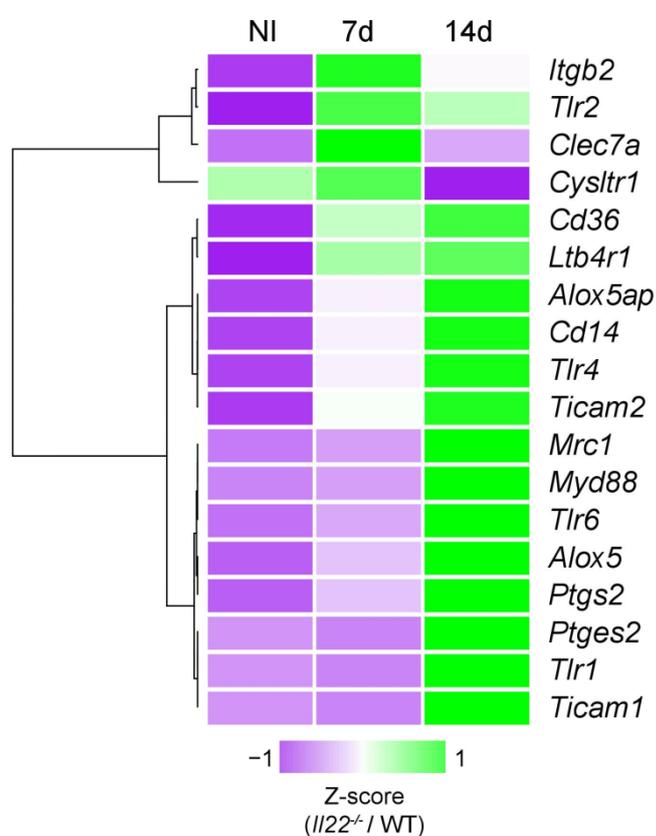


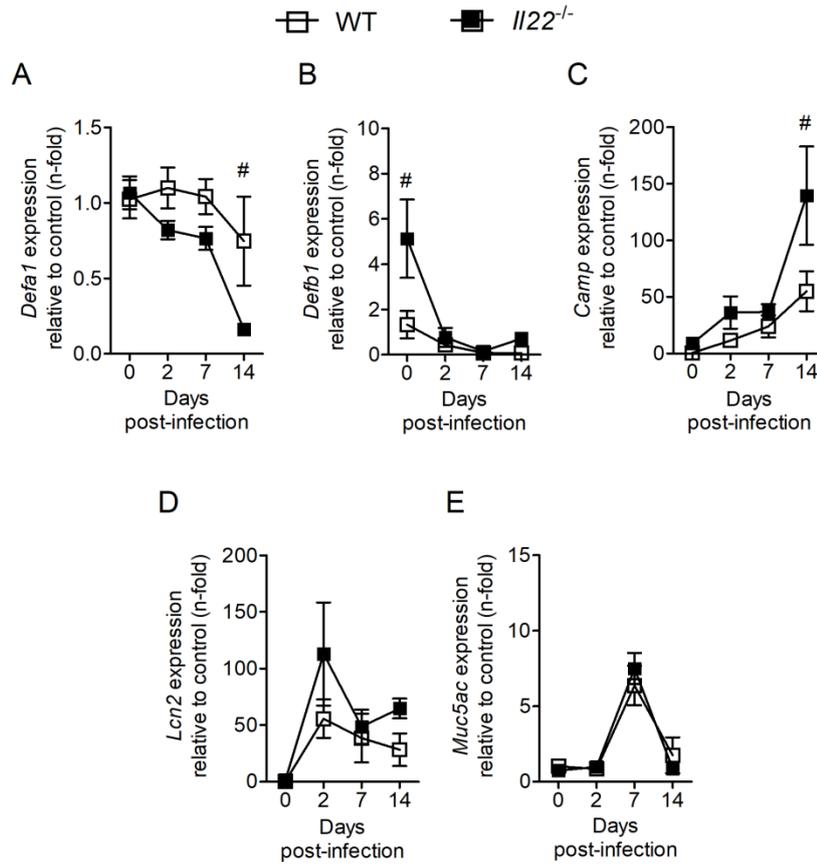
## IL-22 promotes IFN- $\gamma$ -mediated immunity against *Histoplasma capsulatum* infection

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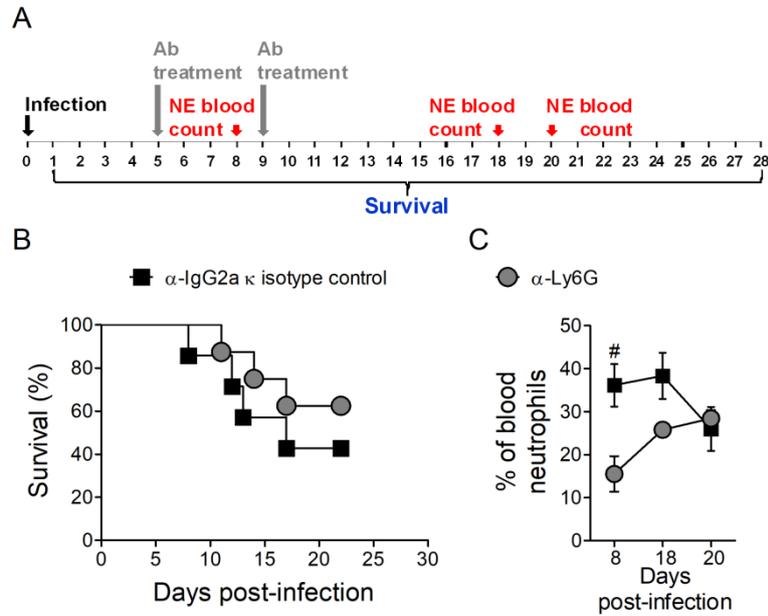
### Supplementary Material



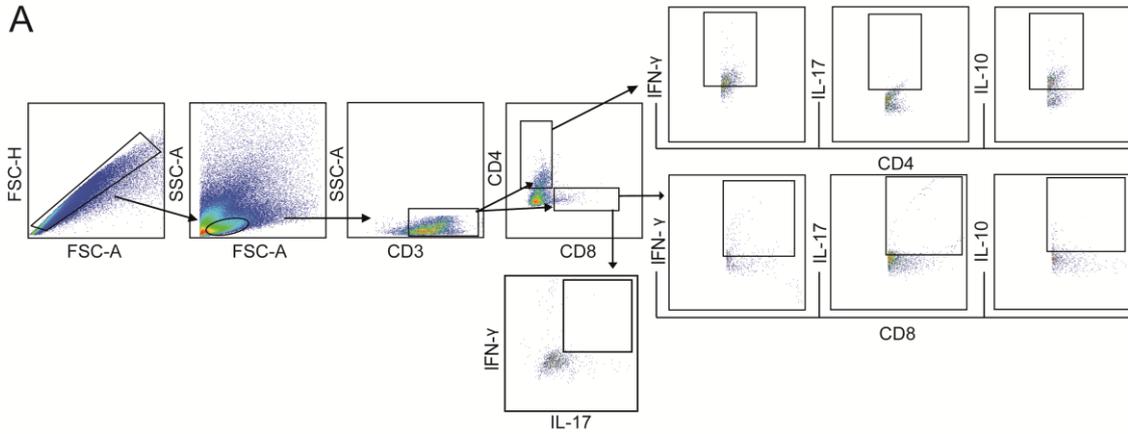
**Figure S1. Gene expression of *Il22*<sup>-/-</sup>/WT mice during *H. capsulatum* infection.** A 96-well precoated plate (Mouse Custom Plates RT<sup>2</sup> Profiler PCR Array; Qiagen – Beverly, MA, EUA) together with the RT<sup>2</sup> SYBR Green qPCR master mixes (330523; Qiagen – Beverly, MA, EUA) was used to examine the relative expression set of of *Itgb2*, *Tlr2*, *Clec7a*, *Cysltr1*, *Cd36*, *Ltb4r1*, *Alox5ap*, *Cd14*, *Tlr4*, *Ticam2*, *Mrc1*, *Myd88*, *Tlr6*, *Alox5*, *Ptgs2*, *Ptges2*, *Tlr1* and *Ticam1* genes in lung samples from eight-weeks-old *Il22*<sup>-/-</sup> and WT mice infected with *H. capsulatum* and respective controls (Eppendorf Mastercycler ep Realplex – Hamburg, Alemanha). The cycle thresholds (Ct) were used to calculate the relative expression of the target genes, which were normalized to the housekeeping gene *Tbp* (TATA-box-binding protein) and analyzed with the  $2^{-\Delta\Delta Ct}$  method. The heat map was generated using the R Language and Environment for Statistical Computing v.3.5.0 and the package *gplots*. Hierarchical clustering was performed with the package *amap* using the correlation method and ward linkage. NI= non-infected; 7d and 14d = 7<sup>th</sup> and 14<sup>th</sup> days post-infection.



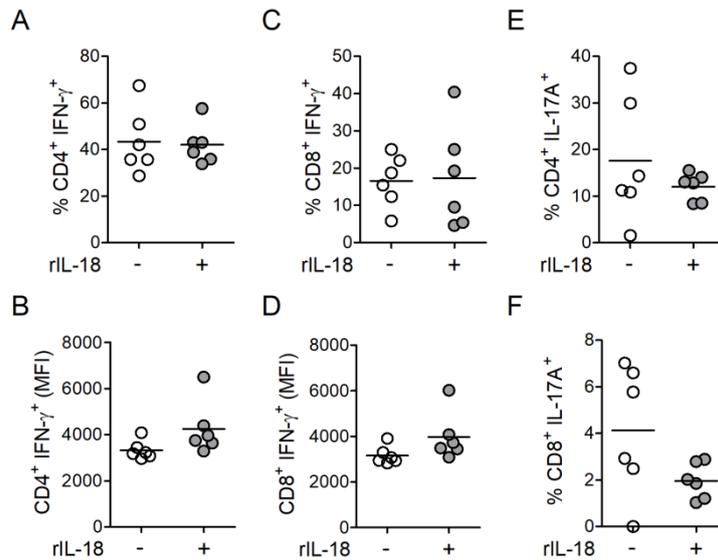
**Figure S2. Lung transcriptional expression of *Defa1*, *Defb1*, *Camp*, *Lcn2* and *Muc5ac* during *H. capsulatum* infection.** Eight-week-old WT or *Il22*<sup>-/-</sup> mice were infected i.t. with lethal inoculum ( $1 \times 10^6$  yeast) of *H. capsulatum*. The left lower lobe was removed and the RNA extracted for analysis of the gene expression of *Defa1* (A), *Defb1* (B), *Camp* (C), *Lcn2* (D) and *Muc5ac* (E) by RT qPCR using Taqman primers (Mm02524428\_g1; Mm00432803\_m1; Mm00438285\_m1; Mm01324470\_m1 and Mm01276718\_m1). *Gapdh* (Mm99999915\_g1) was used as reference genes and to normalize expression levels by  $\Delta\Delta C_t$  method. The expression data was presented as n-fold difference relative to the control group. Data were expressed as mean  $\pm$  SEM of a representative experiment ( $n = 4-7$ ) and analyzed by the Two-Way ANOVA test and Bonferroni post-test. Data were considered statistically significant when  $p < 0.05$ , and #WT + *Hc* vs. *Il22*<sup>-/-</sup> + *Hc*.



**Figure S3. Depletion of circulating neutrophils did not enhance survival of *Il22*<sup>-/-</sup> mice infected with *H. capsulatum*.** Schematic design of the neutrophil depletion experiment. Eight-weeks-old *Il22*<sup>-/-</sup> mice were infected with lethal inoculum ( $1 \times 10^6$  yeasts) of *H. capsulatum* and treated at days five and nine post-infection with 120  $\mu$ g of specific-Ab (i.p.) (A). The animals were observed daily to determine the survival curve (B). Data were analyzed by Log-rank test and Mantel-Cox in the post-test ( $n = 7-8$ ). Data were considered statistically significant when  $p < 0.05$ , <sup>#</sup>*Il22*<sup>-/-</sup> + *Hc* +  $\alpha$ -Ly6G vs. *Il22*<sup>-/-</sup> + *Hc* +  $\alpha$ -IgG2a  $\kappa$  isotype control. At 8<sup>th</sup>, 18<sup>th</sup> and 20<sup>th</sup> days post infection a sample of blood from tail vein were used for differential count of circulating cells using panoptic stain (C). Data were expressed as mean  $\pm$  SEM of a representative experiment ( $n = 4-8$ ) and analyzed by the Two-Way ANOVA test and Bonferroni post-test. Data were considered statistically significant when  $p < 0.05$ , and <sup>#</sup>*Il22*<sup>-/-</sup> + *Hc* +  $\alpha$ -Ly6G vs. *Il22*<sup>-/-</sup> + *Hc* +  $\alpha$ -IgG2a  $\kappa$  isotype control.



**Figure S4. Schematic illustration of the gating hierarchy used for lymphocyte analysis.** Pseudocolor plots show representative flow cytometric data of CD4 and CD8 T lymphocytes intracellularly stained for IFN- $\gamma$ , IL-17A or IL-10 production.



**Figure S5. In vitro stimulation of lung parenchyma with IL-18 did not increase polarization of Th1 or Th17 cells.** Eight-week-old WT or *Il22*<sup>-/-</sup> mice were infected i.t. with lethal inoculum ( $1 \times 10^6$  yeast) of *H. capsulatum*. The upper left lobe was removed, disrupted and stimulated or not with 50 ng.mL<sup>-1</sup> of recombinant mouse IL-18, PMA, ionomycin and brefeldin A for 12 hours. The cells were then evaluated for the percentage of Th1 (CD3<sup>+</sup>CD4<sup>+</sup>IFN- $\gamma$ <sup>+</sup>) (A), Tc1 (CD3<sup>+</sup>CD8<sup>+</sup>IFN- $\gamma$ <sup>+</sup>) (C), Th17 (CD3<sup>+</sup>CD4<sup>+</sup>IL-17A<sup>+</sup>) and Tc17 (CD3<sup>+</sup>CD8<sup>+</sup>IL-17A<sup>+</sup>) by flow cytometry. The median of fluorescence intensity (MFI) of IFN- $\gamma$ <sup>+</sup> CD4<sup>+</sup> (B) or CD8<sup>+</sup> (D) T cells were either evaluated. Data were analyzed by the Two-Way ANOVA test and Bonferroni in the post-test and considered significant when  $p < 0.05$ , <sup>#</sup>WT + *Hc* vs. WT + *Hc* + rIL-18 or <sup>#</sup>*Il22*<sup>-/-</sup> + *Hc* vs. *Il22*<sup>-/-</sup> + *Hc* + rIL-18 ( $n = 4-6$ ) from a representative experiment.