

Review

# Hairy Root Cultures as a Source of Polyphenolic Antioxidants: Flavonoids, Stilbenoids and Hydrolyzable Tannins

Janusz Malarz <sup>1</sup>, Klaudia Michalska <sup>1</sup>, Yulia V. Yudina <sup>2</sup> and Anna Stojakowska <sup>1,\*</sup>

<sup>1</sup> Maj Institute of Pharmacology, Polish Academy of Sciences, Smętna Street 12, 31-343 Kraków, Poland; malarzj@if-pan.krakow.pl (J.M.); klaudiaz@if-pan.krakow.pl (K.M.)

<sup>2</sup> National Technical University, Kharkiv Polytechnic Institute, Kyrpychova Street 2, 61002 Kharkiv, Ukraine; eco3557@gmail.com

\* Correspondence: stoja@if-pan.krakow.pl; Tel.: +48-126623254

**Abstract:** Due to their chemical properties and biological activity, antioxidants of plant origin have gained interest as valuable components of the human diet, potential food preservatives and additives, ingredients of cosmetics and factors implicated in tolerance mechanisms against environmental stress. Plant polyphenols are the most prominent and extensively studied, albeit not only group of, secondary plant (specialized) metabolites manifesting antioxidative activity. Because of their potential economic importance, the productive and renewable sources of the compounds are desirable. Over thirty years of research on hairy root cultures, as both producers of secondary plant metabolites and experimental systems to investigate plant biosynthetic pathways, brought about several spectacular achievements. The present review focuses on the *Rhizobium rhizogenes*-transformed roots that either may be efficient sources of plant-derived antioxidants or were used to elucidate some regulatory mechanisms responsible for the enhanced accumulation of antioxidants in plant tissues.

**Keywords:** *Agrobacterium rhizogenes*; chalcone; ellagitannin; flavone; flavonol; isoflavonoid; proanthocyanidin; resveratrol; *Rhizobium rhizogenes*



**Citation:** Malarz, J.; Michalska, K.; Yudina, Y.V.; Stojakowska, A. Hairy Root Cultures as a Source of Polyphenolic Antioxidants: Flavonoids, Stilbenoids and Hydrolyzable Tannins. *Plants* **2022**, *11*, 1950. <https://doi.org/10.3390/plants11151950>

Academic Editors: Angélica Román-Guerrero and Maria Elena Estrada-Zuñiga

Received: 20 June 2022

Accepted: 25 July 2022

Published: 27 July 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

The interest in hairy root (HR) cultures as fast-growing, genetically stable axenic roots that are effective producers of both biomass and specialized plant metabolites started in the 1980s [1]. Since then, numerous culture systems have been established to produce compounds of economic value or potential economic interest to produce recombinant proteins to obtain transgenic plants of altered phenotype to obtain artificial seeds and perform metabolic engineering of plants [1–6]. The ability of *Rhizobium rhizogenes* (also known as *Agrobacterium rhizogenes*) [7] to transfer and permanently incorporate a foreign genetic material into the plant genomic DNA has also been used to study the regulatory mechanisms of plant metabolism [6,8,9]. Additionally, the enzymatic activity of HRs may be utilized for biotransformation and phytoremediation processes [10–14].

In contrast to other fast-growing types of plant tissue cultures, HRs are usually cultivated in nutrient media with a reduced concentration of macronutrients and without the addition of growth regulators that lower the costs of biotechnological processes. Several strategies were employed to scale up HR cultures from the laboratory to industrial applications, including various bioreactor designs [15,16], e.g., airlifts, bubble columns and nutrient mist, etc. The production of biologically active natural products and proteins by HR cultures has been a subject of numerous patents and shows potential for successful industrial applications.

Until now, the main limitations that hamper the industrial use of HR cultures are still unsatisfactory upscaling results (the need for large-volume bioreactors) and unsatisfactory cost/profit ratios.

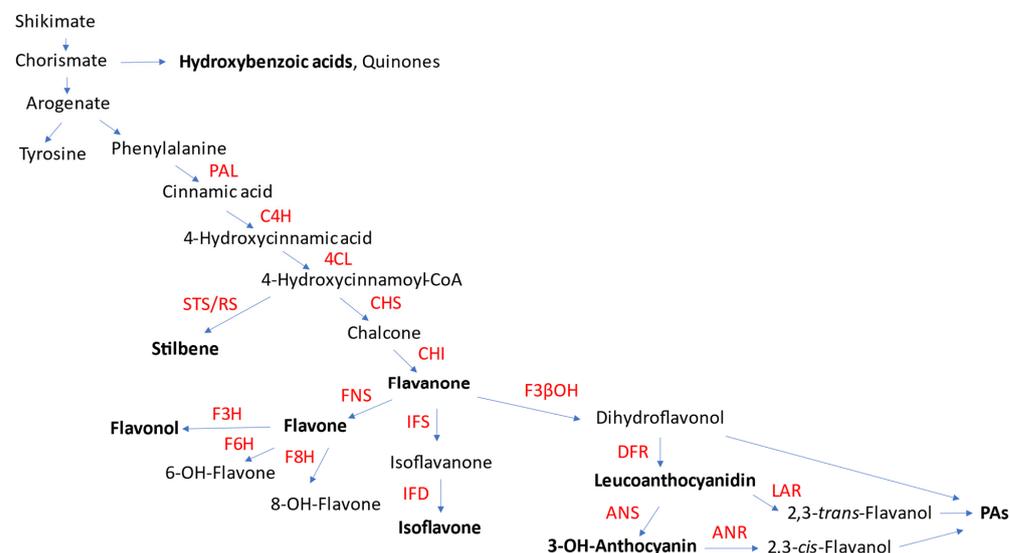
Nearly every class of plant-specialized metabolite includes some compounds of antioxidative activity [17–20], but the most effective antioxidants, acting by direct free-radical scavenging, originate mostly from phenolics and terpenoids [21,22]. Numerous natural products known as antioxidants are also recognized as potent chemopreventive and anti-cancer agents, influencing various intracellular signaling pathways [23,24]. Antioxidants of plant origin may also be useful as protective factors against dysfunctions of nervous system connected with aging [25].

Plant metabolites with antioxidant activity, which are the constituents of widely grown food plants, can be isolated from post-production waste or obtained as side products in the process of food manufacturing [26–29]. Some natural antioxidant products are a common occurrence and their isolation from either cultivated or wild plants seem to be the optimal solution. The yields of these natural products may be improved by bioengineering using the transformation by *R. rhizogenes*. In the case of threatened or overexploited plant species, HR cultures offer an opportunity to produce rare and valuable compounds with no harm to the natural habitats of the plant.

The present review is based on the results published in journals indexed in the Web of Science and Scopus databases. Due to the large number of studies dealing with polyphenolic metabolites of HRs, we decided to limit the scope of the review to flavonoids, stilbenoids and hydrolyzable tannins.

## 2. Polyphenolic Antioxidants in HRs

Phenolics are one of the largest classes of plant-specialized metabolites. They all originate from the shikimate pathway leading to aromatic amino acids phenylalanine and tyrosine and also share the phenylpropanoid pathway (Figure 1). The compounds are divided into several subclasses, including flavonoids, tannins, stilbenoids, lignans, hydroxycinnamates and phenylethanoids, to mention a few of the most popular subclasses.



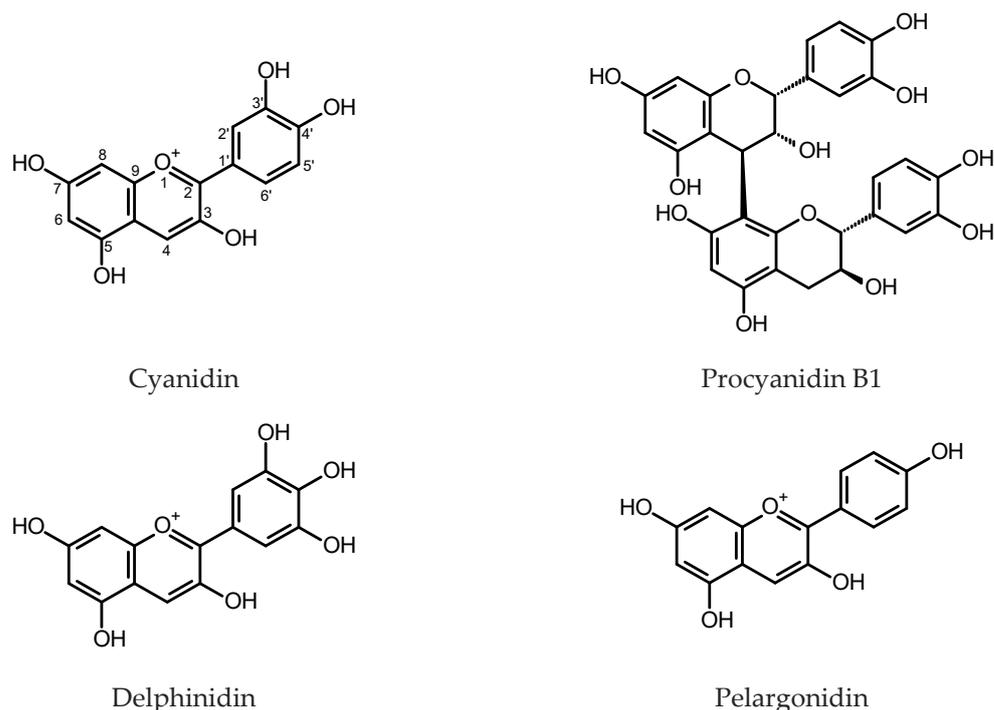
**Figure 1.** Simplified scheme of biosynthesis of selected plant polyphenols and positions of biosynthetic enzymes mentioned in the review (PAL—phenylalanine ammonia lyase; C4H—cinnamate 4-hydroxylase; 4CL—4-coumarate:CoA ligase; STS—stilbene synthase; RS—resveratrol synthase; CHS—chalcone synthase; CHI—chalcone isomerase; FNS—flavone synthase; IFS—isoflavone synthase; F3βOH—flavanone 3-β-hydroxylase; F3H—flavone 3-hydroxylase; F6H—flavone 6-hydroxylase; F8H—flavone 8-hydroxylase; IFD—isoﬂavone dehydratase; DFR—dihydroﬂavonol 4-reductase; ANS—anthocyanidin synthase; ANR—anthocyanidin reductase; LAR—leucoanthocyanidin reductase). Hydroxybenzoic acids group include gallic acid, a precursor of hydrolyzable tannins; PAs—proanthocyanidins).

## 2.1. Flavonoids

### 2.1.1. Anthocyanins and Proanthocyanidins (PAs)

Numerous factors may influence anthocyanin and PA production by HRs. The most frequently studied were the effects of light, plant growth regulators, sucrose concentration and the *R. rhizogenes* strain used in the experiment. The overexpression or heterologous expression of genes-encoding transcription factors implicated in the regulation of flavonoid biosynthesis (especially members of *myb* gene family) or the overexpression of genes-encoding biosynthetic enzymes (*DFR*, *LAR*, *ANR*) are strategies often used to increase anthocyanins and PAs content in the HRs or transgenic plants regenerated from the roots.

Morris and Robbins [30] observed that the HRs of *Lotus corniculatus* L. (Fabaceae), maintained in the dark, accumulated insoluble tannins, which presented cyanin and delphinidin at a ratio like that found in the intact plant after hydrolysis. Supplementation with auxins reduced the content of tannins in the roots. On the contrary, in the HR cultures of *Leontopodium alpinum* Cass. (Asteraceae), the addition of 6-benzylaminopurine (BAP) to the culture medium 14 days prior to harvest increased the anthocyanins content [31]. An effect of auxin supplementation on anthocyanin contents in the HRs of Tartary buckwheat (*Fagopyrum tataricum* (L.) Gaertn., Polygonaceae) cultivar “Hokkai T10” was also studied [32]. Indole-3-butyric acid (IBA) added to a nutrient medium at a concentration of 4.92  $\mu$ M, caused enhanced accumulation of cyanidin 3-*O*-glucoside and cyanidin 3-*O*-rutinoside in the roots (for structures, see Figure 2). The anthocyanin content reached 0.89 and 1.15 mg/g dry weight (DW), respectively, and was about three times higher than that found in the control cultures.



**Figure 2.** Chemical structures of selected anthocyanins and procyanidin B1.

Nishiyama and Yamakawa investigated effects of the culture illumination, composition of nutrient medium and sucrose content on the anthocyanin production in the HRs of *Ipomea batatas* (L.) Lam. (Convolvulaceae) [33]. They found that light and high sucrose concentration in the medium (5%) favored anthocyanin production. The beneficial effect of light on cyanidin 3-*O*-rutinoside accumulation was also observed in Tartary buckwheat HRs [34]. Motomori et al. [35] studied polyphenolics accumulation in the HRs of *Fragaria x ananassa* Duch. cv. Reikou (Rosaceae). In optimum conditions, the root culture produced

up to 8 mg/g DW of procyanidin B-3, but the roots of the field-grown plant contained three times as much of the compound (24.2 mg/g DW).

The expression of full-length cDNA encoding dihydroflavonol 4-reductase (DFR) from *Antirrhinum majus* L. (common snapdragon, Plantaginaceae) in the HRs of *Lotus corniculatus* L. led to several high-producing clones. The maximum content of condensed tannins in the control roots was estimated as 0.62 mg/g fresh weight (FW), whereas the transgenic roots produced up to 1.06 mg/g FW of the compounds [36]. The proportions of subunits (procyanidin, prodelphinidin and propelargonidin) yielded on the hydrolysis of condensed tannins were also measured, and significant differences in propelargonidin accumulation were observed in the transgenic clones in comparison with the controls. The plasmid 121.Sn carrying the maize gene *Sn*, which is responsible for the transactivation of the anthocyanin pathway in different tissues, was introduced into the genomes of ten different plant species using the transformation with *R. rhizogenes*. Over 50% of the obtained *Medicago sativa* L. (Fabaceae) and *Lotus angustissimus* L. accession CPI 113587 HR clones showed pigmentation. On the other hand, the HRs of *Lotus corniculatus*, *L. japonicus* (Regel) K. Larsen and *L. angustissimus* accession CPI 113590 produced no pigment. The results suggested the transcription factor *Sn* derived from monocotyledonous plant may function in several dicot species [37].

The HRs of *Medicago truncatula* Gaertn. (Fabaceae) and *Vitis vinifera* L. (Vitaceae) were used to investigate the mechanisms of anthocyanin biosynthesis regulation by transcription factors encoded by the members of *myb* gene family [38–45]. The expression of *Arabidopsis* TT2 (Transparent Testa 2) transcription factor from the MYB family in the HRs of *M. truncatula* led to the induction of the genes for flavonoid and proanthocyanidin (PA) biosynthesis pathway and enhanced accumulation of PAs in the roots. The average soluble PAs content in TT2-carrying transformants was over ten times higher than that in the control HRs transfected with an “empty” vector. Insoluble PAs content in TT2-expressing roots was over 24-fold higher [38]. The induction of genes for PAs biosynthetic pathway was also true when the transcription factors: GmTT2A, GmTT2B or GmMYB5A were ectopically expressed in soybean (*Glycine max* L.) HRs [46]. Expression of *AtTT2* in chickpea HRs resulted in a high accumulation of soluble PAs (>1200 µg catechin equivalents (eq.)/g FW) [47]. MtPAR MYB transcription factor expressed in *M. truncatula* HRs dramatically increased soluble PAs content (up to 100-fold higher than that in the control roots) but did not influence insoluble PAs accumulation and diminished the anthocyanin content. Maximum content of soluble PAs reached 10 µM of (–) epicatechin eq. per g FW [40]. Li et al. [48] found that transgenic *M. truncatula* HRs that over-expressed MtPAR were characterized by enhanced accumulation of soluble PAs and diminished production of isoflavones. The role of MtPAR in the distribution of precursors to flavonoid, isoflavonoid, anthocyanin and PA pathways was emphasized by the same study. The HRs of *M. truncatula* overexpressing MYB2 demonstrated suppression of the expression of genes-encoding dihydroflavonol 4-reductase (DFR1) and anthocyanidin synthase (ANS). Anthocyanin accumulation characteristic of wild-type roots was ceased in the MYB2-overexpressing roots [43]. Wild-type HRs of *M. truncatula* with the ectopic expression of MtTT8 transcription factor showed enhanced expression of the flavonoid biosynthesis pathway genes and elevated production of PAs and anthocyanins. Contents of anthocyanins (as cyanidin 3-O-glucoside eq.), soluble PAs (as epicatechin eq.) and insoluble PAs (as proanthocyanidin B1 eq.) in the MtTT8-expressing roots reached 600 µg/g FW, 27 µg/g FW and 180 µg/g FW, respectively [44]. Heterologous expression of GhMYB36—a gene for TT2-type MYB transcription factor from a tetraploid *Gossypium hirsutum* L. (Malvaceae), in the HRs of *M. truncatula* made it possible to achieve 100 µg/g FW of total soluble PAs (epicatechin eq.) and over 1600 µg/g FW of total insoluble PAs (proanthocyanidin B1 eq.) [44]. VvMybPA1 and VvMybPA2 are transcription factors involved in the regulation of PA biosynthesis in *V. vinifera* seeds, leaves and exocarp of young berries. The ectopic expression of the genes-encoding these transcription factors in the HRs of *V. vinifera* caused activation of the enzymes of the flavonoid pathway and enhanced accumulation of PAs (up to 8 mg/g FW).

Moreover, the *VvMybPA1*- and *VvMybPA2*-expressing transformed roots started to produce PAs with trihydroxylated ring B [39].

The overexpression of *VvMYBC2-L1* in grapevine HRs negatively affected PAs biosynthesis, as it can be expected for a member of subgroup 4 MYB transcription factors [41]. A similar effect was achieved in *V. vinifera* HRs by an overexpression of *VvMYBC2-L3* [42]. Transgenic HRs of *Litchi chinensis* Sonn. (Sapindaceae), overexpressing *LcMYB1*, contained over 3 mg/g FW of anthocyanidins and nearly 15 mg/g FW of PAs. Wild-type lychee HRs accumulated about 5 mg/g FW PAs and minute amounts of anthocyanidins [49]. Regulatory functions of *PpMYB10.1* and MYB 182 transcription factors in anthocyanidin biosynthesis were also investigated using peach and poplar HR cultures, respectively [50,51]. Expression of *Camellia sinensis* (L.) Kuntze-derived genes-encoding leucoanthocyanidin reductase (LAR) and anthocyanidin reductase (ANR) in the HRs of *M. truncatula* resulted in slightly elevated content of insoluble PAs. The ectopic expression of *CsLAR* enhanced the accumulation of anthocyanins, whereas *CsANRs* expressing roots tend to accumulate more soluble PAs than the control roots [52].

HRs of Tartary buckwheat were frequently used to investigate the effects of different factors on anthocyanins accumulation. Thwe et al. [53] examined seven wild-type *R. rhizogenes* strains to assess transformation efficiency and anthocyanin content in regenerated hairy root clones of *F. tataricum* cv. "Hokkai T10". The R1000 strain, except for the highest transformation efficiency and growth rate of roots, provided high transcript levels for most genes of the flavonoid biosynthetic pathway and the highest contents of cyanidin 3-O-glucoside (800 µg/g DW) and cyanidin 3-O-rutinoside (2410 µg/g DW). A plant growth regulator—ethephon—at the concentration of 0.5 mg/L, significantly increased cyanidin 3-O-glucoside and cyanidin 3-O-rutinoside accumulation in hairy roots of buckwheat. Moreover, upregulation of the genes involved in flavonoid biosynthesis upon ethephon treatment was observed [54]. Overexpression of *FtMYB1*, *FtMYB2*, *FtMYB3* and *FtMYB-like* in the HRs of Tartary buckwheat up-regulated the genes of the phenylpropanoid biosynthetic pathway and enhanced anthocyanin production. *FtMYB18* overexpression negatively influenced anthocyanin biosynthesis in the roots [9,55].

The black or purple carrot (*Daucus carota* L. ssp. *sativus* var. *atrorubens* Alef, Apiaceae) belongs to the Eastern group of domesticated carrots and is cultivated and commonly eaten in India, Pakistan, Turkey and Afghanistan. In contrast to the Western orange carrots, the roots of the plant accumulated anthocyanins. Acylated anthocyanidin triglycosides produced by the plant, due to their chemical stability, are used as food colorants [56]. The HRs of black carrot were obtained by the infection of taproot and hypocotyl explants of black carrot inbred line "43" and taproot explants of cv. "Night Bird". The maximum total monomeric anthocyanin content, spectrophotometrically measured, exceeded 3 mg/g DW and was found in the HRs derived from hypocotyl explants, cultivated in  $\frac{1}{2}$  MS [57] liquid medium at a photoperiod (12/12 h, light:dark). The content was elevated by the addition of ethephon (200 mg/L) at the 10th day after the inoculation in the fresh medium. The more accurate measurement of anthocyanins content, by UHPLC-PDA, revealed that the black carrot HRs derived from roots of the inbred line "43" accumulated over 4 mg/g DW of the pigments and upon the treatment with ethephon up to 8 mg/g DW. Eight anthocyanins, derivatives of cyanidin and pelargonidin, were identified and quantified using UHPLC-PDA-TOF MS [56]. Supplementation of the nutrient medium with 60 g/L sucrose, increased dry mass and anthocyanin accumulation in the culture. Elicitation with 200 µM of H<sub>2</sub>O<sub>2</sub>, on 12th day of culture resulted in about 20% higher anthocyanin content [58].

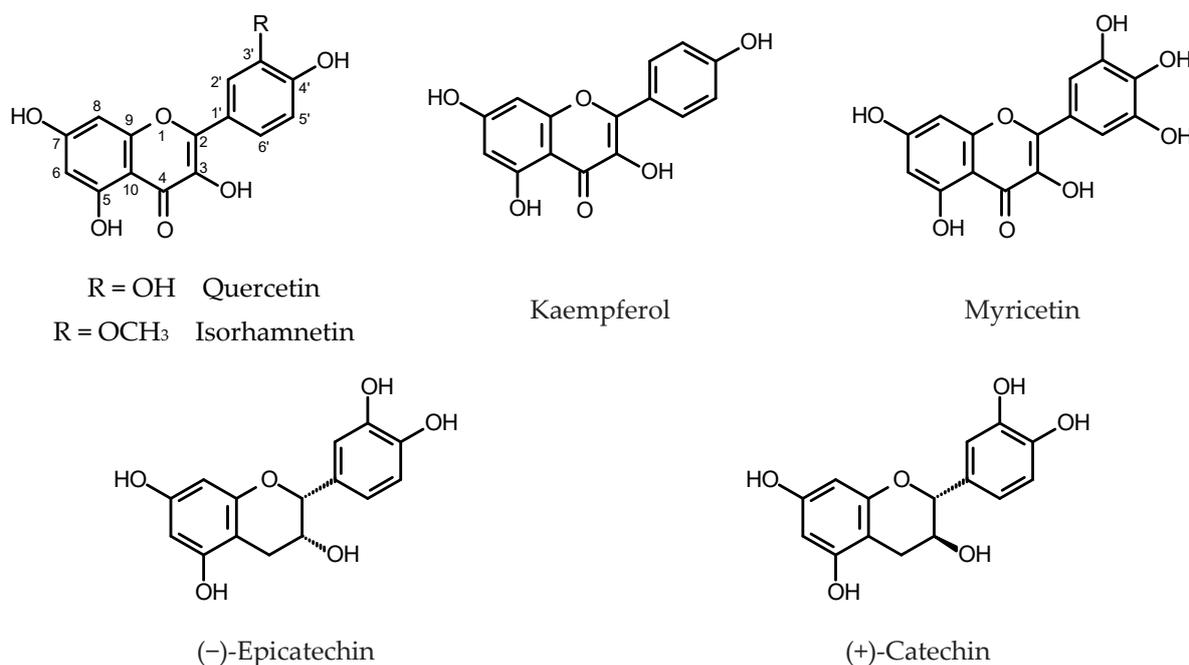
Two gene-encoding transcription factors: a bHLH gene *AmDelila* and an R2R3-MYB gene *AmRosea1* were concomitantly introduced into the HRs of common snapdragon and induced expression of anthocyanin-related genes in the roots. Further research demonstrated that *AmRosea1* alone was enough to start anthocyanin accumulation in the root tissues. The total anthocyanin content in the roots expressing *AmRosea1* reached 2 mg/g FW versus 0.3 mg/g FW in the control HRs [59].

Overexpression of PAP1 (production of anthocyanin pigment 1) gene in *Panax ginseng* C.A. Meyer (Araliaceae) HRs induced expression of phenylpropanoid and flavonoid biosynthetic pathway genes and led to 191- to 341-fold increase in anthocyanins production in comparison to the controls. The increase of anthocyanin accumulation was accompanied by the rise in the antioxidant and radical scavenging activity of *PAP1*-overexpressing roots and their improved anti-melanogenic activity [60]. Further investigation suggested improved antimicrobial and anti-elastase activities of *PAP1*-overexpressing ginseng HRs [61].

### 2.1.2. Flavonols and Flavanols

Except for the optimization of culture conditions (light, growth regulators, sucrose concentration, bacterial strain used to initiate the culture) and overexpression of genes from the *myb* gene family, elicitation with abiotic (UV-B irradiation) and biotic (yeast extract, chitosan, inactivated fungal preparations) elicitors was used to enhance production of flavonols and flavanols in the HRs. Plant hormones (methyl jasmonate, MeJa) and plant growth regulators (ethephon), in some studies classified as abiotic elicitors [16], were also applied to stimulate biosynthesis of the compounds.

HRs of buckwheat (*Fagopyrum esculentum* Moench) and Tartary buckwheat, except for anthocyanins and PAs, accumulated substantial amounts of flavonols (kaempferol, quercetin, rutin) and flavanols (catechin, epicatechin). Trotin et al. [62] estimated contents of (+)-catechin, (–)-epicatechin and (–)-epicatechin 3-*O*-gallate (for structures, see Figure 3) in a HR culture of buckwheat obtained by transformation with *R. rhizogenes* 15834. (–)-Epicatechin gallate was the major flavanol found in the roots. Its content reached 10 mg/g DW after 21 days of culture and was like that estimated in normal root culture. The HRs produced twice as much of (+)-catechin as normal roots (up to 8 mg/g DW) but the content of (–)-epicatechin in the transformed roots was significantly lower. It is worth noting, that after 21 days in culture dry weight of roots reached 720 mg DW per flask and 180 mg DW per flask for hairy and normal roots, respectively. The HRs of buckwheat accumulated higher yields of rutin (quercetin 3-*O*-rutinoside, 1.3 mg/g DW) than did untransformed roots (0.5 mg/g DW) [63]. Overexpression of *Arabidopsis* transcription factor *AtMYB12* in buckwheat HRs led to the increased expression levels of flavonoid biosynthetic enzymes and enhanced rutin production in comparison with the control HRs [64]. Roots of Tartary buckwheat cv. “Hokkai T8” and “Hokkai T10”, obtained by infection of hypocotyl explants with *R. rhizogenes* strain R1000 were cultivated in both light and dark conditions. The roots contained higher amounts of flavonols when exposed to light. Quercetin content reached its maximum (nearly 1 mg/g DW) 5–10 days after the transfer of roots to the fresh medium and then dropped, whereas the highest content of rutin was found after 15 days of culture (59 mg/g DW) [35]. Catechin and rutin contents in the HRs of Tartary buckwheat differed depending on the *R. rhizogenes* strain that was used to obtain the culture. The quercetin content remained unchanged irrespective of the bacterial strain used [53]. The ethephon treatment (0.5–2.0 mg/mL) significantly increased rutin accumulation in the roots [54]. A similar effect was achieved with overexpression of FtMYB6, an SG7 R2R3-MYB transcription factor induced by light [65]. Overexpression of FtUGT73BE5, an UDP-glycosyltransferase, in the HRs of Tartary buckwheat caused increase in rutin content in the biomass from 0.77 to 1.29-fold when compared with the control (up to 80 mg/g DW) [66]. Choi et al. [67] investigated expression of 11 genes of the phenylpropanoid biosynthetic pathway in seedling roots, adventitious roots and HRs of *F. tataricum*. Five of the examined genes were highly expressed in the studied HR clones. The HRs accumulated significantly higher amounts of (+)-catechin, (–)-epicatechin and rutin than those found in the untransformed axenic roots and roots of the seedlings.



**Figure 3.** Chemical structures of selected flavonols (upper row) and flavanols.

Tusevski and coworkers [68], by means of HPLC-PAD-ESI-MS<sup>n</sup>, identified and quantified numerous phenolic compounds produced by HRs of *Hypericum perforatum* L. (Hypericaceae) obtained by a transformation with *R. rhizogenes* A4. Flavonols (quercetin 6-C-glucoside, unidentified isorhamnetin-*O*-hexoside, rutin, kaempferol), flavanol (catechin) and two proanthocyanidin dimers were present in the dark grown roots in small quantities (<1 mg/g DW). Contents of epicatechin and one proanthocyanidin dimer exceeded 1 mg/g DW. The phenolic profile changed when the roots were exposed to light [69]. Selected lines of *H. perforatum* HRs, transformed with *R. rhizogenes* A4M70GUS, accumulated up to 4.7 mg/g DW catechin, negligible amounts of procyanidin dimers, and up to 5.9 mg/g DW of quercetin and kaempferol derivatives [70].

Overexpression of *LcMYB1* in lychee HRs led to the twofold increase in total flavonol content, in reference to the control roots [49]. The HRs of *Linum album* Kotschy ex Boiss. (Linaceae) transformed with *R. rhizogenes* LBA9402, upon elicitation with a cell wall preparation from *Piriformospora indica* (1%, *v/v*; 48–120 h treatment), demonstrated enhanced expression of genes-encoding several enzymes of phenylpropanoid biosynthetic pathway and enhanced flavonols and flavanols accumulation. The roots accumulated up to 0.1 mg/g DW catechin, kaempferol (<3.6 µg/mg DW) and myricetin (<2.4 µg/mg DW). The total flavonoid content did not exceed 0.15 mg/g DW [71–73]. According to Thiruvengadam et al. [74] HRs of *Polygonum multiflorum* Thunb. (Polygonaceae) transformed with *R. rhizogenes* strain KCTC 2703 produced rutin, myricetin, quercetin and kaempferol, although in low quantities (0.35, 0.40, 0.29 and 0.11 mg/g DW, respectively). In the HR cultures of *P. multiflorum* transformed with *R. rhizogenes* KCCM 11,879 quercetin was the most abundant flavonol (up to 4.59 mg/g DW). Elicitation with MeJa (50 µM) caused a 3.83-fold increase in quercetin content [75].

A series of papers on HR cultures of *Momordica charantia* L., *Momordica dioica* Roxb. ex Willd., *Cucumis anguria* L. (Cucurbitaceae), *Brassica rapa* L. ssp. *rapa* and *Brassica rapa* L. ssp. *pekinensis* (Brassicaceae) [76–81] described the quantification of flavonols by an UPLC method with UV detection, using a set of commercially available standards comprising: rutin, myricetin, quercetin, kaempferol and catechin. The estimated contents of flavonols were low (<0.8 mg/g DW), except for *M. charantia* HRs that produced over 1 mg/g DW of catechin.

HRs of *Isatis tinctoria* L. (Brassicaceae), obtained by inoculation with *R. rhizogenes* LBA9402 and cultivated in the optimum conditions, produced rutin (94 µg/g DW), quercetin

(53 µg/g DW), kaempferol (134 µg/g DW) and isorhamnetin (84 µg/g DW). Contents of the flavonols were higher than those found in the roots of two-year-old *I. tinctoria* field-grown plants [82]. Elicitation with 150 mg/L of chitosan, for 36 h, led to a significantly higher flavonol accumulation (rutin—812 µg/g DW; quercetin—733 µg/g DW; kaempferol—285 µg/g DW; isorhamnetin—618 µg/g DW) [83]. A co-culture of the HRs with an immobilized *Aspergillus niger* caused oxidative stress in the plant tissue and triggered up-regulation of the flavonoid biosynthetic pathway. As a result, under the optimum conditions (ca. 10<sup>4</sup> spores/mL, 30 °C, initial pH value 7.0, 72 h treatment), higher contents of all flavonols but rutin were detected in the examined *I. tinctoria* HRs (quercetin—up to 1500 µg/g DW; isorhamnetin—up to 700 µg/g DW; kaempferol—up to 1000 µg/g DW) [84]. The roots exposed to UV-B radiation (108 kJ/m<sup>2</sup>) demonstrated substantial up-regulation of the chalcone synthase gene and increased accumulation of all monitored flavonols (rutin—up to 1500 µg/g DW; quercetin—up to 1700 µg/g DW; isorhamnetin—up to 2000 µg/g DW; kaempferol—up to 2200 µg/g DW) [85].

Ghimire et al. [86] found that elicitation either with yeast extract (100 mg/L) or with MeJa (100 µM) significantly increased flavonol accumulation in the HRs of *Aster scaber* (Asteraceae, full botanical name not provided). Upon elicitation with MeJa, biomass of the roots contained: 2.07 mg/g DW of myricetin, 0.69 mg/g DW of quercetin, 0.26 mg/g DW of kaempferol and 0.19 mg/g DW of rutin. The HRs of *Ligularia fischeri* Turcz. f. *spiciformis* (Nakai) (Asteraceae), obtained by inoculation with *R. rhizogenes* KCTC 2703, were found to produce myricetin (2.38 mg/g DW), quercetin (0.51 mg/g DW), kaempferol (0.12 mg/g DW) and rutin (0.14 mg/g DW) [87].

HRs may be a source of new, previously unknown natural products. Two unusual, biologically active, derivatives of kaempferol and quercetin (4'-O-methylkaempferol-3-O-[(4''→13''')-2''',6''',10''',14'''-tetramethylhexadecan-13'''-olyl]-β-D-glucopyranoside and 3',4'-di-O-methylquercetin-7-O-[(4''→13''')-2''',6''',10''',14'''-tetramethylhexadec-13'''-ol-14'''-enyl]-β-D-glucopyranoside) were isolated from the HRs of *Catharanthus roseus* (L.) G. Don (Apocynaceae) [88].

### 2.1.3. Flavones and Flavanones

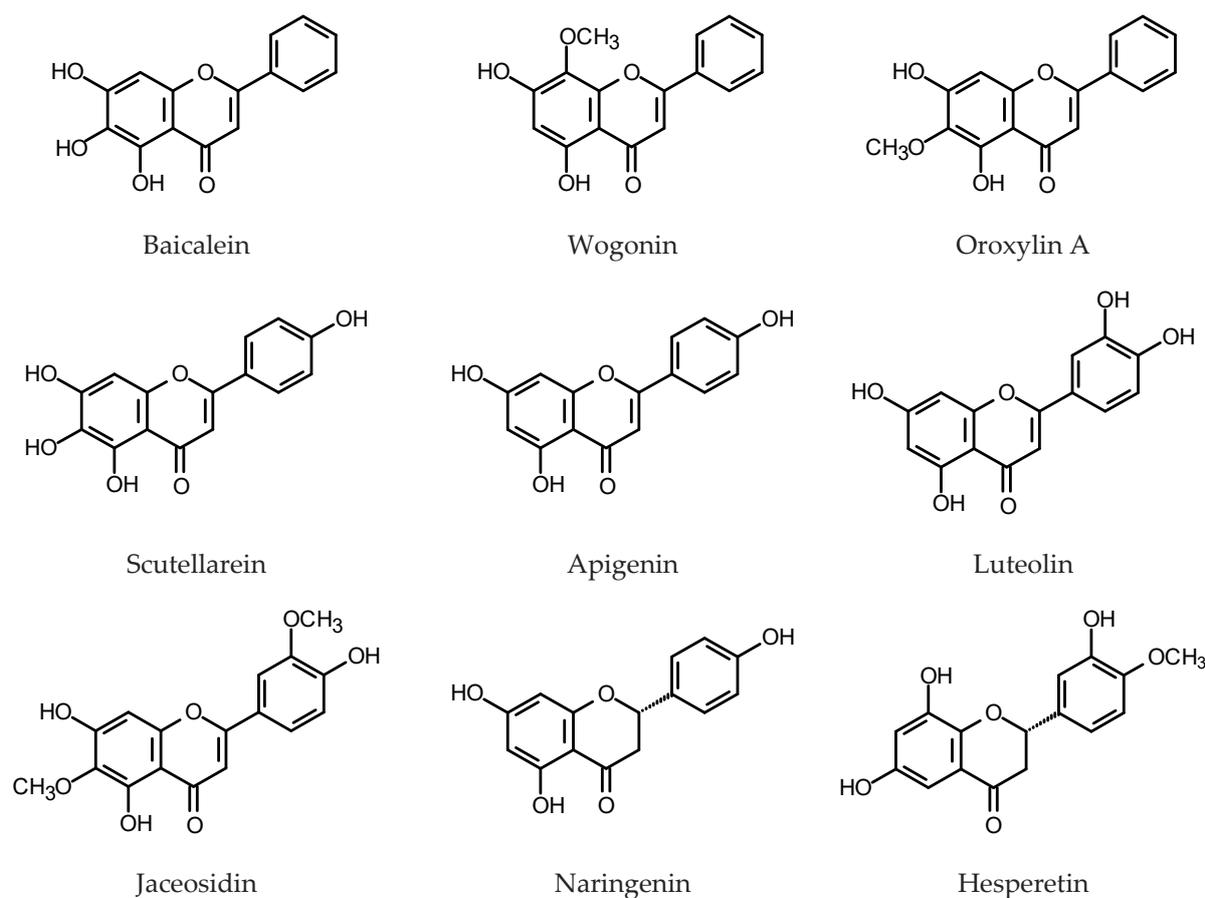
The procedures employed to enhance the production of flavones and flavanones in the HRs were similar to those described earlier for the other groups of flavonoids and include overexpression of the genes-encoding biosynthetic enzymes (*CHI*, *PAL*, *C4H* and *4CL*), elicitation (UV-B irradiation, MeJa, yeast extract, bacterial lysates, β-cyclodextrin, iron oxide nanoparticles) and heterologous expression of transcription factors.

One of the most spectacular examples of a plant that accumulate substantial amounts of flavones is *Scutellaria baicalensis* Georgi (Lamiaceae). Roots of the plant contain up to 27% DW of flavones, mainly glycosidic derivatives of baicalein, wogonin and oroxylin A [89] (for structures, see Figure 4). *S. baicalensis* is valued for its medicinal properties [90,91], including the antioxidant activity of its major constituents [92]. First attempts to develop a HR culture system to produce pharmacologically active flavones of *S. baicalensis* were made in the 1990s [93–95]. Those studies, apart from the identification of over twenty known phenolic compounds synthesized by the roots, including flavones and phenylethanoids, led to the isolation and identification of two formerly unknown flavones. However, the yields of biologically active flavones from the cultures were unsatisfactory (up to 1.2% DW of baicalein 7-O-glucuronide). The HRs that emerged as a result of *S. baicalensis* transformation with *R. rhizogenes* LBA 9402, in optimum culture conditions produced 6.9% DW of total baicalein glycosides and 1.5% DW of total wogonin glycosides, measured by HPLC after the acidic hydrolysis of the native compounds [96]. Kuzovkina et al. [97,98], by a transformation of *S. baicalensis* with *R. rhizogenes* A4 obtained HRs that produced 5% DW of flavonoids (spectrophotometrically measured). The content increased 1.8-fold after the 72 h of treatment with 100 µM of MeJa. More accurate quantification by HPLC revealed 1.27% DW of baicalin (baicalein 7-O-glucuronide), 1.07% DW of wogonoside (wogonin 7-O-glucuronide) and 0.38% DW of their aglycones in the analyzed roots. Maximum yields

of baicalin and wogonoside after MeJa treatment reached 10 mg/g DW and 24 mg/g DW, respectively [98,99]. Hirotani et al. [100] found that accumulation of mRNA for an enzyme UDP-glucose:baicalein 7-*O*-glucosyltransferase (UBGT) in the HRs of *S. baicalensis* was induced in response to wounding and salicylic acid treatment. Overexpression of chalcone isomerase gene (*SbCHI*) in the HRs of Baikal skullcap resulted in the increased accumulation of baicalin (42–60 mg/g DW), baicalein (8.9–12.1 mg/g DW) and wogonin (2.2–5.4 mg/g DW) in reference to GUS-control HR line. As expected, *SbCHI*-silenced HRs produced less flavones than the controls [101]. The HRs overexpressing phenylalanine ammonia-lyase genes (*SbPAL1*, *SbPAL2* or *SbPAL3*) produced significantly higher amounts of flavone aglycones (baicalein: 11–29 mg/g DW; wogonin 2.5–6.7 mg/g DW) than the GUS-control HRs and wild type field-grown roots. Baicalin content in PAL-overexpressing roots (57–136 mg/g DW) was slightly lower than that in wild type field-grown roots (146 mg/g *SbCYP82D2*DW) but much higher than that found in the GUS-control HRs [102]. Also, overexpression of cinnamate 4-hydroxylase (C4H) and 4-coumaroyl CoA ligase (4CL) increased flavone production in *S. baicalensis* HRs [103]. Heterologous expression of the transcription factor Lc from *Zea mays* in the HRs of Baikal skullcap led to the enhanced accumulation of baicalin, baicalein and wogonin in the roots, but higher yields were achieved when the *Arabidopsis* PAP1 transcription factor was overexpressed in the same experimental system (up to 102 mg/g DW of baicalin) [104]. Some improvement in yields of flavones from the cultures was achieved as well by an optimization of the nutrient medium composition [96,105,106]. Aglycones:glucuronides ratios in the cultures may be at least partially controlled by the activities of the endogenous  $\beta$ -glucuronidase of Baikal skullcap (baicalinase, sGUS) and baicalein 7-*O*-glucuronosyltransferase (UBGAT) [107,108]. Hydroxylation of chrysin is the step in the biosynthesis of baicalein and wogonin that requires the respective CYP450 enzymes: flavone 6-hydroxylase (F6H) and flavone 8-hydroxylase (F8H). Two enzymes: *SbCYP82D1.1* (F6H) and *SbCYP82D2* (F8H) from *S. baicalensis* were described and the role of *SbCYP82D2* in the wogonin biosynthesis was confirmed by its silencing in the HRs of the plant [109]. Baikal skullcap collected in the Dauria region (shared by Mongolia, Russian Federation and China) demonstrated high contents of polymethoxylated flavones. The HRs obtained from the plants retained their metabolic profile [110].

Several other species of the *Scutellaria* genus like *S. lateriflora* L., *S. andrachnoides* Vved., *S. bornmuelleri* Hausskn. ex Bornm. *S. przewalskii* Juz. and *S. pycnoclada* Juz. Bunge were also investigated in respect of their prospective use as a source of biologically active flavones [111–117]. American skullcap (*S. lateriflora*) is the second most popular species of *Scutellaria* used for medicinal purposes. Wilczańska-Barska et al. [111] obtained HRs of *S. lateriflora* by a transformation with *R. rhizogenes* strain A4. The roots produced a phenylethanoid glycoside—acteoside (19 mg/g DW) and flavones (scutellarin 0.6 mg/g; baicalin 14.5 mg/g; wogonoside 12.0 mg/g; wogonin 11.5 mg/g; chrysin 0.1 mg/g DW). Except for the wogonin, flavone contents were lower than those in *S. baicalensis* HRs [111,112]. The root culture of *S. lateriflora* upon elicitation with yeast extract (YE, 200 mg/L) demonstrated increased biomass accumulation and enhanced acteoside production. The optimum for flavones accumulation was 50 mg/L YE (flavone total content ca. 50 mg/g DW). Bacterial lysates applied as elicitors were less effective [111]. The HRs of the American skullcap transformed with *R. rhizogenes* ATCC 15834 were cultivated under continuous illumination or in the dark and elicited with  $\beta$ -cyclodextrin and MeJa [113]. Maximum contents of baicalein and wogonin (5.4 mg/g DW and 0.71 mg/g DW, respectively) were found in the cultures maintained in the dark, treated with 15 mM of  $\beta$ -cyclodextrin. Maximum contents of flavone glycosides—scutellarin (scutellarein 7-*O*-glucuronide) and wogonoside (ca. 0.5 mg/g DW each) were accumulated in the roots grown in the light using the same treatment.  $\beta$ -Cyclodextrin (15 mM) turned out to be more effective than MeJa (100  $\mu$ M) as an inducer of flavone accumulation, after 24 h treatment. Better yields of the flavones (baicalin—22.5 mg/g DW; wogonin—5.4 mg/g DW) were achieved by Tuan et al. [114] after 72–96 h treatment with 100  $\mu$ M MeJa. The HRs used in the experiment were obtained by

the transformation with *R. rhizogenes* strain R1000. Stepanova et al. [115] investigated HRs of *S. baicalensis*, *S. lateriflora*, *S. przewalskii* and *S. pycnoclada* derived from the transformation of the plant material with wild-type *R. rhizogenes* A4. The roots of *S. baicalensis* and *S. przewalskii* showed the best growth indices when cultured in a liquid nutrient medium. Baicalin was the major flavone accumulated in all examined cultures. The HRs of *S. przewalskii* were characterized by a high content of flavones (ca. 33 mg/g DW in total) compared to the HRs of *S. baicalensis* (ca. 17 mg/g DW) and HRs of the two remaining species (about 5–13 mg/g DW). The HRs of *S. andrachnoides* and *S. bornmuelleri* produced low amounts of flavones [116,117]. The HRs of *S. viscidula* Bunge, *S. orientalis* L. and *S. araxensis* Grossh. have not been examined in respect of their flavone content yet [118,119].



**Figure 4.** Chemical structures of selected flavones and flavanones.

*Dracocephalum kotschy* Boiss (Lamiaceae) is a rare plant of medicinal properties that synthesizes polymethoxylated flavones and flavonols of antioxidative and anti-inflammatory activity [120]. The HRs of the plant induced by inoculation with *R. rhizogenes* LBA 9402 produced: apigenin, cirsimaritin, isokaempferid, penduletin, xanthomicrol and calycopterin, but the contents of individual compounds varied widely depending on the root clone. The HRs were a better source of flavonoids than the roots of the intact plant. The leaves of *D. kotschy*, however, contained nearly eight times more flavonoids (1.7 mg/g DW) than the most productive HR clone [121]. A HR culture of *D. kotschy* transformed with *R. rhizogenes* ATCC 15834 accumulated 0.19 mg/g FW of apigenin. Upon elicitation with iron oxide nanoparticles (75 mg/L, 24 h) the apigenin content increased up to 0.37 mg/g FW [122].

The genus *Saussurea* comprises ca. 400 plant species. Some of the plants from the genus have been traditionally used as medicines [123,124]. The HRs of *S. medusa* Maxim, transformed with *R. rhizogenes* strain R1601 and maintained in the liquid N6 medium [125], produced ca. 6.1 mg/g DW jaceosidin [126] in a 24–28-day culture. Overexpression of

chalcone isomerase gene from *S. medusa* in the HRs of *S. involucrata* Kar. et Kir. ex Maxim., led to the increased accumulation of apigenin (ca. 2.6 mg/g DW) and a higher content of total flavonoids [127].

*Erigeron breviscapus* (Vaniot) Hand.-Mazz. (Asteraceae) synthesizes pharmacologically active flavones apigenin and scutellarein together with their glycosides. The major flavone accumulated by the plant is scutellarein. The HRs of *E. breviscapus* were obtained by transformation with *R. tumefaciens* strain C58C1 harboring pRiA4 plasmid or with the same bacterial strain modified by introducing a construct containing *EbCHI* gene encoding chalcone isomerase. The roots overexpressing *EbCHI* produced 2.2 mg/g DW of scutellarin, whereas the roots of the intact plant contained 0.21 mg/g DW of the compound (whole plant 2.6 mg/g DW). Elicitation with MeJa induced expression of several genes engaged in the flavonoid biosynthesis, including the genes-encoding chalcone synthase (CHS) and chalcone isomerase (CHI). The HRs of *E. breviscapus* treated with MeJa transiently accumulated up to 4.7 mg/g scutellarin [128].

HRs of *Catharanthus roseus* (L.) G. Don (Apocynaceae) harboring the gene encoding tryptophan feedback-resistant anthranilate synthase holoenzyme (*AS $\alpha\beta$* ) produced naringin (naringenin 7-*O*-neohesperidoside) as the major phenolic metabolite (0.65–0.86 mg/g DW). Hesperidin (hesperetin 7-*O*-rutinoside) was accumulated in the roots in smaller amounts (0.11–0.23 mg/g DW) [129].

Silencing of two flavone synthase II (FNSII) genes: *GmFNSII 1* and *GmFNSII -2*, in the HRs of soybean cultivar Hefeng 47, substantially decreased apigenin accumulation and reduced tolerance to salt stress but increased isoflavonoid production in the investigated root clones [130]. Glucose, mannitol, MeJa and NaCl significantly increased expression of *GmFNSII 1* and *GmFNSII -2* in the HRs of soybean. The motifs responsive to MeJa and glucose were found in the *GmFNSII 1* and *GmFNSII -2* promoter sequences. It was suggested that the oxidative damage induced by the salt stress may be mitigated by the flavone accumulation [131].

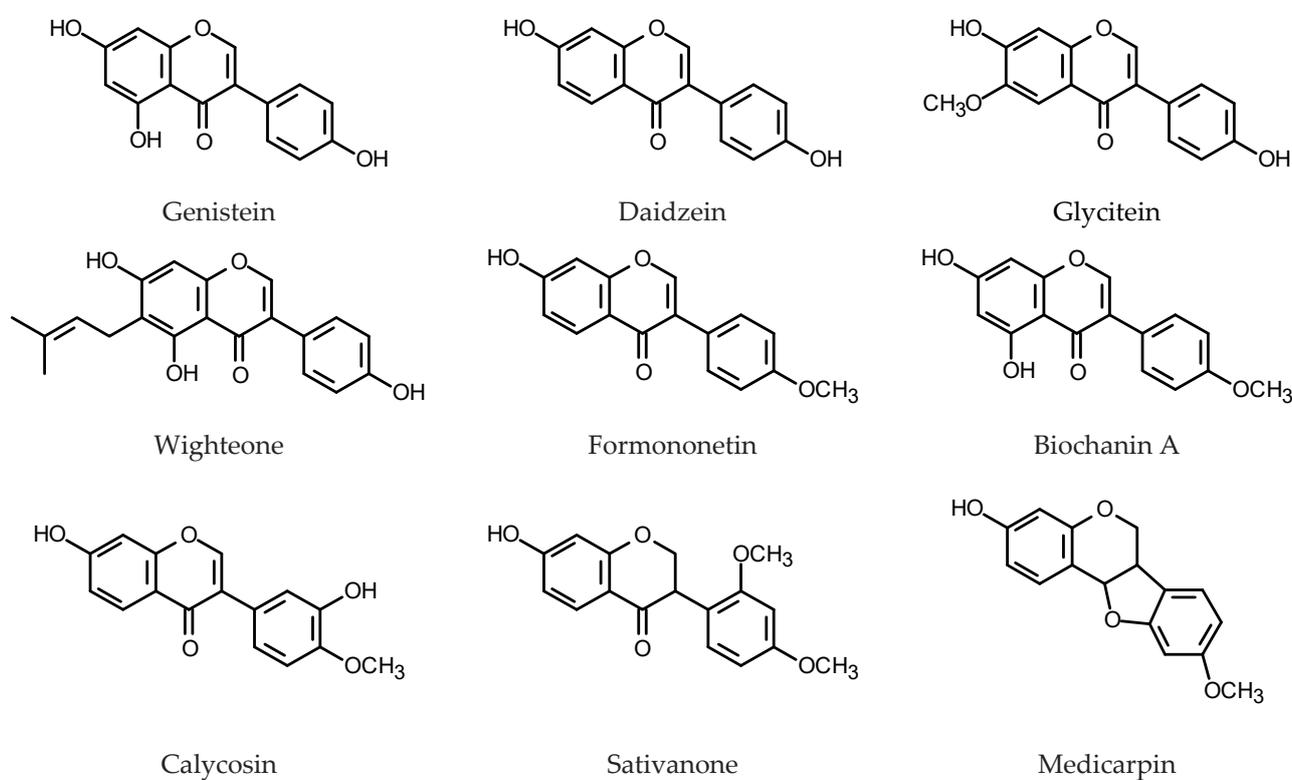
Components of the flavonoid fraction were analyzed in roots, HRs and a cell suspension culture derived from roots of *M. truncatula* [132]. *M. truncatula* HRs were obtained by inoculation with *R. rhizogenes* strain Arqua1. The HRs contained three luteolin glycosides, chrysoeriol and two flavanones: naringenin and liquiritigenin. Extracts from roots of the intact plant analyzed by the same method revealed the presence of seven flavone glycosides, derivatives of luteolin and chrysoeriol. Flavonoid aglycones, as well as flavanones, were not detected in the roots of the plant. Five flavones: orientin (luteolin 8-*C*-glucoside), vitexin (apigenin 8-*C*-glucoside), isovitexin (apigenin 6-*C*-glucoside), luteolin and apigenin were quantified in seeds, leaves, roots, calli and HRs of *Cajanus cajan* (L.) Millsp. (Fabaceae). The contents of orientin, vitexin and isovitexin in the HRs were higher than those in the roots of the intact plant but much lower than those detected in the plant leaves. Apigenin and luteolin contents in the roots of pigeon pea plants were significantly higher than those in the HRs [133]. An effect of UV-B irradiation on accumulation of phenolic constituents in *C. cajan* HRs was investigated by Gai et al. [134]. Contents of orientin, vitexin, isovitexin, luteolin and apigenin increased after 2–8 h UV-B treatment and were 1.27–4.44-fold higher than those in the control roots. However, the yields of the individual compounds were low. The content of the major flavone, luteolin, after 2 h of UV-B irradiation reached 19  $\mu\text{g/g}$  DW.

#### 2.1.4. Isoflavonoids

Isoflavonoids, like the remaining flavonoids, are regarded as dietary antioxidants, i.e., compounds that may protect against oxidative stress linked to inflammation and the risk of macromolecule damage by free radicals. The group of compounds includes isoflavones, isoflavanones, isoflavans, rotenoids and pterocarpanes. Although they are reported from many plant families, isoflavonoids are particularly abundant in Leguminosae plants. The most known dietary source of the compounds are the yellow-skin seeds of *Glycine max* [135,136]. While the isoflavones are normally present in relatively low amounts

in mature soybean tissues, several attempts were made to develop soybeans that accumulate much higher contents of isoflavones than those found in the wild-type seed.

To study the role of isoflavonoids in the plant resistance to fungal infection, HRs were initiated from two soybean genotypes with different susceptibility to the SDS disease (sudden death syndrome) caused by the soil-borne fungal pathogen, *Fusarium solani* f. sp. *glycines*. Daidzein derivatives predominated in the isoflavone fraction extracted both from the soybean hairy roots and intact soybean. The principal isoflavones (genistin, daidzin, their malonyl conjugates and aglycones) and isoflavonoid phytoalexins (coumestrol, coumestrol conjugates and glyceollin) (for structures, see Figure 5) were determined in extracts from the *Fusarium*-inoculated and non-inoculated hairy roots. Inoculation with *F. solani* negatively affected accumulation of all monitored isoflavonoids except for glyceollin. The compound demonstrated better antifungal activity against the investigated *Fusarium* species than the remaining isoflavonoids used in the assay (daidzin, daidzein, genistin, genistein, glycitin, glycitein) [137].



**Figure 5.** Chemical structures of selected isoflavonoids.

Simultaneous silencing of flavone synthase II genes (*GmFNSII-1* and *GmFNSII-2*) in the HRs of soybean led to the reduction or cessation of apigenin biosynthesis. At the same time, increased accumulation of genistein was observed [130]. Synchronous silencing of *FNSII* and flavanone-3-hydroxylase (*F3H*) genes caused an increase of daidzein content in soybean HRs up to 1.55 mg/g DW (0.9 mg/g DW in control roots) [138]. Overexpression of *GmMYB100*, a gene encoding a R2R3 MYB transcription factor, in *G. max* HRs resulted in diminished expression of the genes-encoding enzymes engaged in flavonoid biosynthesis and diminished isoflavone content. Silencing of the *GmMYB100* did not affect expression of the genes for the biosynthetic enzymes (*GmCHS7*, *GmCHS8*, *GmCHI*, *GmIFS*, *GmF3H*) but significantly increased isoflavonoid content [139]. Soybean cotyledon HRs harboring *GmIFS1* showed better tolerance to salt stress and increased isoflavone content under salt stress conditions [140]. *GmMYB58* and *GmMYB205* are seed-specific flavonoid biosynthesis activators. Their overexpression in soybean HRs increased transcription of: *GmCHS8*, *GmCHI*, *GmIFS2*, *GmFLS1*, *UGT73F2* (isoflavone UDP-glucosyltransferase gene)

and *IF7Mat* (isoflavone 7-*O*-glucoside-6''-*O*-malonyltransferase gene). An unidentified daidzein derivative, which dominated the isoflavonoid profile of the control roots (1 mg/g DW), was excessively produced by the transgenic HRs. The daidzein derivative content reached 3.2 mg/g DW in *GmMYB58*-overexpressing HRs and 6.7 mg/g DW in the roots overexpressing *GmMYB205* [141]. Soybean isoflavonoids are mainly glycoconjugates and their biosynthesis is catalyzed by different UDP-glycosyltransferases (UGT). Six genes for GmUGTs were overexpressed in soybean HRs. In the transgenic root lines overexpressing UGT72Z3, UGT73C20 and UGT88E19, the total content of isoflavonoids increased 1.1- to 1.6-fold in reference to the control and reached over 7 mg/g DW [142]. Increased resistance to *Phytophthora sojae* infection was observed in *G. max* HRs overexpressing *GmCHI1A* (chalcone isomerase gene from the soybean cv. Nannong 10-1). This observation agreed with experimentally proven induction of *GmCHI1A* expression and enhancement of daidzein accumulation by *P. sojae* [143]. Fungal infection in soybean seedlings enhance also glycitein production. The last step in the biosynthesis of this compound is methylation catalyzed by isoflavone *O*-methyltransferase (IOMT). *IOMT*, when co-expressed with the flavonoid 6-hydroxylase gene (*F6H*, normally not expressed in the HRs) increased the content of glycitein-related metabolites by ca. 100% compared to the control [144]. The HRs overexpressing *GmMaT2* and *GmMaT4*, genes-encoding malonyl-CoA:flavonoid acyltransferases, produced more malonyldaidzin, malonylgenistin and malonylglucitin than the control. The major isoflavone, malonyldaidzin, content reached ca. 6.3 mg/g FW in *GmMaT2* overexpressing HRs. *GmMaT2* but not *GmMaT4* knockdown resulted in the reduced accumulation of malonylated isoflavonoid glycosides and reduced nodule numbers. *GmMaT2* is also upregulated by the rhizobial infection what implicates its participation in the nodulation process and malonylated isoflavone secretion into the rhizosphere [145]. An attempt was made to enhance isoflavonoid production in soybean HRs by elicitation with MeJa (100 µM), salicylic acid (SA, 200 µM), sonication and vacuum infiltration. Treatment with MeJa (72 h) resulted in 10.67-fold higher total isoflavonoid content (53.16 mg/g DW) than that in the untreated control. Daidzin content reached ca. 33.9 mg/g DW. Elicitation with SA, after 96 h, caused 5.78- and 65-fold increase in total isoflavonoid and genistein contents, respectively. Maximum isoflavonoid production, 75.26 mg/g DW, was achieved with 2 min sonication and subsequent 2 min vacuum infiltration of HRs on 30th day of the culture [146].

Pigeon pea HRs produced genistin (0.11 mg/g DW) and genistein (0.04 mg/g DW) [133]. Their accumulation in the HRs was enhanced by an exposition to UV-B radiation [134]. Leaves of *Lotus japonicus* cv. Miyakojima line MG-20 seedlings supplied with genistein produced isoprenylated isoflavonoid, wighteone. This observation led to the identification of novel prenyltransferase gene *LjG6DT*. The enzyme encoded by the gene worked only with genistein as a substrate and its expression was induced by the reduced glutathione (GSH), MeJa and SA. Overexpression of *LjG6DT* in *L. japonicus* HRs, together with GSH and genistein supplementation, induced enhanced wighteone production (0.18–0.24 µg/g DW) [147]. Twenty isoflavonoids, derivatives of 2'-hydroxyformononetin, afrormosin, biochanin A, daidzein, formononetin, genistein and irisolidone were detected in the HRs of *M. truncatula*. In the roots of the intact plant 32 isoflavonoids were detected and tentatively identified. In contrast to the plant roots HRs preferably accumulated isoflavonoid aglycones [132].

Red clover (*Trifolium pratense* L., Fabaceae) is a forage legume producing formononetin, biochanin A, daidzein and genistein as major isoflavone constituents [148]. Kumar et al. [149] described stable production of isoflavones by HRs of *T. pratense*. One of the fast-growing HRs clones, displayed a high accumulation of all four pharmaceutically important isoflavones: daidzein (8.56 mg/g DW), genistein (2.45 mg/g DW), formononetin (15.23 mg/g DW) and biochanin A (1.10 mg/g DW). The HRs of *T. pratense* var. URS-BRS Mesclador were obtained by inoculation with *R. rhizogenes* strain A4TC24 and 23 isoflavonoids were tentatively identified as their metabolites. The isoflavonoids were, putatively, derivatives of: daidzein, pseudobaptigenin, pratensin, biochanin, irilone, formononetin and 3',7-di-*O*-methylorobol. Isoflavonoid contents were monitored in the selected clones of roots up to five months of cul-

ture. One of the examined clones (6HR) accumulated 5.5 mg/g DW biochanin A, 20.8 mg/g DW formononetin and ca. 0.7 mg/g genistein after 90 days of culture. The HRs were elicited, 7 days after subculture, using either elevated sucrose content in the medium (60 g/L) or SA (10 and 30 mg/L). Sucrose (60 g/L) increased accumulation of isoflavonoids in the HRs. After 3.5 days of treatment, roots of clone 4HR contained 12.8 mg/g DW daidzein, 0.7 mg/g DW genistein, 13.4 mg/g DW formononetin and 1.1 mg/g DW biochanin A, whereas the roots of clone 8HR produced 1.9 mg/g DW daidzein, 0.4 mg/g DW genistein, 14.8 mg/g DW formononetin and 4.1 mg/g DW biochanin A. Untreated roots (controls) accumulated 1.2–2.9 mg/g DW daidzein, 0.1–1.3 mg/g DW genistein, 2.1–3.0 mg/g DW formononetin and 0.7–4.4 mg/g DW biochanin A. Elicitation with SA was less effective than the treatment with sucrose [150].

*Astragalus membranaceus* (Fisch.) Bunge (proper name *A. propinquus* Schischkin, Fabaceae) has a long history of use in traditional Chinese medicine. Roots of the plant are also commercially available in Europe and USA as dietary supplements and functional foodstuffs. Calycosin and calycosin 7-*O*-glucoside were the major isoflavone components of the HRs derived from the plant. The two compounds, together with ononin (formononetin 7-*O*-glucoside), formononetin and astraisoflavan 7-*O*-glucoside were quali-quantitatively determined by LC-MS/MS. Under optimum conditions, the total isoflavonoid content in 34-day old *A. membranaceus* HRs reached 0.24 mg/g DW. This yield was significantly higher compared to that of three-year-old field-grown roots (0.19 mg/g DW) [151,152]. The HRs exposed to UV-B (86.4 kJ/m<sup>2</sup>) synthesized more isoflavonoids (up to 0.53 mg/g DW). All investigated genes involved in isoflavonoid biosynthesis were also up-regulated following the UV-B irradiation. *PAL* and *C4H* were found to be the key genes implicated in the control of the process [153]. Elicitation of 34 days old HRs of *A. membranaceus* with MeJa (283 μM, 33.75 h treatment) raised the content of isoflavonoids to 2.25 mg/g DW and upregulated the genes related to isoflavone biosynthesis [154]. A co-culture of *A. membranaceus* HRs with immobilized *Aspergillus niger* enhanced accumulation of calycosin (0.73 mg/g DW) and formononetin (1.12 mg/g DW) in the roots [155].

*Psoralea corylifolia* L. (Babchi, Fabaceae) is a plant used in traditional medicine of China and India. The plant accumulates coumarins, flavonoids and terpenophenols, including pharmacologically active bakuchiol, which gained some popularity as a component of cosmetics [156,157]. The first study on isoflavonoid production in the HRs of *Psoralea* spp. was published in 1999 [158]. HR lines derived from seven distinct species were examined in respect of their daidzein content. The examination revealed that *P. leucantha* F. Muell. and *P. lachnostachys* F. Muell HRs were the best isoflavonoid producers. One, highly productive line (daidzein 10.2 mg/g DW, coumestrol 0.48 mg/g DW) was chosen for further investigation. The roots were elicited with 30 mg/L of chitosan at the end of the exponential phase of growth (21st day). After the addition of chitosan, the content of daidzein in biomass dropped to ca. 8 mg/g DW due to the release of the compound into the culture medium. Coumestrol and genistein contents in the roots increased from ca. 0.40 to 0.52 mg/g DW and from ca. 0.03 to 0.07 mg/g DW, respectively. Coumestrol was also liberated to the medium after the elicitation with chitosan. HR cultures of *P. corylifolia* L., transformed with *R. rhizogenes* ATCC 15834, similarly to the control roots, accumulated daidzein as a major isoflavonoid. The compound was accompanied with smaller amounts of formononetin glucoside, genistin and daidzein. Bakuchiol was absent from the cultures [159]. Clones of *P. corylifolia* HRs, obtained by inoculation with *R. rhizogenes* LBA 9402, in optimum conditions produced up to 20.6 mg/g DW daidzein and up to 3.7 mg/g DW genistein [160].

*Pueraria* spp. (Fabaceae) due to isoflavonoid content is traditionally used to relieve menopausal symptoms [161]. The HRs of *Pueraria phaseoloides* (Roxb.) Benth induced by *R. rhizogenes* ATCC 15834 contained ca. 1 mg/g DW puerarin (daidzein 8-*C*-glucoside) [162]. The culture was scaled up to a 2.5 L bioreactor and the culture medium was modified to achieve better yield. In optimum conditions, roots cultivated in the bioreactor produced ca. 5.6 mg/g DW of puerarin and the product was partially liberated to the nutrient medium [163]. A HR culture of *P. candollei* Wall. Ex Benth. was established using

*R. rhizogenes* ATCC 15834. The total isoflavonoid content in the roots reached 36.48 mg/g DW. Daidzin was the major product (29.9 mg/g DW), and the puerarin content was estimated as 3.4 mg/g DW [164]. Chitosan (50, 100 and 150 mg/L), MeJa (50, 100 and 200  $\mu$ M), SA (50, 100 and 200  $\mu$ M), YE (0.5, 1 and 2 mg/mL) and autoclaved *Rhizobium* culture (1, 2 and 3% *v/v*) were applied as elicitors to enhance isoflavonoid production in the HRs. Though all of the applied treatments increased isoflavonoid accumulation, the yeast extract addition (0.5 mg/mL, 3 days) proved to be the most effective (60.5 mg/g DW total isoflavonoids) [165]. *P. candollei* var. *myrifica* (Airy Shaw & Suvat.) Niyomdham, except for the isoflavonoids, synthesizes another phytoestrogen, deoxymiroestrol. The HRs of the plant produced up to 7 mg/g DW isoflavonoids (mainly daidzin and genistin) and up to 77  $\mu$ g/g DW of deoxymiroestrol. The contents were higher than those found in the roots of the intact plant (3.5 mg/g DW and 15  $\mu$ g/g DW, respectively). Chitosan, YE and MeJa were applied to study effects of elicitation on the active metabolite content. The HRs elicited with MeJa (200  $\mu$ M) accumulated puerarin (0.54 mg/g DW), daidzin (8.68 mg/g DW), genistin (5.27 mg/g DW), daidzein (0.45 mg/g DW), genistein (0.16 mg/g DW), kwakhurin (1.39 mg/g DW) and deoxymiroestrol (0.25 mg/g DW), after 6 days of the treatment. The remaining elicitors also induced productivity of the roots, though to a lesser extent [166]. *P. candollei* Grah. ex. Benth. var. *candollei* was genetically transformed using two different *R. rhizogenes* strains (ATCC 15834 and 43,057) to obtain several clones of HRs.

The effect of the inoculum size (1 and 2% *w/v*) and temperature (25 or 32 °C) on the growth of the obtained HRs and on their isoflavonoid content was investigated. The maximum flavonoid accumulation (31 mg/g DW with daidzein and puerarin as major compounds) was found in the cultures started with 1% inoculum and cultivated at 32 °C. Moreover, it was found that cultivation of the HRs at higher temperature reduced browning of the tissue [167]. Kim et al. [168] obtained HRs from two different lines of *P. lobata* (Willd.) Ohwi (kudzu), collected in two different regions of Korea. The roots accumulated more puerarin and daidzin than the respective callus cultures, but the contents of the compounds in the tubers of the intact plants were higher. *P. lobata* C-glycosyltransferase (PIUGT43) uses daidzein and genistein as substrates. The overexpression of PIUGT43 in soybean hairy roots that synthesize daidzein, but not puerarin, led to the production of puerarin in the transgenic roots [169].

The transformed root cultures of *Ononis spinosa* L. and *Ononis arvensis* L. (Fabaceae) were obtained by Nóra Gampe et al. [170]. The most abundant compounds in the HRs were medicarpin, sativanone and pseudobaptigenin glucosides (16.9–28.9 mg/g DW, 1.3–11.4 mg/g DW and 0.9–2.0 mg/g DW, respectively). Formononetin and onogenin derivatives were present in smaller amounts. Two new phenolic compounds were found in the HRs (bulatlactone 2''-O-glucoside and ononilactone). The total isoflavonoid production in the cultures was comparable to that in the wild-grown *O. arvensis* and approximately twofold higher than that in wild-grown *O. spinosa* samples.

#### 2.1.5. Miscellaneous Flavonoids

Though the HRs of *Glycyrrhiza glabra* L. (Fabaceae) do not produce glycyrrhizic acid, a sweet-tasting saponin responsible for the major pharmacological activity of the liquorice root (peptic ulcer healing), they synthesize vast array of biologically active polyphenols of unique structures [171,172] (see Figure 6). Two new compounds, licoagrochalcone and licoagrocarpin (chalcone- and pterocarpan-type compounds), were isolated from the HRs of licorice transformed with a *Rhizobacterium* strain harboring pRi15834 and pBI121 (GUS). Eight known compounds were also found in the HRs, including three chalcones, four prenylated flavanones and one prenylated flavanol [173]. Further investigation of the same plant material led to the isolation of licoagrodione, another new compound, together with five known flavonoids. The antimicrobial activities of the isolated compounds were assessed by the disc diffusion method. Glyinflanin K was the only compound that did not show activity against the bacteria and fungi used in the experiment [174]. The continuation of this research allowed for the isolation of unusual prenylated bioaurone, licoagrone, together

with five known flavonoids (kanzonol D, afrormosin, odoratin, phaseol, echinatin) [175]. Li et al. [176] isolated and described a new biflavonoid, licoagrodin and another four new flavonoids (licoagrochalcone B, licoagrochalcone C, licoagrochalcone D and licoagroaurone), as well as four known flavonoids. More polar fractions separated from the HRs gave licoagroside A, ononin, calycosin 7-*O*-glucoside, wistin, vicenin-2, afrormosin 7-*O*-(6''-malonylglucoside) and isoschaftoside. Licoagroisoflavon and licoagrosides C–F were isolated as new compounds from the HRs of *G. pallidiflora* Maxim. The compounds were accompanied by eleven known flavonoids [177,178]. In the HRs of *G. uralensis* Fisch. two-fold increase in total flavonoid content (up to 30 mg/g DW) was observed following the combined 48 h of treatment with YE (0.1%) and polyethylene glycol (PEG8000, 2%) [179]. Tween 80, added to 20-day-old HRs of *G. uralensis*, caused enhanced production of licochalcone A accompanied by elevated mRNA levels for PAL, 4CL (4-coumarate:coenzyme A ligase) and C4H. After 15 days of the treatment, the roots yielded over 3 mg of licochalcone A per flask, and the product was almost entirely liberated to the culture medium. The control HRs accumulated up to 0.35 mg/flask of the compound in their biomass [180]. The overexpression of *CHS* in *G. uralensis* HRs resulted in the increased production of chalcones and liquiritigenin in the transgenic roots. The number of *CHS* copies in the examined clones of HRs was determined as 9, 10, 11, 13 and 18. The root clone with nine copies of *CHS* accumulated the highest contents of flavonoids [181].

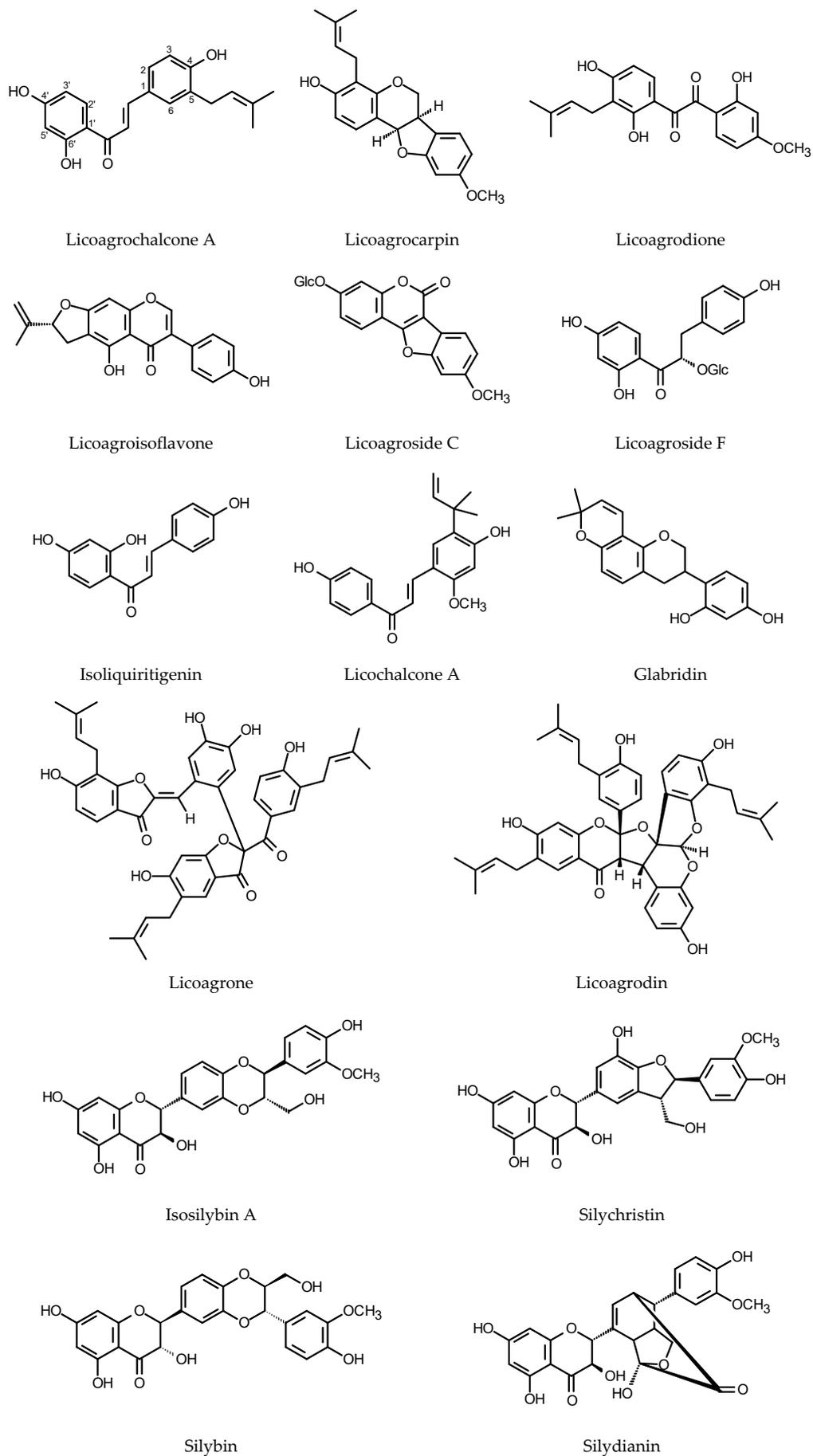
The *Genista tinctoria* L. HRs synthesized neither isoflavone, derivatives of daidzein and genistein, nor the derivatives of apigenin and luteolin characteristic of the intact plant. Instead, the roots accumulated isoliquiritigenin (daidzein precursor, 23 mg/g DW) absent from the roots of the intact plant. Abscisic acid (ABA, 37.8  $\mu$ M), added to the culture on the 42nd day of growth, induced the release of the product (80%) to the culture medium [182].

Alikaridis et al. [183] investigated roots obtained by a transformation of *Silybum marianum* Gaertner with *R. rhizogenes* ATCC 15834. The roots contained only minute amounts (0.4  $\mu$ g/g DW) of silychristin and silydianin. Silybins A and B were not detected, whereas the contents of isosilybins A and B were higher than those in the untransformed roots (isosilybin A—0.01 mg/g DW). The *S. marianum* HRs examined by Rahnama et al. [184] contained up to 0.14 mg/g DW silychristin. The other flavonolignans were present in smaller amounts.

## 2.2. Stilbenoids

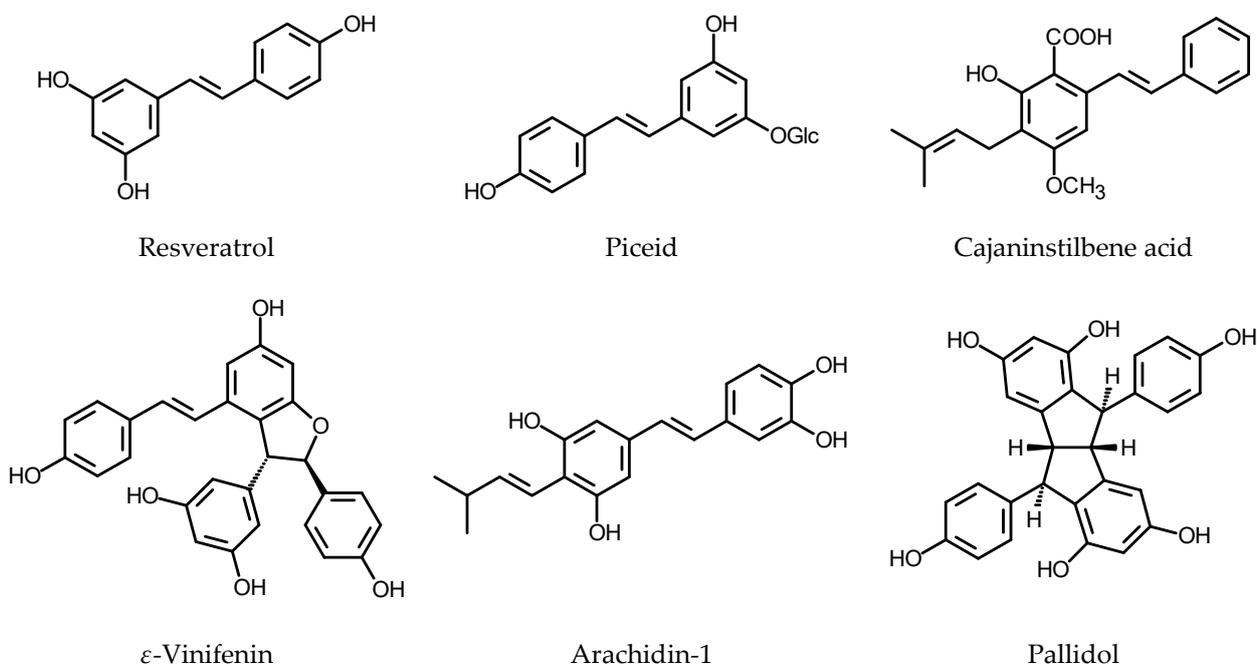
Stilbenoids are a class of polyphenolic plant constituents that have the general structural formula C<sub>6</sub>-C<sub>2</sub>-C<sub>6</sub> and share an initial part of their biosynthetic pathway with flavonoids. The compounds could be found in various plant species, including peanut (Fabaceae), grapevine (Vitaceae), berries (Ericaceae), pine (Pinaceae) and tomato (Solanaceae). Their main function is the protection of the host plant against pathogen infestation and oxidative stress generated by different environmental stimuli [185]. Stilbenoids, due to their biological activity, may find application as protective agents in cardiovascular disease, diabetes, neurodegeneration, obesity and other ailments [186].

The compounds were primarily detected in elicited HRs and the procedures used to enhance the productivity of the cultured roots included elicitation (MeJa, paraquat, H<sub>2</sub>O<sub>2</sub>) with concomitant use of permeabilizing agent (methyl- $\beta$ -cyclodextrin) to liberate the products into the culture medium. R2R3-MYB-type transcription factors are engaged in the up-regulation of stilbenoid biosynthesis, and this may find an application in genetic engineering.



**Figure 6.** Chemical structures of selected rare and atypical flavonoids found in HRs.

Medina-Boliwar et al. [187] found that HRs obtained by the transformation of *Arachis hypogaea* L. cv. Andru II (Fabaceae) with *R. rhizogenes* ATCC 15834, upon elicitation with sodium acetate, produced stilbenoids that are excreted into the culture medium. *Trans*-resveratrol and *trans*-pterostilbene (for structures, see Figure 7) were identified as metabolites present in the elicited cultures. *Trans*-resveratrol, *trans*-arachidin-1 and *trans*-arachidin-3 were isolated from the HRs of peanut cv. Hull elicited with 10.2 mM sodium acetate. Among the three compounds, arachidin-1 was the most active as an inhibitor of lipoprotein oxidation. In the applied assay, arachidin-1 was more active than butylhydroxytoluene (BHT). In contrast to resveratrol, arachidins at a dose of 55  $\mu$ M showed some cytotoxicity towards HeLa and RAW 264.7 cells [188]. The same culture was used to investigate the effects of medium optimization, and the age of the elicited roots on growth performance and root culture phenotype. The effect of elicitation on biosynthesis and liberation of stilbenoids into the culture medium was also studied. The HRs elicited on the ninth day of the culture produced *trans*-resveratrol (ca. 90  $\mu$ g/g DW), *trans*-arachidin-1 (ca. 33  $\mu$ g/g DW), and *trans*-arachidin-3 (ca. 37  $\mu$ g/g DW) when cultivated in modified MS instead of Gamborg's B5 nutrient medium. As the stilbenoids were secreted into the medium, the biomass of HRs contained only minute amounts of the compounds (ca. 1  $\mu$ g/g DW, ca. 0.75  $\mu$ g/g DW and ca. 0.71  $\mu$ g/g DW, respectively) [189]. In a search for the optimum elicitation method, hydrogen peroxide (10 mM), MeJa (100  $\mu$ M), methyl- $\beta$ -cyclodextrin (9 g/L), and combination of MeJa with methyl- $\beta$ -cyclodextrin were used as elicitors, instead of sodium acetate. The application of either H<sub>2</sub>O<sub>2</sub> or cyclodextrin caused secretion of piceatannol, another stilbenoid formerly not found in the HRs of *A. hypogaea* cv. Hull. The contents of stilbenoids in the culture medium were monitored until 96 h after elicitation. Individual compounds reached their maximum content in the analyzed medium at different time intervals. The best results were achieved with HRs treated with the combination of MeJa and cyclodextrin. Sixty hours after elicitation, the roots produced resveratrol (ca. 5.3 mg/g DW), piceatannol (ca. 0.3 mg/g DW), arachidin-1 (ca. 4 mg/g DW) and arachidin-3 (ca. 17.1 mg/g DW) [190].



**Figure 7.** Chemical structures of selected stilbenoids produced by HRs.

Yang et al. [191] found that the prenyl subunit of the prenylated stilbenoids like arachidins originated from the plastidic terpenoid pathway. Moreover, they purified and described membrane-bound stilbenoid-specific prenyltransferase from peanut HRs.

Elicitor-treated *A. hypogaea* HRs were used to discover genes-encoding stilbenoid prenyltransferases. Transcripts encoding five enzymes were identified and two of the enzymes were characterized, including AhR4DT-1 that catalyzes prenylation of resveratrol at C-4 to form arachidin-2 [192]. The prenylated stilbenoids of the peanut manifest an array of interesting biological activities (e.g., anti-inflammatory, antiviral, antioxidant and cytotoxic towards human cancer cells) and have better bioavailability than resveratrol [193–195]. To achieve higher yields of the prenylated stilbenoids from peanut HRs, the elicitation procedure was further optimized using 125  $\mu$ M MeJa (signaling molecule implicated in secondary metabolism regulation), 18 g/L cyclodextrin (permeabilizing agent that may trap the product and prevent feedback inhibition), 3 mM H<sub>2</sub>O<sub>2</sub> (inducer of piceatannol production in the HRs of the peanut) and 1 mM MgCl<sub>2</sub> (Mg<sup>2+</sup> as a co-factor of resveratrol prenyltransferases) [194]. The formerly applied treatment [190] allowed for ca. 56 mg/L of arachidin-1 and ca. 148 mg/L of arachidin-3 to be obtained. With the new procedure, the yields increased to 227.4 mg/L and 370.6 mg/L, respectively. Moreover, arachidin-2 (83.1 mg/L) and arachidin-5 (68.4 mg/L) were produced by the HRs. The new optimized elicitation method was used to induce stilbenoid production in *A. ipaensis* and *A. duranensis* HRs, obtained in the same way as HRs of the peanut cv. Hull. The HRs of the wild relatives of peanut, upon elicitation synthesized less arachidin-1 and arachidin-3, but more arachidin-2 and arachidin-5 [194].

The HRs of *A. hypogaea* cv. Tainan9 were obtained by inoculation with *R. rhizogenes* strain K599 (NCPB 2659). The production of stilbenoids in the roots was induced by the treatment of nine-day-old HRs with 100  $\mu$ M MeJa in combination with 6.87 mM methyl- $\beta$ -cyclodextrin. After 24 h of treatment, resveratrol (72.0  $\mu$ g/g DW), arachidin-1 (179.3  $\mu$ g resveratrol eq./g DW) and arachidin-3 (21  $\mu$ g resveratrol eq./g DW) were found in the culture medium. MeJa applied alone did not induce the liberation of stilbenoids into the medium. Liquid chromatography with tandem mass spectrometry (LC-MS/MS) was applied to analyze the metabolites present in the spent medium from the elicited cultures. Except for the phenolic acids (hydroxybenzoic acid and caffeic acid), the following stilbenoids were detected in the medium; *trans*-piceatannol, *trans*- and *cis*-resveratrol, three isomers of *trans*-arachidin-1, 4-isopentadienyl-3,5,3',4'-tetrahydroxystilbene, two isomers of *trans*-arachidin-3, two isomers of arahypin 7 and *trans*-3'-isopentadienyl-3,5,4'-trihydroxystilbene [196].

Another elicitation strategy was used with the HRs of *A. hypogaea* cv. Kalasin2 transformed with *R. rhizogenes* K599. Paraquat (PQ), a broad-spectrum herbicide that generates reactive oxygen species (ROS) in the plant tissue, was used in combination with MeJa and cyclodextrin. As was in the case of MeJa, PQ alone did not induce the liberation of stilbenoids from the roots. The most effective elicitation procedure that utilized 24 h of pretreatment with 500  $\mu$ M PQ followed by the induction with 100  $\mu$ M MeJa in combination with 6.87 mM methyl- $\beta$ -cyclodextrin caused a sharp increase in stilbenoid biosynthesis. The elicited roots produced *trans*-resveratrol (1.3 mg/g DW, after 120 h), *trans*-arachidin-1 (180.1 mg/g DW, after 192 h) and *trans*-arachidin-3 (444.2 mg/g DW, after 120 h). LC-MS/MS analysis of the extract from the spent nutrient medium revealed the presence of hydroxybenzoic acid, two *trans*-piceatannol isomers, *trans*- and *cis*-resveratrol, jasmonic acid, four *trans*-arachidin-1 isomers, *trans*-arachidin-3, two isomers of arahypin 7 and arahypins 5 and 6 [197].

Wongshaya et al. [198] investigated an effect of mechanical stress (cutting) and light on stilbenoid biosynthesis in *A. hypogaea* cv. Tainan9 HRs. Mechanical stress increased the total phenolic content and antioxidant capacity of the cultures in both light and dark conditions. Roots cultivated in the dark showed a better response to elicitation than those grown in the light. Maximum yields of stilbenoids that were achieved using the elicitation procedure described earlier [197] were as follows: *trans*-resveratrol > 4 mg/g DW (uncut roots, dark, 72 h), *trans*-arachidin-1 ca. 240 mg/g DW (uncut roots, dark, 72 h), *trans*-arachidin-3 ca. 250 mg/g DW (uncut roots, dark, 72 h). The same elicitation procedure was applied to the *A. hypogaea* cv. Tainan9 HRs cultivated in a 5 L capacity stirred tank bioreactor or in

500 mL Erlenmayer flasks. Two densities of inoculum were evaluated: 5 g/L and 20 g/L. The optimum production of arachidins was achieved using 20 g/L inoculum grown in a 500 mL flask. Upon elicitation (72 h treatment), the HRs yielded ca. 1700 mg/L of *trans*-arachidin-1 and ca. 4800 mg/L of *trans*-arachidin-3. The roots grown in the bioreactor showed maximum productivity with 20 g/L inoculum, after 48 h treatment [199].

Three cultivars of peanut, including “Tifrunner,” “Hull,” and “Georgia Green” were transformed with *R. rhizogenes* ATCC 15843. The obtained HRs were treated with 125 µM MeJa, 18 g/L cyclodextrin, 3 mM H<sub>2</sub>O<sub>2</sub> and 1 mM MgCl<sub>2</sub> for 168 h. The extracts from the spent culture media were qualitatively and quantitatively analyzed to assess the content of stilbenoids. The HRs of the “Tifrunner” cultivar were found to be rich in arachidins 1, 2, 5 and 6, whereas the HRs of “Hull” cultivar preferably synthesized resveratrol and arachidin-3. The yields of the analyzed stilbenoids ranged from ca. 5 mg/L (arachidin-5 in “Georgia Green”) to ca. 170 mg/L (arachidin-1 in “Tifrunner”) [200].

Muscadine grape (*Vitis rotundifolia* Michx.), a grapevine species native to the south-eastern part of North America, is a source of polyphenols that possess antioxidative and antimicrobial activity [201]. The HRs of *V. rotundifolia* (21 days old) were treated with 100 µM MeJa for 24 h. Extracts prepared from the root biomass and from the spent nutrient medium were subsequently analyzed in a search for stilbenoid metabolites. Resveratrol, piceid (resveratrol 3-*O*-glucoside) and ε-viniferin (resveratrol dehydrodimer) were present mainly in the biomass, whereas piceatannol was detected exclusively in the culture medium. Growth regulators IBA (0.05 mg/L) and BAP (0.05 mg/L) added to the culture medium did not affect yields of stilbenoids. Piceatannol and ε-viniferin were the best antioxidants among the analyzed HRs metabolites [202].

Nopo-Olazabal et al. [203] used the HRs of muscadine grape to study the biochemical and molecular regulation of stilbenoid biosynthesis upon treatment with either 100 µM MeJa or with 10 mM H<sub>2</sub>O<sub>2</sub>, over a 96 h period. Both treatments induced the transcription of *PAL*, *STS* and resveratrol synthase gene (*RS*) as soon as 3 h after elicitation. Resveratrol, piceid, and ε-viniferin were identified in the control and in the elicited HRs. Except for resveratrol, the stilbenoids were accumulated in the roots. After the elicitation with MeJa, piceid content in the roots increased from 164 µg/g DW (0 h) to 337 µg/g DW at the end of experiment. The increase, however, was similar to that in the control roots. In the culture medium, piceid was undetectable until 12 h of the treatment and reached 4.7 µg/g DW at the end of the experiment. Maximum accumulation of resveratrol took place at 12 h after elicitation (106 µg/g DW) and then the content of the compound in the roots declined. The resveratrol content in the medium reached maximum at 18 h after exposition to MeJa (48 µg/g DW) and subsequently decreased to 6.4 µg/g DW at the 96 h. ε-Viniferin accumulated in the MeJa elicited roots up to 379 µg/g DW at the end of the experiment. The content was over twofold higher than that in the control roots. The culture medium contained up to 11.8 µg/g DW ε-viniferin. Hydrogen peroxide treatment was less effective as an inductor of stilbenoid biosynthesis. The piceid and resveratrol contents in the H<sub>2</sub>O<sub>2</sub> treated roots increased to a lesser extent than it was observed for MeJa treatment. ε-Viniferin accumulated in the biomass (up to 434 µg/g DW, 96 h) and only ca. 5 µg/g DW of the compound was found in the nutrient medium. At 24 and 96 h after the treatment with H<sub>2</sub>O<sub>2</sub>, ca. 72.5 µg/g DW and 24.6 µg/g DW of resveratrol, respectively, were found in the nutrient medium. In the studied cultures, an increase in stilbenoid content correlated with an increased antioxidant capacity. The HRs of *V. vinifera* Pinot Noir cv. PN40024, obtained by inoculation with *R. rhizogenes* ATCC 15834 were cultivated in  $\frac{1}{2}$  SH (Schenk and Hildebrandt) medium supplemented with 2% sucrose. Piceid, resveratrol and two resveratrol dehydrodimers: ε-viniferin and δ-viniferin were constitutively present in the roots. δ-Viniferin was the major component of the stilbenoid fraction. The total stilbenoid content reached 217 µg/flask at the stationary phase of culture. The elicitation with MeJa (100 µM or 200 µM) on the 18th day of the culture (before the end of the exponential growth phase) increased an excretion rate of stilbenoids from 11% (control roots, after 10 days of the experiment) to 37% (200 µM MeJa, 10-day treatment). The control roots

contained ca. 2.42 mg/g DW of stilbenoids at the end of experiment. The HRs elicited with 100 and 200  $\mu$ M MeJa accumulated 6.98 mg/g DW and 4.34 mg/g DW of stilbenoids, respectively. The maximum stilbenoid content in the nutrient medium (19 mg/L) was found 10 days after the addition of MeJa (final concentration 200  $\mu$ M). The concomitant use of MeJa (100  $\mu$ M, added on the 18th day of culture) and methyl- $\beta$ -cyclodextrins (30, 50 and 70 mM, added to the fresh culture medium before autoclaving) resulted in the increased stilbenoid contents in both root biomass and the culture medium. Maximum total stilbenoid contents, measured four days after the addition of jasmonate were 6.4 mg/g DW in the roots (30 mM cyclodextrins) and 165 mg/L in the medium (50 mM cyclodextrins). The combined treatment with MeJa and cyclodextrins caused enhanced liberation of stilbenoids (80–90%) into the nutrient medium [204]. Tisserant et al. [205], based on  $^{13}$ C and  $^1$ H NMR data and results of LC-MS analysis, identified major metabolites of *V. vinifera* Pinot Noir HRs. The main polyphenols found in the culture were stilbenoids (*trans*-resveratrol, *trans*-piceatannol, pallidol,  $\epsilon$ -viniferin, scirpusin A, vitisin B and maackin) and flavanones (eriodictyol and naringenin).

Höll et al. [206] found that two R2R3-MYB-type transcription factors from grapevines regulated the stilbenoid biosynthetic pathway by the activation of the promoters of genes-encoding stilbene synthases (STS). One of the transcription factors, MYB15, expressed in the HRs of *V. vinifera* cv. Chardonnay increased levels of STS and PAL transcription. The content of *trans*-piceid in the MYB15 transgenic HR lines was fivefold higher than that in the control roots. Accumulation of the remaining stilbenoids was less affected.

*V. vinifera* subsp. *sylvestris*, accessions W2 and W16 and cv. Rasha, were transformed using three different strains of *R. rhizogenes*: ArA4, Ar318 and LBA 9402. Although all the three bacterial strains induced HRs from the grapevine explants, HR lines obtained with the strain ArA4 showed the most vigorous growth. The highest resveratrol contents, 2 to 31 times higher than that found in the control roots, were found in the HRs obtained from internodal explants. The HR cultures originated from cv. Rasha, accession W2 and accession W16 contained up to 95, 256 and 273  $\mu$ g/g DW of resveratrol, respectively. The elicitation with MeJa or sodium acetate caused approximately a twofold higher production of the stilbenoid and enhanced release of the product into the culture medium [207]

Ectopic expression of the genes-encoding enzymes and transcription factors engaged in stilbenoid biosynthesis and metabolism in the HRs of tobacco led to the production of biologically active stilbenoids. *V. vinifera* resveratrol-*O*-methyltransferase (VvROMT) and human cytochrome P450 hydroxylase 1B1 (HsCYP1B1) catalyze methylation of *trans*-resveratrol to *trans*-pterostilbene and hydroxylation of *trans*-resveratrol to *trans*-piceatannol. Tobacco leaf segments were inoculated with *R. rhizogenes* harboring the pRiA4 plasmid alone or pRiA4 with the binary plant expression vector pK7WG2\_CYP1B1 or pJCN52\_ROMT for the HsCYP1B1 or VvROMT genes, respectively. The transgenic HRs expressing CYP1B1 converted exogenously added *trans*-resveratrol into piceatannol, especially when treated with methyl- $\beta$ -cyclodextrin as the permeabilizing agent. Biotransformation rates were: 0.4% in wildtype HRs and 1.4–1.6% in the transgenic HRs lines. In contrast to the transgenic HRs, piceid was the major biotransformation product in the wild-type HRs. The yields of piceatannol from the transgenic HR cultures carrying CYP1B1 and wild type HRs were 1888  $\mu$ g/L and 819  $\mu$ g/L, respectively. Addition of cyclodextrin to the medium may enhance the productivity up to 7 mg/L. Transgenic HRs carrying VvROMT metabolized exogenously added *trans*-resveratrol into *trans*-pterostilbene, piceid and piceatannol. Pterostilbene was found both in the roots and the culture medium. Maximum accumulation of the compound (2.6  $\mu$ g/L) was observed 24 h after resveratrol feeding [208]. Tobacco HRs carrying VvSTS (gene for stilbene synthase from *V. vinifera*) and AtMYB12 (gene for the transcription factor from *Arabidopsis thaliana*) produced over 450  $\mu$ g/L stilbenoids and more flavonoids than the wild type HRs [8].

The *R. rhizogenes*-transformed roots of pigeon pea occurred to be an excellent source of cajaninstilbene acid, superior to seeds, leaves and roots of the parent plant [209]. The elicitation of HRs by 10 h of exposure to UV-B radiation resulted in over a twofold in-

creased accumulation of cajaninstilbene acid (up to 6.6 mg/g DW). The radiation induced oxidative stress and caused damage to the roots, which, in turn, enhanced the production of signal molecule, salicylic acid, implicated in the regulation of the secondary metabolism biosynthetic pathways [134].

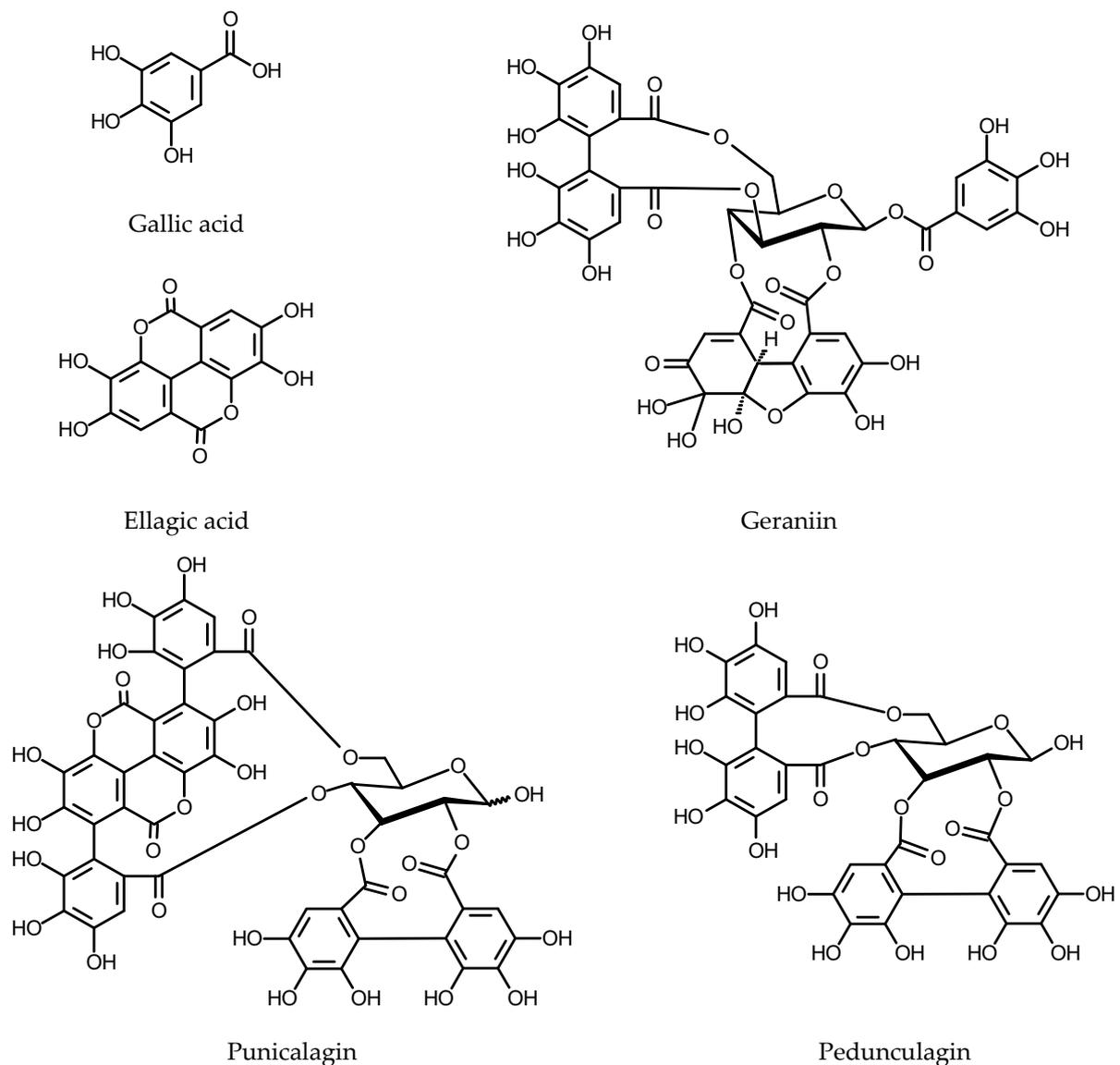
The HRs of pigeon pea induced by infection with *R. rhizogenes* strain K599 showed good biomass increase (17.3 g DW/L was obtained in 18 days of culture). The roots were elicited using the method described earlier for peanut HRs [194]. In 12-day-old cultures, the standard nutrient medium was replaced by the elicitation medium containing 125  $\mu$ M MeJA, 18 g/L methyl- $\beta$ -cyclodextrin, 3 mM H<sub>2</sub>O<sub>2</sub> and 1 mM MgCl<sub>2</sub> and the HRs were cultivated in the dark for another 7 days. After 144 h of treatment, the total content of cajaninstilbene acid in the culture reached over 8 mg/g DW. Almost 96% of the compound was secreted into the medium; thus, 7.7 mg/g DW of the product was extracted from the medium and the remaining 0.35 mg/g DW originated from the biomass. In the control roots, content of cajaninstilbene acid was 0.03 mg/g DW and the product could not be detected in the extract from the nutrient medium [210].

### 2.3. Hydrolyzable Tannins

Plants that accumulate considerable amounts of hydrolysable tannins are traditionally used as anti-diarrhea medicines. They are also used topically as hemostatic, antiphlogistic and astringent agents. Hydrolyzable tannins (for structures, see Figure 8) are components of numerous food plants including the strawberry, raspberry, pomegranate and walnuts, but their bioavailability is poor. When ingested, hydrolyzable tannins are metabolized by gut microbiota to urolithins that are much better absorbed and are found in plasma mostly as glucuronides. The tannin metabolites have gained much interest as potential anticancer agents and estrogen receptor modulators of potential use in cardiovascular disorders, osteoporosis and several hormone-dependent diseases [211–213]. Hydrolyzable tannins production in the HRs has been studied chiefly to elucidate the regulation of the biosynthesis of these compounds.

*Sanguisorba officinalis* L. (Rosaceae) has been traditionally used as hemostatic and wound healing medicine. The HRs of the plant were obtained by inoculation with *R. rhizogenes* strain A4. Five phenolic compounds were identified in the investigated roots: gallic acid, 1,2,3,6-tetra-*O*-galloyl- $\beta$ -D-glucose, 1,2,3,4,6-penta-*O*-galloyl- $\beta$ -D-glucose, sanguin H-6 and sanguin H-11. Five lines of the studied HRs preferably synthesized sanguin H-6 (0.217–0.569% FW) and one, showing the fastest growth, accumulated mainly 1,2,3,6-tetra-*O*-galloyl- $\beta$ -D-glucose (0.322% FW) and sanguin H-11 (0.221% FW). An intact plant contained 0.206% FW sanguin H-6. The contents of hydrolysable tannins in the HRs were similar to those found in adventitious roots; however, the biomass of HRs obtained in one four-week growth cycle was up to six times larger than that produced by adventitious roots [214].

The HRs of *Geranium thunbergii* Siebold ex Lindl. & Paxt. (Geraniaceae), transformed with *R. rhizogenes* strain A4 synthesized: gallic acid, ellagic acid, (+)-catechin,  $\beta$ -glucogallin (1-*O*-galloyl- $\beta$ -D-glucopyranoside), 1,6-di-*O*-galloyl- $\beta$ -D-glucose, 1,2,3,6-tetra-*O*-galloyl- $\beta$ -D-glucose, 1,2,3,4,6-penta-*O*-galloyl- $\beta$ -D-glucose, corilagin and geraniin. The HRs grown in  $\frac{1}{2}$  MS medium accumulated mainly 1,2,3,4,6-penta-*O*-galloyl- $\beta$ -D-glucose whereas Gamborg's B5 medium favored biosynthesis of geraniin [215]. *Phyllanthus niruri* L. (Phyllanthaceae) partly owes its medicinal properties to a high content of corilagin. The HRs of *P. niruri* produced neither corilagin nor geraniin but yielded several phenolic metabolites including gallic acid, (–)-epicatechin 3-*O*-gallate, (+)-gallo catechin and (–)-epigallocatechin 3-*O*-gallate [216]. *Fragaria ananassa* cv. Reikou infected by *R. rhizogenes* strain ATCC 15834 developed hairy roots that produced pedunculagin (2,3,4,6-tetra-*O*-galloyl- $\beta$ -D-glucose). The tannin was present in the HRs grown in MS medium in the initial phase of growth (0.6% DW) and then its content declined (0.4% in 3-week culture). After 5 weeks of culture pedunculagin was not detected in the HRs. Two other nutrient media used in the study were less favorable for both biomass and pedunculagin production [35].



**Figure 8.** Chemical structures of gallic acid, ellagic acid and selected hydrolyzable tannins.

*Lawsonia inermis* L. (Lythraceae) is a source of henna, a pigment used to dye hair, skin and fingernails. A naphthoquinone, lawsone, is the compound responsible for dyeing properties as well as for bacteriostatic activity. Except for lawsone, gallic acid, flavonoids, coumarins and xanthenes are known metabolites of the plant. The HRs of *L. inermis* were derived from the explants infected with *R. rhizogenes* strain NCIB 8196. The optimum growth of roots was achieved using MS medium. The HRs grown in the dark accumulated 1,2,3,6-tetra-*O*-galloyl- $\beta$ -D-glucose (0.43% DW), 1,2,3,4,6-penta-*O*-galloyl- $\beta$ -D-glucose (3.04% DW) and (+)-catechin. Normal roots contained up to 0.5% galloylglucoses, but up to 60 times more (+)-catechin [217].

*Punica granatum* L. (Lythraceae) cv. Wonderful was used to establish pomegranate HR culture. Three wild-type *R. rhizogenes* strains: MSU440, ATCC 15834 and A4, were transformed with a binary vector containing YFP (yellow fluorescent protein). The modified bacterial strains were used to initiate HRs. Punicalagin  $\alpha$  and  $\beta$  were produced by the HRs with better efficiency than that achieved by in vitro cultured normal roots. The roots of the wild-type seedlings, however, contained more punicalagins.

The glycosylation of gallic acid to  $\beta$ -glucogallin is the first step in punicalagin biosynthesis. The reaction is catalyzed by a specific UDP-glycosyltransferase (UGT). Transcriptional analysis of pomegranate fruit peel (rich in tannins) allowed for the identification of

32 putative UGTs. By comparison with the results of transcript level analyses made for other tissues, two candidate genes were identified (*PgUGT1* and *PgUGT13*) that correlated with the tissue-specific abundance of tannins. To verify whether the two genes as well as the genes for the other UGTs are expressed in the HRs, RT-PCR was performed using RNA extracted from fruit peel, wild type roots and HRs. Expression of 26 putative UGTs was confirmed in all of the investigated tissue types [218]. Four candidate UGTs from pomegranate were cloned and biochemically characterized of which only two (UGT84A23 and UGT84A24) catalyzed formation of  $\beta$ -glucogallin. The overexpression or knockdown of the gene encoding one of the two UGTs in the HRs did not cause any obvious alterations in punicalagin accumulation. Double-knockdown HR lines accumulated less punicalagin but started to synthesize galloyl glucosides (ether-linked gallic acid and glucose). The new metabolites of the double-knockdown HR lines were gallic acid 3-O-glucopyranoside, gallic acid 4-O-glucopyranoside and digalloyl glucose conjugate. The results suggested that in the HRs, gallic acid may be utilized by an unknown UGTs for glucoside formation [219]. In a search for the UGTs engaged in the synthesis of gallic acid glycosides 11 candidate UGTs were identified by the comparison of UGTs expression in control root lines and double knockdown lines of pomegranate HRs. Of the 11 candidate UGTs, only one was active toward gallic acid and catalyzed the formation of a single product: gallic acid 4-O-glucoside [220].

### 3. Conclusions

Table 1 summarizes the most spectacular achievements of the research on highly productive hairy root cultures or genetically modified plants containing polyphenolic antioxidants from the flavonoid, stilbenoid and hydrolyzable tannin group. The high yields of the polyphenolic products were obtained using procedures like optimization of culture conditions, selection of the plant material and bacterial strain used to initiate the HRs, elicitation, permeabilization of plant tissue, adding the enzyme cofactors and genetic engineering (overexpression, heterologous expression or silencing of genes-encoding the regulatory transcription factors and biosynthetic enzymes). It should be considered, however, that high contents of polyphenols in the HRs may be not accompanied by the vigorous root growth. This can hamper applications of the cultures that accumulate products in the cells.

**Table 1.** Hairy root cultures that produced high yields of antioxidative natural products.

Plant Species	Hairy Roots	Natural Product	Maximum Content in the Biomass	Lit.
<i>Fragaria x ananassa</i> Duch cv. Reikou	Wild type	Procyanidin B-3	8 mg/g DW	[35]
<i>Litchi chinensis</i> Sonn.	Overexpressing <i>LcMYB1</i>	Anthocyanins Proanthocyanidins	3 mg/g FW 15 mg/g FW	[48]
<i>Vitis vinifera</i> L.	Overexpressing <i>VvMybP1</i> , <i>VvMybP2</i>	Proanthocyanidins	8 mg/g FW	[38]
<i>Fagopyrum tataricum</i> (L.) Moench. cv. Hokkai T10	Wild type	Cyanidin 3-O-glucoside Cyanidin 3-O-rutinoside	0.8 mg/g DW 2.4 mg/g DW	[52]
<i>Daucus carota</i> L. ssp. <i>sativus</i> var. <i>atrorubens</i> Alef	Wild type, treated with ethephon	Anthocyanins	8 mg/g DW	[9]
<i>Antirrhinum majus</i> L.	Overexpressing <i>AmRosea1</i>	Anthocyanins	2 mg/g FW	[58]
<i>Fagopyrum esculentum</i> Moench	Wild type	(-)-Epicatechin 3-O-gallate (+)-Catechin	10 mg/g DW 8 mg/g DW	[61]

Table 1. Cont.

Plant Species	Hairy Roots	Natural Product	Maximum Content in the Biomass	Lit.
<i>Fagopyrum tataricum</i> (L.) Moench cv. Hokkai T10	Wild type	Rutin	59 mg/g DW	[34]
<i>Fagopyrum tataricum</i>	Overexpressing <i>FtUGT73BE5</i>	Rutin	Up to 80 mg/g DW	[65]
<i>Polygonum multiflorum</i> Thunb.	Wild type, elicited with MJ (50 µM)	Quercetin	Up to 14 mg/g DW	[74]
<i>Isatis tinctoria</i> L.	Wild type, exposed to UV-B	Rutin Quercetin Isorhamnetin Kaempferol	Up to 1.5 mg/g DW Up to 1.7 mg/g DW Up to 2.0 mg/g DW Up to 2.2 mg/g DW	[84]
<i>Ligularia fischeri</i> (Ledeb.) Turcz.	Wild type	Myricetin	2.4 mg/g DW	[86]
<i>Scutellaria baicalensis</i> Georgi	Wild type	Baicalein derivatives Wogonin derivatives Oroxylin A derivatives	68.7 mg/g DW 15.1 mg/g DW 11.9 mg/g DW	[96]
<i>Scutellaria baicalensis</i> Georgi	Overexpressing <i>SbPAL</i>	Baicalin Baicalein Wogonin	136 mg/g DW 29 mg/g DW 6.7 mg/g DW	[102]
<i>Scutellaria lateriflora</i> L.	Wild type	Baicalin Wogonoside Wogonin	14.5 mg/g DW 12.0 mg/g DW 11.5 mg/g DW	[111]
<i>Saussurea medusa</i> Maxim	Wild type	Jaceosidin	6.1 mg/g DW	[126]
<i>Saussurea involucrata</i> Kar. et Kir. ex Maxim	Overexpressing <i>SmCHI</i>	Apigenin	2.6 mg/g DW	[127]
<i>Erigeron breviscapus</i> (Vaniot) Hand.-Mazz.	Overexpressing <i>EbCHI</i>	Scutellarin	2.2 mg/g DW	[128]
<i>Glycine max</i> (L.) Merr.	Overexpressing <i>GmMYB205</i>	Daidzein derivative	6.7 mg/g DW	[141]
<i>Glycine max</i> (L.) Merr.	Overexpressing <i>GmMaT2</i>	Malonyldaidzin	6.3 mg/g FW	[145]
<i>Glycine max</i> (L.) Merr.	Wild type, elicited with MeJa (100 µM)	Daidzin	33.9 mg/g DW	[146]
<i>Trifolium pratense</i> L.	Wild type	Daidzein Genistein Formononetin Biochanin A	8.6 mg/g DW 2.5 mg/g DW 15.2 mg/g DW 1.1 mg/g DW	[149]
<i>Astragalus membranaceus</i> (Fisch.) Bunge	Wild type, co-cultivated with the immobilized <i>Aspergillus niger</i>	Calycosin Formononetin	0.7 mg/g DW 1.1 mg/g DW	[155]
<i>Psoralea lachnostachys</i> F. Muell	Wild type	Daidzein Coumestrol	10.2 mg/g DW 0.5 mg/g DW	[158]
<i>Psoralea corylifolia</i> L.	Wild type	Daidzin	Over 10 mg/g DW	[159]
<i>Pueraria candollei</i> Wall. Ex Benth	Wild type	Daidzin Puerarin	Up to 29.9 mg/g DW Up to 3.4 mg/g DW	[164]
<i>Ononis spinosa</i> L.	Wild type	Medicarpin glucoside Sativanone glucoside Pseudobaptigenin glucoside	22.3–28.9 mg/g DW 5.6–11.4 mg/g DW 1.2–2.0 mg/g DW	[170]
<i>Arachis hypogaea</i> L. cv. Hull	Wild type, elicited with MeJa (100 µM) and cyclodextrin (9 g/L)	Resveratrol Piceatannol Arachidin-1 Arachidin-3	5.3 mg/g DW 0.3 mg/g DW 4.0 mg/g DW (56 mg/L) 17.1 mg/g DW (148 mg/L)	[190]

Table 1. Cont.

Plant Species	Hairy Roots	Natural Product	Maximum Content in the Biomass	Lit.
<i>Arachis hypogaea</i> L. cv. Hull	Wild type, elicited with MeJa (125 µM), cyclodextrin (18 g/L), H <sub>2</sub> O <sub>2</sub> (3 mM), and MgCl <sub>2</sub> (1 mM)	Arachidin-1 Arachidin-2 Arachidin-3 Arachidin-5	227.4 mg/L 83.1 mg/L 370.6 mg/L 68.4 mg/L	[194]
<i>Arachis hypogaea</i> L. cv. Kalasin2	Wild type, elicited with paraquat (500 µM), MeJa (100 µM) and cyclodextrin (6.87 mM)	Resveratrol Arachidin-1 Arachidin-3	1.3 mg/g DW 180.1 mg/g DW 444.2 mg/g DW	[197]
<i>Arachis hypogaea</i> L. cv. Tainan9	Wild type, elicited with paraquat (500 µM), MeJa (100 µM) and cyclodextrin (6.87 mM)	Arachidin-1 Arachidin-3	1700 mg/L 4800 mg/L	[199]
<i>Vitis rotundifolia</i> Michx.	Wild type, elicited with MeJa (100 µM)	Piceid ε-Viniferin	0.34 mg/g DW 0.38 mg/g DW	[203]
<i>Vitis rotundifolia</i> Michx	Wild type, elicited with H <sub>2</sub> O <sub>2</sub> (10 mM)	ε-Viniferin	0.43 mg/g DW	[203]
<i>Vitis vinifera</i> Pinot Noir cv. PN40024	Wild type, elicited with MeJa (100 µM)	Total stilbenoids	6.98 mg/g DW	[204]
<i>Vitis vinifera</i> Pinot Noir cv. PN40024	Wild type, elicited with MeJa (100 µM) and cyclodextrin (50 mM)	Total stilbenoids	165 mg/L (medium) 6.4 mg/g DW	[204]
<i>Vitis vinifera</i> subsp. <i>sylvestris</i> acc. W16	Wild type	Resveratrol	0.27 mg/g DW	[207]
<i>Cajanus cajan</i> (L.) Millsp.	Wild type, irradiated with UV-B	Cajanin stilbene acid	Up to 6.6 mg/g DW	[134]
<i>Cajanus cajan</i> (L.) Millsp.	Wild type, elicited with MeJa (125 µM), cyclodextrin (18 g/L), H <sub>2</sub> O <sub>2</sub> (3 mM), and MgCl <sub>2</sub> (1 mM)	Cajanin stilbene acid	Over 8 mg/g DW	[210]

HR cultures provide a valuable tool for the rapid characterization of regulatory mechanisms in plant polyphenol biosynthesis, and elucidation of the roles played by the particular biosynthetic enzymes or transcription factors. The use of HRs has led to the identification and description of several enzymes implicated in polyphenol biosynthesis. Nowadays, it seems that HR culture is primarily a useful tool in the genetic engineering of plants. The implementation of the industrial process using HRs to produce antioxidant natural products looks like a distant prospect.

**Author Contributions:** Conceptualization, A.S.; methodology, A.S and J.M.; resources, A.S., J.M. and Y.V.Y.; data curation, A.S.; writing—original draft preparation, A.S. and Y.V.Y.; writing—review and editing, A.S., J.M., K.M. and Y.V.Y.; visualization, K.M.; supervision, A.S.; project administration, A.S.; funding acquisition, A.S. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

4CL	4-coumarate:CoA ligase (4-coumaroyl CoA ligase)
ANR	anthocyanidin reductase
ANS	anthocyanidin synthase
BAP	6-benzylaminopurine
bHLH	basic helix-loop-helix
BHT	butylhydroxytoluen
C4H	cinnamate 4-hydroxylase
CHI	chalcone isomerase
CHS	chalcone synthase
DFR	dihydroflavonol 4-reductase
DW	dry weight
F3H	flavone 3-hydroxylase
F3 $\beta$ OH	flavanone 3- $\beta$ -hydroxylase
F6H	flavone 6-hydroxylase
F8H	flavone 8-hydroxylase
FNS	flavone synthase
FW	fresh weight
GUS	$\beta$ -glucuronidase
HPLC-PAD-ESI-MS <sup>n</sup>	high-performance liquid chromatography with photodiode detection, electrospray ionization, and multi-stage mass spectrometry
HR	hairy roots
IBA	indole-3-butyric acid
IF7Mat	isoflavone 7-O-glucoside-6''-O-malonyltransferase
IFD	isoflavone dehydratase
IFS	isoflavone synthase
IOMT	isoflavone-O-methyltransferase
LAR	leucoanthocyanidin reductase
LC-MS/MS	liquid chromatography with tandem mass spectrometry
MaT	malonyl CoA:flavonoid acyltransferase
MeJa	methyl jasmonate
MS	Murashige and Skoog
PA	proanthocyanidin
PAL	phenylalanine ammonia lyase
PAP1	production of anthocyanin pigment 1 gene
PQ	paraquat
RS	resveratrol synthase
SA	salicylic acid
SH	Schenk and Hildebrandt
STS	stilbene synthase
TOF-MS	time-of-flight mass spectroscopy
TT2	transparent testa 2 transcription factor
UBGT	UDP-glucose:baicalein 7-O-glucosyltransferase
UGT	UDP-glucosyltransferase
UHPLC-PDA	ultra-high-performance liquid chromatography with photodiode detection
YE	yeast extract

## References

1. Hamill, J.D.; Lidgett, A.J. Hairy root cultures—opportunities and key protocols for studies in metabolic engineering. In *Hairy Roots. Culture and Applications*, 1st ed.; Doran, P., Ed.; Harwood Academic Publishers: Amsterdam, The Netherlands, 1997; pp. 1–29.
2. Giri, A.; Narasu, M.L. Transgenic hairy roots: Recent trends and applications. *Biotechnol. Adv.* **2000**, *18*, 1–22. [[CrossRef](#)]
3. Guillon, S.; Trémouillaux-Guiller, J.; Pati, P.K.; Rideau, M.; Gantet, P. Harnessing the potential of hairy roots: Dawn of a new era. *Trends Biotechnol.* **2006**, *24*, 403–409. [[CrossRef](#)] [[PubMed](#)]
4. Ono, N.N.; Tian, L. The multiplicity of hairy root cultures: Prolific possibilities. *Plant Sci.* **2011**, *180*, 439–446. [[CrossRef](#)]

5. Gutierrez-Valdes, N.; Häkkinen, S.T.; Lemasson, C.; Guillet, M.; Oksman-Caldentey, K.M.; Ritala, A.; Cardon, F. Hairy root cultures—a versatile tool with multiple applications. *Front. Plant Sci.* **2020**, *11*, 33. [[CrossRef](#)]
6. Shi, M.; Liao, P.; Nile, S.H.; Georgiev, M.I.; Kai, G. Biotechnological exploration of transformed root culture for value-added products. *Trends Biotechnol.* **2021**, *39*, 137–149. [[CrossRef](#)] [[PubMed](#)]
7. Young, J.M.; Kuykendall, L.D.; Martínez-Romero, E.; Kerr, A.; Sawada, H. A revision of *Rhizobium* Frank 1889, with an emended description of the genus, and the inclusion of all species of *Agrobacterium* Conn 1942 and *Allorhizobium undicola* de Lajudie et al. 1998 as new combinations: *Rhizobium radiobacter*, *R. rhizogenes*, *R. rubi*, *R. undicola* and *R. vitis*. *Int. J. Syst. Evol. Microbiol.* **2001**, *51*, 89–103. [[CrossRef](#)]
8. Hidalgo, D.; Georgiev, M.; Marchev, A.; Bru-Martinez, R.; Cusido, R.; Corchete, P.; Palazon, J. Tailoring tobacco hairy root metabolism for the production of stilbenes. *Sci. Rep.* **2017**, *7*, 17976. [[CrossRef](#)]
9. Dong, Q.; Zhao, H.; Huang, Y.; Chen, Y.; Wan, M.; Zeng, Z.; Yao, P.; Li, C.; Wang, X.; Chen, H.; et al. FtMYB18 acts as a negative regulator of anthocyanin/proanthocyanidin biosynthesis in Tartary buckwheat. *Plant Mol. Biol.* **2020**, *104*, 309–325. [[CrossRef](#)]
10. Sosa Alderete, L.G.; Talano, M.A.; Ibáñez, S.G.; Purro, S.; Agostini, E.; Milrad, S.R.; Medina, M.I. Establishment of transgenic tobacco hairy roots expressing basic peroxidases and its application for phenol removal. *J. Biotechnol.* **2009**, *139*, 273–279. [[CrossRef](#)]
11. Ibáñez, S.; Talano, M.; Ontañón, O.; Suman, J.; Medina, M.I.; Macek, T.; Agostini, E. Transgenic plants and hairy roots: Exploiting the potential of plant species to remediate contaminants. *New Biotechnol.* **2016**, *33*, 625–635. [[CrossRef](#)]
12. Jha, P.; Sen, R.; Jobby, R.; Sachar, S.; Bhatkalkar, S.; Desai, N. Biotransformation of xenobiotics by hairy roots. *Phytochemistry* **2020**, *176*, 112421. [[CrossRef](#)] [[PubMed](#)]
13. Gantait, S.; Mukherjee, E. Hairy root culture technology: Applications, constraints and prospect. *Appl. Microbiol. Biotechnol.* **2021**, *105*, 35–53. [[CrossRef](#)] [[PubMed](#)]
14. Lystvan, K.; Lystvan, V.; Shcherbak, N.; Kuchuk, M. Rhizoextraction potential of *Convolvulus tricolor* hairy roots for Cr<sup>6+</sup>, Ni<sup>2+</sup>, and Pb<sup>2+</sup> removal from aqueous solutions. *Appl. Biochem. Biotechnol.* **2021**, *193*, 1215–1230. [[CrossRef](#)]
15. Srikantan, C.; Srivastava, S. Bioreactor design and analysis for large-scale plant cell and hairy root cultivation. In *Hairy Roots An Effective Tool of Plant Biotechnology*, 1st ed.; Srivastava, V., Mehrotra, S., Mishra, S., Eds.; Springer Nature Singapore PteLtd.: Singapore, 2018; pp. 147–182.
16. Hussain, M.J.; Abbas, Y.; Nazli, N.; Fatima, S.; Drouet, S.; Hano, C.; Abbasi, B.H. Root cultures, a boon for the production of valuable compounds: A comparative review. *Plants* **2022**, *11*, 439. [[CrossRef](#)] [[PubMed](#)]
17. Macáková, K.; Afonso, R.; Saso, L.; Mladěnka, P. The influence of alkaloids on oxidative stress and related signaling pathways. *Free Radic. Biol. Med.* **2019**, *134*, 429–444. [[CrossRef](#)] [[PubMed](#)]
18. Williamson, G.; Faulkner, K.; Plumb, G.W. Glucosinolates and phenolics as antioxidants from plant foods. *Eur. J. Cancer Prev.* **1998**, *7*, 17–21.
19. Cabello-Hurtado, F.; Gicquel, M.; Esnault, M.-A. Evaluation of the antioxidant potential of cauliflower (*Brassica oleracea*) from a glucosinolate content perspective. *Food Chem.* **2012**, *132*, 1003–1009. [[CrossRef](#)]
20. Wang, X.; Liu, R.; Yang, Y.; Zhang, M. Isolation, purification and identification of antioxidants in an aqueous aged garlic extract. *Food Chem.* **2015**, *187*, 37–43. [[CrossRef](#)]
21. Brewer, M.S. Natural antioxidants: Sources, compounds, mechanisms of action, and potential applications. *Compr. Rev. Food Sci. Food Saf.* **2011**, *10*, 221–247. [[CrossRef](#)]
22. Krishnajah, D.; Sarbatly, R.; Nithyanandam, R. A review of the antioxidant potential of medicinal plant species. *Food Bioprod Process.* **2011**, *89*, 217–233. [[CrossRef](#)]
23. Wink, M.; Ashour, M.L.; El-Readi, M.Z. Secondary metabolites from plants inhibiting ABC transporters and reversing resistance of cancer cells and microbes to cytotoxic and antimicrobial agents. *Front. Microbiol.* **2012**, *3*, 130. [[CrossRef](#)]
24. Choudhari, A.S.; Mandave, P.C.; Deshpande, M.; Ranjekar, P.; Prakash, O. Phytochemicals in cancer treatment: From preclinical studies to clinical practice. *Front. Pharmacol.* **2020**, *10*, 1614. [[CrossRef](#)] [[PubMed](#)]
25. Cui, X.; Lin, Q.; Liang, Y. Plant-derived antioxidants protect the nervous system from aging by inhibiting oxidative stress. *Front. Aging Neurosci.* **2020**, *12*, 209. [[CrossRef](#)] [[PubMed](#)]
26. Duba, K.S.; Casazza, A.A.; Mohamed, H.B.; Perego, P.; Fiori, L. Extraction of polyphenols from grape skins and defatted grape seeds using subcritical water: Experiments and modeling. *Food Bioprod. Process.* **2015**, *94*, 29–38. [[CrossRef](#)]
27. Sorrenti, V.; Randazzo, C.L.; Caggia, C.; Ballistreri, G.; Romeo, F.V.; Fabroni, S.; Timpanaro, N.; Raffaele, M.; Vanella, L. Beneficial effects of pomegranate peel extract and probiotics on pre-adipocyte differentiation. *Front. Microbiol.* **2019**, *10*, 660. [[CrossRef](#)]
28. Cova, C.M.; Boffa, L.; Pistocchi, M.; Giorgini, S.; Luque, R.; Cravotto, G. Technology and process design for phenols recovery from industrial chicory (*Chicorium intybus*) leftovers. *Molecules* **2019**, *24*, 2681. [[CrossRef](#)]
29. Khalili, F.; Dinani, S.T. Extraction of phenolic compounds from olive-waste cake using ultrasonic process. *J. Food Meas. Charact.* **2018**, *12*, 974–981. [[CrossRef](#)]
30. Morris, P.; Robbins, M.P. Condensed tannin formation by *Agrobacterium rhizogenes* transformed root and shoot organ cultures of *Lotus corniculatus*. *J. Exp. Bot.* **1992**, *43*, 221–231. [[CrossRef](#)]
31. Hook, I. Secondary metabolites in hairy root cultures of *Leontopodium alpinum* Cass. (Edelweiss). *Plant Cell Tiss. Org. Cult.* **1994**, *38*, 321–326. [[CrossRef](#)]

32. Park, C.H.; Thwe, A.A.; Kim, S.J.; Park, J.S.; Arasu, M.V.; Al-Dhabi, N.A.; Park, N.I.; Park, S.U. Effect of auxins on anthocyanin accumulation in hairy root cultures of Tartary buckwheat cultivar Hokkai T10. *Nat. Prod. Commun.* **2016**, *11*, 1283–1286. [[CrossRef](#)]
33. Nishiyama, Y.; Yamakawa, T. Effect of medium composition on the production of anthocyanins by hairy root cultures of *Ipomea batatas*. *Plant Biotechnol.* **2004**, *21*, 411–414. [[CrossRef](#)]
34. Thwe, A.A.; Kim, Y.; Li, X.; Kim, Y.B.; Park, N.I.; Kim, H.H.; Kim, S.-J.; Park, S.U. Accumulation of phenylpropanoids and correlated gene expression in hairy roots of Tartary buckwheat under light and dark conditions. *Appl. Biochem. Biotechnol.* **2014**, *174*, 2537–2547. [[CrossRef](#)] [[PubMed](#)]
35. Motomori, Y.; Shimomura, K.; Mori, K.; Kunitake, H.; Nakashima, T.; Tanaka, M.; Miyazaki, S.; Ishimaru, K. Polyphenol production in hairy root cultures of *Fragaria x ananassa*. *Phytochemistry* **1995**, *40*, 1425–1428. [[CrossRef](#)]
36. Bavage, A.D.; Davies, I.G.; Robbins, M.P.; Morris, P. Expression of an Antirrhinum dihydroflavonol reductase gene results in changes in condensed tannin structure and accumulation in root cultures of *Lotus corniculatus* (bird's foot trefoil). *Plant Mol. Biol.* **1997**, *35*, 443–458. [[CrossRef](#)]
37. Damiani, F.; Paolocci, F.; Consonni, G.; Crea, F.; Tonelli, C.; Arcioni, S. A maize anthocyanin transactivator induces pigmentation in hairy roots of dicotyledonous species. *Plant Cell Rep.* **1998**, *17*, 339–344. [[CrossRef](#)]
38. Pang, Y.; Peel, G.J.; Sharma, S.B.; Tang, Y.; Dixon, R.A. A transcript profiling approach reveals an epicatechin-specific glucosyltransferase expressed in the seed coat of *Medicago truncatula*. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 14210–14215. [[CrossRef](#)] [[PubMed](#)]
39. Terrier, N.; Torregrosa, L.; Ageorges, A.; Violet, S.; Verriès, C.; Cheynier, V.; Romieu, C. Ectopic expression of VvMybPA2 promotes proanthocyanidin biosynthesis in grapevine and suggests additional targets in the pathway. *Plant Physiol.* **2009**, *149*, 1028–1041. [[CrossRef](#)]
40. Verdier, J.; Zhao, J.; Torres-Jerez, I.; Ge, S.; Liu, C.; He, X.; Mysore, K.S.; Dixon, R.A.; Udvardi, M.K. MtPAR MYB transcription factor acts as an on switch for proanthocyanidin biosynthesis in *Medicago truncatula*. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 1766–1771. [[CrossRef](#)]
41. Huang, Y.-F.; Violet, S.; Guiraud, J.-L.; Torregrosa, L.; Bertrand, Y.; Cheynier, V.; This, P.; Terrier, N. A negative MYB regulator of proanthocyanidin accumulation, identified through expression quantitative locus mapping in the grape berry. *New Phytol.* **2014**, *201*, 795–809. [[CrossRef](#)]
42. Cavallini, E.; Matus, J.T.; Finezzo, L.; Zenoni, S.; Loyola, R.; Guzzo, F.; Schlechter, R.; Ageorges, A.; Arce-Johnson, P.; Tornielli, G.B. The phenylpropanoid pathway is controlled at different branches by a set of R2R3-MYB C2 repressors in grapevine. *Plant Physiol.* **2015**, *167*, 1448–1470. [[CrossRef](#)]
43. Jun, J.H.; Liu, C.; Xiao, X.; Dixon, R.A. The transcriptional repressor MYB2 regulates both spatial and temporal patterns of proanthocyanidin and anthocyanin pigmentation in *Medicago truncatula*. *Plant Cell* **2015**, *27*, 2860–2879. [[PubMed](#)]
44. Li, P.; Chen, B.; Zhang, G.; Chen, L.; Dong, Q.; Wen, J.; Mysore, K.S.; Zhao, J. Regulation of anthocyanin and proanthocyanidin biosynthesis by *Medicago truncatula* bHLH transcription factor MtTT8. *New Phytol.* **2016**, *210*, 905–921. [[CrossRef](#)] [[PubMed](#)]
45. Lu, N.; Roldan, M.; Dixon, R.A. Characterization of two TT2-type MYB transcription factors regulating proanthocyanidin biosynthesis in tetraploid cotton, *Gossypium hirsutum*. *Planta* **2017**, *246*, 323–335. [[CrossRef](#)] [[PubMed](#)]
46. Lu, N.; Rao, X.; Li, Y.; Jun, J.H.; Dixon, R.A. Dissecting the transcriptional regulation of proanthocyanidin and anthocyanin biosynthesis in soybean (*Glycine max*). *Plant Biotechnol. J.* **2021**, *19*, 1429–1442. [[CrossRef](#)]
47. Aggarwal, P.R.; Nag, P.; Choudhary, P.; Chakraborty, N.; Chakraborty, S. Genotype-independent *Agrobacterium rhizogenes*-mediated root transformation of chickpea: A rapid and efficient method for reverse genetics studies. *Plant Methods* **2018**, *14*, 55. [[CrossRef](#)]
48. Li, P.; Dong, Q.; Ge, S.; He, X.; Verdier, J.; Li, D.; Zhao, J. Metabolic engineering of proanthocyanidin production by repressing the isoflavone pathways and redirecting anthocyanidin precursor flux in legume. *Plant Biotechnol. J.* **2016**, *14*, 1604–1618. [[CrossRef](#)]
49. Qin, Y.; Wang, D.; Fu, J.; Zhang, Z.; Qin, Y.; Hu, G.; Zhao, J. *Agrobacterium rhizogenes*-mediated hairy root transformation as an efficient system for gene function analysis in *Litchi chinensis*. *Plant Methods* **2021**, *17*, 103. [[CrossRef](#)]
50. Xu, S.; Lai, E.; Zhao, L.; Cai, Y.; Ogutu, C.; Cherono, S.; Han, Y.; Zheng, B. Development of a fast and efficient root transgenic system for functional genomics and genetic engineering in peach. *Sci. Rep.* **2020**, *10*, 2836. [[CrossRef](#)]
51. Yoshida, K.; Ma, D.; Constabel, C.P. The MYB182 protein down-regulates proanthocyanidin biosynthesis in poplar by repressing both structural and regulatory flavonoid genes. *Plant Physiol.* **2015**, *167*, 693–710. [[CrossRef](#)]
52. Pang, Y.; Abeysinghe, I.S.B.; He, J.; He, X.; Huhman, D.; Mewan, K.M.; Sumner, L.W.; Yun, J.; Dixon, R.A. Functional characterization of proanthocyanidin pathway enzymes from tea and their application for metabolic engineering. *Plant Physiol.* **2013**, *161*, 1103–1116. [[CrossRef](#)]
53. Thwe, A.; Arasu, M.V.; Li, X.; Park, C.H.; Kim, S.J.; Al-Dhabi, N.A.; Park, S.U. Effect of different *Agrobacterium rhizogenes* strains on hairy root induction and phenylpropanoid biosynthesis in Tartary buckwheat (*Fagopyrum tataricum* Gaertn). *Front. Microbiol.* **2016**, *7*, 318. [[CrossRef](#)] [[PubMed](#)]
54. Li, X.; Thwe, A.A.; Park, C.H.; Kim, S.J.; Arasu, M.V.; Al-Dhabi, N.A.; Lee, S.Y.; Park, S.U. Ethephon-induced phenylpropanoid accumulation and related gene expression in tartary buckwheat (*Fagopyrum tataricum* (L.) Gaertn.) hairy root. *Biotechnol. Biotechnol. Equip.* **2017**, *31*, 304–311. [[CrossRef](#)]
55. Li, X.; Sathasivam, R.; Park, N.I.; Wu, Q.; Park, S.U. Enhancement of phenylpropanoid accumulation in Tartary buckwheat hairy roots by overexpression of MYB transcription factors. *Ind. Crop. Prod.* **2020**, *156*, 112887. [[CrossRef](#)]

56. Barba-Espin, G.; Chen, S.-T.; Agnolet, S.; Nymark Hegelund, J.; Stanstrup, J.; Christensen, J.H.; Müller, R.; Lütken, H. Ethephon-induced changes in antioxidants and phenolic compounds in anthocyanin-producing black carrot hairy root cultures. *J. Exp. Bot.* **2020**, *71*, 7030–7045. [[CrossRef](#)] [[PubMed](#)]
57. Murashige, T.; Skoog, F. A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol. Plant.* **1962**, *15*, 473–497. [[CrossRef](#)]
58. Barba-Espin, G.; Martínez-Jiménez, C.; Izquierdo-Martínez, A.; Acosta-Motos, J.R.; Hernández, J.A.; Díaz-Vivancos, P. H<sub>2</sub>O<sub>2</sub>-elicitation of black carrot hairy roots induces a controlled oxidative burst leading to increased anthocyanin production. *Plants* **2021**, *10*, 2753. [[CrossRef](#)]
59. Piao, C.; Wu, J.; Cui, M.-L. The combination of R2R3-MYB gene *AmRosea1* and hairy root culture is a useful tool for rapidly induction and production of anthocyanins in *Antirrhinum majus* L. *AMB Expr.* **2021**, *11*, 128. [[CrossRef](#)]
60. Jin, S.; Hyun, T.K. Ectopic expression of Production of Anthocyanin Pigment 1 (PAP1) improves the antioxidant and anti-melanogenic properties of ginseng (*Panax ginseng* C.A. Meyer) hairy roots. *Antioxidants* **2020**, *9*, 922. [[CrossRef](#)]
61. Jin, S.; Bang, S.; Ahn, M.-A.; Lee, K.; Kim, K.; Hyun, T.K. Overproduction of anthocyanin in ginseng hairy roots enhances their antioxidant, antimicrobial, and anti-elastase activities. *J. Plant Biotechnol.* **2021**, *48*, 100–105. [[CrossRef](#)]
62. Trotin, F.; Moumou, Y.; Vasseur, J. Flavanol production by *Fagopyrum esculentum* hairy and normal root cultures. *Phytochemistry* **1993**, *32*, 929–931. [[CrossRef](#)]
63. Kim, Y.K.; Xu, H.; Park, W.T.; Park, N.I.; Lee, S.Y.; Park, S.U. Genetic transformation of buckwheat (*Fagopyrum esculentum* M.) with *Agrobacterium rhizogenes* and production of rutin in transformed root cultures. *Aust. J. Crop Sci.* **2010**, *4*, 485–490.
64. Park, N.I.; Li, X.; Thwe, A.A.; Lee, S.Y.; Kim, S.G.; Wu, Q.; Park, S.U. Enhancement of rutin in *Fagopyrum esculentum* hairy root cultures by the *Arabidopsis* transcription factor AtMYB12. *Biotechnol. Lett.* **2012**, *34*, 577–583. [[CrossRef](#)] [[PubMed](#)]
65. Yao, P.; Huang, Y.; Dong, Q.; Wan, M.; Wang, A.; Chen, Y.; Li, C.; Wu, Q.; Chen, H.; Zhao, H. FtMYB6, a light-induced SG7 R2R3-MYB transcription factor, promotes flavonol biosynthesis in Tartary buckwheat (*Fagopyrum tataricum*). *J. Food Agric. Chem.* **2020**, *68*, 13685–13696. [[CrossRef](#)]
66. Yin, Q.; Han, X.; Han, Z.; Chen, Q.; Shi, Y.; Gao, H.; Zhang, T.; Dong, G.; Xiong, C.; Song, C.; et al. Genome-wide analyses reveals a glucosyltransferase involved in rutin and emodin glucoside biosynthesis in tartary buckwheat. *Food Chem.* **2020**, *318*, 126478. [[CrossRef](#)] [[PubMed](#)]
67. Choi, M.; Sathasivam, R.; Nguyen, B.V.; Park, N.I.; Woo, S.-H.; Park, S.U. Expression analysis of phenylpropanoid pathway genes and metabolomic analysis of phenylpropanoid compounds in adventitious, hairy, and seedling roots of Tartary buckwheat. *Plants* **2021**, *11*, 90. [[CrossRef](#)]
68. Tusevski, O.; Petreska Stanoeva, J.; Stefova, M.; Kungulovski, D.; Pancevska, N.A.; Sekulovski, N.; Panov, S.; Gadzovska Simic, S. Hairy roots of *Hypericum perforatum* L.: A promising system for xanthone production. *Cent. Eur. J. Biol.* **2013**, *8*, 1010–1022. [[CrossRef](#)]
69. Tusevski, O.; Petreska Stanoeva, J.; Stefova, M.; Gadzovska Simic, S. Phenolic profile of dark-grown and photoperiod-exposed *Hypericum perforatum* L hairy root cultures. *Sci. World J.* **2013**, 602752. [[CrossRef](#)]
70. Tusevski, O.; Vinterhalter, B.; Kristić Milošević, D.; Soković, M.; Ćirić, A.; Vinterhalter, D.; Zdravković Korać, S.; Petreska Stanoeva, J.; Gadzovska Simic, S. Production of phenolic compounds, antioxidant and antimicrobial activities in hairy root and shoot cultures of *Hypericum perforatum* L. *Plant Cell Tissue Organ Cult.* **2017**, *128*, 589–605. [[CrossRef](#)]
71. Tashackori, H.; Sharifi, M.; Chashmi, N.A.; Safaie, N.; Behmanesh, M. Induced-differential changes on lignan and phenolic acid compounds in *Linum album* hairy roots by fungal extract of *Piriformospora indica*. *Plant Cell Tissue Organ Cult.* **2016**, *127*, 187–194. [[CrossRef](#)]
72. Tashackori, H.; Sharifi, M.; Chashmi, N.A.; Behmanesh, M.; Safaie, N. *Piriformospora indica* cell wall modulates gene expression and metabolite profile in *Linum album* hairy roots. *Planta* **2018**, *248*, 1289–1306. [[CrossRef](#)]
73. Tashackori, H.; Sharifi, M.; Chashmi, N.A.; Behmanesh, M.; Safaie, N.; Sagharyan, M. Physiological, biochemical, and molecular responses of *Linum album* to digested cell wall of *Piriformospora indica*. *Physiol. Mol. Biol. Plants* **2021**, *27*, 2695–2708. [[CrossRef](#)]
74. Thiruvengadam, M.; Praveen, N.; Kim, E.-H.; Kim, S.-H.; Chung, I.-M. Production of antraquinones, phenolic compounds and biological activities from hairy root cultures of *Polygonum multiflorum* Thunb. *Protoplasma* **2014**, *251*, 555–566. [[CrossRef](#)] [[PubMed](#)]
75. Ho, T.-T.; Lee, J.-D.; Ahn, M.-S.; Kim, S.-W.; Park, S.-Y. Enhanced production of phenolic compounds in hairy root cultures of *Polygonum multiflorum* and its metabolite discrimination using HPLC and FT-IR methods. *Appl. Microbiol. Biotechnol.* **2018**, *102*, 9563–9575. [[CrossRef](#)]
76. Thiruvengadam, M.; Praveen, N.; John, K.M.M.; Yang, Y.-S.; Kim, S.-H.; Chung, I.-M. Establishment of *Momordica charantia* hairy root cultures for the production of phenolic compounds and determination of their biological activities. *Plant Cell Tissue Organ Cult.* **2014**, *118*, 545–557. [[CrossRef](#)]
77. Thiruvengadam, M.; Rekha, K.; Chung, I.-M. Induction of hairy roots by *Agrobacterium rhizogenes*-mediated transformation of spine gourd (*Momordica dioica* Roxb. ex. Willd) for the assessment of phenolic compounds and biological activities. *Sci. Hortic.* **2016**, *198*, 132–141. [[CrossRef](#)]
78. Yoon, J.-Y.; Chung, I.-M.; Thiruvengadam, M. Evaluation of phenolic compounds, antioxidant and antimicrobial activities from transgenic hairy root cultures of gherkin (*Cucumis anguria* L.). *S. Afr. J. Bot.* **2015**, *100*, 80–86. [[CrossRef](#)]

79. Chung, I.-M.; Rekha, K.; Rajakumar, G.; Thiruvengadam, M. Production of glucosinolates, phenolic compounds and associated gene expression profiles of hairy root cultures in turnip (*Brassica rapa* ssp. *rapa*). *3 Biotech* **2016**, *6*, 175. [[CrossRef](#)] [[PubMed](#)]
80. Chung, I.-M.; Rekha, K.; Rajakumar, G.; Thiruvengadam, M. Production of bioactive compounds and gene expression alterations in hairy root cultures of chinese cabbage elicited by copper oxide nanoparticles. *Plant Cell Tissue Organ Cult.* **2018**, *134*, 95–106. [[CrossRef](#)]
81. Chung, I.-M.; Rekha, K.; Rajakumar, G.; Thiruvengadam, M. Influence of silver nanoparticles on the enhancement and transcriptional changes of glucosinolates and phenolic compounds in genetically transformed root cultures of *Brassica rapa* ssp. *rapa*. *Bioprocess Biosyst. Eng.* **2018**, *41*, 1665–1677. [[CrossRef](#)]
82. Gai, Q.-Y.; Jiao, J.; Luo, M.; Wei, Z.-F.; Zu, Y.-G.; Ma, W.; Fu, Y.-J. Establishment of hairy root cultures by *Agrobacterium rhizogenes* mediated transformation of *Isatis tinctoria* L. for the efficient production of flavonoids and evaluation of antioxidant activities. *PLoS ONE* **2015**, *10*, e0119022. [[CrossRef](#)]
83. Jiao, J.; Gai, Q.-Y.; Wang, X.; Qin, Q.-P.; Wang, Z.-Y.; Liu, J.; Fu, Y.-J. Chitosan elicitation of *Isatis tinctoria* L. hairy root cultures for enhancing flavonoid productivity and gene expression and related antioxidant activity. *Ind. Crops Prod.* **2018**, *124*, 28–35. [[CrossRef](#)] [[PubMed](#)]
84. Jiao, J.; Gai, Q.-Y.; Wang, W.; Zang, Y.-P.; Niu, L.-L.; Fu, Y.-J.; Wang, X. Remarkable enhancement of flavonoid production in a co-cultivation system of *Isatis tinctoria* L. hairy root cultures and immobilized *Aspergillus niger*. *Ind. Crops Prod.* **2018**, *112*, 252–261. [[CrossRef](#)] [[PubMed](#)]
85. Jiao, J.; Gai, Q.-Y.; Yao, L.-P.; Niu, L.-L.; Zang, Y.-P.; Fu, Y.-J. Ultraviolet radiation for flavonoid augmentation in *Isatis tinctoria* L. hairy root cultures mediated by oxidative stress and biosynthetic gene expression. *Ind. Crops Prod.* **2018**, *118*, 347–354. [[CrossRef](#)]
86. Ghimire, B.K.; Thiruvengadam, M.; Chung, I.-M. Identification of elicitors enhances the polyphenolic compounds and pharmacological potential in hairy root cultures of *Aster scaber*. *S. Afr. J. Bot.* **2019**, *125*, 92–101. [[CrossRef](#)]
87. Ansari, M.A.; Chung, I.-M.; Rajakumar, G.; Alzohairy, M.A.; Almatroudi, A.; Khanna, V.G.; Thiruvengadam, M. Evaluation of polyphenolic compounds and pharmacological activities in hairy root cultures of *Ligularia fischeri* Turcz. f. *spiciformis* (Nakai). *Molecules* **2019**, *24*, 1586. [[CrossRef](#)]
88. Chung, I.-M.; Ahmad, A.; Ali, M.; Lee, O.-K.; Kim, M.-Y.; Kim, J.-H.; Yoon, D.-Y.; Peebles, C.A.M.; San, K.Y. Flavonoid glucosides from the hairy roots of *Catharanthus roseus*. *J. Nat. Prod.* **2009**, *72*, 613–620. [[CrossRef](#)] [[PubMed](#)]
89. Tomimori, T.; Jin, H.; Miyaichi, Y.; Toyofuku, S.; Namba, T. Studies on the constituents of *Scutellaria* species. VI. On the flavonoid constituents of the root of *Scutellaria baicalensis* Georgi (5). Quantitative analysis of flavonoids in *Scutellaria* roots by high-performance liquid chromatography. *Yakugaku Zasshi* **1985**, *105*, 148–155. [[CrossRef](#)] [[PubMed](#)]
90. Shang, X.; He, X.; He, X.; Li, M.; Zhang, R.; Fan, P.; Zhang, Q.; Jia, Z. The genus *Scutellaria* an ethnopharmacological and phytochemical review. *J. Ethnopharmacol.* **2010**, *128*, 279–313. [[CrossRef](#)]
91. Dinda, B.; Dinda, S.; DasSharma, S.; Banik, R.; Chakraborty, A.; Dinda, M. Therapeutic potentials of baicalin and its aglycone, baicalein against inflammatory disorders. *Eur. J. Med. Chem.* **2017**, *131*, 68–80. [[CrossRef](#)]
92. Gao, Z.; Huang, K.; Yang, X.; Xu, H. Free radical scavenging and antioxidant activities of flavonoids extracted from the radix of *Scutellaria baicalensis* Georgi. *Biochim. Biophys. Acta Gen. Subj.* **1999**, *1472*, 643–650. [[CrossRef](#)]
93. Zhou, Y.; Hirotani, M.; Yoshikawa, T.; Furuya, T. Flavonoids and phenylrthanoids from hairy root cultures of *Scutellaria baicalensis*. *Phytochemistry* **1997**, *44*, 83–87. [[CrossRef](#)]
94. Nishikawa, K.; Ishimaru, K. Flavonoids in root cultures of *Scutellaria baicalensis*. *J. Plant Physiol.* **1997**, *151*, 633–636. [[CrossRef](#)]
95. Nishikawa, K.; Furukawa, H.; Fujioka, T.; Fujii, H.; Mihashi, K.; Shimomura, K.; Ishimaru, K. Flavone production in transformed root cultures of *Scutellaria baicalensis* Georgi. *Phytochemistry* **1999**, *52*, 885–890. [[CrossRef](#)]
96. Stojakowska, A.; Malarz, J. Flavonoid production in transformed root cultures of *Scutellaria baicalensis*. *J. Plant Physiol.* **2000**, *156*, 121–125. [[CrossRef](#)]
97. Kuzovkina, I.N.; Guseva, A.V.; Alterman, I.E.; Karnachuk, N.A. Flavonoid production in transformed *Scutellaria baicalensis* roots and ways of its regulation. *Russ. J. Plant. Physiol.* **2001**, *48*, 448–452. [[CrossRef](#)]
98. Kuzovkina, I.N.; Guseva, A.V.; Kovács, D.; Szöke, É.; Vdovitchenko, M.Y. Flavones in genetically transformed *Scutellaria baicalensis* roots and induction of their synthesis by elicitation with methyl jasmonate. *Russ. J. Plant Physiol.* **2005**, *52*, 77–82. [[CrossRef](#)]
99. Kovács, G.; Kuzovkina, I.N.; Szöke, É.; Kursinszki, L. HPLC Determination of flavonoids in hairy-root cultures of *Scutellaria baicalensis* Georgi. *Chromatographia* **2004**, *60*, S81–S85. [[CrossRef](#)]
100. Hirotani, M.; Kuroda, R.; Suzuki, H.; Yoshikawa, T. Cloning and expression of UDP-glucose:flavonoid 7-O-glucosyltransferase from hairy root cultures of *Scutellaria baicalensis*. *Planta* **2000**, *210*, 1006–1013. [[CrossRef](#)]
101. Park, N.I.; Xu, H.; Li, X.; Kim, S.-J.; Park, S.U. Enhancement of flavone levels through overexpression of chalcone isomerase in hairy root cultures of *Scutellaria baicalensis*. *Funct. Integr. Genom.* **2011**, *11*, 491–496. [[CrossRef](#)]
102. Park, N.I.; Xu, H.; Li, X.; Kim, Y.S.; Lee, M.Y.; Park, S.U. Overexpression of phenylalanine ammonia-lyase improves flavones production in transgenic hairy root cultures of *Scutellaria baicalensis*. *Process Biochem.* **2012**, *47*, 2575–2580. [[CrossRef](#)]
103. Kim, Y.S.; Kim, Y.B.; Kim, Y.J.; Lee, M.Y.; Park, S.U. Overexpression of cinnamate 4-hydroxylase and 4-coumaroyl CoA ligase prompted flavone accumulation in *Scutellaria baicalensis* hairy roots. *Nat. Prod. Commun.* **2014**, *9*, 803–807. [[CrossRef](#)] [[PubMed](#)]
104. Park, C.H.; Xu, H.; Yeo, H.J.; Park, Y.E.; Hwang, G.-S.; Park, N.I.; Park, S.U. Enhancement of the flavone contents of *Scutellaria baicalensis* hairy roots via metabolic engineering using maize Lc and *Arabidopsis* PAP1 transcription factors. *Metab. Eng.* **2021**, *64*, 64–73. [[CrossRef](#)] [[PubMed](#)]

105. Kim, Y.S.; Li, X.; Park, W.T.; Uddin, M.R.; Park, N.I.; Kim, Y.B.; Lee, M.Y.; Park, S.U. Influence of media and auxins on growth and flavone production in hairy root cultures of baikal skullcap, *Scutellaria baicalensis*. *Plant Omics J.* **2012**, *5*, 24–27.
106. Park, C.H.; Kim, Y.S.; Li, X.; Kim, H.H.; Arasu, M.V.; Al-Dhabi, N.A.; Lee, S.Y.; Park, S.U. Influence of different carbohydrates on flavonoid accumulation in hairy root cultures of *Scutellaria baicalensis*. *Nat. Prod. Commun.* **2016**, *11*, 799–802. [[CrossRef](#)]
107. Dikaya, V.S.; Solovyeva, A.I.; Sidorov, R.A.; Solovyev, P.A.; Stepanova, A.Y. The relationship between endogenous  $\beta$ -glucuronidase activity and biologically active flavones-aglycone contents in hairy roots of Baikal skullcap. *Chem. Biodivers.* **2018**, *15*, e1700409. [[CrossRef](#)] [[PubMed](#)]
108. Solov'eva, A.I.; Evsyukov, S.V.; Sidorov, R.A.; Stepanova, A.Y. Correlation of endogenous  $\beta$ -glucuronidase activity with differentiation of in vitro cultures of *Scutellaria baicalensis*. *Acta Physiol. Plant.* **2020**, *42*, 169. [[CrossRef](#)]
109. Zhao, Q.; Cui, M.-Y.; Levsh, O.; Yang, D.; Liu, J.; Li, J.; Hill, L.; Yang, L.; Hu, Y.; Weng, J.-K.; et al. Two CYP82D enzymes function as flavone hydroxylases in the biosynthesis of root-specific 4'-deoxyflavones in *Scutellaria baicalensis*. *Mol. Plant* **2018**, *11*, 135–148. [[CrossRef](#)]
110. Elkin, Y.N.; Kulesh, N.I.; Stepanova, A.Y.; Solovieva, A.I.; Kargin, V.M.; Manyakhin, A.Y. Methylated flavones of the hairy root culture *Scutellaria baicalensis*. *J. Plant Physiol.* **2018**, *231*, 277–280. [[CrossRef](#)]
111. Wilczańska-Barska, A.; Królicka, A.; Głód, D.; Majdan, M.; Kawiak, A.; Krauze-Baranowska, M. Enhanced accumulation of secondary metabolites in hairy root cultures of *Scutellaria lateriflora* following elicitation. *Biotechnol. Lett.* **2012**, *34*, 1757–1763. [[CrossRef](#)]
112. Kim, J.K.; Kim, Y.S.; Kim, Y.J.; Uddin, M.R.; Kim, Y.B.; Kim, H.H.; Park, S.Y.; Lee, M.Y.; Chung, S.O.; Park, S.U. Comparative analysis of flavonoids and polar metabolites from hairy roots of *Scutellaria baicalensis* and *Scutellaria lateriflora*. *World J. Microbiol. Biotechnol.* **2014**, *30*, 887–892. [[CrossRef](#)]
113. Marsh, Z.; Yang, T.; Nopo-Olazabal, L.; Wu, S.; Ingle, T.; Joshee, N.; Medina-Bolivar, F. Effect of light, methyl jasmonate and cyclodextrin on production of phenolic compounds in hairy root cultures of *Scutellaria lateriflora*. *Phytochemistry* **2014**, *107*, 50–60. [[CrossRef](#)] [[PubMed](#)]
114. Tuan, P.A.; Kim, Y.S.; Kim, Y.; Thwe, A.A.; Li, X.; Park, C.H.; Lee, S.Y.; Park, S.U. Molecular characterization of flavonoid biosynthetic genes and accumulation of baicalin, baicalein, and wogonin in plant and hairy root of *Scutellaria lateriflora*. *Saudi J. Biol. Sci.* **2018**, *25*, 1639–1647. [[CrossRef](#)] [[PubMed](#)]
115. Stepanova, A.Y.; Solov'eva, A.I.; Malunova, M.V.; Salamaikina, S.A.; Panov, Y.M.; Lelishentsev, A.A. Hairy roots of *Scutellaria* spp. (Lamiaceae) as promising producers of antiviral flavones. *Molecules* **2021**, *26*, 3927. [[CrossRef](#)]
116. Kuzovkina, I.N.; Prokof'eva, M.Y.; Umralina, A.R.; Chernysheva, T.P. Morphological and biochemical characteristics of genetically transformed roots of *Scutellaria andrachnoides*. *Russ. J. Plant Physiol.* **2014**, *61*, 697–706. [[CrossRef](#)]
117. Gharari, Z.; Bagheri, K.; Danafar, H.; Sharafi, A. Enhanced flavonoid production in hairy root cultures of *Scutellaria bornmuelleri* by elicitor induced over-expression of MYB7 and FNSII2 genes. *Plant Physiol. Biochem.* **2020**, *148*, 35–44. [[CrossRef](#)]
118. Lei, W.; Tang, S.-H.; Luo, K.-M.; Sun, M. Molecular cloning and expression profiling of a chalcone synthase gene from hairy root cultures of *Scutellaria viscidula* Bunge. *Genet. Mol. Biol.* **2010**, *33*, 285–291. [[CrossRef](#)] [[PubMed](#)]
119. Gharari, Z.; Bagheri, K.; Sharafi, A. Establishment of in vitro genetically engineered cultures in *Scutellaria orientalis* and *S. araxensis*. *Biologia* **2020**, *75*, 2383–2393. [[CrossRef](#)]
120. Heydari, P.; Yavari, M.; Adibi, P.; Asghari, G.; Ghanadian, S.-M.; Dida, G.O.; Khamesipour, F. Medicinal Properties and Active Constituents of *Dracocephalum kotschy* and Its Significance in Iran: A Systematic Review. *Evid.-Based Complement. Altern. Med.* **2019**, *2019*, 9465309. [[CrossRef](#)]
121. Fattahi, M.; Nazeri, V.; Torras-Claveria, L.; Sefidkon, F.; Cusido, R.M.; Zamani, Z.; Palazon, J. A new biotechnological source of rosmarinic acid and surface flavonoids: Hairy root cultures of *Dracocephalum kotschy* Boiss. *Ind. Crops Prod.* **2013**, *50*, 256–263. [[CrossRef](#)]
122. Nourozi, E.; Hosseini, B.; Maleki, R.; Mandoulakani, B.A. Iron oxide nanoparticles: A novel elicitor to enhance anticancer flavonoid production and gene expression in *Dracocephalum kotschy* hairy-root cultures. *J. Sci. Food Agric.* **2019**, *99*, 6418–6430. [[CrossRef](#)]
123. Wang, Y.-F.; Ni, Z.-Y.; Dong, M.; Cong, B.; Shi, Q.-W.; Gu, Y.-C.; Kiyota, H. Secondary metabolites of plants from the genus *Saussurea*: Chemistry and biological activity. *Chem. Biodiv.* **2010**, *7*, 2623–2659. [[CrossRef](#)] [[PubMed](#)]
124. Zhao, T.; Li, S.-J.; Zhang, Z.-X.; Zhang, M.-L.; Shi, Q.-W.; Gu, Y.-C.; Dong, M.; Kiyota, H. Chemical constituents from the genus *Saussurea* and their biological activities. *Heterocycl. Commun.* **2017**, *23*, 331–358. [[CrossRef](#)]
125. Chu, C.; Wang, C.; Sun, C.; Hsu, C.; Yin, C.; Chu, C.; Bi, F. Establishment of an efficient medium for anther culture of rice through comparative experiments on the nitrogen sources. *Sci. Sin.* **1975**, *18*, 223–231.
126. Zhao, D.; Fu, C.; Chen, Y.; Ma, F. Transformation of *Saussurea medusa* for hairy roots and jaceosidin production. *Plant Cell Rep.* **2004**, *23*, 468–474. [[CrossRef](#)]
127. Li, F.-X.; Jin, Z.-P.; Zhao, D.-X.; Cheng, L.-Q.; Fu, C.-X.; Ma, F. Overexpression of the *Saussurea medusa* chalcone isomerase gene in *S. involucrata* hairy root cultures enhances their biosynthesis of apigenin. *Phytochemistry* **2006**, *67*, 553–560. [[CrossRef](#)]
128. Chen, R.; Chen, X.; Zhu, T.; Liu, J.; Xiang, X.; Yu, J.; Tan, H.; Gao, S.; Li, Q.; Fang, Y.; et al. Integrated transcript and metabolite profiles reveal that EbCHI plays an important role in scutellarin accumulation in *Erigeron breviscapus* hairy roots. *Front. Plant Sci.* **2018**, *9*, 789. [[CrossRef](#)]

129. Chung, I.-M.; Hong, S.-B.; Peebles, C.A.M.; Kim, J.-A.; San, K.Y. Effect of the engineered indole pathway on accumulation of phenolic compounds in *Catharanthus roseus* hairy roots. *Biotechnol. Prog.* **2007**, *23*, 327–332. [[CrossRef](#)] [[PubMed](#)]
130. Jiang, Y.N.; Wang, B.; Li, H.; Yao, L.M.; Wu, T.L. Flavonoid production is effectively regulated by RNAi interference of two flavone synthase genes from *Glycine max*. *J. Plant Biol.* **2010**, *53*, 425–432. [[CrossRef](#)]
131. Yan, J.; Wang, B.; Jiang, Y.; Cheng, L.; Wu, T. GmFNSII-Controlled soybean flavone metabolism responds to abiotic stresses and regulates plant salt tolerance. *Plant Cell Physiol.* **2014**, *55*, 74–86. [[CrossRef](#)]
132. Staszaków, A.; Swarcewicz, B.; Banasiak, J.; Muth, D.; Jasiński, M.; Stobiecki, M. LC/MS profiling of flavonoid glycoconjugates isolated from hairy roots, suspension root cell cultures and seedling roots of *Medicago truncatula*. *Metabolomics* **2011**, *7*, 604–613. [[CrossRef](#)]
133. Gai, Q.-Y.; Jiao, J.; Wang, X.; Fu, Y.-J.; Lu, Y.; Liu, J.; Wang, Z.-Y.; Xu, X.-J. Simultaneous quantification of eleven bioactive phenolic compounds in pigeon pea natural resources and in vitro cultures by ultra-high performance liquid chromatography coupled with triple quadrupole mass spectrometry (UPLC-QqQ-MS/MS). *Food Chem.* **2021**, *335*, 127602. [[CrossRef](#)] [[PubMed](#)]
134. Gai, Q.-Y.; Lu, Y.; Jiao, J.; Fu, J.-X.; Xu, X.-J.; Yao, L.; Fu, Y.-J. Application of UV-B radiation for enhancing the accumulation of bioactive phenolic compounds in pigeon pea [*Cajanus cajan* (L.) Millsp.] hairy root cultures. *J. Photochem. Photobiol. B Biol.* **2022**, *228*, 112406. [[CrossRef](#)] [[PubMed](#)]
135. Miadoková, E. Isoflavonoids—an overview of their biological activities and potential health benefits. *Interdiscip. Toxicol.* **2009**, *2*, 211–218. [[CrossRef](#)] [[PubMed](#)]
136. Sohn, S.L.; Pandian, S.; Oh, Y.J.; Kang, H.J.; Cho, W.S.; Cho, Y.S. Metabolic engineering of isoflavones: An updated overview. *Front. Plant Sci.* **2021**, *12*, 670103. [[CrossRef](#)] [[PubMed](#)]
137. Lozovaya, V.V.; Lygin, A.V.; Zernova, O.V.; Li, S.; Hartman, G.L.; Widholm, J.M. Isoflavonoid accumulation in soybean hairy roots upon treatment with *Fusarium solani*. *Plant Physiol. Biochem.* **2004**, *42*, 671–679. [[CrossRef](#)]
138. Jiang, Y.; Hu, Y.; Wang, B.; Wu, T. Bivalent RNA interference to increase isoflavone biosynthesis in soybean (*Glycine max*). *Braz. Arch. Biol. Technol.* **2014**, *57*, 163–170. [[CrossRef](#)]
139. Yan, J.; Wang, B.; Zhong, Y.; Yao, L.; Cheng, L.; Wu, T. The soybean R2R3 MYB transcription factor GmMYB100 negatively regulates plant flavonoid biosynthesis. *Plant Mol. Biol.* **2015**, *89*, 35–48. [[CrossRef](#)]
140. Jia, T.; An, J.; Liu, Z.; Yu, B.; Chen, J. Salt stress induced soybean *GmIFS1* expression and isoflavone accumulation and salt tolerance in transgenic soybean cotyledon hairy roots and tobacco. *Plant Cell Tissue Organ Cult.* **2017**, *128*, 469–477. [[CrossRef](#)]
141. Han, X.; Yin, Q.; Liu, J.; Jiang, W.; Di, S.; Pang, Y. GmMYB58 and GmMYB205 are seed-specific activators for isoflavonoid biosynthesis in *Glycine max*. *Plant Cell Rep.* **2017**, *36*, 1889–1902. [[CrossRef](#)]
142. Yin, Q.; Shen, G.; Di, S.; Fan, C.; Chang, Z.; Pang, Y. Genome-wide identification and functional characterization of UDP-glucosyltransferase genes involved in flavonoid biosynthesis in *Glycine max*. *Plant Cell Physiol.* **2017**, *58*, 1558–1572. [[CrossRef](#)]
143. Zhou, Y.; Huang, J.; Zhang, X.; Zhu, L.; Wang, X.; Guo, N.; Zhao, J.; Xing, H. Overexpression of chalcone isomerase (CHI) increases resistance against *Phytophthora sojae* in soybean. *J. Plant Biol.* **2018**, *61*, 309–319. [[CrossRef](#)]
144. Uchida, K.; Sawada, Y.; Ochiai, K.; Sato, M.; Inaba, J.; Hirai, M.Y. Identification of a unique type of isoflavone O-methyltransferase, GmIOMT1, based on multi-omics analysis of soybean under biotic stress. *Plant Cell Physiol.* **2020**, *61*, 1974–1985. [[CrossRef](#)] [[PubMed](#)]
145. Ahmad, M.Z.; Zhang, Y.; Zeng, X.; Li, P.; Wang, X.; Benedito, V.A.; Zhao, J. Isoflavone malonyl-CoA acyltransferase GmMaT2 is involved in nodulation of soybean by modifying synthesis and secretion of isoflavones. *J. Exp. Bot.* **2021**, *72*, 1349–1369. [[CrossRef](#)]
146. Theboral, J.; Sivanandhan, G.; Subramanyam, K.; Arun, M.; Selvaraj, N.; Manickavasagam, M.; Ganapathi, A. Enhanced production of isoflavones by elicitation in hairy root cultures of Soybean. *Plant Cell Tissue Organ Cult.* **2014**, *117*, 477–481. [[CrossRef](#)]
147. Liu, J.; Jiang, W.; Xia, Y.; Wang, X.; Shen, G.; Pang, Y. Genistein-specific *G6DT* gene for the inducible production of wightone in *Lotus japonicus*. *Plant Cell Physiol.* **2018**, *59*, 128–141. [[CrossRef](#)]
148. Saviranta, N.M.; Julkunen-Tiitto, R.; Oksanen, E.; Karjalainen, R.O. Red clover (*Trifolium pratense* L.) isoflavones: Root phenolic compounds affected by biotic and abiotic stress factors. *J. Sci. Food Agric.* **2010**, *90*, 418–423. [[CrossRef](#)]
149. Kumar, M.A.; Pammi, S.S.S.; Sukanya, M.S.; Giri, A. Enhanced production of pharmaceutically important isoflavones from hairy root rhizoclones of *Trifolium pratense* L. *In Vitro Cell Dev. Biol.* **2018**, *54*, 94–103. [[CrossRef](#)]
150. Reis, A.; Boutet-Mercey, S.; Massot, S.; Ratet, P.; Zuanazzi, J.A.S. Isoflavone production in hairy root cultures and plantlets of *Trifolium pratense*. *Biotechnol. Lett.* **2019**, *41*, 427–442. [[CrossRef](#)] [[PubMed](#)]
151. Park, Y.J.; Thwe, A.A.; Li, X.; Kim, Y.J.; Kim, J.K.; Arasu, M.V.; Al-Dhabi, N.A.; Park, S.U. Triterpene and flavonoid biosynthesis and metabolic profiling of hairy roots, adventitious roots, and seedling roots of *Astragalus membranaceus*. *J. Agric. Food Chem.* **2015**, *63*, 8862–8869. [[CrossRef](#)]
152. Jiao, J.; Gai, Q.-Y.; Luo, M.; Peng, X.; Zhao, C.-J.; Fu, Y.-J.; Ma, W. Direct determination of astragalosides and isoflavonoids from fresh *Astragalus membranaceus* hairy root cultures by high speed homogenization coupled with cavitation-accelerated extraction followed by liquid chromatography-tandem mass spectrometry. *RSC Adv.* **2015**, *5*, 34672–34681. [[CrossRef](#)]
153. Jiao, J.; Gai, Q.-Y.; Wang, W.; Luo, M.; Gu, C.-B.; Fu, Y.-J.; Ma, W. Ultraviolet radiation-elicited enhancement of isoflavonoid accumulation, biosynthetic gene expression, and antioxidant activity in *Astragalus membranaceus* hairy root cultures. *J. Agric. Food Chem.* **2015**, *63*, 8216–8224. [[CrossRef](#)] [[PubMed](#)]

154. Gai, Q.-Y.; Jiao, J.; Luo, M.; Wang, W.; Gu, C.-B.; Fu, Y.-J.; Ma, W. Tremendous enhancements of isoflavonoid biosynthesis, associated gene expression and antioxidant capacity in *Astragalus membranaceus* hairy root cultures elicited by methyl jasmonate. *Process Biochem.* **2016**, *51*, 642–649. [[CrossRef](#)]
155. Jiao, J.; Gai, Q.-Y.; Niu, L.-L.; Wang, X.-Q.; Guo, N.; Zang, Y.-P.; Fu, Y.-J. Enhanced production of two bioactive isoflavone aglycones in *Astragalus membranaceus* hairy root cultures by combining deglycosylation and elicitation of immobilized edible *Aspergillus niger*. *J. Agric. Food Chem.* **2017**, *65*, 9078–9086. [[CrossRef](#)]
156. Cui, Y.; Taniguchi, S.; Kuroda, T.; Hatano, T. Constituents of *Psoralea corylifolia* fruits and their effects on methicillin-resistant *Staphylococcus aureus*. *Molecules* **2015**, *20*, 12500–12511. [[CrossRef](#)] [[PubMed](#)]
157. Huang, Y.; Liu, X.; Wu, Y.; Li, Y.; Guo, F. Meroterpenes from *Psoralea corylifolia* against *Pyricularia oryzae*. *Planta Med.* **2014**, *80*, 1298–1303. [[CrossRef](#)] [[PubMed](#)]
158. Bourgaud, F.; Bouque, V.; Guckert, A. Production of flavonoids by *Psoralea* hairy root cultures. *Plant Cell Tissue Organ Cult.* **1999**, *56*, 97–104. [[CrossRef](#)]
159. Abhyankar, G.; Reddy, V.D.; Giri, C.C.; Rao, K.V.; Lakshmi, V.V.S.; Prabhakar, S.; Vairamani, M.; Thippeswamy, B.S.; Bhattacharya, P.S. Amplified fragment length polymorphism and metabolomic profiles of hairy roots of *Psoralea corylifolia* L. *Phytochemistry* **2005**, *66*, 2441–2457. [[CrossRef](#)]
160. Shinde, A.N.; Malpathak, N.; Fulzele, D.P. Impact of nutrient components on production of the phytoestrogens daidzein and genistein by hairy roots of *Psoralea corylifolia*. *J. Nat. Med.* **2010**, *64*, 346–353. [[CrossRef](#)]
161. Cherdshewasart, W.; Sutjit, W. Correlation of antioxidant activity and major isoflavonoid contents of the phytoestrogen-rich *Pueraria mirifica* and *Pueraria lobata* tubers. *Phytomedicine* **2008**, *15*, 38–43. [[CrossRef](#)]
162. Shi, H.-P.; Kintzios, S. Genetic transformation of *Pueraria phaseoloides* with *Agrobacterium rhizogenes* and puerarin production in hairy roots. *Plant Cell Rep.* **2003**, *21*, 1103–1107. [[CrossRef](#)]
163. Kintzios, S.; Makri, O.; Pistola, E.; Matakias, T.; Shi, H.P.; Economou, A. Scale-up production of puerarin from hairy roots of *Pueraria phaseoloides* in an airlift bioreactor. *Biotechnol. Lett.* **2004**, *26*, 1057–1059. [[CrossRef](#)] [[PubMed](#)]
164. Udomsuk, L.; Jarukamjorn, K.; Tanaka, H.; Putalun, W. Isoflavonoid Production in a Hairy Roots Culture of *Pueraria candollei*. *Z. Naturforsch.* **2009**, *64*, 687–691. [[CrossRef](#)] [[PubMed](#)]
165. Udomsuk, L.; Jarukamjorn, K.; Tanaka, H.; Putalun, W. Improved isoflavonoid production in *Pueraria candollei* hairy root cultures using elicitation. *Biotechnol. Lett.* **2011**, *33*, 369–374. [[CrossRef](#)] [[PubMed](#)]
166. Udomsin, O.; Yusakul, G.; Kraithong, W.; Udomsuk, L.; Kitisripanya, T.; Juengwatanatrakul, T.; Putalun, W. Enhanced accumulation of high-value deoxymiroestrol and isoflavonoids using hairy root as a sustainable source of *Pueraria candollei* var. *mirifica*. *Plant Cell Tissue Organ Cult.* **2019**, *136*, 141–151. [[CrossRef](#)]
167. Danphitsanuparn, P.; Boonsongcheep, P.; Boriboonkaset, T.; Chintapakorn, Y.; Prathanturug, S. Effects of *Agrobacterium rhizogenes* strains and other parameters on production of isoflavonoids in hairy roots of *Pueraria candollei* Grah. ex Benth. var. *candollei*. *Plant Cell Tissue Organ Cult.* **2012**, *111*, 315–322. [[CrossRef](#)]
168. Kim, S.; Cha, M.-S.; Lee, E.; Kim, I.; Kwon, J.E.; Kang, S.C.; Park, T.-H. In vitro induction of hairy root from isoflavones-producing Korean wild arrowroot *Pueraria lobata*. *J. Plant Biotechnol.* **2012**, *39*, 205–211. [[CrossRef](#)]
169. Wang, X.; Li, C.; Zhou, C.; Li, J.; Zhang, Y. Molecular characterization of the C-glucosylation for puerarin biosynthesis in *Pueraria lobata*. *Plant J.* **2017**, *90*, 535–546. [[CrossRef](#)]
170. Gampe, N.; Szakács, Z.; Darcsi, A.; Boldizsár, I.; Szóke, É.; Kuzovkina, I.; Kursinszki, L.; Béni, S. Qualitative and quantitative phytochemical analysis of *Ononis* hairy root cultures. *Front. Plant Sci.* **2021**, *11*, 622585. [[CrossRef](#)]
171. Lee, M.R. Liquorice (*Glycyrrhiza glabra*): The journey of the sweet root from Mesopotamia to England. *J. R. Coll. Physicians Edinb.* **2018**, *48*, 378–382. [[CrossRef](#)]
172. Asl, M.N.; Hosseinzadeh, H. Review of pharmacological effects of *Glycyrrhiza* sp. and its bioactive compounds. *Phytother. Res.* **2008**, *22*, 709–724. [[CrossRef](#)]
173. Asada, Y.; Li, W.; Yoshikawa, T. Isoprenylated flavonoids from hairy root cultures of *Glycyrrhiza glabra*. *Phytochemistry* **1998**, *47*, 389–392. [[CrossRef](#)]
174. Li, W.; Asada, Y.; Yoshikawa, T. Antimicrobial flavonoids from *Glycyrrhiza glabra* hairy root cultures. *Planta Med.* **1998**, *64*, 746–747. [[CrossRef](#)] [[PubMed](#)]
175. Asada, Y.; Li, W.; Yoshikawa, T. The first prenylated bioaurone, licoagrone from hairy root cultures of *Glycyrrhiza glabra*. *Phytochemistry* **1999**, *50*, 1015–1019. [[CrossRef](#)]
176. Li, W.; Asada, Y.; Yoshikawa, T. Flavonoid constituents from *Glycyrrhiza glabra* hairy root cultures. *Phytochemistry* **2000**, *55*, 447–456. [[CrossRef](#)]
177. Li, W.; Asada, Y.; Koike, K.; Hirotsu, M.; Rui, H.; Yoshikawa, T.; Nikaido, T. Flavonoids from *Glycyrrhiza pallidiflora* hairy root cultures. *Phytochemistry* **2001**, *58*, 595–598. [[CrossRef](#)]
178. Li, W.; Koike, K.; Asada, Y.; Hirotsu, M.; Rui, H.; Yoshikawa, T.; Nikaido, T. Flavonoids from *Glycyrrhiza pallidiflora* hairy root cultures. *Phytochemistry* **2002**, *60*, 351–355. [[CrossRef](#)]
179. Zhang, H.-C.; Liu, J.-M.; Lu, H.-Y.; Gao, S.-L. Enhanced flavonoid production in hairy root cultures of *Glycyrrhiza uralensis* Fisch by combining the over-expression of chalcone isomerase gene with the elicitation treatment. *Plant Cell Rep.* **2009**, *28*, 1205–1213. [[CrossRef](#)]

180. Zhang, H.-C.; Liu, J.-M.; Chen, H.-M.; Gao, C.-C.; Lu, H.-Y.; Zhou, H.; Li, Y.; Gao, S.-L. Up-regulation of licochalcone A biosynthesis and secretion by Tween 80 in hairy root cultures of *Glycyrrhiza uralensis* Fisch. *Mol. Biotechnol.* **2011**, *47*, 50–56. [[CrossRef](#)]
181. Yin, Y.-C.; Hou, J.-M.; Tian, S.-K.; Yang, L.; Zhang, Z.-X.; Li, W.-D.; Liu, Y. Overexpressing chalcone synthase (CHS) gene enhanced flavonoids accumulation in *Glycyrrhiza uralensis* hairy roots. *Bot. Lett.* **2020**, *167*, 219–231. [[CrossRef](#)]
182. Luczkiewicz, M.; Kokotkiewicz, A. *Genista tinctoria* hairy root cultures for selective production of isoliquiritigenin. *Z. Naturforsch.* **2005**, *60*, 867–875. [[CrossRef](#)]
183. Alikaridis, F.; Papadakis, D.; Pantelia, K.; Kephelas, T. Flavonolignan production from *Silybum marianum* transformed and untransformed root cultures. *Fitoterapia* **2000**, *71*, 379–384. [[CrossRef](#)]
184. Rahnama, H.; Hasanloo, T.; Shams, M.R.; Sepehrifar, R. Silymarin production by hairy root culture of *Silybum marianum* (L.) Gaertn. *Iran. J. Biotechnol.* **2008**, *6*, 113–118.
185. Chong, J.; Poutaraud, A.; Huguency, P. Metabolism and roles of stilbenes in plants. *Plant Sci.* **2009**, *177*, 143–155. [[CrossRef](#)]
186. Akinwumi, B.C.; Bordun, K.-A.M.; Anderson, H.D. Biological activities of stilbenoids. *Int. J. Mol. Sci.* **2018**, *19*, 792. [[CrossRef](#)]
187. Medina-Bolivar, F.; Condori, J.; Rimando, A.M.; Hubstenberger, J.; Shelton, K.; O’Keefe, S.F.; Bennett, S.; Dolan, M.C. Production and secretion of resveratrol in hairy root cultures of peanut. *Phytochemistry* **2007**, *68*, 1992–2003. [[CrossRef](#)] [[PubMed](#)]
188. Abbott, J.A.; Medina-Bolivar, F.; Martin, E.M.; Engelberth, A.S.; Villagarca, H.; Clausen, E.C.; Carrier, D.J. Purification of resveratrol, arachidin-1, and arachidin-3 from hairy root cultures of peanut (*Arachis hypogaea*) and determination of their antioxidant activity and cytotoxicity. *Biotechnol. Prog.* **2010**, *26*, 1344–1351. [[CrossRef](#)] [[PubMed](#)]
189. Condori, J.; Sivakumar, G.; Hubstenberger, J.; Dolan, M.C.; Sobolev, V.S.; Medina-Bolivar, F. Induced biosynthesis of resveratrol and the prenylated stilbenoids arachidin-1 and arachidin-3 in hairy root cultures of peanut: Effects of culture medium and growth stage. *Plant Physiol. Biochem.* **2010**, *48*, 310–318. [[CrossRef](#)]
190. Yang, T.; Fang, L.; Nopo-Olazabal, C.; Condori, J.; Nopo-Olazabal, L.; Balmaceda, C.; Medina-Bolivar, F. Enhanced production of resveratrol, piceatannol, arachidin-1, and arachidin-3 in hairy root cultures of peanut co-treated with methyl jasmonate and cyclodextrin. *J. Agric. Food Chem.* **2015**, *63*, 3942–3950. [[CrossRef](#)]
191. Yang, T.; Fang, L.; Rimando, A.M.; Sobolev, V.; Mockaitis, K.; Medina-Bolivar, F. A Stilbenoid-specific prenyltransferase utilizes dimethylallyl pyrophosphate from the plastidic terpenoid pathway. *Plant Physiol.* **2016**, *171*, 2483–2498. [[CrossRef](#)]
192. Yang, T.; Fang, L.; Sanders, S.; Jayanthi, S.; Rajan, G.; Podicheti, R.; Thallapuranam, S.K.; Mockaitis, K.; Medina-Bolivar, F. Stilbenoid prenyltransferases define key steps in the diversification of peanut phytoalexins. *J. Biol. Chem.* **2018**, *293*, 28–46. [[CrossRef](#)]
193. Mohammadhosseinpour, S.; Ho, L.-C.; Fang, L.; Xu, J.; Medina-Bolivar, F. Arachidin-1, a prenylated stilbenoid from peanut, induces apoptosis in triple-negative breast cancer cells. *Int. J. Mol. Sci.* **2022**, *23*, 1139. [[CrossRef](#)] [[PubMed](#)]
194. Fang, L.; Yang, T.; Medina-Bolivar, F. Production of prenylated stilbenoids in hairy root cultures of peanut (*Arachis hypogaea*) and its wild relatives *A. ipaensis* and *A. duranensis* via an optimized elicitation procedure. *Molecules* **2020**, *25*, 509. [[CrossRef](#)]
195. Brents, L.K.; Medina-Bolivar, F.; Seely, K.A.; Nair, V.; Bratton, S.M.; Nopo-Olazabal, L.; Patel, R.Y.; Liu, H.; Doerksen, R.J.; Prather, P.L.; et al. Natural prenylated resveratrol analogs arachidin-1 and -3 demonstrate improved glucuronidation profiles and have affinity for cannabinoid receptors. *Xenobiotica* **2012**, *42*, 139–156. [[CrossRef](#)]
196. Pilaisangsuree, V.; Somboon, T.; Tonglairoom, P.; Keawracha, P.; Wongsa, T.; Kongbangkerd, A.; Limmongkon, A. Enhancement of stilbene compounds and anti-inflammatory activity of methyl jasmonate and cyclodextrin elicited peanut hairy root. *Plant Cell Tissue Organ Cult.* **2018**, *132*, 165–179. [[CrossRef](#)]
197. Somboon, T.; Chayjarung, P.; Pilaisangsuree, V.; Keawracha, P.; Tonglairoom, P.; Kongbangkerd, A.; Wongkrajang, K.; Limmongkon, A. Methyl jasmonate and cyclodextrin-mediated defense mechanism and protective effect in response to paraquat-induced stress in peanut hairy root. *Phytochemistry* **2019**, *163*, 11–22. [[CrossRef](#)] [[PubMed](#)]
198. Wongshaya, P.; Chayjarung, P.; Tothong, C.; Pilaisangsuree, V.; Somboon, T.; Kongbangkerd, A.; Limmongkon, A. Effect of light and mechanical stress in combination with chemical elicitors on the production of stilbene compounds and defensive responses in peanut hairy root culture. *Plant Physiol. Biochem.* **2020**, *157*, 93–104. [[CrossRef](#)]
199. Eungsuwan, N.; Chayjarung, P.; Pankam, J.; Pilaisangsuree, V.; Wongshaya, P.; Kongbangkerd, A.; Sriphannam, C.; Limmongkon, A. Production and antimicrobial activity of trans-resveratrol, trans-arachidin-1 and trans-arachidin-3 from elicited peanut hairy root cultures in shake flasks compared with bioreactors. *J. Biotechnol.* **2021**, *326*, 28–36. [[CrossRef](#)] [[PubMed](#)]
200. Gajurel, G.; Hasan, R.; Medina-Bolivar, F. Antioxidant assessment of prenylated stilbenoid-rich extracts from elicited hairy root cultures of three cultivars of peanut (*Arachis hypogaea*). *Molecules* **2021**, *26*, 6778. [[CrossRef](#)] [[PubMed](#)]
201. Xu, C.; Yagiz, Y.; Hsu, W.-Y.; Simonne, A.; Lu, J.; Marshall, M.R. Antioxidant, antibacterial, and antibiofilm properties of polyphenols from muscadine grape (*Vitis rotundifolia* Michx.) pomace against selected foodborne pathogens. *J. Agric. Food Chem.* **2014**, *62*, 6640–6649. [[CrossRef](#)]
202. Nopo-Olazabal, C.; Hubstenberger, J.; Nopo-Olazabal, L.; Medina-Bolivar, F. Antioxidant activity of selected stilbenoids and their bioproduction in hairy root cultures of muscadine grape (*Vitis rotundifolia* Michx.). *J. Agric. Food Chem.* **2013**, *61*, 11744–11758. [[CrossRef](#)]
203. Nopo-Olazabal, C.; Condori, J.; Nopo-Olazabal, L.; Medina-Bolivar, F. Differential induction of antioxidant stilbenoids in hairy roots of *Vitis rotundifolia* treated with methyl jasmonate and hydrogen peroxide. *Plant Physiol. Biochem.* **2014**, *74*, 50–69. [[CrossRef](#)] [[PubMed](#)]

204. Tisserant, L.-P.; Aziz, A.; Jullian, N.; Jeandet, P.; Clément, C.; Courot, E.; Boitel-Conti, M. Enhanced stilbene production and excretion in *Vitis vinifera* cv Pinot Noir hairy root cultures. *Molecules* **2016**, *21*, 1703. [[CrossRef](#)]
205. Tisserant, L.-P.; Hubert, J.; Lequart, M.; Borie, N.; Maurin, N.; Pilard, S.; Jeandet, P.; Aziz, A.; Renault, J.-H.; Nuzillard, J.-M.; et al. <sup>13</sup>C NMR and LC-MS profiling of stilbenes from elicited grapevine hairy root cultures. *J. Nat. Prod.* **2016**, *79*, 2846–2855. [[CrossRef](#)] [[PubMed](#)]
206. Höll, J.; Vannozzi, A.; Czemmel, S.; D’Onofrio, C.; Walker, A.R.; Rausch, T.; Lucchin, M.; Boss, P.K.; Dry, I.B.; Bogs, J. The R2R3-MYB transcription factors MYB14 and MYB15 regulate stilbene biosynthesis in *Vitis vinifera*. *Plant Cell* **2013**, *25*, 4135–4149. [[CrossRef](#)]
207. Hosseini, S.M.; Bahramnejad, B.; Baneh, H.D.; Emamifar, A.; Goodwin, P.H. Hairy root culture optimization and resveratrol production from *Vitis vinifera* subsp. *sylvestris*. *World J. Microbiol. Biotechnol.* **2017**, *33*, 67. [[CrossRef](#)] [[PubMed](#)]
208. Hidalgo, D.; Martínez-Márquez, A.; Moyano, E.; Bru-Martínez, R.; Corchete, P.; Palazon, J. Bioconversion of stilbenes in genetically engineered root and cell cultures of tobacco. *Sci. Rep.* **2017**, *7*, 45331. [[CrossRef](#)]
209. Jiao, J.; Gai, Q.-Y.; Wang, X.; Liu, J.; Lu, Y.; Wang, Z.-Y.; Xu, X.-J.; Fu, Y.-J. Effective Production of Phenolic Compounds with Health Benefits in Pigeon Pea [*Cajanus cajan* (L.) Millsp.] Hairy Root Cultures. *J. Agric. Food Chem.* **2020**, *68*, 8350–8361. [[CrossRef](#)]
210. Gajurel, G.; Ñopo-Olazabal, I.; Hendrix, E.; Medina-Bolivar, F. Production and secretion of cajanin stilbene acid in hairy root cultures of pigeon pea (*Cajanus cajan*) co-treated with multiple elicitors. *Planta* **2022**, *11*, 834. [[CrossRef](#)]
211. Espín, J.C.; Larrosa, M.; García-Conesa, M.T.; Tomás-Barberán, F. Biological significance of urolithins, the gut microbial ellagic acid-derived metabolites: The evidence so far. *Evid.-Based Complement. Altern. Med.* **2013**, *2013*, 270418. [[CrossRef](#)]
212. Al-Harbi, S.A.; Abdulrahman, A.O.; Zamzami, M.A.; Khan, M.I. Urolithins: The gut based polyphenol metabolites of ellagitannins in cancer prevention, a review. *Front. Nutr.* **2021**, *8*, 647582. [[CrossRef](#)]
213. Vini, R.; Azeez, J.M.; Remadevi, V.; Susmi, T.R.; Ayswarya, R.S.; Sujatha, A.S.; Muraleedharan, P.; Lathika, L.M.; Sreeharshan, S. Urolithins: The colon microbiota metabolites as endocrine modulators: Prospects and perspectives. *Front. Nutr.* **2022**, *8*, 800990. [[CrossRef](#)] [[PubMed](#)]
214. Ishimaru, K.; Hirose, M.; Takahashi, K.; Koyama, K.; Shimomura, K. Tannin production in hairy root cultures of *Sanguisorba officinalis* L. *Plant Tissue Cult. Lett.* **1991**, *8*, 114–117. [[CrossRef](#)]
215. Ishimaru, K.; Shimomura, K. Tannin production in hairy root culture of *Geranium thunbergii*. *Phytochemistry* **1991**, *30*, 825–828. [[CrossRef](#)]
216. Ishimaru, K.; Yoshimatsu, K.; Yamakawa, T.; Kamada, H.; Shimomura, K. Phenolic constituents in tissue cultures of *Phyllanthus niruri*. *Phytochemistry* **1992**, *31*, 2015–2018. [[CrossRef](#)]
217. Bakkali, A.-T.; Jaziri, M.; Ishimaru, K.; Tanaka, N.; Shimomura, K.; Yoshimatsu, K.; Homes, J.; Vanhaelen, M. Tannin production in hairy root cultures of *Lawsonia inermis*. *J. Plant. Physiol.* **1997**, *151*, 505–508. [[CrossRef](#)]
218. Ono, N.N.; Bandaranayake, P.C.G.; Tian, L. Establishment of pomegranate (*Punica granatum*) hairy root cultures for genetic interrogation of the hydrolyzable tannin biosynthetic pathway. *Planta* **2012**, *236*, 931–941. [[CrossRef](#)] [[PubMed](#)]
219. Ono, N.N.; Qin, X.; Wilson, A.E.; Li, G.; Tian, L. Two UGT84 family glycosyltransferases catalyze a critical reaction of hydrolyzable tannin biosynthesis in pomegranate (*Punica granatum*). *PLoS ONE* **2016**, *11*, e0156319. [[CrossRef](#)]
220. Chang, L.; Wu, S.; Tian, L. Effective genome editing and identification of a regiospecific gallic acid 4-O-glycosyltransferase in pomegranate (*Punica granatum* L.). *Hortic. Res.* **2019**, *6*, 123. [[CrossRef](#)]