

Article

Yield, Physiological Performance, and Phytochemistry of Basil (*Ocimum basilicum* L.) under Temperature Stress and Elevated CO₂ Concentrations

T. Casey Barickman ^{1,*}, Omolayo J. Olorunwa ¹, Akanksha Sehgal ², C. Hunt Walne ², K. Raja Reddy ² and Wei Gao ³

¹ North Mississippi Research and Extension Center, Mississippi State University, Verona, MS 38879, USA; ojo26@msstate.edu

² Department of Plant and Soil Sciences, Mississippi State University, Mississippi State, MS 39762, USA; as5002@msstate.edu (A.S.); chw148@msstate.edu (C.H.W.); krreddy@pss.msstate.edu (K.R.R.)

³ USDA UVB Monitoring and Research Program, Natural Resource Ecology Laboratory, Department of Ecosystem Science and Sustainability, Colorado State University, Fort Collins, CO 80523, USA; wei.gao@colostate.edu

* Correspondence: t.c.barickman@msstate.edu; Tel.: +1-(662)-566-2201

Abstract: Early season sowing is one of the methods for avoiding yield loss for basil due to high temperatures. However, basil could be exposed to sub-optimal temperatures by planting it earlier in the season. Thus, an experiment was conducted that examines how temperature changes and carbon dioxide (CO₂) levels affect basil growth, development, and phytonutrient concentrations in a controlled environment. The experiment simulated temperature stress, low (20/12 °C), and high (38/30 °C), under ambient (420 ppm) and elevated (720 ppm) CO₂ concentrations. Low-temperature stress prompted the rapid closure of stomata resulting in a 21% decline in net photosynthesis. Chlorophylls and carotenoids decreased when elevated CO₂ interacted with low-temperature stress. Basil exhibited an increase in stomatal conductance, intercellular CO₂ concentration, apparent quantum yield, maximum photosystem II efficiency, and maximum net photosynthesis rate when subjected to high-temperature stress. Under elevated CO₂, increasing the growth temperature from 30/22 °C to 38/30 °C markedly increased the antioxidants content of basil. Taken together, the evidence from this research recommends that varying the growth temperature of basil plants can significantly affect the growth and development rates compared to increasing the CO₂ concentrations, which mitigates the adverse effects of temperature stress.

Keywords: Genovese cultivar; photosynthesis; stomatal conductance; chlorophyll; carotenoids; antioxidant defense metabolites



Citation: Barickman, T.C.; Olorunwa, O.J.; Sehgal, A.; Walne, C.H.; Reddy, K.R.; Gao, W. Yield, Physiological Performance, and Phytochemistry of Basil (*Ocimum basilicum* L.) under Temperature Stress and Elevated CO₂ Concentrations. *Plants* **2021**, *10*, 1072. <https://doi.org/10.3390/plants10061072>

Academic Editor: James Bunce

Received: 8 April 2021

Accepted: 22 May 2021

Published: 27 May 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Climate change remains an important challenge affecting the attainment of global food security as it negatively impacts the growth and development of crops. Several studies have demonstrated higher atmospheric carbon dioxide (CO₂) concentrations, extreme temperature conditions, and other extreme weather events as evidence of climate change [1,2]. Global atmospheric CO₂ is rising (above 415 ppm in 2020). It is projected by climate models to reach the range of 540 to 970 ppm by 2100 because of human activities, declining carbon sinks, and natural global cycles [3,4]. Recent climate models have also predicted that global air temperature may experience increments in the range of 1.5 and 4.5 °C in the next century due to the increasing levels of atmospheric CO₂ and other greenhouse gases at an alarming rate [1,5]. Atmospheric CO₂ and temperature are critical in the photosynthesis, physiological, and developmental processes that occur in many crops, especially C₃ crops [6,7]. Thus, anticipated increasing atmospheric CO₂ and temperature will affect several crops' growth and development, including basil (*Ocimum basilicum*). Increased

environmental fluctuations resulting from climate change have been forecasted in many agricultural regions [8], and this poses a bane to achieving sustainable food production. Hence, it is pertinent to understand the mechanisms associated with basil's response to elevated atmospheric CO₂ and temperature stress to managing future production.

Basil is an important herbaceous aromatic plant with a noteworthy contribution to enhancing cuisine nutrition, healthy living, and landscape aesthetics. Globally, a large proportion of high-quality basil is cultivated for its essential oil, dry leaves, and flowers [9,10]. Studies have revealed different curative properties of basil, such as lowering blood pressures and fevers, reducing glucose and cholesterol levels in the blood, suppressing muscle spasms and inflammation, and strengthening the body's natural activity [9].

In general, basil is widely adapted and grown throughout the globe. However, it is most suitable for warmer temperatures [11,12]. The optimum temperature for basil growth is in the range of 25 and 30 °C, while the minimum temperature at which basil can survive is 10.9 °C [11,13]. Recent evidence suggests that temperature significantly affects the growth and development of basil plants. For example, Chang et al. [11] indicated that increasing the growth temperature of basil to 30 °C culminated in the maximum rate of net photosynthesis (P_n), transpiration rate (E), and stomatal conductance (g_s), with resultant benefits on basil yields. Though basil is considered heat-tolerant, a temperature above 38 °C has been noted to cause detrimental effects on the yield, especially during the reproductive stages of development [11,14]. Temperature stress perturbs plant metabolism due to its damaging impact on the binding affinity and structure of proteins and enzymes, thereby resulting in the build-up of undesirable toxic intermediates, disjoining of diverse reactions, and increased levels of reactive oxygen species (ROS) [15]. Thus, many farmers usually resort to growing basil plants early in the growing season to avoid harmful environmental stress factors such as heat and drought, which can cause the plants to produce elevated ROS in tissues. Elevated levels of ROS can have a devastating effect on the growth, development, and production of phytonutrients. However, early-season planting increases the risk of exposing basil crops to low-temperature stress conditions.

Basil is sensitive to low temperatures, mostly below 10 °C, resulting in damage to growth and developmental processes [16]. Chilling causes brown discoloration of interveinal leaf areas, increased leaf blade thickening, decreased plant growth, reduced postharvest shelf life, and deterioration of quality and marketability [11,16]. Many studies show that cold temperature stress can also have harmful effects on the physiological traits of basil. Kalisz et al. [17] demonstrated that cold temperatures have negative impacts on basil P_n , E , and g_s , which impaired plant growth and development. Additionally, the negative impact was identified to decrease photosystem II (PSII) activity (F_v'/F_m') and variable-to-initial chlorophyll fluorescence (F_v/F_o) after a 16-day low-temperature treatment [17].

Moreover, basil leaves subjected to low-temperature stress were identified to experience chlorosis due to rapid degradation of carotenoid, chlorophyll, and antioxidant content [11,18]. It is important to note that the alteration of basil leaf pigments due to chilling stress could damage the photosynthetic apparatus, with detrimental impacts on plant growth and development. Previous studies have revealed that basil's greenness is an indicator of chlorophyll content, usually used by consumers to determine its productivity [19]. Therefore, further research is required to quantify basil's physiological response to low-temperature stress to promote the breeding of cold-tolerant genotypes of basil.

Several studies have indicated that increasing CO₂ concentrations positively impact plant growth and development, primarily because of the significant role CO₂ plays in respiration and photosynthesis [6,20]. Al Jaouni et al. [2] reported that biomass production increased by 40% along with the photosynthetic and respiratory rate of basil, which significantly improved by 80% when atmospheric CO₂ was increased from 360 to 620 ppm. The improved photosynthetic rate was attributed to the role of elevated atmospheric CO₂ in repressing the oxygenation reaction of Rubisco, leading to improved carbon gains [20]. Previous studies by Gillig et al. [21] also found that basil grown under 1500 ppm of CO₂

had a significant increase in their chlorophyll concentrations due to the accumulation of large grains of starch and non-structural carbohydrates. The levels of antioxidant defense metabolites, such as fumarate, glutamine, glutathione (GSH), ascorbic acid (ASC), phyloquinone (vitamin K1), anthocyanins (Anth), and most flavonoids and minerals were significantly improved by elevated CO₂ [2]. Hence, the beneficial impacts of elevated atmospheric CO₂ can counteract the adverse impacts of low- and high-temperature stress on basil physiology and phytochemistry. Even more than that, the benefits of improved growth rate in basil due to elevated CO₂ could facilitate harvesting early, which will aid more crop cycles each year and contribute positively to food security.

Studies have been conducted to find out the influence of elevated CO₂ and temperature stress on basil's growth and photosynthesis. However, there has been a lack of information on the interactive impacts of CO₂ and temperature on basil plants' physiological and phytochemical performance. Hence, this study aims to evaluate the interactive impacts of elevated CO₂ and temperature stress on photosynthesis parameters, carotenoids, and chlorophylls content of basil plants. Moreover, we evaluated important biochemical parameters acting as enzymatic and nonenzymatic antioxidant defense metabolites responsible for cellular osmotic adjustments in stressed plants.

2. Results

2.1. Gas Exchange Parameters

The results showed that temperature and its interaction with CO₂ significantly ($p < 0.001$) affected the P_n of the basil plant (Table 1). However, elevated CO₂ had no significant effect ($p > 0.05$) on P_n. Under ambient CO₂, high-temperature stress increased P_n by 72%, while low-temperature stress decreased the P_n by 38%. Correspondingly, under elevated CO₂, high-temperature stress increased the P_n of basil by 45%, while low-temperature stress decreased the P_n by 21%, compared to the control treatments.

Table 1. Interactive effects of temperature stress and CO₂ on net photosynthesis (P_n), stomatal conductance to water vapor (g_s), intercellular CO₂ concentration (C_i), electron transport rate (ETR), leaf transpiration rate (E), intercellular/ambient CO₂ ratio (C_i/C_a), and the maximal quantum yield of photosystem II photochemistry (F_v'/F_m'), of basil plants. Measurements were taken on the fourth/fifth fully expanded leaf of plants grown without temperature stress (Control), with low-temperature stress, and high-temperature stress at 420 and 720 ppm of CO₂ concentration between 37 and 38 days of treatment.

Treatment	P _n (μmol·m ⁻² ·s ⁻¹)	g _s (mol·m ⁻² ·s ⁻¹)	C _i (μmol·m ⁻² ·s ⁻¹)	ETR (μmol m ⁻² ·s ⁻¹)	E (mmol·m ⁻² ·s ⁻¹)	C _i /C _a ¹	F _v '/F _m '
420 PPM							
Control	24.48 c	0.38 b	295.09 d	187.33 ab	6.79 c	0.70 b	0.47 b
High Temperature	42.22 a	0.71 a	303.47 d	205.94 a	15.66 a	0.72 b	0.52 a
Low Temperature	15.20 d	0.14 c	237.64 e	146.46 bc	1.91 d	0.54 c	0.42 c
720 PPM							
Control	31.51 b	0.31 b	530.71 b	184.97 ab	6.67 c	0.74 b	0.51 a
High Temperature	35.51 b	0.63 a	597.80 a	183.19 abc	14.39 b	0.83 a	0.49 ab
Low Temperature	19.46 d	0.13 c	444.50 c	130.16 c	1.63 d	0.62 c	0.47 b
Treatment ^{2,3}	***	***	***	*	***	***	**
CO ₂	NS	NS	***	NS	NS	**	NS
Trt *CO ₂	***	NS	*	NS	NS	NS	*

¹ The measured intercellular CO₂/ambient CO₂ of LI-6400XT leaf cuvette. ² SE— standard error of the mean; P_n = 1.5044; g_s = 0.03683; C_i = 15.4158; ETR = 17.876; E = 0.3881; C_i/C_a = 0.02751; F_v'/F_m' = 0.01425. ³ NS represents non-significant $p > 0.05$; *, **, *** represent significance levels at $p \leq 0.05$, $p \leq 0.01$, and $p \leq 0.001$ respectively; within columns, values followed by the same letters are not significantly different.

Additionally, temperature treatments significantly impacted the g_s and E of basil plants (Table 1). Specifically, the g_s and E of the basil plant were significantly increased due to exposure to high-temperature stress. Under low-temperature stress, g_s and E

were reduced by 63% and 72%, respectively. Although elevated CO₂ had no significant effects ($p > 0.05$) on g_s and E , there was a decreasing trend of g_s and E at elevated CO₂ (Table 1). In contrast, the C_i concentrations of basil plants were significantly affected by both temperature and CO₂ ($p < 0.001$), with significantly higher C_i at elevated CO₂ as compared to the ambient CO₂.

The C_i/C_a was significantly different from temperature and CO₂ stresses, but there was no difference from the control treatment when CO₂ interacted with temperature stress. The photosynthetic ETR of basil reduced by 22% when exposed to low-temperature treatment and increased by 10% under high-temperature treatment compared to the control treatment. Similarly, the quantum efficiency (F_v'/F_m') was affected by temperature stress and its interaction with CO₂ (Table 1). The F_v'/F_m' was decreased at low-temperature stress at ambient CO₂, whereas F_v'/F_m' increased when interacted with elevated CO₂ at both low- and high-temperature stresses.

2.2. Chlorophyll Content and Total Carotenoids

Compared to the control treatments, low-temperature stress when interacted with elevated CO₂ caused a loss of pigment content in basil leaves by decreasing Chl a, b, and total chlorophyll (a + b) content by 1%, 12%, and 2%, respectively (Figure 1). While high-temperature stress at elevated CO₂ significantly increased Chl a, b, and total chlorophyll (a + b) by 35%, 18%, and 33%, respectively. However, temperature stress only had significant effects ($p > 0.001$) on the total xanthophyll of the basil plant (Table 2). At ambient CO₂, the total xanthophyll content was higher under heat stress, whereas it was lower under cold stress than in the control treatments. Compared to the control treatment, the carotenoid cycle (violaxanthin (V) + antheraxanthin (A) + zeaxanthin (Z)) decreased significantly under both low- and high-temperature stresses. Analogous results were observed for ZA/ZAV at elevated CO₂. Elevated CO₂ and its interaction with temperature stress significantly decreased the proportion of both lutein and neoxanthin when compared to the control treatments. However, temperature stress and its interaction with CO₂ had no significant ($p < 0.05$) effects on the β -carotene. β -carotene was atypically much lower (24%) under elevated CO₂ than in the ambient CO₂ in the present study. The concentration of violaxanthin revealed no effects with temperature or elevated CO₂ treatments.

Table 2. Interactive effects of temperature stress and CO₂ on carotenoids concentration of basil leaf tissue. Leaf samples were taken from basil plants grown without temperature stress (control), with low-temperature stress and high-temperature stress at 420 and 720 ppm of CO₂ concentration between 37 and 38 days of treatment.

Treatment	Concentration ($\mu\text{g}\cdot\text{g}^{-1}$ Dry Mass)							
	Neo ^a	Viol	Anth	Zea	Lut	β -car	Total Xan	ZA/ZAV ^b
420 ppm								
Control	276.43 a	204.25 a	68.76 ab	163.94 a	793.09 a	509.66 a	436.95 a	0.53 a
High Temperature	252.16 abc	215.89 a	58.23 b	101.84 bc	710.64 ab	464.48 ab	375.97 bc	0.43 c
Low Temperature	264.59 ab	239.62 a	74.12 a	161.19 a	669.04 b	506.33 a	474.93 a	0.50 ab
720 ppm								
Control	222.36 c	208.03 a	74.29 a	157.75 a	561.09 c	384.95 bc	440.08 a	0.53 a
High Temperature	265.46 ab	226.79 a	43.14 c	78.38 c	687.31 b	390.65 bc	348.31 c	0.35 d
Low Temperature	235.20 bc	236.26 a	62.88 ab	119.79 b	552.75 c	342.73 c	418.93 ab	0.44 bc
Treatment ^{c,d}	NS	NS	***	***	NS	NS	***	***
CO ₂	*	NS	NS	*	***	***	NS	*
Trt *CO ₂	*	NS	*	NS	*	NS	NS	NS

^a Neo—neoxanthin; Vio—violaxanthin; Anth—antheraxanthin; Zea—zeaxanthin; Lut—lutein; β -car—beta carotene; Xan—xanthophylls.

^b Xanthophyll cycle ratio = zeaxanthin to antheraxanthin/zeaxanthin to antheraxanthin to violaxanthin. ^c The standard error of mean was: Neo—14.10; Vio—14.55; Anth—7.17; Zea—11.27; Lut—35.85; Bcar—32.55; Total Xan—19.09; ZA/ZAV—0.025. ^d NS represents non-significant $p > 0.05$; *, *** represent significance levels at $p \leq 0.05$ and $p \leq 0.001$, respectively; within columns, values followed by the same letters are not significantly different.

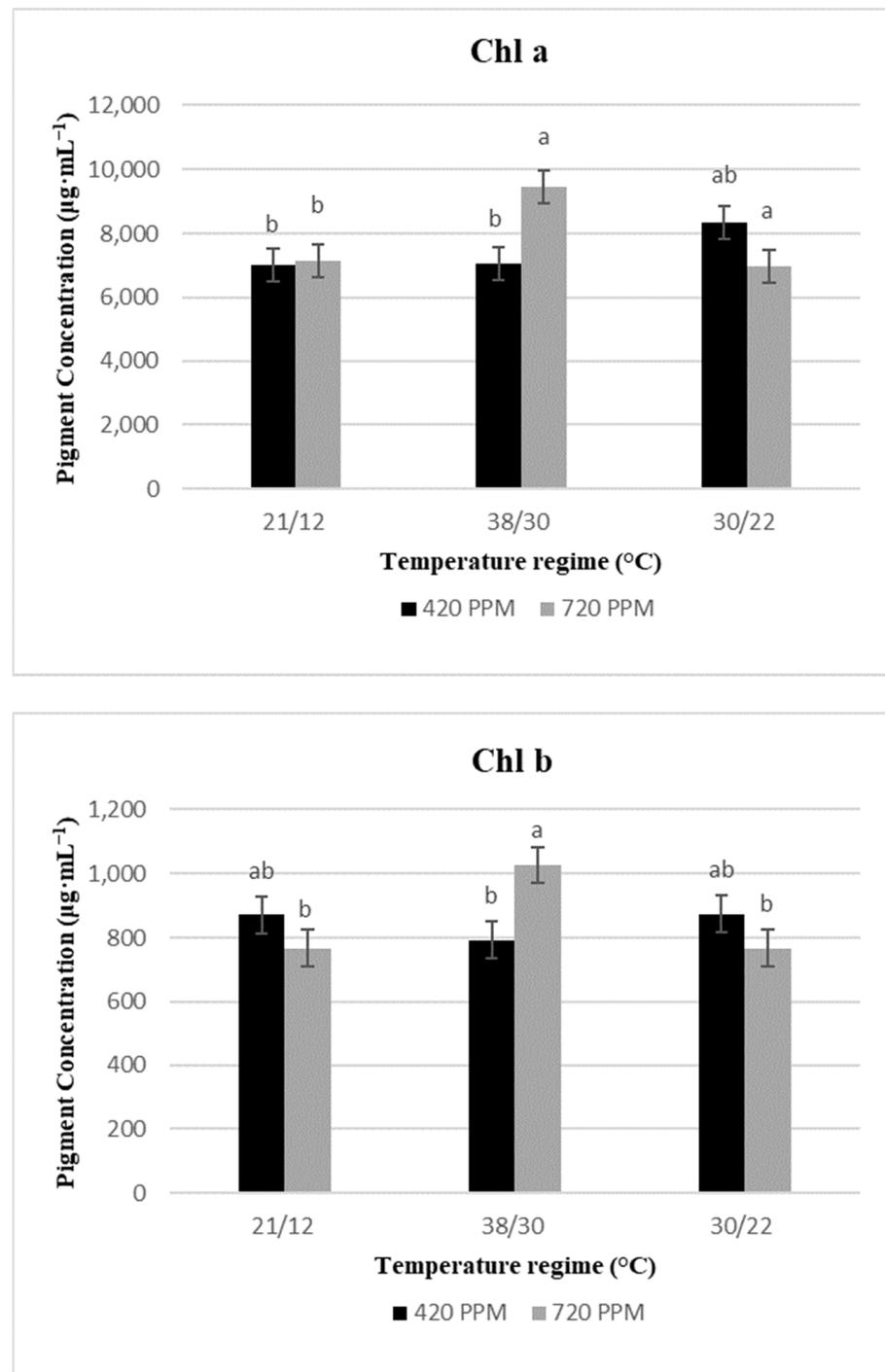


Figure 1. Cont.

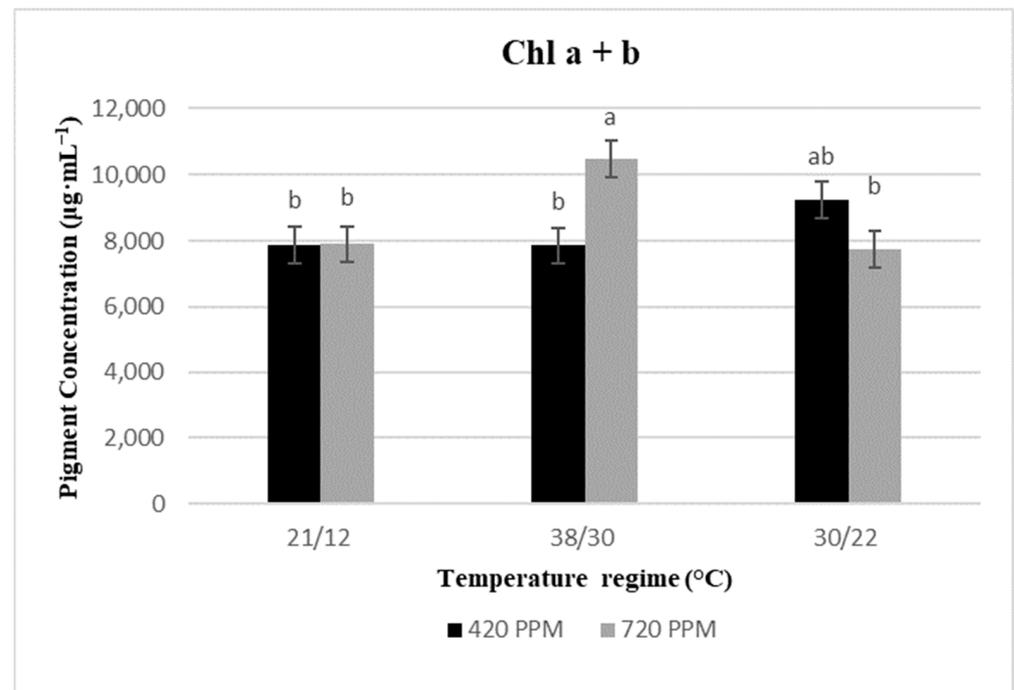


Figure 1. Chlorophyll a (Chl a), and chlorophyll b (Chl b), and total chlorophyll (Chl a + b), concentrations of basil plants under no temperature stress (Control), low-temperature stress, and high-temperature stress at 420 and 720 ppm of CO₂ concentration. The standard error mean was Chl a = 506.33, Chl b = 57.19, and Chl a + b = 547.56. Different low case letters indicate significant difference at $p < 0.05$ by least significant difference.

2.3. Total Phenolics

Conflicting with the pattern of changes of chlorophyll contents, total phenolics reduced (7%) in basil plants subjected to the interactions of high-temperature stress and elevated CO₂ (Figure 2), whereas a significant increase of 10% was observed under low temperature at elevated CO₂.

2.4. Epicuticular Wax

The basil plants showed a significant reduction in leaf wax content when subjected to both low- and high-temperature stresses and elevated CO₂ (Figure 3). However, there was no interaction effect between CO₂ and temperature treatments on basil leaf wax content.

2.5. Antioxidant and Oxidative Parameters

Interactions between temperature and CO₂ significantly affected the content of MDA and GSH only (Table 3). Basil grown at low temperature under elevated CO₂ significantly increased the MDA content by 150%, whereas the MDA content decreased by 43% under elevated CO₂ at high temperature. In contrast, the total GSH levels of basil were markedly increased by 43% under high-temperature stress at elevated CO₂. It decreased by 2% when subjected to low-temperature treatment at elevated CO₂ related to the control. There was only a small difference between the GSH content under low-temperature stress at ambient CO₂ and low-temperature stress at elevated CO₂. Compared to the control treatments, the SOD content increased significantly under elevated CO₂ at low and high-temperature stresses. Additionally, elevated CO₂ alone was discovered to increase the ASC and TRE content of basil substantially by 89% and 41%, respectively, compared to the control treatments. However, temperature and elevated CO₂ treatments and their interactions had no significant effects on H₂O₂ content.

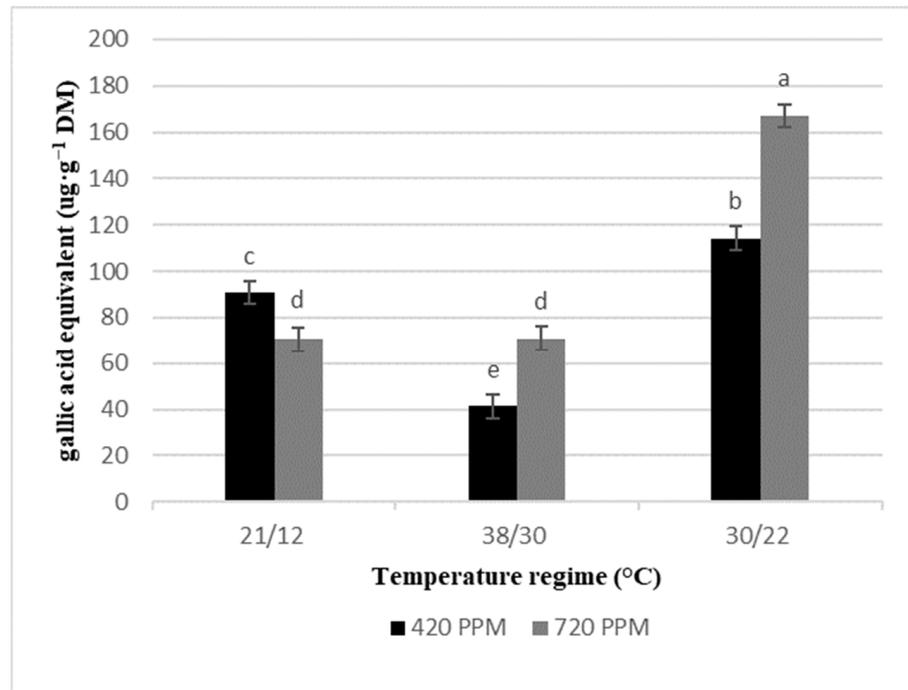


Figure 2. Total phenolic of basil leaf tissue subjected to no temperature stress (Control), low-temperature stress, and high-temperature stress at 420 and 720 ppm of CO₂ concentration. Total phenolic content is presented as gallic acid equivalent concentration $\mu\text{g}\cdot\text{g}^{-1}$ dry mass (DM). The standard error mean was total phenolic = 5.133. Different low case letters indicate significant difference at $p < 0.05$ by least significant differences.

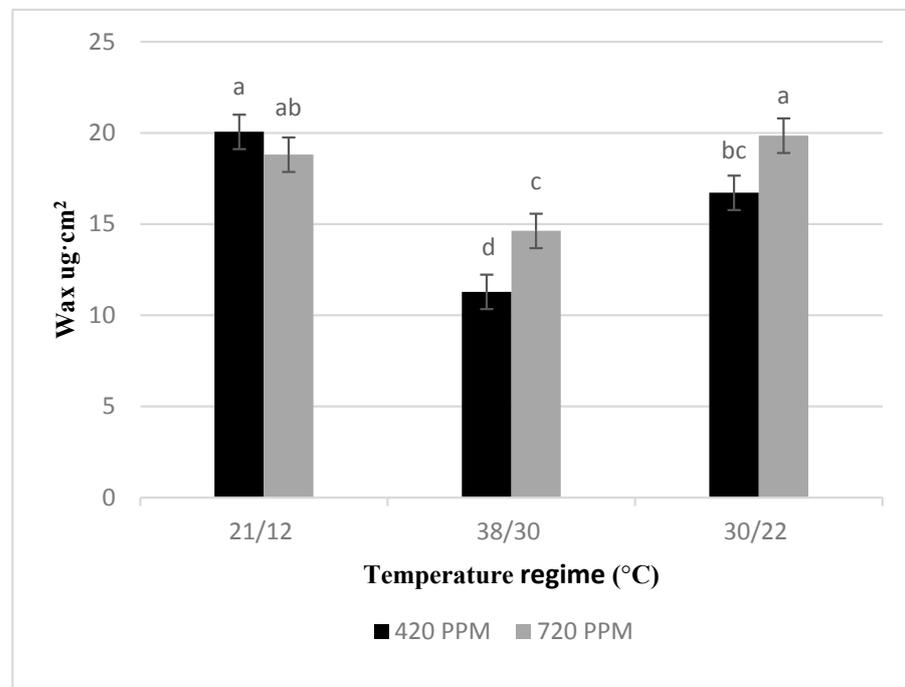


Figure 3. Average epicuticular wax content for basil plants grown without temperature stress (Control), with low-temperature stress and high-temperature stress at 420 and 720 ppm of CO₂ concentration after 34 days of treatment. The standard error mean for wax was 0.9463. Different low case letters indicate a significant difference at $p < 0.05$ by the least significant difference.

Table 3. Interactive effects of temperature stress and CO₂ on metabolites of basil leaf tissues. Leaf samples were taken from basil plants grown without temperature stress (Control), with low-temperature stress and high-temperature stress at 420 and 720 ppm of CO₂ concentration between 37 and 38 days of treatment.

Treatment	Concentration					
	nmol·g ⁻¹ DM	μmol·g ⁻¹ DM	Units/mg Protein	nmol·g ⁻¹ DM	μmol·g ⁻¹ DM	nmol·g ⁻¹ DM
	MDA ^a	H ₂ O ₂	SOD	ASC	TRE	GSH
420 ppm						
Control	0.0080 b	0.191 a	0.0304 c	0.101 b	0.0892 bc	0.192 b
High Temperature	0.0076 b	0.206 a	0.0306 c	0.123 ab	0.103 abc	0.188 b
Low Temperature	0.0074 b	0.198 a	0.0396 ab	0.106 b	0.0834 c	0.189 b
720 ppm						
Control	0.0078 b	0.183 a	0.0392 ab	0.191 a	0.126 ab	0.177 b
High Temperature	0.0046 b	0.206 a	0.0330 bc	0.150 ab	0.116 abc	0.275 a
Low Temperature	0.0200 a	0.266 a	0.0466 a	0.175 a	0.133 a	0.188 b
Treatment ^{b,c}	**	NS	**	NS	NS	*
CO ₂	NS	NS	*	**	**	NS
Trt *CO ₂	**	NS	NS	NS	NS	*

^a MDA—malondialdehyde; H₂O₂—peroxide; SOD—superoxide dismutase; ASC—ascorbic acid; TRE—trehalose; GSH—glutathione. ^b The standard error of mean was MDA = 0.001976; H₂O₂ = 0.03045; SOD = 0.00272; ASC = 0.02317; TRE = 0.01383; GSH = 0.01776. ^c NS represents non-significant $p > 0.05$; *, ** represent significance levels at $p \leq 0.05$ and $p \leq 0.01$ respectively; within columns, values followed by the same letters are not significantly different.

3. Discussion

Abiotic stress as a result of increased fluctuations of temperature and atmospheric CO₂ affects the productivity of many important crops, including basil [17,22]. In particular, extreme temperature stress during seedling growth reduced the yield of basil. Early season sowing is one of the methods for avoiding yield loss in basil due to high temperatures. However, when producers plant basil earlier in the season, the crop can be exposed to sub-optimal temperatures. Previous research has indicated that elevated CO₂ is beneficial to C3 crop growth and development [6,23]. The connection to whether increasing CO₂ from the ambient concentrations is beneficial in mitigating the adverse impacts of temperature stress in basil is yet to be determined. Hence, we evaluated the physiological and phytochemistry of basil subjected to varying temperature stress and CO₂ concentrations.

The current research indicated that P_n and g_s reduced with low-temperature stress and increased when subjected to high-temperature stress. These observations agree with the results of Ribeiro et al. [16], Kalisz et al. [17], and Balasooriya et al. [24], who revealed a negative impact of chilling temperatures on basil photosynthesis. Basil plants lower the g_s of their leaves to adapt adequately to low-temperature stress. The lower g_s are linked to preventing leaf water loss (wilting), which are directly related to decreases in P_n and C_i concentration. However, in the current study, there were no observations of wilting plants due to decreased g_s. The lower g_s and E and decreased P_n and C_i further complements the results of Saibo et al. [25], who proposed that reduced g_s are an essential factor influencing P_n decrease compared to non-stomatal limitations including reduced -Rubisco activity and -energy consumption. However, C_i/C_a was observed to reduce under low temperature, suggesting that P_n reduction could also be due to decreased C_i/C_a. The accelerated chlorophyll pigment degradation of basil plants observed when elevated CO₂ interacted with low-temperature stress could also be responsible for decreased P_n. The results of this current study match those observed in basil by Gillig et al. [21]. Basil plants subjected to low-temperature stress exhibit lower gas exchange, decreased g_s, and E, thereby explaining a decrease in the metabolic synthesis of metabolites, cell wall formation, and overall growth. Conversely, when basil plants are subjected to high-temperature stress, there is an increase in metabolic processes, gas exchanges, and morphological changes such as plant height.

It is important to remember that fluctuations in temperature have a dramatic effect on plants' metabolic functions that can lead to morphological and biochemical changes that can ultimately decrease crop yields.

Importantly, the adverse effects of the chilling temperature on P_n were ameliorated when interacted with elevated CO_2 . The increased CO_2 inside the leaf would increase leaf photosynthesis through increased activity of the rubisco enzyme and reduced photorespiration [6,24]. In contrast, the high-temperature stress was observed to increase the g_s , E , C_i , apparent quantum yield, maximum photosystem II efficiency, and finally, maximum P_n of basil plants. This observation could be linked to the positive impacts of high-temperature treatments detected on the morphological parameters of basil plants [26]. Comparable results were accounted for by Chang et al. [11]. However, elevated CO_2 did not have significant effects on g_s of basil plants in this study. Hence, proposing that the interactive impacts of elevated CO_2 and temperature stress on P_n were not linked with changes in g_s under temperature stress because reduced g_s were already instigated by low-temperature stress. Photosynthetic ETR and F_v'/F_m' also demonstrated a similar pattern with P_n and g_s . The results additionally propose that the raised P_n and g_s under high-temperature stress could be responsible for the increased ETR chain under temperature stress. F_v'/F_m' is sensitive to chilling conditions and therefore decreased F_v'/F_m' can show stress via photo-inhibition. Moreover, the occurrence of photoinhibition in basil due to chilling stress could result to increased production of ROS in basil, which causes oxidative damage in plants. Kalisz et al. [17] attributed the F_v'/F_m' reduction to the presence of photochemically inactive reaction centers and reduced ETR, which was also observed in this present study. On the contrary, elevated CO_2 interacted with temperature stress to remarkably attenuated the damage caused by chilling stress on P_n and promoted F_v'/F_m' by maintaining proper redox balance. Generally, photosynthesis is perhaps the most susceptible parameter to physiological processes, especially when C3 (basil) plants are subjected to low-temperature stress [27]. Thus, when basil plants are exposed to chilling stress, a decrease of P_n , g_s , E , and metabolites such as chlorophylls and carotenoids can have a considerable impact on plant growth and development.

Similar to most of our results, increased growth temperature from 30/22 °C to 38/30 °C under elevated CO_2 produced more chlorophyll content of basil plants. Specifically, Chl a and Chl b increased 18% and 35%, respectively. These results suggest that subjecting basil to heat stress and elevated CO_2 does not cause a drastic loss in Chl content, and this matches those observed in thermophilic plants [28,29]. Zhou et al. [30] reported that in tomato plants, a significant increase of 35% and 31% in Chl a and Chl b was observed, respectively, when the growth temperature increased from 25/20 °C to 38/30 °C. On the other hand, cold temperature stress under elevated CO_2 significantly decreased the concentrations of basil Chl a with no change in the concentration of Chl b. Corroborating the present results, Kalisz et al. [17] and Liu et al. [31] reported a significant reduction in cold-sensitive crops when subjected to chilling stress. Contrary to previous research, it can be conceivably hypothesized that elevated CO_2 does not mitigate the adverse effects of chilling stress on basil chlorophyll concentrations. The data also suggests that reduction in chlorophyll under low-temperature stress may be due to decreased metabolic functions that induce growth and development and production of metabolites to protect the plants. Conversely, under high-temperature stress conditions, growth and metabolic processes increase due to elevated P_n , g_s , and E .

Temperature is an important factor that regulates several compounds in basil that have health benefits to humans, including carotenoids, chlorophylls, phenolics, and epicuticular wax [10]. Previous research by Al-Huqail et al. [18] found that basil under any stress conditions caused a decrease in its carotenoid content. For instance, both cold and heat stress impair the xanthophyll cycle pigments of basil plants in the current study. Likewise, elevated CO_2 caused a decrease in the xanthophyll cycle of basil plants compared to control treatments. Loladze et al. [32] ascribed the negative impacts of CO_2 levels on the xanthophyll cycle of C3 plants to reduce non-photochemical quenching (NPQ) integral

photoprotection. Thus, these results may be due to more of the energy produced by increasing CO₂ levels is directed toward photosynthesis and less toward heat dissipation. However, subjecting basil plants to low-temperature stress led to a significant increase in total phenolics content. While under high-temperature treatment, total phenolics of basil decreased considerably. These findings contradict many studies [18,33,34] because increased phenolics are usually associated with thermophilic plant defense's mechanism against heat stress. Moreover, epicuticular wax decreased significantly both under heat and cold stress conditions, which is unexpected because increased wax content is always utilized as a physiological trait for selecting thermophilic plants [35].

Furthermore, the decreased Chl, P_n, g_s, E, and F_v'/F_m' observed in this study could promote a significant proportion of the light energy utilized during photosynthesis to induce excessive accumulation of ROS in basil tissues, thereby impairing the overall growth of the plant. Previous studies have shown that temperature stress increases ROS production (e.g., H₂O₂ and O₂), which causes oxidative stress in plants [36]. However, temperature and elevated CO₂ treatments and their interactions had no significant effects on H₂O₂ content in this study. Thus, suggesting that heat stress did not induce oxidative damage in basil plants, which could further support basil's tolerance to heat stress. Moreover, Al Jaouni et al. [2] demonstrated that elevated CO₂ positively impacts basil antioxidant compounds. Correspondingly, results from this study indicated that increasing the CO₂ concentration from 420 ppm to 720 ppm ameliorated the adverse effects of ROS by increasing TRE and ASC content. An increment in antioxidant enzyme activity has been described to be linked with plant resilience to heat stress [37]. It is important to note that the maximum increase was found in SOD content among the antioxidant enzymes, indicating that when basil was subjected to elevated CO₂, this enzyme had played a critical part in decreasing the damaging consequences of ROS. SOD is considered an enzymatic antioxidant curbing the harmful effects of elevated ROS by catalyzing the dismutation of superoxide radicals to H₂O₂ and O₂ [36]. Hence, the increased contents of SOD in basil plants under elevated CO₂ would reduce the toxic effects of elevated ROS levels. Additionally, GSH is a nonenzymatic antioxidant, which minimizes the damage caused by ROS in plants and protects the photosynthetic apparatus from oxidative damage [27]. Hence, basil's tolerance to heat stress could be linked to the increased GSH antioxidant levels observed in this study when high-temperature stress interacted with elevated CO₂.

Low-temperature stress frequently induces harm to cell membranes. MDA is a crucial indicator of membrane system injuries and cellular metabolism deterioration [31]. Basil grown at low temperature under elevated CO₂ significantly increased the MDA content, whereas the MDA content decreased under high temperature at elevated CO₂. This result suggests that the combination of low-temperature stress and elevated CO₂ treatments is more damaging than the interaction of heat stress and elevated CO₂. Thus, indicating increased resistance of basil to heat stress due to low levels of MDA observed.

4. Materials and Methods

4.1. Growth Condition and Plant Material

Basil 'Genovese' (Johnny's Selected Seeds, Winslow, ME) seeds were planted in polyvinyl-chloride pots (15.2 cm diameter by 30.5 cm height). The lower part of each pot was loaded up with 500 g gravel while the upper parts were filled with a mixture of sand and soil (3:1 VV) in the soil-plant-atmosphere-research (SPAR) units at the Rodney Foil Plant Science research facility of Mississippi State University, Mississippi State, MS, USA, June-July 2019. The SPAR units can control environmental conditions, including temperature and CO₂ concentration levels, at estimated set points. More information on the SPAR chamber subtleties were earlier portrayed by Reddy et al. [38] and Wijewardana et al. [39].

Six seeds previously selected by size and quality were planted in each pot, and approximately 14 days after sowing (DAS), the plants were thinned to one plant per pot. Throughout the experiment, basil plants were irrigated with full-strength Hoagland's

nutrient solution [40] three times daily (7 am, 12 pm, and 5 pm) via an automated computer-controlled drip system.

The experiment was organized in a randomized complete block design within a three by two factorial arrangement with temperature and CO₂ treatments. A total of six SPAR chambers represented three blocks with ten replications. Each SPAR chamber consisted of three rows of pots with ten pots per row in each SPAR chamber. All environmental growing conditions, except for temperature and CO₂, were kept the same throughout the experiment.

4.2. Temperature and CO₂ Treatments

Basil plants were randomly assigned to each chamber consisting of 20/12 (day/night), 30/22, and 38/30 °C in combination with ambient (420 ppm) or elevated (720 ppm) CO₂ concentrations. The day- and night-time temperatures were respectively initiated at dawn and one hour after nightfall. Table 4 shows the average environmental conditions in which the experiment was conducted. During the period of this experiment, three temperature treatments, 20/12, 30/22, and 38/30 °C, were regarded as low, optimum, and high temperatures, respectively, for basil growth and development.

Table 4. Temperature stress treatments based on the percentage of daily evapotranspiration (ET) imposed at 14 days after sowing, mean day/night temperature, mean day chamber CO₂ concentration, mean day/night vapor pressure deficit (VPD), and mean day/night evapotranspiration (ET) during the experimental period 38 days for each treatment.

Treatments		Measured Temperature (°C)	CO ₂ (ppm)	VPD (kPa)	Mean ET (H ₂ O L·d ⁻¹)
		Day/night	Day	Day/night	Day/night
Control	30/22 °C, 420 ppm	26.27 ± 0.02	430.47 ± 0.98	1.82 ± 0.01	14.64 ± 1.41
Control + High CO ₂	30/22 °C, 720 ppm	26.34 ± 0.01	731.21 ± 1.52	1.98 ± 0.01	12.60 ± 1.27
High Temperature	38/30 °C, 420 ppm	32.16 ± 0.49	434.19 ± 1.21	2.80 ± 0.07	8.74 ± 0.64
Low Temperature	20/12 °C, 420 ppm	19.53 ± 0.56	431.08 ± 0.66	0.89 ± 0.08	8.59 ± 0.47
High Temperature + High CO ₂	38/30 °C, 720 ppm	32.09 ± 0.49	728.79 ± 0.83	2.87 ± 0.07	18.41 ± 1.86
Low Temperature + High CO ₂	20/12 °C, 720 ppm	19.56 ± 0.57	724.78 ± 0.35	0.95 ± 0.09	6.39 ± 0.37

4.3. Physiology and Gas Exchange Measurements

The OJIP fluorescence readings were taken utilizing a FluorPen FP 100 (Photon Systems Instruments, Drasov, Czech Republic) on the second, most developed basil leaf. The minimal fluorescence (F_o), which was estimated at 50 μs when all PSII reaction centers are open, maximal fluorescence (F_m) when all PSII response focuses are shut, and the steady-state state fluorescence (F_s) were recorded in each plant at 17 DAT.

Analogous to the leaf chlorophyll readings, the photosynthesis and fluorescence parameters of basil leaf subjected to different treatments were recorded between 10 am and 12 pm with LI-6400XT portable photosynthesis system (LiCor Biosciences, Inc., Lincoln, NE, USA) at 17 DAT. These parameters include P_n, E, g_s, internal CO₂ concentration (C_i), electron transport rate (ETR), and the quantum efficiency (F_v'/F_m'). The internal to external CO₂ ratio was calculated by the relationship C_i/C_a. The conditions of the leaf chamber were set at light intensity (PAR) of 1500 μmol m⁻² s⁻¹, the relative humidity of 50%, the CO₂ concentration of 410 μmol mol⁻¹, and the flow rate through the chamber was regulated to 500 mol s⁻¹. The temperature of the chamber was set at the current temperatures (22, 30, or 38 °C) the readings were taken.

4.4. Carotenoid and Chlorophyll Analysis

Carotenoid and chlorophyll pigments were extracted and analyzed from freeze-dried basil tissues, according to Kopsell et al. [41,42], with few changes as portrayed in Barickman et al. [43].

4.5. Epicuticular Wax Content Determination

The epicuticular leaf waxes were extracted and quantitatively analyzed in accordance with the method of Ebercon et al. [44] with minor modifications as described by Singh and Reddy [45].

4.6. Antioxidant and Oxidative Analysis

4.6.1. Malondialdehyde (MDA)

The basil leaf samples were analyzed for MDA content by adapting the procedure used by Heath and Packer [46]. Fresh leaf tissue (500 mg) was homogenized in 0.1% trichloroacetic acid (TCA). The homogenate was centrifuged at $11,320 \times g$ for 5 min, and a 1 mL aliquot of the supernatant was treated with 4 mL 0.5% thiobarbituric acid in 20% TCA; the mixture was heated at 958°C for 30 min and afterward immediately cooled in an ice bath. After centrifugation at $5700 \times g$ for 10 min, the absorbance of the supernatant was recorded at 532 nm. The MDA content was determined by its extinction coefficient of $155 \text{ mM}^{-1} \text{ cm}^{-1}$ and expressed as $\text{nmol g}^{-1} \text{ DW}$.

4.6.2. Hydrogen Peroxide (H_2O_2)

The content of H_2O_2 was estimated according to the previously reported procedure of Mukherjee and Choudhuri [47]. Fresh leaf tissue (500 mg) was homogenized in 5 mL chilled acetone (80%) and filtered through Whatman filter paper, and 4 mL titanium reagent was added, followed by 5 mL ammonia solution. The mixture was centrifuged at $5030 \times g$, and the supernatant was disposed of. The residue was dissolved with 1 M H_2SO_4 , and the absorbance was recorded at 410 nm. The extinction coefficient of H_2O_2 is $0.28 \text{ mmol}^{-1} \text{ cm}^{-1}$. The content of H_2O_2 in samples was acquired from a standard curve using pure H_2O_2 and expressed as $\mu\text{mol g}^{-1} \text{ DW}$.

4.6.3. Superoxide Dismutase (SOD)

The SOD activity was estimated using the method of Dhindsa et al. [48] with minor modifications reported by Awasthi et al. [49].

4.6.4. Ascorbic Acid (ASC)

ASC was assessed following the method of Mukherjee and Choudhuri [47]. Details of extraction and analysis were described in Awasthi et al. [49].

4.6.5. Glutathione (GSH)

Basil leaf samples were analyzed for reduced GSH according to the method of Griffith [50], with few changes detailed in Awasthi et al. [49].

4.6.6. Trehalose (TRE)

Trehalose concentration was estimated according to the method of Trevelyan and Harrison [51] and the Anthrone method of Brin [52]. The enzymes associated with TRE metabolism were assayed as per the procedures of Pramanik and Imai [53], with few changes. Trehalose-6-phosphate synthase (TPS) activity was assayed, according to Hottiger et al. [54], which determined the release of UDP from UDP-glucose, involving glucose-6-phosphate. Trehalose-6-phosphate phosphatase (TPP) activity was assayed according to the method of Klutts et al. [55] by measuring the release of inorganic phosphate from trehalose-6-phosphate. Trehalase activity was determined by the activation of phosphorylation using cAMP (cyclic adenosine monophosphate) and assayed by measuring the glucose concentration [56].

5. Data Analysis

The experimental design was a randomized complete block in a factorial arrangement with three temperature treatments, two CO_2 treatments, three-block, and ten replications. Data were analyzed using the PROC GLIMMIX analysis of variance (ANOVA) followed by

mean separation. Statistical analysis of the data was performed using SAS (version 9.4; SAS Institute, Cary, NC, USA). The standard errors were based on the pooled error term from the ANOVA table. Duncan's multiple range test ($p \leq 0.05$) was used to differentiate between treatment classifications when F-values were significant for the main effects. Model-based values were reported rather than the unequal standard error from a data-based calculation because pooled errors reflected the statistical testing. Diagnostic tests were conducted to ensure that treatment variances were statistically equal before pooling.

6. Conclusions

The interaction of temperature stress and elevated CO₂ significantly impacted the physiological processes and phytonutrient concentrations of basil plants. Decreasing the basil's growth temperature to 20/12 °C significantly reduced P_n and g_s, with detrimental impacts on the basil plants' growth. Furthermore, low-temperature stress-induced excessive ROS production, which caused harm to the photosynthetic apparatus, as confirmed by reduced F_v'/F_m', low ETR, and altered oxidized and reduced states of PSII and PSI. Additionally, the accelerated chlorophyll and carotenoid pigment degradation of basil plants were observed when elevated CO₂ interacted with low-temperature stress.

Contrarily, elevated CO₂ remarkably ameliorated the damage caused by low-temperature stress to photosynthetic apparatus. Thus, elevated CO₂ promoted leaf C_i/C_a and increased SOD, TRE, and ASC antioxidant levels. Basil, being a thermophilic plant, was observed to increase its chlorophyll and carotenoid concentration, apparent quantum yield, and maximum photosystem II efficiency when subjected to high-temperature stress. Likewise, increased growth temperature for basil under elevated CO₂ produced more antioxidant content. The findings of this study recommend that varying the growth temperature of basil plants would significantly affect the growth and development rates of basil compared to increasing the CO₂ concentrations, which mitigated the constraining effects of temperature stress.

Author Contributions: T.C.B., Conceptualization, Methodology, Validation, Formal Analysis, Investigation, Resources, Data curation, Writing—Original Draft, Writing—Review and Editing, Visualization, Supervision, Project Administration, Funding Acquisition; O.J.O., Formal Analysis, Writing—Original Draft, Writing—Review and Editing; A.S., Methodology, Validation, Investigation; C.H.W., Methodology, Validation, Formal Analysis, Investigation; K.R.R., Conceptualization, Methodology, Validation, Formal Analysis, Investigation, Resources, Data curation, Writing—Review and Editing, Visualization, Supervision, Project Administration, Funding Acquisition; W.G., Conceptualization, Methodology, Validation, Resources, Funding Acquisition. All authors have read and agreed to the published version of the manuscript.

Funding: This material is based on the work supported by the USDA-NIFA Hatch Project under accession number 149,210 and the National Institute of Food and Agriculture, 2019-34263-30552, and MIS 043,050 funded this research.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Acknowledgments: We thank David Brand and Thomas Horgan for their technical assistance and graduate students at the Vegetable Physiology Laboratory and the Environmental Plant Physiology Laboratory for helping during data collection.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Field, C.B.; Barros, V.R.; Dokken, D.J.; Mach, K.J.; Mastrandrea, M.D.; Bilir, T.E.; Chatterjee, M.; Ebi, K.L.; Estrada, Y.O.; Genova, R.C.; et al. *IPCC. Climate Change 2014: Impacts, Adaptation, and Vulnerability*; Part A: Global and Sectoral Aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change; Field, C.B., Barros, V.R., Dokken, D.J., Mach, K.J., Mastrandrea, M.D., Bilir, T.E., Chatterjee, M., Ebi, K.L., Estrada, Y.O., Genova, R.C., et al., Eds.; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2014; p. 1132.
- Al Jaouni, S.; Saleh, A.M.; Wadaan, M.A.M.; Hozzein, W.N.; Selim, S.; AbdElgawad, H. Elevated CO₂ induces a global metabolic change in basil (*Ocimum Basilicum* L.) and peppermint (*Mentha Piperita* L.) and improves their biological activity. *J. Plant Physiol.* **2018**, *224–225*, 121–131. [[CrossRef](#)] [[PubMed](#)]
- Prentice, I.C.; Farquhar, G.D.; Fasham, M.J.R.; Goulden, M.L.; Heimann, M.; Jaramillo, V.J.; Khesghi, H.S.; Le Quéré, C.; Scholes, R.J.; Wallace, D.W.R. The carbon cycle and atmospheric carbon dioxide. In *Climate Change 2001: The Scientific Basis*; Cambridge University Press: Cambridge, UK, 2001; pp. 183–237.
- Stocker, T.F.; Qin, D.; Plattner, G.-K.; Tignor, M.; Allen, S.K.; Boschung, J.; Nauels, A.; Xia, Y.; Bex, V.; Midgley, P.M. *Climate Change 2013: The Physical Science Basis*; Intergovernmental Panel on Climate Change, Working Group I Contribution to the IPCC Fifth Assessment Report (AR5); Cambridge University Press: New York, NY, USA, 2013; p. 25.
- USGCRP. *Impacts, Risks, and Adaptation in the United States: Fourth National Climate Assessment, Volume II*; Reidmiller, D.R., Avery, C.W., Easterling, D.R., Kunkel, K.E., Lewis, K.L.M., Maycock, T.K., Stewart, B.C., Eds.; U.S. Global Change Research Program: Washington, DC, USA, 2018; p. 1515. [[CrossRef](#)]
- Reddy, A.R.; Reddy, K.R.; Hodges, H.F. Interactive effects of elevated carbon dioxide and growth temperature on photosynthesis in cotton leaves. *Plant Growth Regul.* **1998**, *26*, 33–40. [[CrossRef](#)]
- Dong, J.; Gruda, N.; Li, X.; Tang, Y.; Zhang, P.; Duan, Z. Sustainable vegetable production under changing climate: The impact of elevated CO₂ on yield of vegetables and the interactions with environments—A review. *J. Clean. Prod.* **2020**, *253*, 119920. [[CrossRef](#)]
- Solomon, S.; Manning, M.; Marquis, M.; Qin, D. *Climate Change 2007—The Physical Science Basis: Working Group I Contribution to the Fourth Assessment Report of the IPCC*; Cambridge University Press: Cambridge, UK, 2007; Volume 4.
- Simon, J.E.; Quinn, J.; Murray, R.G. Basil: A Source of Essential Oils. In *Adv. New Crops*; Janick, J., Simon, J.E., Eds.; Timber Press: Portland, OR, USA, 1990; pp. 484–489.
- Kopsell, D.A.; Kopsell, D.E.; Curran-Celentano, J. Carotenoid and chlorophyll pigments in sweet basil grown in the field and greenhouse. *Hort. Sci.* **2005**, *40*, 1230–1233. [[CrossRef](#)]
- Chang, X.; Alderson, P.; Wright, C. Effect of temperature integration on the growth and volatile oil content of basil (*Ocimum Basilicum* L.). *J. Hortic. Sci. Biotechnol.* **2005**, *80*, 593–598. [[CrossRef](#)]
- Hiltunen, R.; Holm, Y. Essential oil of *Ocimum*. In *Basil: The Genus Ocimum*; Hiltunen, R., Holm, Y., Eds.; Harwood Academic Publishers: Amsterdam, The Netherlands, 1999; pp. 113–135.
- Kumar, B.; Gupta, E.; Yadav, R.; Singh, S.C.; Lal, R.K. Temperature effects on seed germination potential of holy basil (*Ocimum tenuiflorum*). *Seed Technol.* **2014**, *36*, 75–79.
- Mortensen, L.M. The effect of air temperature on growth of eight herb species. *Am. J. Plant Sci.* **2014**, *5*, 1542–1546. [[CrossRef](#)]
- Taiz, L.; Zeiger, E.; Møller, I.M.; Murphy, A. *Plant Physiology and Development*, 6th ed.; Sinauer Associates: Sunderland, MA, USA, 2015; ISBN 978-1-60535-255-8.
- Ribeiro, P.; Simon, J.E. Breeding sweet basil for chilling tolerance. In *Issues in New Crops and New Uses*; Janick, J., Whipkey, A., Eds.; ASHS Press: Alexandria, VA, USA, 2007; pp. 302–305.
- Kalisz, A.; Jezdinsky, A.; Pokluda, R.; Sekara, A.; Grabowska, A.; Gil, J. Impacts of chilling on photosynthesis and chlorophyll pigment content in juvenile basil cultivars. *Hortic. Environ. Biotechnol.* **2016**, *57*, 330–339. [[CrossRef](#)]
- Al-Huqail, A.; El-Dakak, R.M.; Sanad, M.N.; Badr, R.H.; Ibrahim, M.M.; Soliman, D.; Khan, F. Effects of climate temperature and water stress on plant growth and accumulation of antioxidant compounds in sweet basil (*Ocimum basilicum* L.) leafy vegetable. *Scientifica* **2020**, 3808909. [[CrossRef](#)]
- Singh, H.; Poudel, M.R.; Dunn, B.L.; Fontanier, C.; Kakani, G. Effect of greenhouse CO₂ supplementation on yield and mineral element concentrations of leafy greens grown using nutrient film technique. *Agronomy* **2020**, *10*, 323. [[CrossRef](#)]
- Watanabe, C.K.; Sato, S.; Yanagisawa, S.; Uesono, Y.; Terashima, I.; Noguchi, K. Effects of elevated CO₂ on levels of primary metabolites and transcripts of genes encoding respiratory enzymes and their diurnal patterns in *Arabidopsis thaliana*: Possible relationships with respiratory rates. *Plant Cell Physiol.* **2014**, *55*, 341–357. [[CrossRef](#)] [[PubMed](#)]
- Gillig, S.; Heinemann, R.; Hurd, G.; Pittore, K.; Powell, D. Response of basil (*Ocimum basilicum*) to increased CO₂ levels. In *E&ES359 Global Climate Change, Johan Varekamp*; Wesleyan University: Middletown, CT, USA, 2008.
- Walters, K.J.; Currey, C.J. Growth and development of basil species in response to temperature. *HortScience* **2019**, *54*, 1915–1920. [[CrossRef](#)]
- Brand, D.; Wijewardana, C.; Gao, W.; Reddy, K.R. Interactive effects of carbon dioxide, low temperature, and ultraviolet-B radiation on cotton seedling root and shoot morphology and growth. *Front. Earth Sci.* **2016**, *10*, 607–620. [[CrossRef](#)]
- Balasoorya, H.N.; Dassanayake, K.B.; Seneweera, S.; Ajlouni, S. Interaction of elevated carbon dioxide and temperature on strawberry (*Fragaria* × *Ananassa*) growth and fruit yield. *Int. J. Biol. Biomol. Agric. Food Biotechnol. Eng. World Acad. Sci. Eng. Technol. Int. Sci. Index* **2018**, *12*, 279–287. [[CrossRef](#)]

25. Saibo, N.J.M.; Lourenço, T.; Oliveira, M.M. Transcription factors and regulation of photosynthetic and related metabolism under environmental stresses. *Ann. Bot.* **2009**, *103*, 609–623. [[CrossRef](#)] [[PubMed](#)]
26. Barickman, T.C.; Olorunwa, O.J.; Sehgal, A.; Walne, C.H.; Reddy, K.R.; Gao, W. Interactive impacts of temperature and elevated CO₂ on Basil (*Ocimum Basilicum* L.) root and shoot morphology and growth. *Horticulturae* **2021**, *7*, 112. [[CrossRef](#)]
27. Yuan, L.; Yuan, Y.; Liu, S.; Wang, J.; Zhu, S.; Chen, G.; Hou, J.; Wang, C. Influence of high temperature on photosynthesis, antioxidative capacity of chloroplast, and carbon assimilation among heat-tolerant and heat-susceptible genotypes of nonheading chinese cabbage. *HortScience* **2017**, *52*, 1464–1470. [[CrossRef](#)]
28. Camejo, D.; Rodríguez, P.; Angeles Morales, M.; Miguel Dell’Amico, J.; Torrecillas, A.; Alarcón, J.J. High temperature effects on photosynthetic activity of two tomato cultivars with different heat susceptibility. *J. Plant Physiol.* **2005**, *162*, 281–289. [[CrossRef](#)]
29. Wahid, A.; Gelani, S.; Ashraf, M.; Foolad, M.R. Heat tolerance in plants: An overview. *Environ. Exp. Bot.* **2007**, *61*, 199–223. [[CrossRef](#)]
30. Zhou, R.; Yu, X.; Li, X.; Mendanha dos Santos, T.; Rosenqvist, E.; Ottosen, C.-O. Combined high light and heat stress induced complex response in tomato with better leaf cooling after heat priming. *Plant Physiol. Biochem.* **2020**, *151*, 1–9. [[CrossRef](#)]
31. Liu, W.; Yu, K.; He, T.; Li, F.; Zhang, D.; Liu, J. The low temperature induced physiological responses of avena nuda L., a cold-tolerant plant species. *Sci. World J.* **2013**, *2013*, 658793. [[CrossRef](#)]
32. Loladze, I.; Nolan, J.M.; Ziska, L.H.; Knobbe, A.R. Rising atmospheric CO₂ lowers concentrations of plant carotenoids essential to human health: A Meta-Analysis. *Mol. Nutr. Food Res.* **2019**, *63*, 1801047. [[CrossRef](#)]
33. Shamloo, M.; Babawale, E.A.; Furtado, A.; Henry, R.J.; Eck, P.K.; Jones, P.J.H. Effects of genotype and temperature on accumulation of plant secondary metabolites in canadian and australian wheat grown under controlled environments. *Sci. Rep.* **2017**, *7*, 9133. [[CrossRef](#)]
34. Sublett, W.L.; Barickman, T.C.; Sams, C.E. Effects of elevated temperature and potassium on biomass and quality of dark red ‘lollo rosso’ lettuce. *Horticulturae* **2018**, *4*, 11. [[CrossRef](#)]
35. Jumrani, K.; Bhatia, V.S. Interactive effect of temperature and water stress on physiological and biochemical processes in soybean. *Physiol. Mol. Biol. Plants* **2019**, *25*, 667–681. [[CrossRef](#)]
36. Gill, S.S.; Tuteja, N. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiol. Biochem.* **2010**, *48*, 909–930. [[CrossRef](#)] [[PubMed](#)]
37. Duan, M.; Feng, H.-L.; Wang, L.-Y.; Li, D.; Meng, Q.-W. Overexpression of thylakoidal ascorbate peroxidase shows enhanced resistance to chilling stress in tomato. *J. Plant Physiol.* **2012**, *169*, 867–877. [[CrossRef](#)] [[PubMed](#)]
38. Reddy, K.; Read, J.J.; McKinion, J.M. Soil-Plant-Atmosphere-Research (SPAR) facility: A tool for plant research and modeling. *Biotronics* **2001**, *30*, 27–50.
39. Wijewardana, C.; Hock, M.; Henry, B.; Reddy, K.R. Screening corn hybrids for cold tolerance using morphological traits for early-season seeding. *Crop Sci.* **2015**, *55*, 851–867. [[CrossRef](#)]
40. Hoagland, D.R.; Arnon, D.I. *The Water-Culture Method for Growing Plants without Soil*, 2nd ed.; Circular 347; California Agricultural Experiment Station: Berkeley, CA, USA, 1950; p. 347.
41. Kopsell, D.A.; Kopsell, D.E.; Lefsrud, M.G.; Curran-Celentano, J.; Dukach, L.E. Variation in lutein, β-carotene, and chlorophyll concentrations among brassica oleracea cultivars and seasons. *HortScience* **2004**, *39*, 361–364. [[CrossRef](#)]
42. Kopsell, D.A.; Kopsell, D.E.; Curran-Celentano, J. carotenoid pigments in kale are influenced by nitrogen concentration and form. *J. Sci. Food Agric.* **2007**, *87*, 900–907. [[CrossRef](#)]
43. Barickman, T.C.; Kopsell, D.A.; Sams, C.E. Abscisic acid impacts tomato carotenoids, soluble sugars, and organic acids. *HortScience* **2016**, *51*, 370–376. [[CrossRef](#)]
44. Ebercon, A.; Blum, A.; Jordan, W.R. A rapid colorimetric method for epicuticular wax content of sorghum leaves. *Crop Sci.* **1977**, *17*, 179–180. [[CrossRef](#)]
45. Singh, S.K.; Reddy, K.R. Regulation of photosynthesis, fluorescence, stomatal conductance and water-use efficiency of cowpea (*Vigna Unguiculata* [L.] Walp.) under drought. *J. Photochem. Photobiol. B Biol.* **2011**, *105*, 40–50. [[CrossRef](#)]
46. Heath, R.L.; Packer, L. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Arch. Biochem. Biophys.* **1968**, *125*, 189–198. [[CrossRef](#)]
47. Mukherjee, S.P.; Choudhuri, M.A. Implications of water stress-induced changes in the levels of endogenous ascorbic acid and hydrogen peroxide in vigna seedlings. *Physiol. Plant.* **1983**, *58*, 166–170. [[CrossRef](#)]
48. Dhindsa, R.S.; Plumb-Dhindsa, P.; Thorpe, T.A. Leaf senescence: Correlated with increased levels of membrane permeability and lipid peroxidation, and decreased levels of superoxide dismutase and catalase. *J. Exp. Bot.* **1981**, *32*, 93–101. [[CrossRef](#)]
49. Awasthi, R.; Gaur, P.; Turner, N.C.; Vadez, V.; Siddique, K.H.M.; Nayyar, H. Effects of individual and combined heat and drought stress during seed filling on the oxidative metabolism and yield of chickpea (*Cicer Arietinum*) genotypes differing in heat and drought tolerance. *Crop Past. Sci.* **2017**, *68*, 823–841. [[CrossRef](#)]
50. Griffith, O.W. Determination of glutathione and glutathione disulfide using glutathione reductase and 2-vinylpyridine. *Anal. Biochem.* **1980**, *106*, 207–212. [[CrossRef](#)]
51. Trevelyan, W.E.; Harrison, J.S. Studies on yeast metabolism. Fractionation and microdetermination of cell carbohydrates. *Biochem. J.* **1952**, *50*, 298–303. [[CrossRef](#)]
52. Brin, M. [89] Transketolase: Clinical aspects. In *Methods in Enzymology*; Academic Press: Cambridge, MA, USA, 1966; Volume 9, pp. 506–514. ISBN 0076-6879.

53. Pramanik, H.R.; Imai, R. Functional identification of a trehalose 6-phosphate phosphatase gene that is involved in transient induction of trehalose biosynthesis during chilling stress in rice. *Plant Mol. Biol.* **2005**, *58*, 751–762. [[CrossRef](#)] [[PubMed](#)]
54. Hottiger, T.; Boller, T.; Wiemken, A. Rapid changes of heat and desiccation tolerance correlated with changes of trehalose content in *Saccharomyces cerevisiae* cells subjected to temperature shifts. *FEBS Lett.* **1987**, *220*, 113–115. [[CrossRef](#)]
55. Klutts, S.; Pastuszak, I.; Edavana, V.K.; Thampi, P.; Pan, Y.-T.; Abraham, E.C.; Carroll, J.D.; Elbein, A.D. Purification, cloning, expression, and properties of mycobacterial trehalose-phosphate phosphatase. *J. Biol. Chem.* **2003**, *278*, 2093–2100. [[CrossRef](#)] [[PubMed](#)]
56. Einig, W.; Hampp, R. Carbon partitioning in Norway spruce: Amounts of Fructose 2, 6-bisphosphate and of intermediates of starch/sucrose synthesis in relation to needle age and degree of needle loss. *Trees* **1990**, *4*, 9–15. [[CrossRef](#)]