1535	C <sub>18</sub> H <sub>33</sub> O <sub>3</sub> <sup>-</sup>	279.9880; 183.0120	[19]	√	√
79	5.17	9-hydroxy-10,12- octadecadienoic acid	295.2276	C <sub>18</sub> H <sub>31</sub> O <sub>3</sub> <sup>-</sup>	277.2178; 195.1393 171.1029	[5]	√	√
80	5.20	Sphingolipid conjugate III	566.3478	C <sub>27</sub> H <sub>53</sub> O <sub>9</sub> NP <sup>-</sup>	281.2497	[19]	√	-
81	5.76	Hydroxy palmitic acid	271.2275	C <sub>16</sub> H <sub>31</sub> O <sub>3</sub> <sup>-</sup>	253.2167; 225.2220	[19]	√	√
82	5.93	Linolenic acid	277.2171	C <sub>18</sub> H <sub>29</sub> O <sub>2</sub> <sup>-</sup>	259.2068; 234.1868	[5]	√	√
83	5.96	Yohimbic acid	339.1999	C <sub>20</sub> H <sub>23</sub> N <sub>2</sub> O <sub>3</sub> <sup>-</sup>	183.0117	[10]	√	√
84	6.15	Linoleic acid	279.2326	C <sub>18</sub> H <sub>31</sub> O <sub>2</sub> <sup>-</sup>	261.2199; 59.0138	[5]	√	√
85	6.32	Palmitic acid	255.2328	C <sub>16</sub> H <sub>31</sub> O <sub>2</sub> <sup>-</sup>	234.9956; 206.9872 166.9912	[19]	√	√
86	6.44	Oleic acid	281.2486	C <sub>18</sub> H <sub>33</sub> O <sub>2</sub> <sup>-</sup>	260.9945; 240.9874	[5]	√	√
87	6.74	Stearic acid	283.2632	C <sub>18</sub> H <sub>35</sub> O <sub>2</sub> <sup>-</sup>	242.9835; 179.0094	[19]	√	√

√, found; -, not found; Al, *A. lucidior*; Ap, *A. procera*

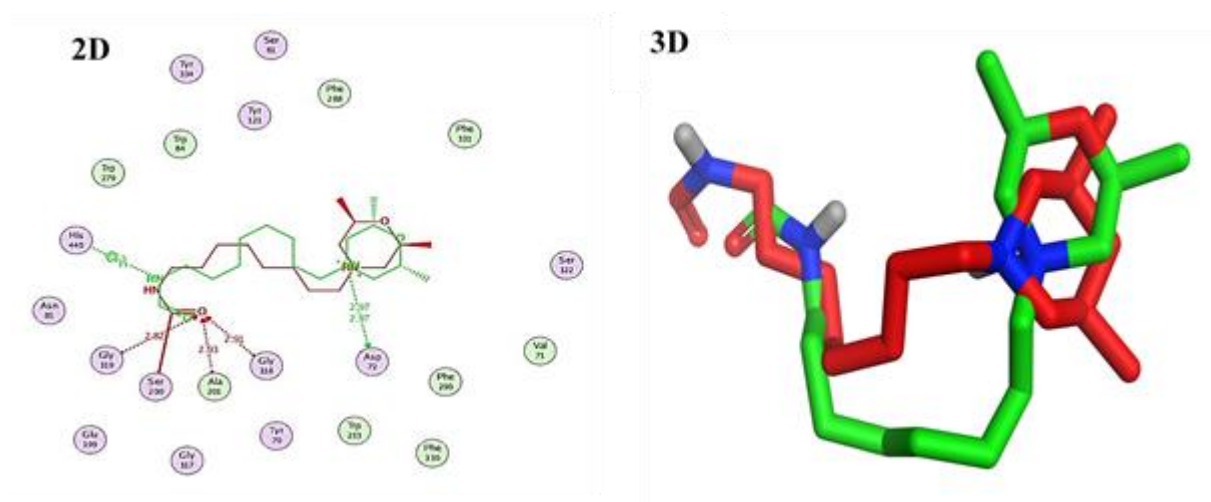


**Figure S1.** GNPS molecular network of two *Albizia* spp. ethanolic extracts in the positive ion mode. Each node is displayed as pie chart representing relative abundance of the metabolite with red and blue colors in the ethanolic extracts of *A. lucidior* and *A. procera*, respectively. The node label represents precursor mass ( $m/z$ ). The node size represents the sum of precursor ion intensity.





**Figure S2.** GNPS molecular network of two *Albizia* spp. ethanolic extracts in the negative ion mode. Each node is displayed as pie chart representing relative abundance of the metabolite with red and blue colors in the ethanolic extracts of *A. lucidior* and *A. procera*, respectively. The node label represents precursor mass ( $m/z$ ). The node size represents the sum of precursor ion intensity.



**Figure S3.** Superimposition of the redocked MF2 700 inhibitor (green) over its native one (red).

## References

1. Kramberger, K.; Barlič-Maganja, D.; Bandelj, D.; Baruca Arbeiter, A.; Peeters, K.; Miklavčič Višnjevec, A.; Jenko Pražnikar, Z. HPLC-DAD-ESI-QTOF-MS determination of bioactive compounds and antioxidant activity comparison of the hydroalcoholic and water extracts from two *Helichrysum italicum* species. *Metabolites* **2020**, *10*, 403.
2. Rosa, W.; da Silva Domingos, O.; de Oliveira Salem, P.P.; Caldas, I.S.; Murgu, M.; Lago, J.H.G.; Sartorelli, P.; Dias, D.F.; Chagas-Paula, D.A.; Soares, M.G. *In vivo* anti-inflammatory activity of Fabaceae species extracts screened by a new *ex vivo* assay using human whole blood. *Phytochem. Anal.* **2021**, *32*, 859-883.
3. Singh, D.; Siew, Y.-Y.; Chong, T.-I.; Yew, H.-C.; Ho, S.S.-W.; Lim, C.S.E.-S.; Tan, W.-X.; Neo, S.-Y.; Koh, H.-L. Identification of phytoconstituents in *Leea indica* (Burm. F.) Merr. leaves by high performance liquid chromatography micro time-of-flight mass spectrometry. *Molecules* **2019**, *24*, 714.
4. Ateba, S.B.; Njamen, D.; Gatterer, C.; Scherzer, T.; Zehl, M.; Kählig, H.; Krenn, L. Rare phenolic structures found in the aerial parts of *Eriosema laurentii* De Wild. *Phytochemistry* **2016**, *128*, 5-11.
5. Kalogiouri, N.P.; Aalizadeh, R.; Dasenaki, M.E.; Thomaidis, N.S. Authentication of Greek PDO kalamata table olives: A novel non-target high resolution mass spectrometric approach. *Molecules* **2020**, *25*, 2919.
6. Shivanagoudra, S.R.; Perera, W.H.; Perez, J.L.; Athrey, G.; Sun, Y.; Wu, C.S.; Jayaprakasha, G.; Patil, B.S. *In vitro* and *in silico* elucidation of antidiabetic and anti-inflammatory

- activities of bioactive compounds from *Momordica charantia* L. *Bioorg. Med. Chem.* **2019**, *27*, 3097-3109.
7. Sobeh, M.; Rezq, S.; Sabry, O.M.; Abdelfattah, M.A.O.; El Raey, M.A.; El-Kashak, W.A.; El-Shazly, A.M.; Mahmoud, M.F.; Wink, M. *Albizia anthelmintica*: HPLC-MS/MS profiling and *in vivo* anti-inflammatory, pain killing and antipyretic activities of its leaf extract. *Biomed. Pharmacother.* **2019**, *115*, 108882.
  8. Mudrić, S.Ž.; Gašić, U.M.; Dramićanin, A.M.; Ćirić, I.Ž.; Milojković-Opsenica, D.M.; Popović-Dorđević, J.B.; Momirović, N.M.; Tešić, Ž.L. The polyphenolics and carbohydrates as indicators of botanical and geographical origin of Serbian autochthonous clones of red spice paprika. *Food Chem.* **2017**, *217*, 705-715.
  9. Abu-Reidah, I.M.; Arráez-Román, D.; Warad, I.; Fernández-Gutiérrez, A.; Segura-Carretero, A. UHPLC/MS<sup>2</sup>-based approach for the comprehensive metabolite profiling of bean (*Vicia faba* L.) by-products: A promising source of bioactive constituents. *Food Res. Int.* **2017**, *93*, 87-96.
  10. Elkousy, R.H.; Said, Z.N.A.; Abd El-Baseer, M.A.; Abu El wafa, S.A. Antiviral activity of castor oil plant (*Ricinus communis*) leaf extracts. *J. Ethnopharmacol.* **2021**, *271*, 113878.
  11. Elez Garofulić, I.; Malin, V.; Repajić, M.; Zorić, Z.; Pedisić, S.; Sterniša, M.; Smole Možina, S.; Dragović-Uzelac, V. Phenolic Profile, Antioxidant Capacity and Antimicrobial Activity of Nettle Leaves Extracts Obtained by Advanced Extraction Techniques. *Molecules* **2021**, *26*, 6153.
  12. Sobeh, M.; Hassan, S.A.; El Raey, M.A.; Khalil, W.A.; Hassan, M.A.; Wink, M. Polyphenolics from *Albizia harveyi* exhibit antioxidant activities and counteract oxidative damage and ultra-structural changes of cryopreserved bull semen. *Molecules* **2017**, *22*, 1993.
  13. Ruan, J.; Yan, J.; Zheng, D.; Sun, F.; Wang, J.; Han, L.; Zhang, Y.; Wang, T. Comprehensive chemical profiling in the ethanol extract of *Pluchea indica* aerial parts by liquid chromatography/mass spectrometry analysis of its silica gel column chromatography fractions. *Molecules* **2019**, *24*, 2784.
  14. Fotirić Akšić, M.; Dabić Zagorac, D.; Sredojević, M.; Milivojević, J.; Gašić, U.; Meland, M.; Natić, M. Chemometric characterization of strawberries and blueberries according to their phenolic profile: Combined effect of cultivar and cultivation system. *Molecules* **2019**, *24*, 4310.
  15. Limwachiranon, J.; Huang, H.; Li, L.; Duan, Z.; Luo, Z. Recovery of lotus (*Nelumbo nucifera* Gaertn.) seedpod flavonoids using polar macroporous resins: The updated understanding on adsorption/desorption mechanisms and the involved intermolecular attractions and bonding. *Food Chem.* **2019**, *299*, 125108.
  16. Huang, Q.; Zhang, F.; Liu, S.; Jiang, Y.; Ouyang, D. Systematic investigation of the pharmacological mechanism for renal protection by the leaves of *Eucommia ulmoides* Oliver using UPLC-Q-TOF/MS combined with network pharmacology analysis. *Biomed. Pharmacother.* **2021**, *140*, 111735.
  17. Chiriac, E.R.; Chițescu, C.L.; Borda, D.; Lupoae, M.; Gird, C.E.; Geană, E.-I.; Blaga, G.-V.; Boscencu, R. Comparison of the polyphenolic profile of *Medicago sativa* L. and *Trifolium*

- pratense* L. sprouts in different germination stages using the UHPLC-Q exactive hybrid quadrupole orbitrap high-resolution mass spectrometry. *Molecules* **2020**, *25*, 2321.
18. Zhang, T.; Qiu, F.; Chen, L.; Liu, R.; Chang, M.; Wang, X. Identification and *in vitro* anti-inflammatory activity of different forms of phenolic compounds in *Camellia oleifera* oil. *Food Chem.* **2021**, *344*, 128660.
  19. Farag, M.A.; Khattab, A.R.; Maamoun, A.A.; Kropf, M.; Heiss, A.G. UPLC-MS metabolome based classification of Lupinus and Lens seeds: A prospect for phyto-equivalency of its different accessions. *Food Res. Int.* **2019**, *115*, 379-392.
  20. Song, Q.; Song, Y.; Zhang, N.; Li, J.; Jiang, Y.; Zhang, K.; Zhang, Q.; Tu, P. Potential of hyphenated ultra-high performance liquid chromatography-scheduled multiple reaction monitoring algorithm for large-scale quantitative analysis of traditional Chinese medicines. *RSC Adv.* **2015**, *5*, 57372-57382.