



Neuroprotective Roles of the Biliverdin Reductase-A/Bilirubin Axis in the Brain

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Abstract: Biliverdin reductase-A (BVRA) is a multi-functional enzyme with a multitude of important roles in physiologic redox homeostasis. Classically, BVRA is well known for converting the heme metabolite biliverdin to bilirubin, which is a potent antioxidant in both the periphery and the brain. However, BVRA additionally participates in many neuroprotective signaling cascades in the brain that preserve cognition. Here, we review the neuroprotective roles of BVRA and bilirubin in the brain, which together constitute a BVRA/bilirubin axis that influences healthy aging and cognitive function.

Keywords: bilirubin; biliverdin reductase; superoxide; lipophilic antioxidant; cognition

1. Introduction

Biliverdin reductase (BVR) was discovered in 1936 when Lemberg and Wyndham observed that crude liver extracts convert green biliverdin, the product of heme metabolism, into a yellow colored substance [1]. They termed this "biliverdin reductase activity." Much later, in 1965, the enzyme responsible for this activity was partially purified from guinea pig liver [2]. Subsequently, in 1981 and 1993, Maines and colleagues purified BVR to homogeneity from rat and human liver, respectively, and fully characterized its enzymatic activity [3,4].

Today, BVR is well known as a key enzyme in heme catabolism. Specifically, after heme oxygenases cleave heme to biliverdin, with a concomitant release of iron and carbon monoxide, BVR rapidly reduces biliverdin to bilirubin in a reaction coupled to the oxidation of the pyridine nucleotide cofactors NADH and NADPH (Figure 1). An intriguing feature of BVR is that it utilizes these dual cofactors at two different pH optima: NADH at pH 6.7 and NADPH at pH 8.7 [3,4]. This dual pH usage allows BVR to function in different cellular compartments in varying physiological states of altered pH [5]. BVR exists as two functional isoforms, BVRA and BVRB, with BVRB expression emerging at 14–15 weeks of age and BVRA expression not until closer to 20 weeks [6]. BVRB is also predominantly expressed in fetal tissues, while BVRA is the predominant form present in adults and the major enzyme for bilirubin production across our lifespan [7,8]. Importantly, bilirubin



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). is cytoprotective, with in vitro and in vivo antioxidant activity [8–15], and BVRA also possesses multiple activities that play additional roles in cellular protection.



Figure 1. Heme metabolism requires NADPH oxidation. Conversion of cytosolic heme to biliverdin by heme oxygenase (HO) and cytochrome P450 reductase (CPR), both tethered to the endoplasmic reticulum membrane, involves NADPH oxidation to NADP+. Subsequent conversion of biliverdin to bilirubin by biliverdin reductase (BVR) also depends on NADPH oxidation to NADP+. BVR plays multiple additional roles in cellular physiology as well.

Although bilirubin is one of the most frequently measured blood metabolites in a clinical setting, its physiological functions are still not completely understood. To date, most studies on bilirubin in the brain have focused on peripheral body systems and the consequences of accumulating toxic levels of bilirubin in the brain, while the role of bilirubin in protecting the brain at normal physiologic levels has been largely ignored. Here, we review the neuroprotective roles of BVRA and bilirubin, with specific focus on cognitive function in neurodegenerative conditions.

2. Bilirubin in the Brain: The Good, the Bad, and the Ugly

The fact that bilirubin production has been retained by a diverse set of organisms despite its high energetic cost suggests important physiologic functions. The production of bilirubin is inducible and tightly controlled, and it exhibits circadian rhythms and is spatially and temporally controlled [16,17]. Upon the breakdown of heme, bilirubin initially exists in its indirect (unconjugated) albumin-bound form, rendering it lipid-soluble and water-insoluble. For bilirubin to be excreted into bile for elimination from the body, it must be converted to its direct (conjugated) water-soluble and less lipid-soluble form. This occurs via conjugation in the liver to glucuronic acid by uridine diphosphate glucuronosyltransferase (UGT1A1) [18]. Also in the liver, microsomal cytochrome P450 2A5 (CYP2A5) acts as an inducible bilirubin oxidase to convert bilirubin to biliverdin [19]. Notably, unconjugated bilirubin is yellow, while conjugated bilirubin is not. Clinically, unconjugated bilirubin is elevated in hemolytic anemia, as well as various inherited disorders such as Gilbert syndrome and Crigler–Najjar syndromes type I and II. Elevated conjugated bilirubin, by contrast, is found in disorders such as obstructive jaundice and biliary atresia. Due to differences in lipid solubility, unconjugated bilirubin passes across the blood-brain barrier (BBB), while conjugated bilirubin cannot.

Bilirubin predominantly circulates in the blood in its unconjugated form, with only a minor (0.01%) conjugated fraction [20]. Historically, increased levels of bilirubin have been traditionally discussed in terms of toxicity. However, normal and mildly elevated levels of bilirubin also serve a less well-recognized normal protective role. For example, mild biliru-

bin elevation in Gilbert syndrome (GS), which is caused by variants in the gene encoding UGT1A1, is correlated with a reduced incidence of kidney and cardiovascular disease, diabetes, metabolic syndrome, and some forms of cancer [21-23]. Additionally, some patients with GS also display improved antioxidant status and decreased levels of pro-inflammatory and pro-oxidant markers, including interleukin 6 (IL-6), IL-1β, apolipoprotein B (Apo B), and C-reactive protein (CRP), compared to study subjects with normal bilirubin levels. The protective effect of bilirubin is especially prominent in the fourth to sixth decade of life [24]. By contrast, the much higher plasma and tissue bilirubin levels in Crigler–Najjar syndrome (CN syndrome), which is characterized by myriad genetic aberrations that result in a near total lack of UGT1A1 activity, are toxic [25,26]. Here, the resulting accumulation of unconjugated bilirubin leads to encephalopathy (kernicterus) and bilirubin-induced neurologic damage (BIND) [27,28]. The consequences can be extreme, such as with CN syndrome type I, which appears shortly after birth, with serum bilirubin approaching 20 to 50 mg/dl, and causes encephalopathy and death. Complications caused by neonatal hyperbilirubinemia, collectively termed kernicterus spectrum disorder (KSD), encompass bilirubin neurotoxicity, acute bilirubin encephalopathy (ABE), chronic bilirubin encephalopathy (CBE), kernicterus, and BIND [27]. The symptoms of KSD generally include impaired motor control and abnormal movements, auditory processing disturbance that may be accompanied by hearing loss, impaired oculomotor function with paralysis of the upward vertical gaze (setting sun sign), and enamel dysplasia of deciduous (baby) teeth [28]. Hyperbilirubinemia can be controlled by aggressive phototherapy to isomerize unconjugated bilirubin to render it excretable from the body, bypassing the need for hepatic conjugation, or by exchange transfusions in neonates [29]. It has been speculated that an isomerization of bilirubin may reduce its lipophilicity and decrease its ability to cross the blood-brain barrier (BBB), but this has not been established experimentally [30-32]. In an informative review by Hansen, the clinical implications of photoisomerization therapy have been extensively discussed [31].

Bilirubin may also exert beneficial effects at physiological concentrations. Studies from our group have shown that the brain actively synthesizes bilirubin with potent neuroprotective effects [8]. For example, bilirubin is a highly effective scavenger of superoxide radicals (O_2^{-}) , both in vitro and in vivo. This scavenging activity plays a central role in N-methyl D-aspartate (NMDA)-mediated neurotransmission by preventing the accumulation of reactive oxygen species (ROS) [8] (Figure 2). Although normal NMDAR signaling is in part dependent on O_2 ⁻⁻, excessive NMDAR activity with an excessive accumulation of O₂⁻⁻ mediates uncontrolled lipid peroxidation and neuronal death [29]. Indeed, pathologic NMDA receptor activation and lipid peroxidation occur in several neurodegenerative diseases [30–33]. Notably, cells and neurons derived from mice lacking BVRA ($Blvra^{-/-}$) are hypersensitive to oxidative stress induced by O2⁻⁻. Blvra^{-/-} cells are more sensitive to several superoxide cyclers, as compared to other oxidants and electrophiles [8]. Thus, mouse embryonic fibroblasts (MEFs) derived from Blvra-/- mice are highly prone to oxidative death and are especially sensitive to the O2⁻⁻ donors pyrogallol, paraquat, and menadione. These MEFs were relatively mildly sensitive to the oxidant hydrogen peroxide (H_2O_2) and the electrophile 4-hydroxynonenal (4-HNE). These MEFs were also found to be less efficient in clearing ROS as compared to wild-type WT MEFs when exposed to menadione, as revealed by the fluorescent redox probe dihydroethidium (DHE) [8]. Additionally, NMDA treatment and O_2^{-} production cause exacerbated excitotoxicity and brain lesions in $Blvra^{-/-}$ mice [8]. These effects were also observed in ex vivo brain slices [34].



Figure 2. Bilirubin is a physiologic antioxidant. Activation of *N*-methyl-D-aspartate receptor (NM-DAR) signaling by glutamate mediates calcium (Ca²⁺) influx and activates downstream signaling pathways involving CaMKII. NMDAR engagement also generates superoxide (O₂⁻⁻), which, if not controlled, will lead to neurotoxicity. Bilirubin directly scavenges O₂⁻⁻ and prevents NMDAR-mediated neurotoxicity in neurons.

Although superoxide dismutases (SODs) scavenge O2^{·-}, these enzymes are specifically localized to soluble compartments in the cell [35]. However, O_2^{--} is often produced in aprotic and membrane-rich regions of cells, where it can be readily acted upon by lipophilic bilirubin. As O_2^{-} has a negative charge, its passive diffusion out of the cell is prevented, and thus, it accumulates in the mitochondrial matrix or intramembranous spaces. Using mitoUnaG, a mitochondrially targeted UnaG that binds bilirubin and fluoresces, we have shown that bilirubin is present in mitochondria, a site responsible for the generation of O2⁻ when electrons escape from the electron transport chain complexes I-III [8,36]. Notably, mitochondrial O_2^{--} play central roles in neuronal death in several conditions, such as ischemic stroke [37]. MEFs derived from $Blvra^{-/-}$ mice are highly sensitive to rotenone, an inhibitor of complex I. $Blvra^{-/-}$ MEFs exhibited higher levels of O₂⁻⁻ as compared to their wild-type controls despite the comparable expression of SODs and component proteins of the electron transport chain [8]. Furthermore, incubating these MEFs with bilirubin substantially decreased O2⁻⁻ levels. Along similar lines, cerebella of Blvra^{-/-} mice stereotactically injected with rotenone exhibited increased lesions and neuronal death, suggesting a neuroprotective role for the BVRA/bilirubin axis. Thus, bilirubin exerts both beneficial and deleterious effects, depending on its concentration.

3. Biliverdin Reductase-A in the Brain: More Than an Enzyme for Bilirubin Synthesis

In addition to its role in converting biliverdin to bilirubin, BVRA has additional protective roles in the brain [4,38]. Moreover, BVR expression is induced in neurons in response to ischemic injury. Specifically, BVR expression was increased in injured and neighboring neurons in a rodent model of stroke within six hours after middle cerebral artery occlusion (MCAO) [39]. This upregulation has been suggested to be related to the neuroprotective role of BVR in counteracting ROS after ischemia.

It has also been demonstrated that knocking down BVRA in rat primary neurons increases oxidative stress and elicits toxicity, further underscoring a role for BVRA in mitigating oxidative stress [14]. The subsequent generation of $Blvra^{-/-}$ mice confirmed

these findings, as these animals were found to be hypersensitive to oxidative stress, especially O_2^{--} , leading to elevated lipid peroxidation and oxidative damage [8,34]. Thus, the depletion or inactivation of BVRA can have deleterious effects.

Indeed, in normal physiology, BVRA is highly versatile in its actions, participating in diverse physiologic processes [40,41]. In addition to its canonical role as the terminal enzyme in heme catabolism, BVRA also functions as both a kinase and a kinase substrate, such as for insulin receptor kinase [5,42-45], and additionally autophosphorylates itself. For example, BVRA phosphorylation is essential for its own reductase activity, which generates bilirubin. In addition, insulin receptor (IR) engagement phosphorylates BVRA at specific tyrosine residues, activating BVRA to function as a dual-specificity serine/threonine and tyrosine kinase that modulates several signaling events, including the signaling mediated by Akt and Erk [5,42]. BVRA also phosphorylates and thereby inhibits the insulin receptor substrate (IRS) to prevent the aberrant activation of IRS as part of a feedback regulatory loop. Furthermore, BVRA participates in a BVRA/glycogen synthase kinase 3ß $(GSK-3\beta)$ /peroxisome proliferator-activated receptor α (PPAR α) axis that regulates hepatic lipid metabolism [46]. Here, GSK-3β phosphorylates its substrate, glycogen synthase 2 (GYS2), which is involved in glycogen production and storage in liver and muscle. GSK- 3β phosphorylates GYS2 and inhibits its activity, leading to decreased glycogen storage [47]. Thus, glycogen storage is activated when GSK-3 β is inhibited, which occurs when Ser 9 is phosphorylated by Akt [48,49]. GYS2 is also regulated at the transcriptional level by PPAR α , a process inhibited by GSK-3 β . GSK-3 β phosphorylates Ser 73 of PPAR α leading to its increased ubiquitination and protein turnover, as well as decreased activity [46]. Accordingly, liver-specific $Blvra^{-/-}$ mice exhibited increased Ser 73 phosphorylation of PPAR α , increased GSK-3 β activity, and decreased glycogen storage, revealing a role for BVRA in inhibiting GSK-3 β . Interestingly, bilirubin has been reported to inhibit many forms of protein phosphorylation that impact important cellular signaling processes [50–53]. How this might affect the above-mentioned feedback loop is an important area of future investigation. BVRA additionally modulates multiple metabolic and cellular pathways, including the control of glucose uptake, Akt signaling, immunosignaling, cellular proliferation, differentiation, and apoptosis [42,50–52,54]. Notably, signaling by BVRA has been shown to be disrupted in several diseases, as discussed below.

3.1. BVRA and Bilirubin in Alzheimer's Disease

Alzheimer's disease (AD) is the largest cause of dementia worldwide, with an estimated 413 million individuals on the AD continuum globally, of which 30 million have AD dementia [53]. AD may arise due to genetic causes or arise sporadically, with familial causes accounting for less than 5% of all cases [55].

The pathological hallmarks of AD include the deposition of intracellular neur ofibrillary tangles and paired helical fibrils composed of the protein tau and the accumulation of extracellular amyloid plaques composed of amyloid β (A β) peptides [56,57]. Tau is a microtubule binding protein that participates in axon outgrowth and neuronal transport [58,59]. Aberrant post-translational modifications on tau, such as hyperphosphorylation and acetylation, cause its dissociation from microtubules and aggregation, leading to neurotoxicity in AD and other forms of neurodegeneration [60–63]. The A β peptides are generated from the amyloid precursor protein (APP). APP can be cleaved at different sites by two major pathways: the nonamyloidogenic pathway and the amyloidogenic pathway. In AD, an increased production of the A β (1–42) fragment occurs by the action of beta-secretase 1 (BACE1) on APP [64–66]. AD has no cure, and existing treatments do not adequately treat disease symptoms. Recently approved therapies that target amyloid peptides and plaques have met with limited clinical success and are complicated by dangerous side effects to patients [67].

One of the earliest studies that indicated the dysregulation of the BVRA/bilirubin axis in AD reported increased bilirubin levels in the cerebrospinal fluid (CSF) of patients [68]. Later, decreased plasma levels of antioxidants, including bilirubin, were identified as additional features of AD [69]. Moreover, the first enzyme in heme catabolism in the brain, heme oxygenase 2 (HO-2), is also dysregulated in AD. In addition, APP interacts with HO-2 and inhibits its activity, causing neurotoxicity [70]. Furthermore, cortical neuronal cultures from APP695swe transgenic mice have lower levels of bilirubin, which has been ascribed to lower HO-2 activity and lower biliverdin accumulation [70]. BVRA activity in AD was later analyzed by Barone and colleagues in 2011, who found decreased phosphorylation and increased oxidative/nitrosative post-translational modifications of BVRA in the hippocampus of patients with AD and mild cognitive impairment (MCI) [71–73]. These same modifications to BVRA were also observed in the plasma of AD patients and associated with increased plasma levels of BVRA and 3-nitrotyrosine-modification but decreased phosphotyrosine levels and BVRA reductase activity [74]. Furthermore, in a longitudinal study in the 3xTg-AD mouse model of AD, Barone and colleagues demonstrated that impaired BVRA activity promoted brain insulin resistance (BIR) and was one of the earliest pathologic events in this model [75]. A mechanism was proposed wherein oxidative-stress-mediated BVRA enzymatic impairment causes prolonged the activation of IRS, which triggers negative feedback mechanisms (such as those involving mTOR) that diminish autophagy and trigger IRS hyperactivity, thereby causing BIR [75–77]. The diminished activity of BVRA was also observed in other animal models of AD to lead to increased BACE1 phosphorylation, A β accumulation, and insulin resistance through downregulation of the insulin receptor (IR) and inhibition of IRS [77]. In line with these studies, the statin atorvastatin, which reduces the risk of AD, has also been shown to elevate the expression, phosphorylation, and activity of BVRA in the parietal cortex of aged beagles, a preclinical model of AD. Moreover, negative correlation between levels of BVRA, the accumulation of oxidative stress markers, and errors in discrimination learning were also observed in this model [78]. Notably, bilirubin treatment of a diet-induced model of obesity in mice reduced body weight and mitigated elevated blood glucose, total cholesterol (TC), leptin, and adiponectin levels, as well as normalized transcripts of sterol regulatory element-binding protein (SREBP-1), insulin receptor (IR), and PPARγ, and improved insulin sensitivity [79]. Independent studies also showed that either bilirubin levels or reductions in BVRA negatively correlated with obesity and its associated sequelae [80,81]. Lastly, signaling links between BVRA and neurodegeneration were established when it was shown that the loss of BVRA impaired the Akt-mediated inhibition of GSK-3 β in response to oxidative stress, leading to tau hyperphosphorylation in early-stage AD [82]. The same study reported similar findings in MCI and also demonstrated that cells lacking BVRA exhibited lower GSK-3β inhibition and increased Tau Ser404 phosphorylation when subjected to oxidative stress. Thus, the BVRA/bilirubin axis is compromised in AD at multiple levels (Figure 3). An increased activation of GSK-3 β has also been reported in the liver-specific *Blvra*^{-/-} mice, as described in Section 3 [46].

The aberrant expression and activity of BVRA have also been observed in several other conditions of impaired cognition. For example, BVRA levels were disrupted in the hippocampus, prefrontal cortex, and temporal lobe in a rat model of postoperative delayed neurocognitive recovery (dNCR) [83]. In addition to BVRA, the disposition of bilirubin is also compromised in several conditions, affecting brain function, as well as in various models of neurodegeneration, summarized in Table 1. Altered bilirubin levels have also been linked to neuropsychiatric disease. For instance, in male subjects with schizophrenia, total serum bilirubin levels were linked to cognition. Male patients with cognitive dysfunction had lower levels of total serum bilirubin [84]. An independent study reported an inverse correlation between lower bilirubin levels and working memory in first episode psychosis [85]. The same study also reported that lower bilirubin levels correlate with the duration of untreated psychosis (DUP). It must be noted, however, that while some studies reported an increase in either bilirubin levels or BVRA expression in brain diseases, others reported a decrease. For example, in a study involving drug-naïve PD patients, higher serum bilirubin levels were associated with better prognosis and outcomes over



a two-year period [86]. Taken together, the date reflects perturbed bilirubin metabolism (Table 1).

Figure 3. BVRA activity is decreased in Alzheimer's disease (AD). Under normal conditions (left), insulin binds to the insulin receptor (IR) and stimulates auto-phosphorylation of IR on Tyr (Y) residues, which activates its kinase activity. Activated IR phosphorylates biliverdin reductase A (BVRA), as well as the insulin receptor substrate (IRS), on specific Y residues and activates them. Activated BVRA then phosphorylates and inhibits IRS itself, which prevents excessive IRS activation in a negative feedback loop. Activated IRS stimulates downstream signaling proteins, including phosphatidylinositol 3 kinase (PI3K), which activates Akt by phosphorylation. Activated Akt then phosphorylates GSK-3β on Ser9 and inhibits its kinase activity. BVRA additionally promotes Akt activation and GSK-3ß inhibition by serving as a scaffold protein. Additionally, BVRA activity generates bilirubin, which scavenges superoxide (O_2^{-}) and prevents oxidative and nitrosative damage. In AD (right), the activity of BVRA is low due to its inactivation by 4-hydroxynonenal (4-HNE) and nitration. This leads to an accumulation of reactive oxygen species (ROS) and reactive nitrogen species (RNS), which affect numerous cellular processes, including mitochondrial function. The loss of BVRA causes IRS hyperactivation and impaired insulin signaling. Downstream of IRS, suboptimal BVRA activity decreases Akt activation and GSK-3ß inhibition, causing hyperphosphorylation of tau that leads to neurofibrillary tangle formation and neurotoxicity. Elevated ROS and RNS also favor tau and amyloid-β deposition. All these processes culminate in neurodegeneration.

 Table 1. Summary of biliverdin reductase-A/bilirubin disposition in brain diseases.

Condition	Status of Bilirubin and BVRA	Reference
Alzheimer's disease (3xTg-AD mouse model)	Decreased BVRA levels and activity, decreased Tyr-phosphorylation. Increased 3-NT modification of BVRA.	[75,87,88]
Alzheimer's disease and MCI patients	Reduced BVR-A Tyr-phosphorylation and elevated 3-NT modification of BVR-A in the hippocampus.	[71,72,89]
Down Syndrome (Ts65dn mouse model)	Reduced BVR-A Tyr-phosphorylation in the frontal cortex (9 months).	[90]

Condition	Status of Bilirubin and BVRA	Reference
Aging (C57BL/6J mice)	Decreased BVRA levels and phosphorylation (12 months) and increased 3-NT modification of BVRA (18 months) in the hippocampus.	[75]
Aging (Canine model)	Decreased BVRA Tyr-phosphorylation (4–12 months), elevated 3-NT modification of BVRA (10–12 months) in the parietal cortex.	[77]
Parkinson's disease patients (without dementia)	Low bilirubin levels in serum of PD patients.	[91]
Parkinson's disease (without dementia)	Higher bilirubin levels in plasma of PD patients.	[92]
Parkinson's disease patients (de novo, drug-naïve)	Higher levels of bilirubin in serum, which correlated with better outcomes in a 2-year longitudinal study.	[86]
Amyotrophic lateral sclerosis (G93A SOD1 rat model)	Higher serum bilirubin levels in the symptomatic stage of ALS.	[93]
ALS patients	Lower serum bilirubin levels in patients with long duration ALS.	[94]
Migraine Multiple sclerosis	Lower serum bilirubin in people with migraine. Lower serum bilirubin in people with MS.	[95] [96]
Multiple sclerosis (relapsing, remitting, or RRMS)	Lower serum bilirubin in people with MS	[97]

Table 1. Cont.

3.2. BVRA and Synaptic Plasticity

Our laboratory has also shown that BVRA modulates synaptic function. First, we demonstrated that Blvra^{-/-} mice exhibited spatial learning deficits as assessed by the Morris water maze, which is dependent on intact hippocampal synaptic plasticity [98]. We also found that $Blvra^{-/-}$ mice were compromised in fear conditioning as compared to wildtype (WT) littermates. Fear conditioning is a form of learning, distinct from spatial learning, in which anticipation of adverse events is learned. In a typical fear conditioning test, mice are trained to respond to a sound followed by a mild foot shock. With increasing tone-shock pair events, mice will increasingly freeze after the sound in anticipation of a foot-shock. The following day, the mice are returned to the same chamber. Mice that have associated the chamber with being shocked will freeze, whereas those that have not learned this association will not. Mice are then placed in a new chamber and exposed to the same sound from the previous day. Mice that have learned that the sound can precede a shock will freeze in fear, whereas those that were unable to learn this will not. In this test, young $Blvra^{-/-}$ mice learned the cues and context as well as WT mice but had poorer recall memory, and aged Blvra^{-/-} mice were impaired in both learning and recall. Related to learning, a role for BVR has been established in synaptic plasticity. Synapses are dynamically modulated and can be structurally strengthened or weakened depending on the signal [99–101]. This plasticity has been linked to learning and memory involving various components of brain circuitry [102]. Under certain conditions, such as high-frequency stimulation, a sustained strengthening in synaptic transmission occurs, termed long-term potentiation (LTP), which regulates learning and memory [103]. An RNASeq analysis of the hippocampi of $Blvra^{-/-}$ and WT mice revealed downregulation of focal adhesion kinase (FAK), which plays a central role in synaptic plasticity (Figure 4). FAKs are dynamic entities that link the intracellular cytoskeleton to the extracellular matrix [104]. FAK is a tyrosine kinase that signals alterations in the structure of focal adhesion complexes to key intracellular signaling hubs, such as phosphatidylinositol 3-kinase (PI3K) and mitogen-activated protein kinases (MAPKs), thereby transducing extracellular stimuli to intracellular signals that regulate synaptic strength [105-108]. BVRA facilitates this process by bridging focal adhesion signaling molecules at the synapse. BVRA also acts as an adaptor or bridge between

the primary focal adhesion signaling kinases FAK and Pyk2 and the effector kinase Src (Figure 4). This activity is independent of BVRA's catalytic activity. In the absence of BVRA, FAK and Pyk2 are unable to bind and stimulate Src, which is then unable to implement the phosphorylation of the NMDA receptor that is required for synaptic plasticity. As Src serves as a molecular hub where several signaling pathways converge to enhance NMDAR-mediated neurotransmission, BVR is positioned at a prominent intersection of synaptic signaling. Not surprisingly, the depletion of BVRA in hippocampal slices causes deficits in electrophysiological responses to stimuli.



Figure 4. BVR-A mediates focal adhesion kinase-mediated synaptic signaling. Left: postsynaptic FAK-mediated signaling pathways during basal conditions. Right: FAK signaling during *N*-methyl-D-aspartate receptor (NMDAR)-mediated neurotransmission. Upon glutamate stimulation of the NMDAR, Ca²⁺ influx activates protein kinase C (PKC). Ca²⁺/PKC activates Pyk2 by stimulating autophosphorylation at Y402 and phosphorylation of integrins. The resulting conformational change in integrins activates autophosphorylation of FAK at Y397. Autophosphorylated Pyk2 and FAK then recruit and activate Src through phosphorylation at Y416, with BVRA acting as an adapter. Next, Src interacts with and phosphorylates the NR2B subunit of the NMDAR at Y1472, which further stimulates Ca²⁺ influx through the NMDAR.

4. Therapeutic Opportunities

As BVRA is unparalleled in its versatility of functions, it also offers opportunities for developing therapeutics at multiple levels. BVRA is positioned at the crossroads of a plethora of signaling cascades in the periphery and the brain and thus may provide a useful therapeutic target for a wide variety of diseases, including various forms of cancer, cardiovascular diseases, immune system disorders, and neurodegenerative diseases.

To date, BVRA-based peptide agonists have been considered for metabolic diseases. Specifically, a 7-residue peptide derived from the BVRA primary sequence can independently activate insulin receptor kinase autophosphorylation, kinase activity, and glucose uptake [50]. In addition to the periphery, BVRA also regulates insulin receptor signaling in the brain and prevents the hyperphosphorylation of tau through the Akt pathway [41,77,82].

Furthermore, BVRA levels are dynamically altered in response to insulin, including intranasal administration used to alleviate brain insulin resistance in AD [77,109]. In agreement with these reports, the loss of BVRA causes elevated oxidative stress, abnormal tau pathology, and impaired autophagy [8,14,34,76,82]. Autophagy impairment is a key feature of neurodegenerative diseases, and loss of BVRA impacts several aspects of autophagy. The reduced activity of AMP-activated protein kinase (AMPK), a key energy sensor that regulates cellular homeostasis and which is also an inhibitor of mTOR, is associated with loss of BVRA [76,110]. In line with these observations, it has been shown that high doses of biliverdin (the substrate of BVRA) activate the mTOR pathway, while rapamycin, an mTOR inhibitor, prevents this activation by biliverdin.

Several strategies may be adopted to augment BVRA levels and/or activity. For instance, peptides that activate the kinase and reductase activity of BVRA may be harnessed to mitigate neurodegeneration [111]. Earlier studies have revealed that BVR-derived peptides modulate functions related to kinase and scaffold activities of this enzyme. Three peptides derived from BVR have been characterized:¹⁶²FGFPAFSG and ²⁷⁵KKRILHC and ²⁹⁰KYCCSRK. While the peptide KKRILHC inhibits BVRA kinase activity, the peptide KYCCSRK activates both the kinase and reductase activities [13,40,50,112]. In a preclinical model of cardiotoxicity, such as the perfused rat heart exposed to isoproterenol (ISO), the peptide KKRILHC (inhibitor) counteracted ISO-mediated BVRA activity and caused apoptosis and left ventricular dysfunction. On the other hand, perfusion with KYCCSRK (activator) augmented BVRA activity, prevented apoptosis, and preserved left ventricular function.

A significant challenge for delivering drugs to the brain involves the blood–brain barrier, and the development of lipid nanoparticles and cell-penetrating peptides derived from BVRA is an area that merits further investigation. As BVRA acts as an adaptor protein and participates in neurotransmission, augmenting the levels and activity of BVRA could optimize related processes as well [98]. In addition to these strategies, the use of bilirubin nanoparticles as therapeutics for metabolic and cardiovascular diseases has been considered [113–115]. In disorders with lowered bilirubin, suppressing the activity of UGT1A1 may provide an alternative approach to boost serum/plasma bilirubin levels [116]. In neurodegenerative disorders such as PD, in which bilirubin offers protection, inducers of BVRA and bilirubin protection are being considered. In a cell culture model of rotenone (an inhibitor of mitochondrial complex I) toxicity, in which bilirubin is depleted, BRUP-1-mediated modulation of bilirubin levels was found to be protective [117]. The use of such inducers of bilirubin production may be beneficial in other diseases such as AD in the future.

Screening for small molecules that upregulate the BVRA/bilirubin pathway could also result in the identification of novel therapeutic compounds. The administration of Atorvastatin (80 mg/day for 14.5 months) was reported to elevate BVR protein level, phosphorylation, and activity in the cortex of aged beagles, a preclinical model of AD [78]. The same study also reported that the induction of BVRA negatively correlated with the level of BACE1 protein and oxidative stress biomarkers, supporting a neuroprotective role for BVRA. Along similar lines with respect to the induction of BVRA protein, ferulic acid, a polyphenol abundant in fruits and vegetables, induces BVRA in theSH-SY5Y human neuroblastoma cell line and prevents oxidative stress-mediated neuronal death [118,119]. The augmentation of BVRA activity may also be beneficial in aging, the biggest risk factor for neurodegenerative diseases [120]. Centenarians, for example, are likely to display increased BVR-A levels in their blood [121]. Thus, stimulating the BVRA or augmenting bilirubin levels may be therapeutic in neurodegenerative diseases involving low bilirubin content.

In conclusion, the BVRA/bilirubin axis in the brain has, until recently, been an underappreciated system of natural neuroprotection that warrants further investigation and could serve to identify novel neuroprotective strategies for patients. **Author Contributions:** B.D.P. and A.A.P. conceptualized and wrote the review. B.D.P. prepared the figures. All authors have read and agreed to the published version of the manuscript.

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References

- 1. Lemberg, R.; Wyndham, R.A. Reduction of biliverdin to bilirubin in tissues. *Biochem. J.* 1936, 30, 1147–1170. [CrossRef] [PubMed]
- 2. Singleton, J.W.; Laster, L. Biliverdin reductase of guinea pig liver. J. Biol. Chem. 1965, 240, 4780–4789. [CrossRef] [PubMed]
- 3. Kutty, R.K.; Maines, M.D. Purification and characterization of biliverdin reductase from rat liver. *J. Biol. Chem.* **1981**, 256, 3956–3962. [CrossRef] [PubMed]
- 4. Maines, M.D.; Trakshel, G.M. Purification and characterization of human biliverdin reductase. *Arch. Biochem. Biophys.* **1993**, 300, 320–326. [CrossRef] [PubMed]
- 5. Kapitulnik, J.; Maines, M.D. Pleiotropic functions of biliverdin reductase: Cellular signaling and generation of cytoprotective and cytotoxic bilirubin. *Trends Pharmacol. Sci.* 2009, *30*, 129–137. [CrossRef] [PubMed]
- O'Brien, L.; Hosick, P.A.; John, K.; Stec, D.E.; Hinds, T.D., Jr. Biliverdin reductase isozymes in metabolism. *Trends Endocrinol. Metab.* 2015, 26, 212–220. [CrossRef] [PubMed]
- Cunningham, O.; Gore, M.G.; Mantle, T.J. Initial-rate kinetics of the flavin reductase reaction catalysed by human biliverdin-IXbeta reductase (BVR-B). *Biochem. J.* 2000, 345 Pt 2, 393–399. [CrossRef]
- 8. Vasavda, C.; Kothari, R.; Malla, A.P.; Tokhunts, R.; Lin, A.; Ji, M.; Ricco, C.; Xu, R.; Saavedra, H.G.; Sbodio, J.I.; et al. Bilirubin Links Heme Metabolism to Neuroprotection by Scavenging Superoxide. *Cell Chem. Biol.* **2019**, *26*, 1450–1460.e7. [CrossRef]
- 9. Stocker, R.; Yamamoto, Y.; McDonagh, A.F.; Glazer, A.N.; Ames, B.N. Bilirubin is an antioxidant of possible physiological importance. *Science* **1987**, *235*, 1043–1046. [CrossRef]
- 10. Mireles, L.C.; Lum, M.A.; Dennery, P.A. Antioxidant and cytotoxic effects of bilirubin on neonatal erythrocytes. *Pediatr. Res.* **1999**, 45, 355–362. [CrossRef]
- 11. McDonagh, A.F. Is bilirubin good for you? Clin. Perinatol. 1990, 17, 359–369. [CrossRef]
- Mancuso, C.; Barone, E.; Guido, P.; Miceli, F.; Di Domenico, F.; Perluigi, M.; Santangelo, R.; Preziosi, P. Inhibition of lipid peroxidation and protein oxidation by endogenous and exogenous antioxidants in rat brain microsomes in vitro. *Neurosci. Lett.* 2012, 518, 101–105. [CrossRef]
- 13. Mancuso, C. Biliverdin as a disease-modifying agent: An integrated viewpoint. *Free Radic. Biol. Med.* **2023**, 207, 133–143. [CrossRef]
- 14. Sedlak, T.W.; Saleh, M.; Higginson, D.S.; Paul, B.D.; Juluri, K.R.; Snyder, S.H. Bilirubin and glutathione have complementary antioxidant and cytoprotective roles. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 5171–5176. [CrossRef]
- 15. Baranano, D.E.; Rao, M.; Ferris, C.D.; Snyder, S.H. Biliverdin reductase: A major physiologic cytoprotectant. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 16093–16098. [CrossRef] [PubMed]
- 16. Kaasik, K.; Lee, C.C. Reciprocal regulation of haem biosynthesis and the circadian clock in mammals. *Nature* **2004**, *430*, 467–471. [CrossRef] [PubMed]
- 17. Falchuk, K.H.; Contin, J.M.; Dziedzic, T.S.; Feng, Z.; French, T.C.; Heffron, G.J.; Montorzi, M. A role for biliverdin IXalpha in dorsal axis development of Xenopus laevis embryos. *Proc. Natl. Acad. Sci. USA* 2002, *99*, 251–256. [CrossRef] [PubMed]

- Tukey, R.H.; Strassburg, C.P. Human UDP-glucuronosyltransferases: Metabolism, expression, and disease. *Annu. Rev. Pharmacol. Toxicol.* 2000, 40, 581–616. [CrossRef] [PubMed]
- 19. Abu-Bakar, A.; Arthur, D.M.; Aganovic, S.; Ng, J.C.; Lang, M.A. Inducible bilirubin oxidase: A novel function for the mouse cytochrome P450 2A5. *Toxicol. Appl. Pharmacol.* 2011, 257, 14–22. [CrossRef] [PubMed]
- Vitek, L.; Ostrow, J.D. Bilirubin chemistry and metabolism; harmful and protective aspects. *Curr. Pharm. Des.* 2009, 15, 2869–2883. [CrossRef] [PubMed]
- Boon, A.C.; Bulmer, A.C.; Coombes, J.S.; Fassett, R.G. Circulating bilirubin and defense against kidney disease and cardiovascular mortality: Mechanisms contributing to protection in clinical investigations. *Am. J. Physiol. Renal Physiol.* 2014, 307, F123–F136. [CrossRef]
- 22. Wagner, K.H.; Wallner, M.; Molzer, C.; Gazzin, S.; Bulmer, A.C.; Tiribelli, C.; Vitek, L. Looking to the horizon: The role of bilirubin in the development and prevention of age-related chronic diseases. *Clin. Sci.* 2015, 129, 1–25. [CrossRef]
- 23. Gazzin, S.; Masutti, F.; Vitek, L.; Tiribelli, C. The molecular basis of jaundice: An old symptom revisited. *Liver Int.* 2017, 37, 1094–1102. [CrossRef]
- Wagner, K.H.; Khoei, N.S.; Hana, C.A.; Doberer, D.; Marculescu, R.; Bulmer, A.C.; Hormann-Wallner, M.; Molzer, C. Oxidative Stress and Related Biomarkers in Gilbert's Syndrome: A Secondary Analysis of Two Case-Control Studies. *Antioxidants* 2021, 10, 1474. [CrossRef]
- 25. Canu, G.; Minucci, A.; Zuppi, C.; Capoluongo, E. Gilbert and Crigler Najjar syndromes: An update of the UDPglucuronosyltransferase 1A1 (UGT1A1) gene mutation database. *Blood Cells Mol. Dis.* 2013, 50, 273–280. [CrossRef]
- Gantla, S.; Bakker, C.T.; Deocharan, B.; Thummala, N.R.; Zweiner, J.; Sinaasappel, M.; Roy Chowdhury, J.; Bosma, P.J.; Roy Chowdhury, N. Splice-site mutations: A novel genetic mechanism of Crigler-Najjar syndrome type 1. *Am. J. Hum. Genet.* 1998, 62, 585–592. [CrossRef]
- Ritter, J.K.; Yeatman, M.T.; Ferreira, P.; Owens, I.S. Identification of a genetic alteration in the code for bilirubin UDPglucuronosyltransferase in the UGT1 gene complex of a Crigler-Najjar type I patient. J. Clin. Investig. 1992, 90, 150–155. [CrossRef]
- 28. Crigler, J.F., Jr.; Najjar, V.A. Congenital familial nonhemolytic jaundice with kernicterus. *Pediatrics* 1952, 10, 169–180. [PubMed]
- Reyes, R.C.; Brennan, A.M.; Shen, Y.; Baldwin, Y.; Swanson, R.A. Activation of neuronal NMDA receptors induces superoxidemediated oxidative stress in neighboring neurons and astrocytes. J. Neurosci. 2012, 32, 12973–12978. [CrossRef] [PubMed]
- Dexter, D.T.; Holley, A.E.; Flitter, W.D.; Slater, T.F.; Wells, F.R.; Daniel, S.E.; Lees, A.J.; Jenner, P.; Marsden, C.D. Increased levels of lipid hydroperoxides in the parkinsonian substantia nigra: An HPLC and ESR study. *Mov. Disord.* 1994, 9, 92–97. [CrossRef] [PubMed]
- Ferrante, R.J.; Browne, S.E.; Shinobu, L.A.; Bowling, A.C.; Baik, M.J.; MacGarvey, U.; Kowall, N.W.; Brown, R.H., Jr.; Beal, M.F. Evidence of increased oxidative damage in both sporadic and familial amyotrophic lateral sclerosis. *J. Neurochem.* 1997, 69, 2064–2074. [CrossRef]
- 32. Subbarao, K.V.; Richardson, J.S.; Ang, L.C. Autopsy samples of Alzheimer's cortex show increased peroxidation in vitro. *J. Neurochem.* **1990**, *55*, 342–345. [CrossRef]
- Wang, R.; Reddy, P.H. Role of Glutamate and NMDA Receptors in Alzheimer's Disease. J. Alzheimers Dis. 2017, 57, 1041–1048. [CrossRef] [PubMed]
- 34. Vasavda, C.; Snyder, S.H.; Paul, B.D. Quantitative measurement of reactive oxygen species in ex vivo mouse brain slices. *STAR Protoc.* **2021**, *2*, 100332. [CrossRef]
- 35. McCord, J.M.; Fridovich, I. Superoxide dismutase. An enzymic function for erythrocuprein (hemocuprein). *J. Biol. Chem.* **1969**, 244, 6049–6055. [CrossRef]
- 36. Murphy, M.P. How mitochondria produce reactive oxygen species. Biochem. J. 2009, 417, 1–13. [CrossRef] [PubMed]
- 37. Kontos, C.D.; Wei, E.P.; Williams, J.I.; Kontos, H.A.; Povlishock, J.T. Cytochemical detection of superoxide in cerebral inflammation and ischemia in vivo. *Am. J. Physiol.* **1992**, *263*, H1234–H1242. [CrossRef] [PubMed]
- 38. Maines, M.D. Multiple forms of biliverdin reductase: Age-related change in pattern of expression in rat liver and brain. *Mol. Pharmacol.* **1990**, *38*, 481–485.
- Panahian, N.; Huang, T.; Maines, M.D. Enhanced neuronal expression of the oxidoreductase--biliverdin reductase--after permanent focal cerebral ischemia. *Brain Res.* 1999, 850, 1–13. [CrossRef]
- 40. Gibbs, P.E.; Tudor, C.; Maines, M.D. Biliverdin reductase: More than a namesake-the reductase, its Peptide fragments, and biliverdin regulate activity of the three classes of protein kinase C. *Front. Pharmacol.* **2012**, *3*, 31. [CrossRef]
- 41. Cimini, F.A.; Perluigi, M.; Barchetta, I.; Cavallo, M.G.; Barone, E. Role of Biliverdin Reductase A in the Regulation of Insulin Signaling in Metabolic and Neurodegenerative Diseases: An Update. *Int. J. Mol. Sci.* **2022**, *23*, 5574. [CrossRef] [PubMed]
- Lerner-Marmarosh, N.; Shen, J.; Torno, M.D.; Kravets, A.; Hu, Z.; Maines, M.D. Human biliverdin reductase: A member of the insulin receptor substrate family with serine/threonine/tyrosine kinase activity. *Proc. Natl. Acad. Sci. USA* 2005, 102, 7109–7114. [CrossRef] [PubMed]
- Lerner-Marmarosh, N.; Miralem, T.; Gibbs, P.E.; Maines, M.D. Regulation of TNF-alpha-activated PKC-zeta signaling by the human biliverdin reductase: Identification of activating and inhibitory domains of the reductase. *FASEB J.* 2007, 21, 3949–3962. [CrossRef] [PubMed]

- 44. Lerner-Marmarosh, N.; Miralem, T.; Gibbs, P.E.; Maines, M.D. Human biliverdin reductase is an ERK activator; hBVR is an ERK nuclear transporter and is required for MAPK signaling. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 6870–6875. [CrossRef] [PubMed]
- 45. Maines, M.D. Biliverdin reductase: PKC interaction at the cross-talk of MAPK and PI3K signaling pathways. *Antioxid. Redox Signal.* **2007**, *9*, 2187–2195. [CrossRef]
- Hinds, T.D., Jr.; Burns, K.A.; Hosick, P.A.; McBeth, L.; Nestor-Kalinoski, A.; Drummond, H.A.; AlAmodi, A.A.; Hankins, M.W.; Vanden Heuvel, J.P.; Stec, D.E. Biliverdin Reductase A Attenuates Hepatic Steatosis by Inhibition of Glycogen Synthase Kinase (GSK) 3beta Phosphorylation of Serine 73 of Peroxisome Proliferator-activated Receptor (PPAR) alpha. J. Biol. Chem. 2016, 291, 25179–25191. [CrossRef]
- 47. Imazu, M.; Strickland, W.G.; Chrisman, T.D.; Exton, J.H. Phosphorylation and inactivation of liver glycogen synthase by liver protein kinases. *J. Biol. Chem.* **1984**, 259, 1813–1821. [CrossRef]
- Cross, D.A.; Alessi, D.R.; Cohen, P.; Andjelkovich, M.; Hemmings, B.A. Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. *Nature* 1995, 378, 785–789. [CrossRef]
- Orena, S.J.; Torchia, A.J.; Garofalo, R.S. Inhibition of glycogen-synthase kinase 3 stimulates glycogen synthase and glucose transport by distinct mechanisms in 3T3-L1 adipocytes. J. Biol. Chem. 2000, 275, 15765–15772. [CrossRef] [PubMed]
- 50. Miralem, T.; Lerner-Marmarosh, N.; Gibbs, P.E.; Jenkins, J.L.; Heimiller, C.; Maines, M.D. Interaction of human biliverdin reductase with Akt/protein kinase B and phosphatidylinositol-dependent kinase 1 regulates glycogen synthase kinase 3 activity: A novel mechanism of Akt activation. *FASEB J.* **2016**, *30*, 2926–2944. [CrossRef] [PubMed]
- Gibbs, P.E.; Lerner-Marmarosh, N.; Poulin, A.; Farah, E.; Maines, M.D. Human biliverdin reductase-based peptides activate and inhibit glucose uptake through direct interaction with the kinase domain of insulin receptor. *FASEB J.* 2014, 28, 2478–2491. [CrossRef]
- Song, S.; Wang, S.; Ma, J.; Yao, L.; Xing, H.; Zhang, L.; Liao, L.; Zhu, D. Biliverdin reductase/bilirubin mediates the anti-apoptotic effect of hypoxia in pulmonary arterial smooth muscle cells through ERK1/2 pathway. *Exp. Cell Res.* 2013, 319, 1973–1987. [CrossRef]
- Wegiel, B.; Gallo, D.; Csizmadia, E.; Roger, T.; Kaczmarek, E.; Harris, C.; Zuckerbraun, B.S.; Otterbein, L.E. Biliverdin inhibits Toll-like receptor-4 (TLR4) expression through nitric oxide-dependent nuclear translocation of biliverdin reductase. *Proc. Natl. Acad. Sci. USA* 2011, 108, 18849–18854. [CrossRef]
- Gustavsson, A.; Norton, N.; Fast, T.; Frolich, L.; Georges, J.; Holzapfel, D.; Kirabali, T.; Krolak-Salmon, P.; Rossini, P.M.; Ferretti, M.T.; et al. Global estimates on the number of persons across the Alzheimer's disease continuum. *Alzheimers Dement.* 2023, 19, 658–670. [CrossRef]
- 55. Karch, C.M.; Goate, A.M. Alzheimer's disease risk genes and mechanisms of disease pathogenesis. *Biol. Psychiatry* 2015, 77, 43–51. [CrossRef]
- Masters, C.L.; Bateman, R.; Blennow, K.; Rowe, C.C.; Sperling, R.A.; Cummings, J.L. Alzheimer's disease. *Nat. Rev. Dis. Primers* 2015, 1, 15056. [CrossRef] [PubMed]
- 57. Lane, C.A.; Hardy, J.; Schott, J.M. Alzheimer's disease. Eur. J. Neurol. 2018, 25, 59–70. [CrossRef] [PubMed]
- 58. Limorenko, G.; Lashuel, H.A. Revisiting the grammar of Tau aggregation and pathology formation: How new insights from brain pathology are shaping how we study and target Tauopathies. *Chem. Soc. Rev.* **2022**, *51*, 513–565. [CrossRef] [PubMed]
- Santarella, R.A.; Skiniotis, G.; Goldie, K.N.; Tittmann, P.; Gross, H.; Mandelkow, E.M.; Mandelkow, E.; Hoenger, A. Surfacedecoration of microtubules by human tau. J. Mol. Biol. 2004, 339, 539–553. [CrossRef] [PubMed]
- Kopke, E.; Tung, Y.C.; Shaikh, S.; Alonso, A.C.; Iqbal, K.; Grundke-Iqbal, I. Microtubule-associated protein tau. Abnormal phosphorylation of a non-paired helical filament pool in Alzheimer disease. *J. Biol. Chem.* 1993, 268, 24374–24384. [CrossRef] [PubMed]
- 61. Davidson, R.; Krider, R.I.; Borsellino, P.; Noorda, K.; Alhwayek, G.; Vida, T.A. Untangling Tau: Molecular Insights into Neuroinflammation, Pathophysiology, and Emerging Immunotherapies. *Curr. Issues Mol. Biol.* **2023**, *45*, 8816–8839. [CrossRef]
- 62. Tracy, T.; Claiborn, K.C.; Gan, L. Regulation of Tau Homeostasis and Toxicity by Acetylation. *Adv. Exp. Med. Biol.* 2019, 1184, 47–55. [CrossRef]
- 63. Shin, M.K.; Vazquez-Rosa, E.; Koh, Y.; Dhar, M.; Chaubey, K.; Cintron-Perez, C.J.; Barker, S.; Miller, E.; Franke, K.; Noterman, M.F.; et al. Reducing acetylated tau is neuroprotective in brain injury. *Cell* **2021**, *184*, 2715–2732.e23. [CrossRef] [PubMed]
- 64. Hampel, H.; Hardy, J.; Blennow, K.; Chen, C.; Perry, G.; Kim, S.H.; Villemagne, V.L.; Aisen, P.; Vendruscolo, M.; Iwatsubo, T.; et al. The Amyloid-beta Pathway in Alzheimer's Disease. *Mol. Psychiatry* **2021**, *26*, 5481–5503. [CrossRef] [PubMed]
- 65. Hardy, J.; Selkoe, D.J. The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* **2002**, 297, 353–356. [CrossRef] [PubMed]
- Li, R.; Lindholm, K.; Yang, L.B.; Yue, X.; Citron, M.; Yan, R.; Beach, T.; Sue, L.; Sabbagh, M.; Cai, H.; et al. Amyloid beta peptide load is correlated with increased beta-secretase activity in sporadic Alzheimer's disease patients. *Proc. Natl. Acad. Sci. USA* 2004, 101, 3632–3637. [CrossRef] [PubMed]
- 67. Granzotto, A.; Sensi, S.L. Once upon a time, the Amyloid Cascade Hypothesis. *Ageing Res. Rev.* 2024, 93, 102161. [CrossRef] [PubMed]
- 68. Kimpara, T.; Takeda, A.; Yamaguchi, T.; Arai, H.; Okita, N.; Takase, S.; Sasaki, H.; Itoyama, Y. Increased bilirubins and their derivatives in cerebrospinal fluid in Alzheimer's disease. *Neurobiol. Aging* **2000**, *21*, 551–554. [CrossRef] [PubMed]

- 69. Kim, T.S.; Pae, C.U.; Yoon, S.J.; Jang, W.Y.; Lee, N.J.; Kim, J.J.; Lee, S.J.; Lee, C.; Paik, I.H.; Lee, C.U. Decreased plasma antioxidants in patients with Alzheimer's disease. *Int. J. Geriatr. Psychiatry* **2006**, *21*, 344–348. [CrossRef]
- 70. Takahashi, M.; Dore, S.; Ferris, C.D.; Tomita, T.; Sawa, A.; Wolosker, H.; Borchelt, D.R.; Iwatsubo, T.; Kim, S.H.; Thinakaran, G.; et al. Amyloid precursor proteins inhibit heme oxygenase activity and augment neurotoxicity in Alzheimer's disease. *Neuron* 2000, 28, 461–473. [CrossRef]
- 71. Barone, E.; Di Domenico, F.; Cenini, G.; Sultana, R.; Coccia, R.; Preziosi, P.; Perluigi, M.; Mancuso, C.; Butterfield, D.A. Oxidative and nitrosative modifications of biliverdin reductase-A in the brain of subjects with Alzheimer's disease and amnestic mild cognitive impairment. *J. Alzheimers Dis.* **2011**, *25*, 623–633. [CrossRef] [PubMed]
- 72. Barone, E.; Di Domenico, F.; Cenini, G.; Sultana, R.; Cini, C.; Preziosi, P.; Perluigi, M.; Mancuso, C.; Butterfield, D.A. Biliverdin reductase—A protein levels and activity in the brains of subjects with Alzheimer disease and mild cognitive impairment. *Biochim. Biophys. Acta* 2011, 1812, 480–487. [CrossRef] [PubMed]
- 73. Barone, E.; Di Domenico, F.; Mancuso, C.; Butterfield, D.A. The Janus face of the heme oxygenase/biliverdin reductase system in Alzheimer disease: It's time for reconciliation. *Neurobiol. Dis.* **2014**, *62*, 144–159. [CrossRef]
- 74. Di Domenico, F.; Barone, E.; Mancuso, C.; Perluigi, M.; Cocciolo, A.; Mecocci, P.; Butterfield, D.A.; Coccia, R. HO-1/BVR-a system analysis in plasma from probable Alzheimer's disease and mild cognitive impairment subjects: A potential biochemical marker for the prediction of the disease. J. Alzheimers Dis. 2012, 32, 277–289. [CrossRef]
- 75. Barone, E.; Di Domenico, F.; Cassano, T.; Arena, A.; Tramutola, A.; Lavecchia, M.A.; Coccia, R.; Butterfield, D.A.; Perluigi, M. Impairment of biliverdin reductase-A promotes brain insulin resistance in Alzheimer disease: A new paradigm. *Free Radic. Biol. Med.* 2016, *91*, 127–142. [CrossRef] [PubMed]
- 76. Lanzillotta, C.; Zuliani, I.; Vasavda, C.; Snyder, S.H.; Paul, B.D.; Perluigi, M.; Di Domenico, F.; Barone, E. BVR-A Deficiency Leads to Autophagy Impairment through the Dysregulation of AMPK/mTOR Axis in the Brain-Implications for Neurodegeneration. *Antioxidants* 2020, 9, 671. [CrossRef]
- 77. Triani, F.; Tramutola, A.; Di Domenico, F.; Sharma, N.; Butterfield, D.A.; Head, E.; Perluigi, M.; Barone, E. Biliverdin reductase-A impairment links brain insulin resistance with increased Abeta production in an animal model of aging: Implications for Alzheimer disease. *Biochim. Biophys. Acta Mol. Basis Dis.* 2018, 1864, 3181–3194. [CrossRef]
- 78. Barone, E.; Mancuso, C.; Di Domenico, F.; Sultana, R.; Murphy, M.P.; Head, E.; Butterfield, D.A. Biliverdin reductase-A: A novel drug target for atorvastatin in a dog pre-clinical model of Alzheimer disease. *J. Neurochem.* **2012**, *120*, 135–146. [CrossRef]
- 79. Liu, J.; Dong, H.; Zhang, Y.; Cao, M.; Song, L.; Pan, Q.; Bulmer, A.; Adams, D.B.; Dong, X.; Wang, H. Bilirubin Increases Insulin Sensitivity by Regulating Cholesterol Metabolism, Adipokines and PPARgamma Levels. *Sci. Rep.* **2015**, *5*, 9886. [CrossRef]
- Kipp, Z.A.; Xu, M.; Bates, E.A.; Lee, W.H.; Kern, P.A.; Hinds, T.D., Jr. Bilirubin Levels Are Negatively Correlated with Adiposity in Obese Men and Women, and Its Catabolized Product, Urobilin, Is Positively Associated with Insulin Resistance. *Antioxidants* 2023, 12, 170. [CrossRef]
- Cimini, F.A.; Arena, A.; Barchetta, I.; Tramutola, A.; Ceccarelli, V.; Lanzillotta, C.; Fontana, M.; Bertoccini, L.; Leonetti, F.; Capoccia, D.; et al. Reduced biliverdin reductase-A levels are associated with early alterations of insulin signaling in obesity. *Biochim. Biophys. Acta Mol. Basis Dis.* 2019, 1865, 1490–1501. [CrossRef] [PubMed]
- Sharma, N.; Tramutola, A.; Lanzillotta, C.; Arena, A.; Blarzino, C.; Cassano, T.; Butterfield, D.A.; Di Domenico, F.; Perluigi, M.; Barone, E. Loss of biliverdin reductase-A favors Tau hyper-phosphorylation in Alzheimer's disease. *Neurobiol. Dis.* 2019, 125, 176–189. [CrossRef] [PubMed]
- Li, Y.; Chen, L.; Li, Z.; Song, Y.; Yuan, Y.; Liu, T.; Hong, J.; Wang, Q.; Chang, H.; Kuang, Z.; et al. Potential Serum Biomarkers for Postoperative Neurocognitive Disorders Based on Proteomic Analysis of Cognitive-Related Brain Regions. *Front. Aging Neurosci.* 2021, 13, 741263. [CrossRef] [PubMed]
- 84. He, S.; Li, Y.; Li, T.; Xu, F.; Zeng, D.; Shi, Y.; Zhao, N.; Zhang, L.; Ma, Y.Z.; Wang, Q.; et al. Sex differences between serum total bilirubin levels and cognition in patients with schizophrenia. *BMC Psychiatry* **2021**, *21*, 396. [CrossRef] [PubMed]
- Becklen, M.; Orhan, F.; Piehl, F.; Cervenka, S.; Sellgren, C.M.; Flyckt, L.; Erhardt, S.; Fatouros-Bergman, H. Plasma bilirubin levels are reduced in first-episode psychosis patients and associates to working memory and duration of untreated psychosis. *Sci. Rep.* 2021, *11*, 7527. [CrossRef] [PubMed]
- 86. Moccia, M.; Picillo, M.; Erro, R.; Longo, K.; Amboni, M.; Santangelo, G.; Palladino, R.; Allocca, R.; Caporale, O.; Triassi, M.; et al. Increased bilirubin levels in de novo Parkinson's disease. *Eur. J. Neurol.* **2015**, *22*, 954–959. [CrossRef] [PubMed]
- Talbot, K.; Wang, H.Y.; Kazi, H.; Han, L.Y.; Bakshi, K.P.; Stucky, A.; Fuino, R.L.; Kawaguchi, K.R.; Samoyedny, A.J.; Wilson, R.S.; et al. Demonstrated brain insulin resistance in Alzheimer's disease patients is associated with IGF-1 resistance, IRS-1 dysregulation, and cognitive decline. *J. Clin. Investig.* 2012, 122, 1316–1338. [CrossRef]
- Barone, E.; Tramutola, A.; Triani, F.; Calcagnini, S.; Di Domenico, F.; Ripoli, C.; Gaetani, S.; Grassi, C.; Butterfield, D.A.; Cassano, T.; et al. Biliverdin Reductase-A Mediates the Beneficial Effects of Intranasal Insulin in Alzheimer Disease. *Mol. Neurobiol.* 2019, 56, 2922–2943. [CrossRef]
- 89. Tramutola, A.; Triplett, J.C.; Di Domenico, F.; Niedowicz, D.M.; Murphy, M.P.; Coccia, R.; Perluigi, M.; Butterfield, D.A. Alteration of mTOR signaling occurs early in the progression of Alzheimer disease (AD): Analysis of brain from subjects with pre-clinical AD, amnestic mild cognitive impairment and late-stage AD. *J. Neurochem.* **2015**, *133*, 739–749. [CrossRef]

- Lanzillotta, C.; Tramutola, A.; Di Giacomo, G.; Marini, F.; Butterfield, D.A.; Di Domenico, F.; Perluigi, M.; Barone, E. Insulin resistance, oxidative stress and mitochondrial defects in Ts65dn mice brain: A harmful synergistic path in down syndrome. *Free Radic. Biol. Med.* 2021, 165, 152–170. [CrossRef]
- Hatano, T.; Saiki, S.; Okuzumi, A.; Mohney, R.P.; Hattori, N. Identification of novel biomarkers for Parkinson's disease by metabolomic technologies. J. Neurol. Neurosurg. Psychiatry 2016, 87, 295–301. [CrossRef] [PubMed]
- Albillos, S.M.; Montero, O.; Calvo, S.; Solano-Vila, B.; Trejo, J.M.; Cubo, E. Plasma acyl-carnitines, bilirubin, tyramine and tetrahydro-21-deoxycortisol in Parkinson's disease and essential tremor. A case control biomarker study. *Park. Relat. Disord.* 2021, 91, 167–172. [CrossRef] [PubMed]
- Aydemir, D.; Surucu, S.; Basak, A.N.; Ulusu, N.N. Evaluation of the Hematological and Serum Biochemistry Parameters in the Pre-Symptomatic and Symptomatic Stages of ALS Disease to Support Early Diagnosis and Prognosis. *Cells* 2022, *11*, 3569. [CrossRef] [PubMed]
- Ilzecka, J.; Stelmasiak, Z. Serum bilirubin concentration in patients with amyotrophic lateral sclerosis. *Clin. Neurol. Neurosurg.* 2003, 105, 237–240. [CrossRef] [PubMed]
- 95. Cao, L.; Xue, L.; Luo, D.M. Lower serum bilirubin concentration in patients with migraine. *Int. J. Clin. Exp. Med.* **2015**, *8*, 13398–13402. [PubMed]
- Peng, F.; Deng, X.; Yu, Y.; Chen, X.; Shen, L.; Zhong, X.; Qiu, W.; Jiang, Y.; Zhang, J.; Hu, X. Serum bilirubin concentrations and multiple sclerosis. J. Clin. Neurosci. 2011, 18, 1355–1359. [CrossRef] [PubMed]
- Ljubisavljevic, S.; Stojanovic, I.; Vojinovic, S.; Milojkovic, M.; Dunjic, O.; Stojanov, D.; Pavlovic, D. Association of serum bilirubin and uric acid levels changes during neuroinflammation in patients with initial and relapsed demyelination attacks. *Metab. Brain Dis.* 2013, *28*, 629–638. [CrossRef]
- Vasavda, C.; Semenza, E.R.; Liew, J.; Kothari, R.; Dhindsa, R.S.; Shanmukha, S.; Lin, A.; Tokhunts, R.; Ricco, C.; Snowman, A.M.; et al. Biliverdin reductase bridges focal adhesion kinase to Src to modulate synaptic signaling. *Sci. Signal.* 2022, 15, eabh3066. [CrossRef]
- Cingolani, L.A.; Goda, Y. Actin in action: The interplay between the actin cytoskeleton and synaptic efficacy. *Nat. Rev. Neurosci.* 2008, 9, 344–356. [CrossRef]
- 100. Huganir, R.L.; Nicoll, R.A. AMPARs and synaptic plasticity: The last 25 years. Neuron 2013, 80, 704–717. [CrossRef]
- 101. Sutton, M.A.; Schuman, E.M. Dendritic protein synthesis, synaptic plasticity, and memory. Cell 2006, 127, 49–58. [CrossRef]
- Mayford, M.; Siegelbaum, S.A.; Kandel, E.R. Synapses and memory storage. *Cold Spring Harb. Perspect. Biol.* 2012, 4, a005751.
 [CrossRef]
- Luscher, C.; Malenka, R.C. NMDA receptor-dependent long-term potentiation and long-term depression (LTP/LTD). Cold Spring Harb. Perspect. Biol. 2012, 4, a005710. [CrossRef] [PubMed]
- Mitra, S.K.; Hanson, D.A.; Schlaepfer, D.D. Focal adhesion kinase: In command and control of cell motility. *Nat. Rev. Mol. Cell Biol.* 2005, *6*, 56–68. [CrossRef] [PubMed]
- Schlaepfer, D.D.; Hanks, S.K.; Hunter, T.; van der Geer, P. Integrin-mediated signal transduction linked to Ras pathway by GRB2 binding to focal adhesion kinase. *Nature* 1994, 372, 786–791. [CrossRef] [PubMed]
- Husi, H.; Ward, M.A.; Choudhary, J.S.; Blackstock, W.P.; Grant, S.G. Proteomic analysis of NMDA receptor-adhesion protein signaling complexes. *Nat. Neurosci.* 2000, *3*, 661–669. [CrossRef] [PubMed]
- 107. Yang, Y.C.; Ma, Y.L.; Chen, S.K.; Wang, C.W.; Lee, E.H. Focal adhesion kinase is required, but not sufficient, for the induction of long-term potentiation in dentate gyrus neurons in vivo. J. Neurosci. 2003, 23, 4072–4080. [CrossRef] [PubMed]
- 108. Babayan, A.H.; Kramar, E.A.; Barrett, R.M.; Jafari, M.; Haettig, J.; Chen, L.Y.; Rex, C.S.; Lauterborn, J.C.; Wood, M.A.; Gall, C.M.; et al. Integrin dynamics produce a delayed stage of long-term potentiation and memory consolidation. *J. Neurosci.* 2012, 32, 12854–12861. [CrossRef] [PubMed]
- 109. Cimini, F.A.; Tramutola, A.; Barchetta, I.; Ceccarelli, V.; Gangitano, E.; Lanzillotta, S.; Lanzillotta, C.; Cavallo, M.G.; Barone, E. Dynamic Changes of BVRA Protein Levels Occur in Response to Insulin: A Pilot Study in Humans. *Int. J. Mol. Sci.* 2023, 24, 7282. [CrossRef]
- 110. Xu, J.; Ji, J.; Yan, X.H. Cross-talk between AMPK and mTOR in regulating energy balance. *Crit. Rev. Food Sci. Nutr.* **2012**, *52*, 373–381. [CrossRef]
- 111. Mancuso, C. Biliverdin reductase as a target in drug research and development: Facts and hypotheses. *Free Radic. Biol. Med.* **2021**, 172, 521–529. [CrossRef]
- 112. Gibbs, P.E.; Miralem, T.; Maines, M.D. Biliverdin reductase: A target for cancer therapy? Front. Pharmacol. 2015, 6, 119. [CrossRef]
- 113. Ai, W.; Bae, S.; Ke, Q.; Su, S.; Li, R.; Chen, Y.; Yoo, D.; Lee, E.; Jon, S.; Kang, P.M. Bilirubin Nanoparticles Protect Against Cardiac Ischemia/Reperfusion Injury in Mice. J. Am. Heart Assoc. 2021, 10, e021212. [CrossRef]
- 114. Hinds, T.D., Jr.; Creeden, J.F.; Gordon, D.M.; Stec, D.F.; Donald, M.C.; Stec, D.E. Bilirubin Nanoparticles Reduce Diet-Induced Hepatic Steatosis, Improve Fat Utilization, and Increase Plasma beta-Hydroxybutyrate. *Front. Pharmacol.* 2020, *11*, 594574. [CrossRef] [PubMed]
- 115. Lee, Y.; Kim, H.; Kang, S.; Lee, J.; Park, J.; Jon, S. Bilirubin Nanoparticles as a Nanomedicine for Anti-inflammation Therapy. *Angew. Chem. Int. Ed. Engl.* **2016**, *55*, 7460–7463. [CrossRef]
- 116. Sundararaghavan, V.L.; Sindhwani, P.; Hinds, T.D., Jr. Glucuronidation and UGT isozymes in bladder: New targets for the treatment of uroepithelial carcinomas? *Oncotarget* 2017, *8*, 3640–3648. [CrossRef] [PubMed]

- 117. Kataura, T.; Saiki, S.; Ishikawa, K.I.; Akamatsu, W.; Sasazawa, Y.; Hattori, N.; Imoto, M. BRUP-1, an intracellular bilirubin modulator, exerts neuroprotective activity in a cellular Parkinson's disease model. *J. Neurochem.* 2020, 155, 81–97. [CrossRef] [PubMed]
- 118. Barone, E.; Calabrese, V.; Mancuso, C. Ferulic acid and its therapeutic potential as a hormetin for age-related diseases. *Biogerontology* **2009**, *10*, 97–108. [CrossRef] [PubMed]
- Catino, S.; Paciello, F.; Miceli, F.; Rolesi, R.; Troiani, D.; Calabrese, V.; Santangelo, R.; Mancuso, C. Ferulic Acid Regulates the Nrf2/Heme Oxygenase-1 System and Counteracts Trimethyltin-Induced Neuronal Damage in the Human Neuroblastoma Cell Line SH-SY5Y. Front. Pharmacol. 2015, 6, 305. [CrossRef] [PubMed]
- 120. Hou, Y.; Dan, X.; Babbar, M.; Wei, Y.; Hasselbalch, S.G.; Croteau, D.L.; Bohr, V.A. Ageing as a risk factor for neurodegenerative disease. *Nat. Rev. Neurol.* 2019, *15*, 565–581. [CrossRef]
- 121. Garagnani, P.; Marquis, J.; Delledonne, M.; Pirazzini, C.; Marasco, E.; Kwiatkowska, K.M.; Iannuzzi, V.; Bacalini, M.G.; Valsesia, A.; Carayol, J.; et al. Whole-genome sequencing analysis of semi-supercentenarians. *elife* **2021**, *10*, e57849. [CrossRef] [PubMed]

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