An OpenCV-Based Approach for Automated Cardiac Rhythm Measurement in Zebrafish from Video Datasets

Ali Farhan

PhD fellow

Chung Yuan Christian University
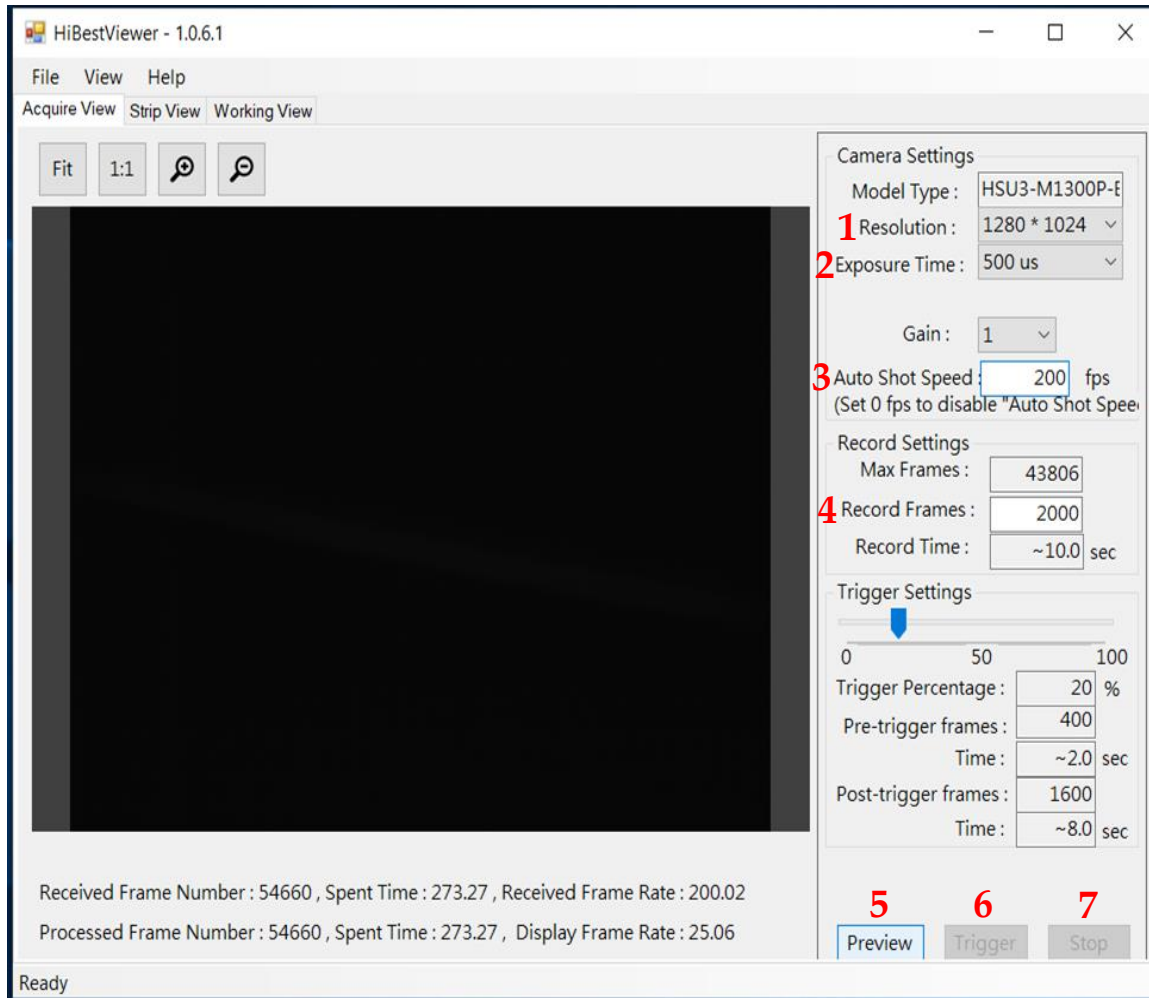
Supervisor: Prof. Chung-Der Hsiao

Introduction

- Using AI-based OpenCV library to detect automatically cardiac rhythm in zebrafish larvae videos

- Advanced features of Python programing language enables the user to perform analysis on 2D video using multiprocessing to select atrium and ventricle region of Interest (ROI) simultaneously.

- Results being validated on the basis of comparison with previously published method of this lab as ImageJ

Setup for automatic cardiac rhythm analysis for zebrafish larvae

➢ 2D video recording using high speed CCD

➢ Installation of Python 3.8 or above version

➢ Download essential computer vision packages

➢ Open video in Python (mp4 format)

➢ Run the program

➢ Select ROI for atrium and ventricle

➢ Analysis data

➢ Results presentation

Video Recording using HiBestViewer Application
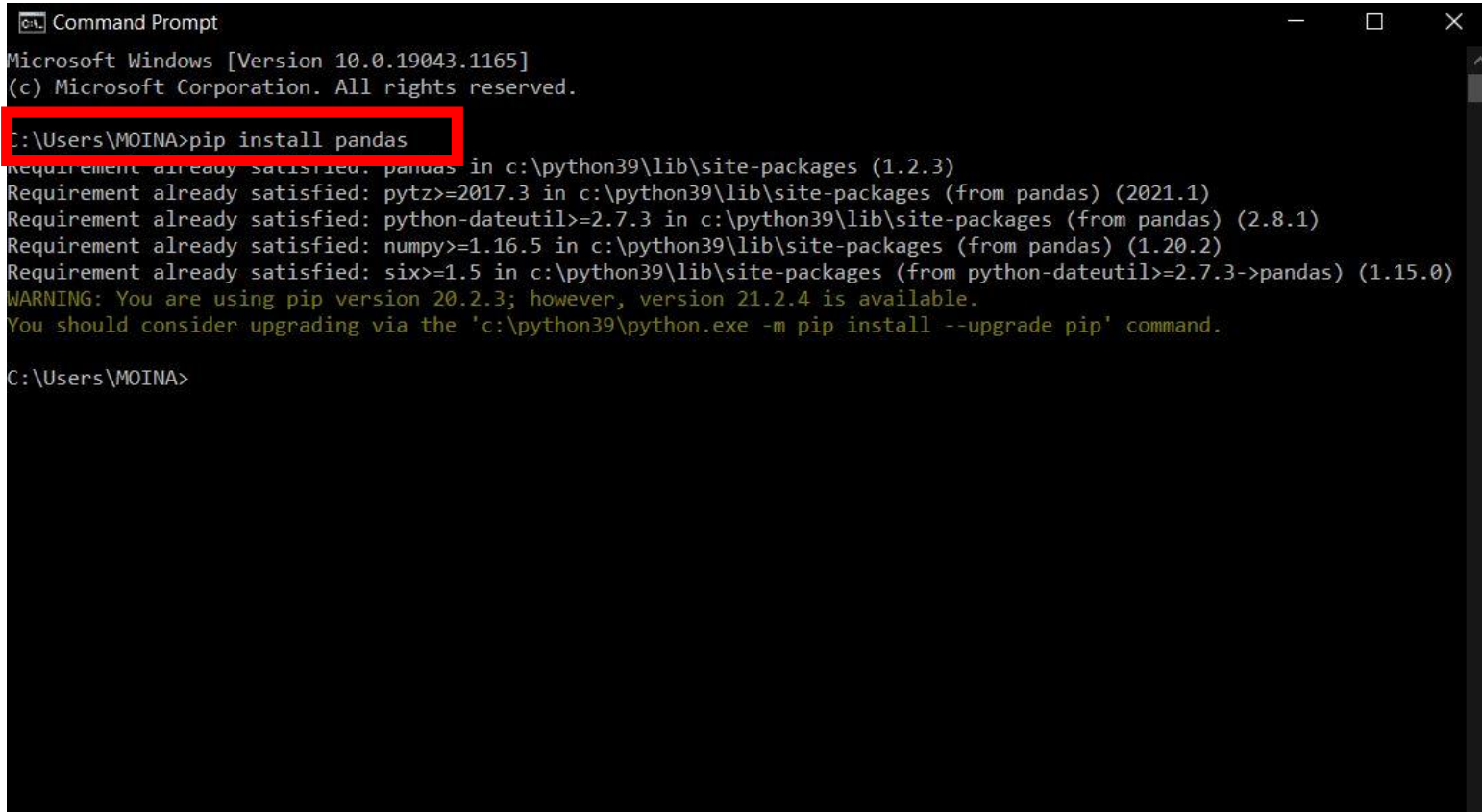## (AZ Instrument, Taiwan)

1. **Resolution :** Size of the video taken with a linear correlation to sharpness and clarity.

2. **Exposure time** camera exposed **:** duration of time in which the sensor within the to light

3. **Auto Shot Speed :** number of frames taken in one second for the recording (fps)

4. **Record Frame :** Number of frame set manually by the user to determine the recording length. The final duration of the video is presented in **Record Time**. This number cannot exceed the **Max Frames** or **0**.

5. **Preview :** Start viewing the current condition of the object of interest under the microscope.

6. **Trigger :** Start recording

7. **Stop :** Stop the recording or the viewing.

Steps to Initialize program

- Put mp4 format video in desired location or folder in system
- Download Python 3.8 version from(https://www.python.org/downloads/)
- Create folder with name as per desired by the user
- Place the zebrafish heart video in the folder and make title as "test.mp4"(at one time one video can be processed)
- Open command prompt window from 'start' panel of system
- Program does not require GPU it can be processed on average CPU
- No need to use anaconda prompt

Necessary commands use to import libraries in command prompt

➤Pip install argparse

➤Pip install imutlis

➤Pip install opencv-python

➤Pip install scipy

➤Pip install numpy

➤Pip install matplotlib

➤Pip install pandas

➤Pip install cydets

➤Pip install rainflow
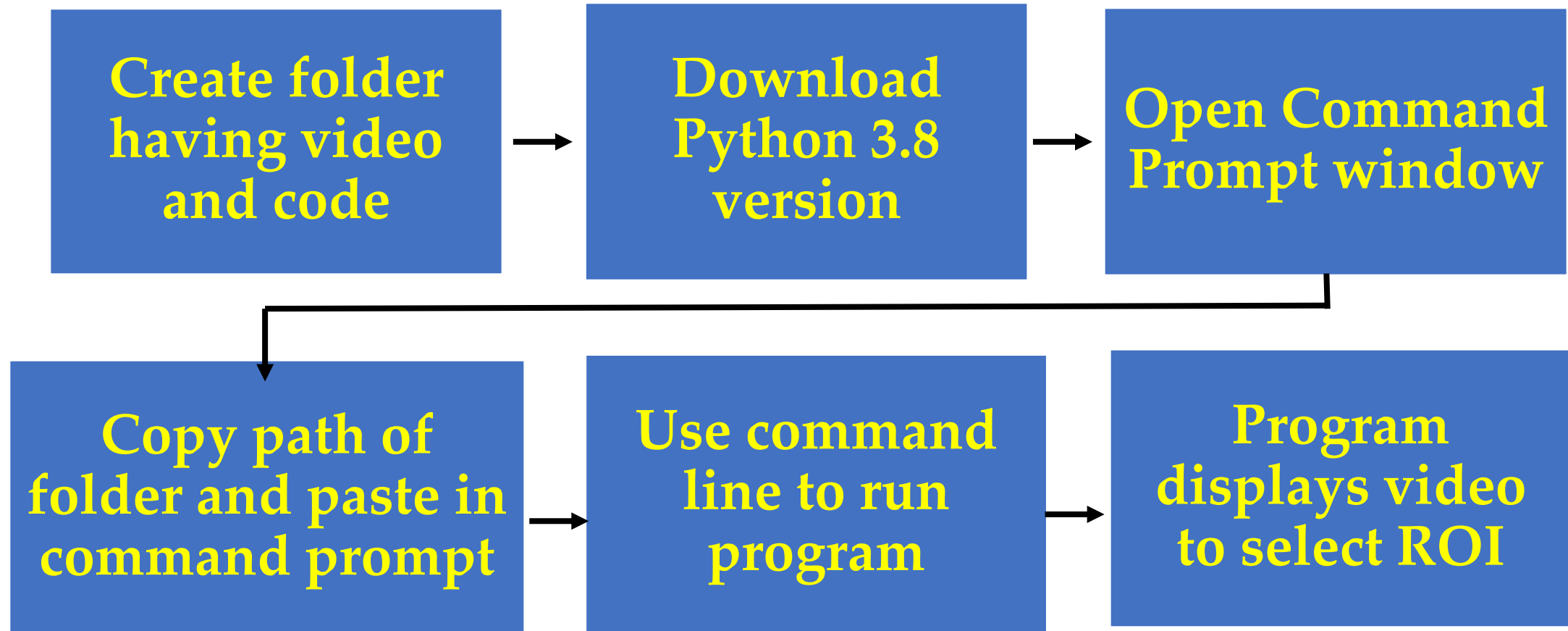
Operational Flowchart for non-expert users

```
┌─────────────────┐      ┌─────────────────┐      ┌─────────────────┐
│  Create folder  │      │    Download     │      │                 │
│  having video   │ ───► │  Python 3.8     │ ───► │  Open Command   │
│    and code     │      │    version      │      │ Prompt window   │
└─────────────────┘      └─────────────────┘      └─────────────────┘

┌─────────────────┐      ┌─────────────────┐      ┌─────────────────┐
│  Copy path of   │      │  Use command    │      │    Program      │
│ folder and paste│ ───► │  line to run    │ ───► │ displays video  │
│ in command      │      │   program       │      │ to select ROI   │
│   prompt        │      │                 │      │                 │
└─────────────────┘      └─────────────────┘      └─────────────────┘
```

Folder having code file and video

Command prompt Window

Paste Path of folder

Command Prompt Window

Copy path of folder

Continued…..

- By using command line to copy from .txt file in folder paste in the command prompt window


```
C:\Windows\System32\cmd.exe

Microsoft Windows [Version 10.0.19043.1237]
(c) Microsoft Corporation. All rights reserved.

C:\Users\MOINA\Desktop\My research work\OpenCv\OpenCv Program Heart Rate>
C:\Users\MOINA\Desktop\My research work\OpenCv\OpenCv Program Heart Rate>python opencv_object.py --video test.mp4
```

```
# Command to run

python opencv_object.py --video test.mp4
```

Continued…..

- After completing the installation requirements and program command lines user does not require to do further any step to 'open the video". Video shall be opened automatically after copy and paste the command line.

- Once video displayed, users have to select ROI(s) in the video and execution will start automatically till message box displays with 'heart rate and peaks will be shown as well'.

Continued…..

- Press Enter to continue
- Program will show message box to select ROI in video file

Selecting ROIs in Zebrafish heart video

Ventricle (V)

Atrium (A)

It is necessary to select accurate ROI (region of interesting), otherwise results will be compromised as Tool cannot detect atrium and ventricle itself

Display of Execution

Window to show peaks for first ROI

Window to show peaks for second ROI

Success rate with execution to time (s)

Calculating Rhythm for different frame rate setting (200 and 60 fps as examples)

```
time_list = [len(s_atri_time), len(s_ventri_time), len(s_atri_heart), len(s_ventri_heart), len(peaks), len(peaks2), len(i
time_len = min(time_list)
s_atri_time = s_atri_time[:time_len]
s_ventri_time = s_ventri_time[:time_len]
s_atri_heart = s_atri_heart[:time_len]
s_ventri_heart = s_ventri_heart[:time_len]
peaks = peaks[1:time_len+1]
peaks2 = peaks2[1:time_len+1]
inv_peaks = inv_peaks[1:time_len+1]
inv_peaks2 = inv_peaks2[1:time_len+1]


final = {
    'Time interval (Atrium)':               s_atri_time,
    'Time interval (Ventricle)':            s_ventri_time,
    'Heart rate (Atrium)':                  s_atri_heart,
    'Heart rate (Ventricle)':               s_ventri_heart,
    'High peaks (Atrium)':                  peaks,
    'High peaks (Ventricle)':               peaks2,
    'Low peaks (Atrium)':                   inv_peaks,
    'Low peaks (Ventricle)':                inv_peaks2
}


df_final = pd.DataFrame(data=final)
df_final.to_csv('result_final.csv', index = False)



get_interval_1(inv_peaks, inv_peaks2, interval_1_cof)
get_interval_2(inv_peaks, inv_peaks2, interval_2_cof)
```
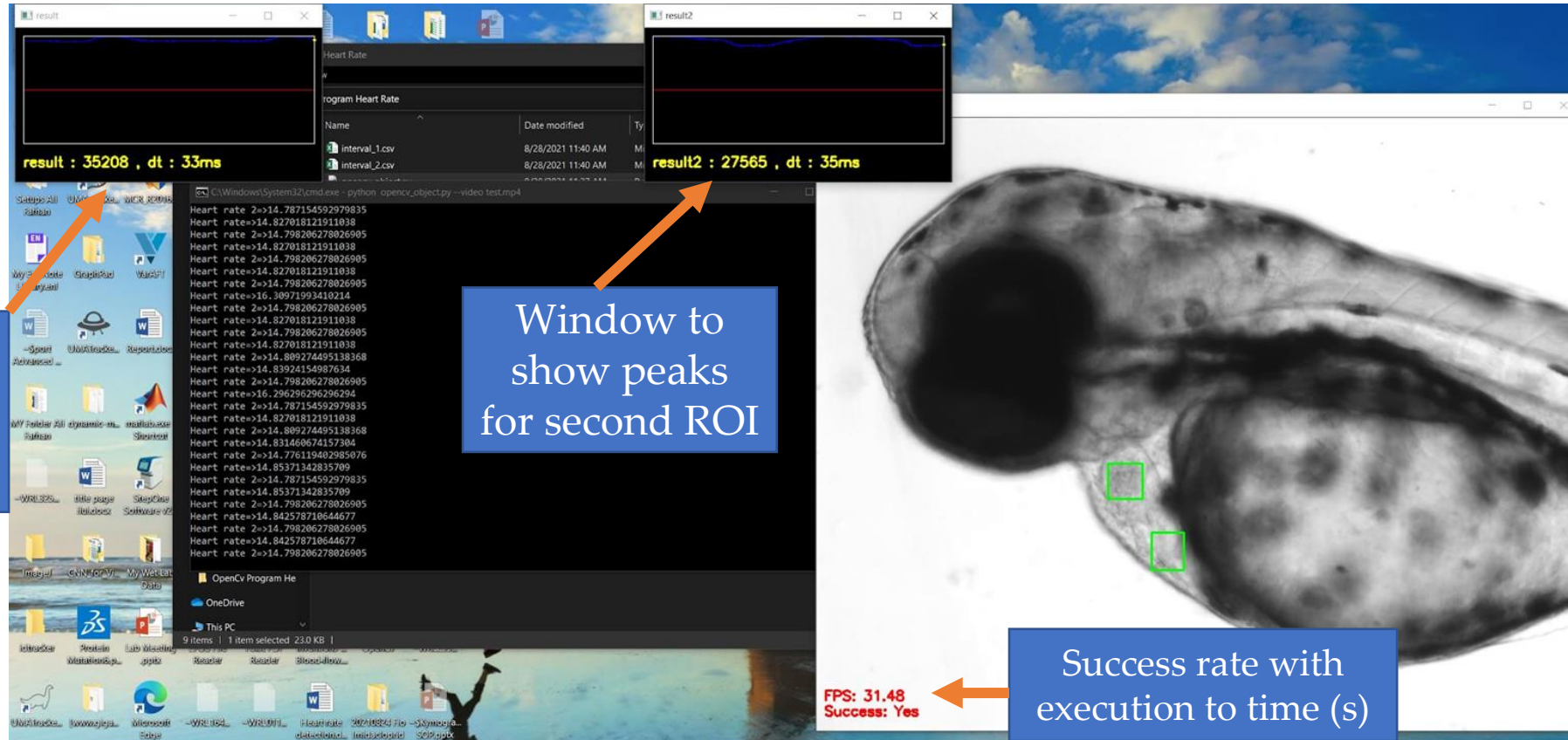
Script first check the length of video then compute inverse peaks

Final data for Time interval for atrium and ventricle in ROIs

15

Calculating Rhythm for different frame rate setting (200 and 60 fps as examples)

```python
inv_atrium_data = -atrium_data
inv_ventricle_data = -ventricle_data

interval_1_cof = 0.005
interval_2_cof = 200
if real_fps > 35:
    peaks = signal.find_peaks(atrium_data, distance=10)[0]
    peaks2 = signal.find_peaks(ventricle_data, distance=10)[0]
    inv_peaks = signal.find_peaks(inv_atrium_data, distance=10)[0]
    inv_peaks2 = signal.find_peaks(inv_ventricle_data, distance=10)[0]
    interval_1_cof = 0.016
    interval_2_cof = 60
elif 20 < real_fps and real_fps < 35:
    peaks = signal.find_peaks(atrium_data, distance=80)[0]
    peaks2 = signal.find_peaks(ventricle_data, distance=80)[0]
    inv_peaks = signal.find_peaks(inv_atrium_data, distance=80)[0]
    inv_peaks2 = signal.find_peaks(inv_ventricle_data, distance=80)[0]

###########################################
# Calculate heartrate and time interval
_, s_atri_time, s_atri_heart = get_heartrate(peaks)
_, s_ventri_time, s_ventri_heart = get_heartrate(peaks2)
```
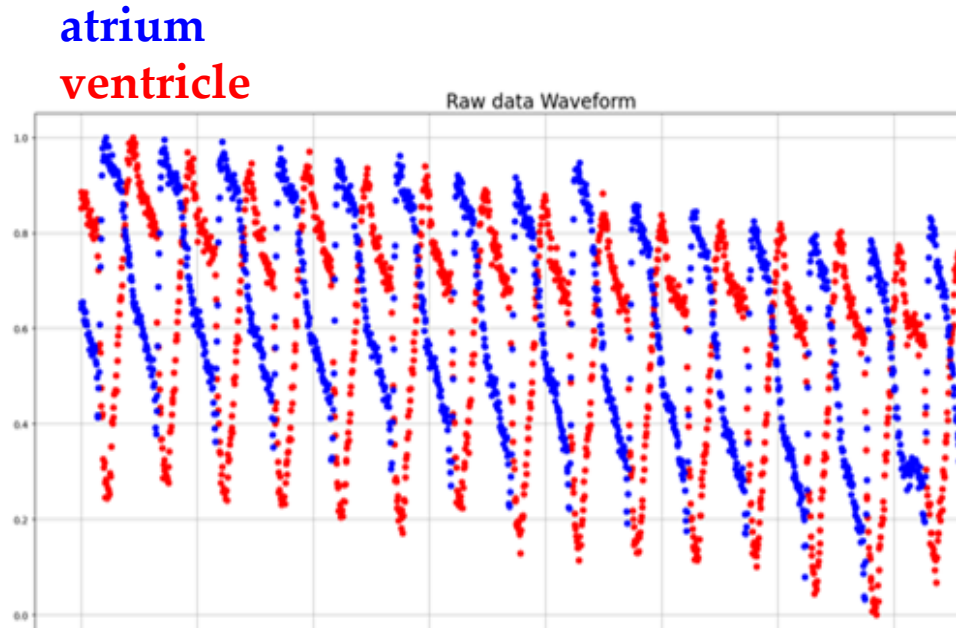
For 200 fps video, 1/200=0.005 sec

For 60 fps video, 1/60=0.016 sec

Raw Data Waveform patterns

atrium
ventricle

Raw data Waveform

Peaks detected display as raw data
(number of frames)

Preprocessed data Waveform

Spline Interpolation used to process the
raw data waveform for smoothing

Smoothing of waveforms

- Using Python scipy-module as interpolate
- Apply multivariate data interpolation
- Griddata (function)
- Check dataset as cubic or linear
- Apply cubic griddata function
- 2D data smoothing with float optional inv
- Call function CloughTocher2DInterpolator

Functions for smoothening

__call__(x, y[, dx, dy, grid])	Evaluate the spline or its derivatives at given positions.
ev(xi, yi[, dx, dy])	Evaluate the spline at points
get_coeffs()	Return spline coefficients.
get_knots()	Return a tuple (tx,ty) where tx,ty contain knots positions of the spline with respect to x-, y-variable, respectively.
get_residual()	Return weighted sum of squared residuals of the spline approximation: sum ((w[i]*(z[i]-s(x[i],y[i])))**2,axis=0)
integral(xa, xb, ya, yb)	Evaluate the integral of the spline over area [xa,xb] x [ya,yb].

Logic design for waveform smoothening

- # #######################################

- # # x_data = Frame, y_data = Atrium

- # # x_data2 = Frame, y_data2 = Ventricle

- # Split time length

- # #######################################

- time_list = [len(s_atri_time), len(s_ventri_time), len(s_atri_heart), len(s_ventri_heart), len(peaks), len(peaks2), len(inv_peaks), len(inv_peaks2)]

- time_len = min(time_list) -1

- s_atri_time = s_atri_time[:time_len]

- s_ventri_time = s_ventri_time[:time_len]

- s_atri_heart = s_atri_heart[:time_len]

- s_ventri_heart = s_ventri_heart[:time_len]

- peaks = peaks[1:time_len+1]

- peaks2 = peaks2[1:time_len+1]

- inv_peaks = inv_peaks[1:time_len+1]

- inv_peaks2 = inv_peaks2[1:time_len+1]

Functions for time interval

Function to extract atrium and ventricle difference (time)

Atrium=atri
Ventrcile=ventri
Defined variables to save data

Module in scipy: to find all peaks in array

Logic design for waveform smoothening

This function takes a 1-D array and finds all local maxima by simple comparison of neighboring values

- **Parameters**

points*ndarray of floats, shape (npoints, ndims); or Delaunay* (Data point coordinates, or a precomputed

Delaunay triangulation)

values*ndarray of float or complex, shape (npoints, …)*

Data values as **fill_value***float, optional*

Value used to fill in for requested points outside of the convex hull of the input points. If not provided

then the default is nan.

tol*float, optional*

- Absolute/relative tolerance for gradient estimation.
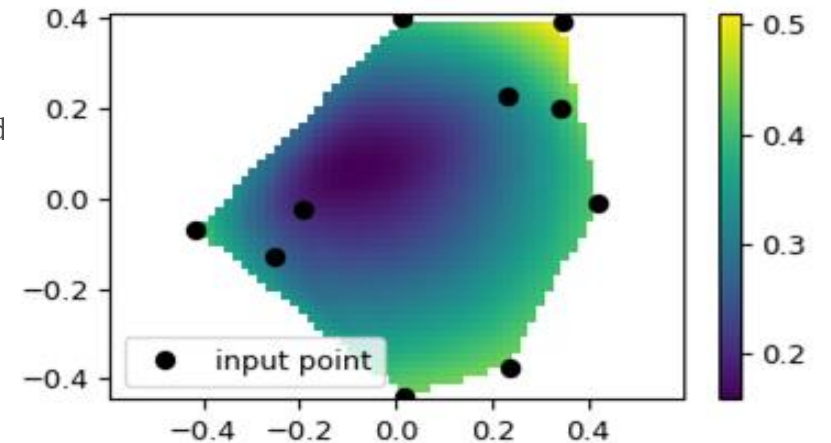
maxiter*int, optional*

- Maximum number of iterations in gradient estimation.

rescale*bool, optional*

- Rescale points to unit cube before performing interpolation.

This is useful if some of the input dimensions have incommensurable units and differ by many orders of
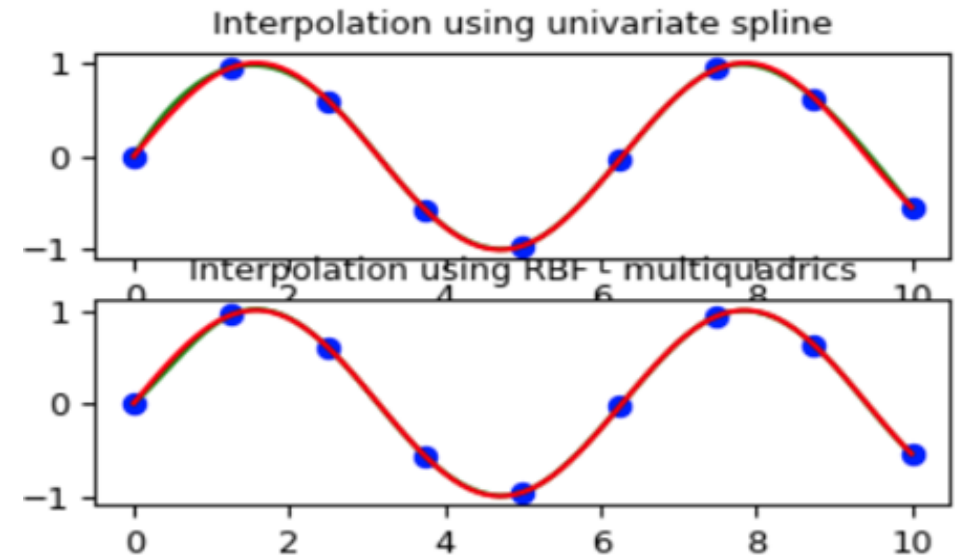
magnitude.

Evaluate parameters

Logic design for waveform smoothening

2-d tests - setup scattered data >>> rng **=** np**.**random**.**default_rng**()** >>> x **=** rng**.**random**(100)*4.0-2.0** >>> y **=** rng**.**random**(100)*4.0-2.0** >>> z **=** x*np**.**exp**(-**x**2**-y**2) >>> edges = np**.**linspace**(-2.0, 2.0, 101)** >>> centers **=** edges**[:-1] +** np**.**diff**(**edges**[:2])[0] / 2.** >>> XI, YI **=** np**.**meshgrid**(**centers, centers**)**

Scipy built-in module for edges

Reference:
https://docs.scipy.org/doc/scipy/reference/tutorial/interpolate.html#d-spline-representation-object-oriented-bivariatespline

Finalize dataplots
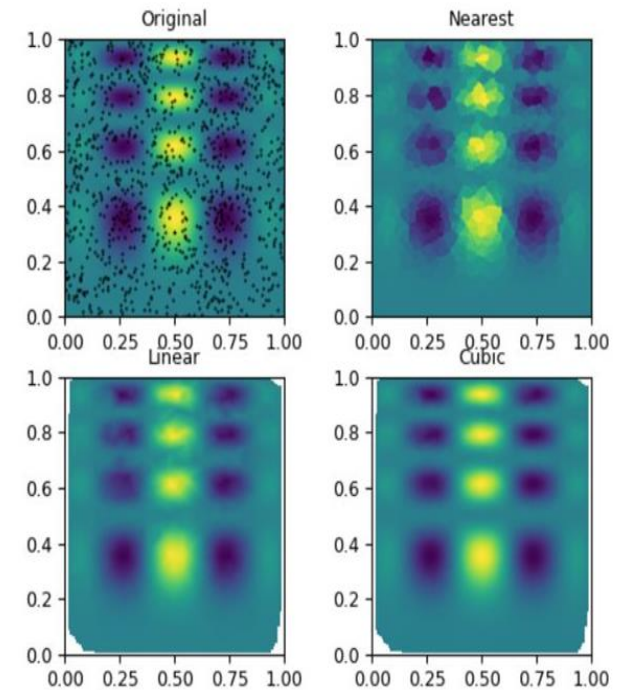
Use **scipy.interpolate.BivariateSpline** ← Module to confined the x, y peaks with spline interpolation

This describes a spline s(x, y) of degrees kx and ky on the rectangle [xb, xe] * [yb, ye] calculated from a given set of data points (x, y, z).

This class is meant to be subclassed, not instantiated directly. To construct these splines, call either **SmoothBivariateSpline** or **LSQBivariateSpline** or **RectBivariateSpline Compare**

```
import matplotlib.pyplot as plt >>> plt.subplot(221) >>> plt.imshow(func(grid_x, grid_y).T, extent=(0,1,0,1), origin='lower') >>> plt.plot(points[:,0], points[:,1], 'k.', ms=1) >>> plt.title('Original') >>> plt.subplot(222) >>> plt.imshow(grid_z0.T, extent=(0,1,0,1), origin='lower') >>> plt.title('Nearest') >>> plt.subplot(223) >>> plt.imshow(grid_z1.T, extent=(0,1,0,1), origin='lower') >>> plt.title('Linear') >>> plt.subplot(224) >>> plt.imshow(grid_z2.T, extent=(0,1,0,1), origin='lower') >>> plt.title('Cubic') >>> plt.gcf().set_size_inches(6, 6) >>> plt.show()
```

Finalize dataplots

- s = UnivariateSpline(x_data, y_data, s=50) ← Class for spline fit data 1D
- yy_data = s(x_data)
- peaks = findPeak(yy_data)

- y_data2 = np.array(pplotpix2)
- x_data2 = np.arange(0, len(y_data2))
- y_data2 = normalize_array(y_data2)
- raw_data2 = std_normalize(y_data2)

- s2 = UnivariateSpline(x_data2, y_data2, s=50)
- yy_data2 = s2(x_data2)
- peaks2 = findPeak(yy_data2)

Script (modified)

Display Raw data

plt.title("Raw data Waveform", fontsize = 20)

- plt.plot(raw_data, 'bo', label = "Atrium")

- plt.plot(raw_data2, 'ro', label = "Ventricle")

- plt.grid()
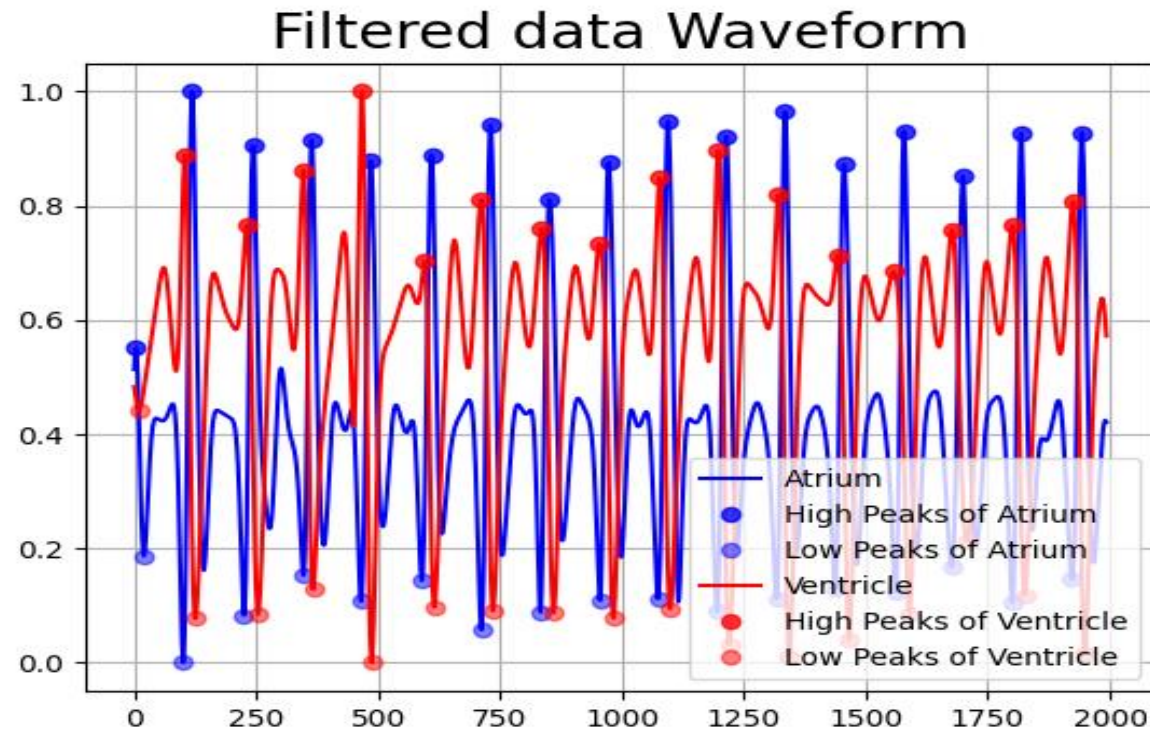
- plt.legend(loc = 4)

- plt.show()

Display splined data

- plt.title("Preprocessed data Waveform", fontsize = 20)

- plt.plot(yy_data, color = "b", label = "Atrium")

- plt.plot(yy_data2, color = "r", label = "Ventricle")

Function(s) to make color as "Blue" for Atrium and Red for "ventricle"

- plt.grid()

- plt.legend(loc = 4)

- plt.show()

Display filtered data

- plt.title("Filtered data Waveform", fontsize = 20)

- plt.plot(atrium_data, color = "b", label = "Atrium")

- plt.plot(peaks, atrium_data[peaks], 'bo', label = "High Peaks of Atrium", alpha = 0.8,)

- plt.plot(inv_peaks, -inv_atrium_data[inv_peaks], 'bo', alpha = 0.5, label = "Low Peaks of Atrium")

- plt.plot(ventricle_data, color = "r", label = "Ventricle")

- plt.plot(peaks2, ventricle_data[peaks2], 'ro', label = "High Peaks of Ventricle", alpha = 0.8,)

- plt.plot(inv_peaks2, -inv_ventricle_data[inv_peaks2], 'ro', alpha = 0.5, label = "Low Peaks of Ventricle")

- plt.grid()

- plt.legend(loc = 4)

- plt.show()

Smoothing of Peaks shown

Filtered data Waveform

Peaks smoothing performed to get clear visualization as blue for atrium and red for ventricle

Final Data saved as .csv file

Name	Date modified	Type	Size
interval_1.csv	8/28/2021 11:55 AM	Microsoft Excel Co...	1 KB
interval_2.csv	8/28/2021 11:55 AM	Microsoft Excel Co...	1 KB
opencv_object.py	8/28/2021 11:37 AM	Python File	24 KB
readme.txt	8/26/2021 1:46 PM	Text Document	1 KB
result_bps.csv	8/28/2021 11:55 AM	Microsoft Excel Co...	11 KB
result_bps2.csv	8/28/2021 11:55 AM	Microsoft Excel Co...	12 KB
result_final.csv	8/28/2021 11:55 AM	Microsoft Excel Co...	1 KB
test.mp4	3/2/2021 12:08 PM	MP4 File	32,445 KB
to_ali.docx	4/15/2021 2:20 PM	Microsoft Word D...	127 KB

A	B	C	D	E	F	G	H	I
Time inter	Time inter	Heart rate	Heart rate	High peaks	High peaks	Low peaks	Low peaks (Ventricle)	
116	80	15.51724	22.5	121	184	226	124	
125	106	14.4	16.98113	246	290	347	254	
118	171	15.25424	10.52632	364	461	467	376	
123	100	14.63415	18	487	561	590	486	
123	96	14.63415	18.75	610	657	712	617	
122	177	14.7541	10.16949	732	834	834	735	
121	240	14.87603	7.5	853	1074	955	860	
121	123	14.87603	14.63415	974	1197	1074	992	
121	123	14.87603	14.63415	1095	1320	1196	1098	
121	182	14.87603	9.89011	1216	1502	1318	1220	
120	121	15	14.87603	1336	1623	1438	1344	
120	126	15	14.28571	1456	1749	1561	1466	
123	128	14.63415	14.0625	1579	1877	1676	1589	
121	108	14.87603	16.66667	1700	1985	1801	1710	

For AV-VA Interval saved in .csv file as interval 1 and 2 respectively

Heart rate measurement for *Daphnia magna*

- In order to determine heart rate of Daphnia as it has single chamber heart the ROI selection was created with bounding box as single ROI selection

- The single chamber heart of Daphnia displays single waveform as compared to zebrafish for two chambers (atrium and ventricle)

Heart rate measurement for *Daphnia magna*

➢ Similar protocol used for *Daphnia magna* heart rate measurement

➢ User will have to write option "Daphnia/Daphnia_35/Daphnia_IMI" after its shown in command prompt
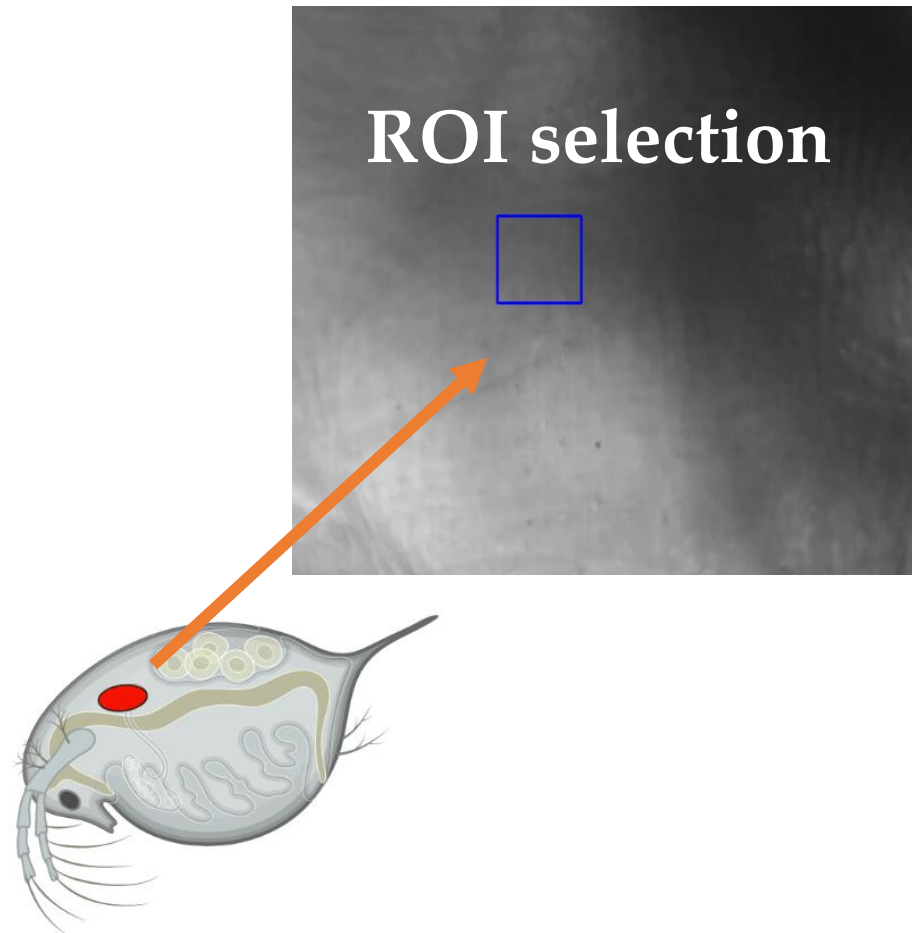
```
C:\Windows\System32\cmd.exe - python opencv_object.py --video test.mp4

Microsoft Windows [Version 10.0.19043.1165]
(c) Microsoft Corporation. All rights reserved.

C:\Users\MOINA\Desktop\OpenCv Program Heart Rate>python opencv_object.py --video test.mp4
Input the animal name ( Fish/Daphnia/Daphnia_35/Daphnia_IMI)  Daphnia
```

➢ Daphnia (For Normal Heart Rate without any toxicity test)
➢ Daphnia_35 for Temperature effect
➢ Daphnia_IMI for IMI induced to check Dysregulation

These options created as algorithm designed for zebrafish here we added scales to validate the results of heartbeat in *Daphnia magna* as additional feature
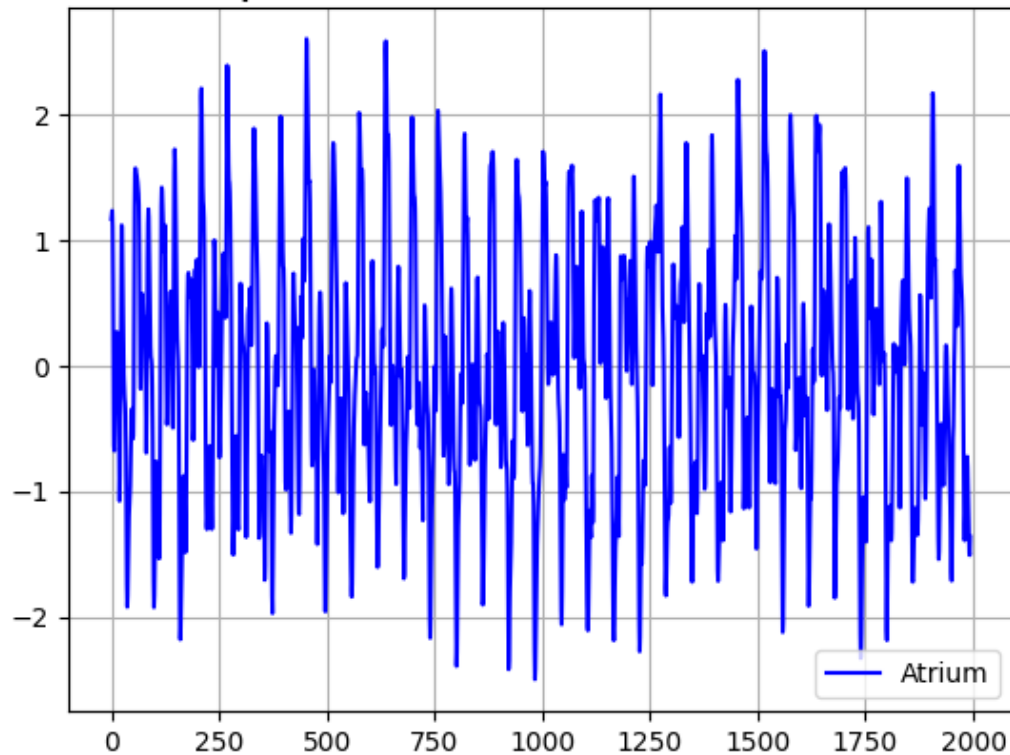
ROI selection in the heart of *Daphnia magna*

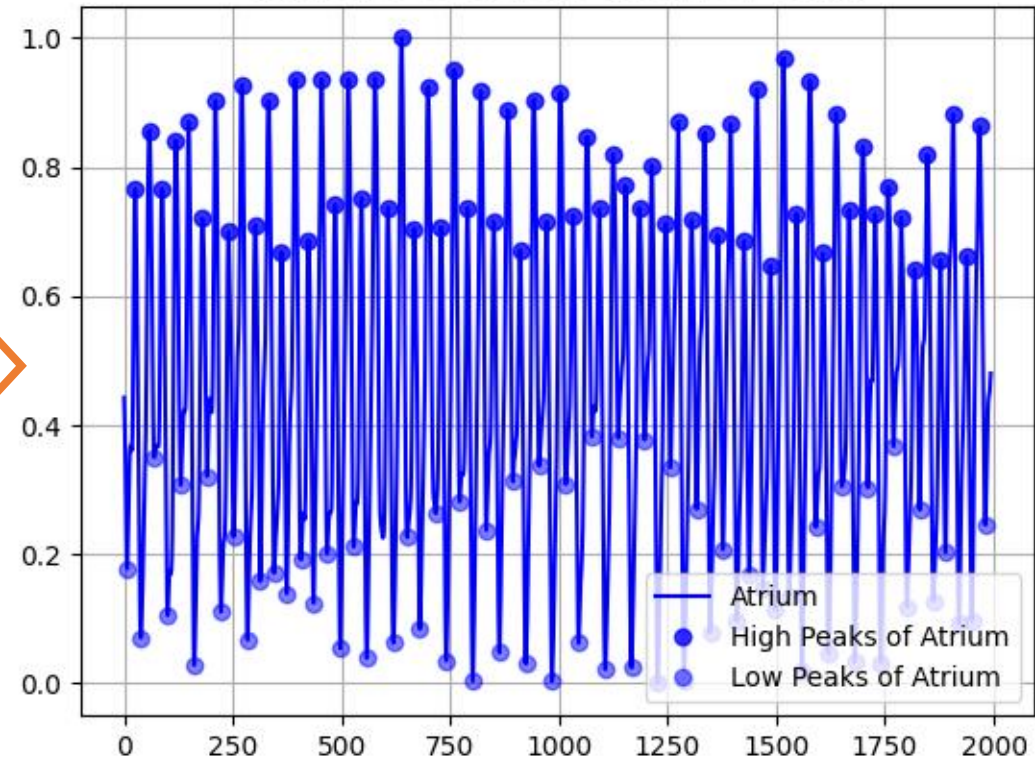| B	C	D	E
Heart rate	High peaks	Low peaks (Atrium)	
54.54545	58	39	
62.06897	87	69	
56.25	119	100	
62.06897	148	130	
56.25	180	161	
60	210	191	
60	240	223	
58.06452	271	252	
58.06452	302	284	
60	332	313	
60	362	345	
56.25	394	374	
60	424	407	
60	454	436	
58.06452	485	467	
58.06452	516	497	
60	546	528	
58.06452	577	558	
60	607	619	
58.06452	638	651	
62.06897	667	680	
56.25	699	717	
60	729	741	
58.06452	760	772	
60	790	802	
60	820	833	
60	850	863	
54.54545	883	894	
64.28571	911	924	
58.06452	942	955	
60	972	984	
58.06452	1003	1016	
60	1033	1045	
56.25	1065	1076	
64.28571	1093	1107	
56.25	1125	1137	
64.28571	1153	1167	
56.25	1185	1197	
64.28571	1213	1227	

Data showed more heartbeat events as compared to zebrafish due to *Daphnia magna* has higher heart rate than zebrafish

Waveform for *Daphnia magna* heartbeat

OpenCV method can also be used to detect ultrafast heartbeat in *Daphnia magna*

Key Features of OpenCV method for heart cardiac rhythm detection

- **No need to do video format conversion**

- **No need any manual calculation for heartbeat**

- **Automatic detection of rhythm and A-V and V-A intervals**

- **Results saved in respective folder using csv format automatically**

- **No need to install or incorporate any external plugins for the analysis**

- **In zebrafish, both atrium and ventricle rhythm can be detected simultaneously**