Chain-selective Isotopic Labeling of the Heterodimeric Type III Secretion Chaperone, Scc4:Scc1, Reveals the Total Structural Rearrangement of the *Chlamydia trachomatis* Bifunctional Protein, Scc4

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Figure S1. Expression and purification of ¹⁵N-labeled Scc4:His₆-Scc1. (A) SDS-PAGE analysis with lanes numbered at the bottom. Lanes 1 and 2 are proteins in the culture before (-IPTG) and after (+IPTG) induction. Lanes 3 and 4 are the pellet and clarified lysate after cell lysis and centrifugation. Lanes 5, 6, and 7 are the lysate that flowed through the Ni-IMAC resin (FT), buffer wash step, and elution of the Scc4:His₆-Scc1 complex using imidazole buffer, respectively. Lane 8 is the Fisher BioReagents EZ-Run Rec pre-stained protein ladder with molecular weights listed to the right in kDa. Lanes 9 and 10 are samples from the SEC purification of the complex shown in (B). The arrow heads indicate the bands corresponding to Scc4 (14.7 kDa) and His₆-Scc1 (18.8 kDa). (B) SEC purification of the Scc4:His₆-Scc1 imidazole elution produced two peaks labeled 1 and 2. The SDS-PAGE analysis of these peaks are shown in lanes 9 and 10, respectively, in (A). The second peak contains the purified Scc4:His₆-Scc1 complex.



Figure S2. Expression and purification of ¹⁵N-labeled His₆-Scc4:Scc1-FLAG. (A) SDS-PAGE analysis with lanes numbered at the bottom. Lanes 1 and 2 are proteins in the culture before (-IPTG) and after (+IPTG) induction. Lanes 3 and 4 are the pellet (Pellet) and clarified lysate (Lysate) after cell lysis and centrifugation. Lanes 5, 6, and 7 are the lysate that flowed through the Ni-IMAC resin (FT), buffer wash step (Wash), and elution of the His₆-Scc4:Scc1-FLAG complex using imidazole buffer (Elution), respectively. Lane 8 is the Bio-Rad Precision Plus protein ladder with molecular weights listed to the right in kDa. The arrow heads indicate the bands corresponding to His₆-Scc4 (15.8 kDa) and Scc1-FLAG (18.2 kDa). (B) SEC purification of the His₆-Scc4:Scc1-FLAG imidazole elution produced two peaks labeled 1 and 2. The second peak contains the purified His₆-Scc4:Scc1-FLAG complex.



Figure S3. 2D ¹H-¹⁵N HSQC spectra of the *in vivo*-associated, ¹⁵N-labeled protein complexes Scc4:His6-Scc1 (blue) and His6-Scc4:Scc1-FLAG (pink) with the (A) full spectra and (B) expanded regions from the dashed box in (A). The complexes were 0.35 mM concentration in 50 mM sodium phosphate, 10 mM DTT, pH 7.3 buffer, and the spectra were collected with 32 scans using a Bruker AVIII 500 MHz spectrometer.



Figure S4. Full gel images of the native gels from Figure 4A. Lane numbers across the bottom correspond to the lane numbers in Figure 4A.



Figure S5. (A-F) ¹H,¹⁵N-HSQC spectra of ¹⁵N-Scc4 titrated with Scc1-FLAG. Relative moles of Scc1-FLAG to ¹⁵N-Scc4 added are (A) 0, (B) 0.25, (C) 0.5, (D) 1.0, (E) 1.5, and (F) 2.0. The spectra were collected on a Bruker AVIII 500 MHz spectrometer with 8 scans for each spectrum, with the exception

of spectrum (F), which was collected with 32 scans. The contour levels were corrected for the number of scans to allow for direct comparison.



Figure S6. ¹H,¹⁵N-HSQC peak changes during the Scc1-FLAG titration of Scc4 mapped onto the Scc4 homology model. The homology model is shown as a ribbon diagram (top) and in all-atom representation (bottom) with three points of view via rotation about the vertical axis shown. The HSQC peaks were identified using the published assignments [1]. Light blue residues indicate peaks that are unassigned [1] or cannot be positively identified or tracked due to overlap during the titration. The disappearance of peaks due to broadening at 0.25 – 0.5X addition of Scc1-FLAG are shaded dark purple and at 1.0 – 1.5X addition of Scc1-FLAG are shaded dark blue. The identified residues with chemical shift perturbations > 0.2 ppm (0.4 ppm maximum) are shaded in medium purple and the perturbations < 0.2 ppm are shaded in medium blue. The color scheme is based on palettes for color blindness and interpretability in grayscale [2].



Figure S7. SDS-PAGE analysis of the His6-Scc1 inclusion body purification. Lanes are numbered at the bottom. Lanes 1 and 2 are proteins in the culture before and after induction with IPTG. Lanes 3 and 4 are the clarified lysate and the pellet after cell lysis. Lanes 5 and 6 are the inclusion body and supernatant from washing the pellet in Lane 4 with 1X BugBuster in Tris buffer. Lanes 7 and 8 are the inclusion body and supernatant from the second wash step with lysozyme in Tris buffer. Lanes 9 and 10 are the inclusion body and supernatant from the last wash step with Tris buffer. Lane 11 is the Bio-Rad Precision Plus protein ladder with molecular weights listed in kDa to the right. The His6-Scc1 (18.8 kDa) band is indicated with an arrow.



Figure S8. Full gel image of the native gel from Figure 6A. Lane numbers across the bottom correspond to the lane numbers in Figure 6A.



Figure S9. SDS-PAGE analysis of the renatured and chain-selectively labeled Scc4:His6-Scc1 complexes: (A) ¹⁵N-Scc4:His6-Scc1 and (B) Scc4:¹⁵N-His6-Scc1. Filled arrow heads indicate the His6-Scc1 bands and open arrow heads indicate the Scc4 bands. The bold font indicates the protein that is ¹⁵N-labeled. (A) Samples of the cultures before and after induction with IPTG and expressing His6-Scc1 (lanes 1 and 2) or ¹⁵N-Scc4 (lanes 4 and 5) are shown. The His6-Scc1 pellet (lane 3) and ¹⁵N-Scc4 clarified lysate (lane 6) were used to make the renatured complex. Lanes 7 and 9 show the purification of the renatured complex by Ni-IMAC and SEC, respectively. Lane 8 is the Bio-Rad Precision Plus protein ladder and lane 10 is the Fisher BioReagents EZ-Run Rec pre-stained protein ladder with molecular weights listed to the right of each lane in kDa. (B) The same samples with opposite isotopic labeling are shown with the same lanes numbers as in (A). The difference between (B) and (A) is the labeling of the proteins with ¹⁵N-His6-Scc1 and unlabeled Scc4.

13 d1xkpc1 Probab=97.69 E-value=0.001 Score=41.63 Aligned_columns=112 Identities=19%

Q	ss_pred		HHHHHCCCCCEEEEEEEEEEEEEEEEEEEEEEEEEEEE		
ñ	Scc4-20190820	٩	EVAVMGVASELEEDADGSVVEDTSSLVRMRVRONADEETTTSAELGET-DASMDTEKAVA	67	(133)
Š	Conconsus	0	f lal L D a C L id il d lip la D	67	(1))
Q	consensus	9	+++ + - - +++ +- ++++- + ++ ++ +++	67	(122)
т	Conconcus	6	A Sal GaB EvADa - GaVbl nTD - any (all ans gave 1) at Planana la demanana la h	61	(121)
÷	d1vkmo1	ć		C1	(121)
+	итхкрст	0		01	. (121)
	ss_dssp		HHHHIICCCCCCC-CCEEEEECC-CEEEEEEIIEEEEEECCCGGGEEIIEECHHHHH		
Т	ss_pred		HHHHCCCCEECCC-CEEEEEEE-EEEEEEECCCEEEEECCCCCC		
Т	ss_conf		7865188300027-74889821-06788605992677323331112461222389999		
Q	ss_pred		ННННННСССССССЕЕЕЕЕСССССЕЕЕЕЕЕССС-ССНННННННН		
Q	ss_conf		9997514576665428998179968999843344-6999999999999999999999986		
Q	Scc4-20190820	68	RMMEGNLFGQETGGAALGLDSDGHAVLVRRVPGE-VSQEDFASYIESVLNYAEAWLEDL	125	(133)
Q	Consensus	68	~LL~~N~~~~t~g~~lgl~~~~~vl~~~~a~-l~~~f~~~l~~Fv~~a~~W~~~l	125	(133)
			+ + ++++ ++++++++++++++++		
Т	Consensus	62	~Llqqv~~W~rryPqalVLDd~G~L~LeARL~L~~LD~~~L~e~l~Q-iaLLE~L~PqL	119	(121)
Т	d1xkpc1	62	SLMQQALAWAKRYPQTLVLDDCGQLVLEARLRLQELDTHGLQEVINQ-LALLEHLIPQL	119	(121)
Т	ss_dssp		HHHHHHHHHTTTCCCEEEECTTSCEEEEEEEGGGCCHHHHHHHHCC-HHHHHHGGGG		
Т	ss_pred		ННННННННННННСССЕЕЕЕЕСССССННННННННСССССНННННН		
Т	ss conf		99999998755289148870233005542230111498999999999-78698760315		

Figure S10. Phyre2 [3] alignment of *CT* Scc4 and *Yersinia pestis* YscB from PDB 1XKP, chain C [4]. Details for interpreting the alignment are available on the Phyre2 web page [5].

12 d1xkpb1 Probab	b=97.60 E-value=0.0038 Score=37.47 Aligned_columns=113 Identities=	17%
Q ss_pred Q ss_conf Q CtScc1-intensi Q Consensus	HHHHHHHHHHCCCC-EEECCCCEEEEEEECCCCEEEEEEEE	(146) (146)
T Consensus	3 in~v~eF~r~mG~~~~~v~L~~e~~G-tL~iE~~~e~L~l~lar~l~~~56	(119)
T d1xkpb1 T ss_dssp T ss_pred T ss_conf	3 IEPIISHFCQDLGVPTSSPLS-PLIQLEMAQSG-TLQLEQHGATLTLWLARSLAWH 56 THHHHHHHHHHHTTCCCCSSCC-SEEEEETTTE-EEEEEETTEEEEEEECCGG HHHHHHHHHHHHHCCCCCCCCCCC-CEEEEEECCCC-EEEEEEECCCCEEEEEE	(119)
Q ss_pred	ССНИНИНИНИННСССССССЕЕЕЕЕЕСССССЕЕЕЕЕЕСИНИССИНИНИНИН	
Q ss_conf	6689999999963455126761899836896099999624444899999999999999999	
Q CtScc1-intensi	64 FRQKIFKAALSINGSPQSNIKGTLGYGEISNQLYLCDRLNMTYLNGEKLARYLVLFSQHA 123	(146)
Q Consensus	64 ~r~~~~~LL~~NI~~~t~g~~lg1~~~~~11L~~~~~~l~~~~l~~~~l~~~~l~~~	(146)
T Consensus	57 -~~~~kal~l~hy~~~~pl~aG~~-ge~~Lvl~~~l~~~~tl~~le~A~~~L~~l~ 114	(119)
T d1xkpb1	57 -RCEDAMVALTLTAÁQSGALPLRAGWL-GESQLVLFVSLDERSLTLPLLHQAFEQLLRLQ 114	(119)
T ss_dssp	-GHHHHHCCTTTTSCCCCSSCCEEEE-TTTEEEEEEEEGGGCCHHHHHHHHHHHHHHHH	
T ss_pred	-СНННННННННННСССССССЕЕЕЕЕС-СССЕЕЕЕЕЕЕСНННССНННННННН	
T ss_conf	-20669999986377778742010026-998199999818443367999999999999999	
0		
Q ss_pred	HH	
Q SS_CONT 0 CtScc1_intensi	124 NT 125 (146)	
	124 Mi $125 (146)$	
ę consciisus	+.	
T Consensus	115 ge 116 (119)	
T d1xkpb1	115 QE 116 (119)	
T ss_dssp	HH	
T ss_pred	HH	
T ss_conf	98	

Figure S11. Phyre2 [3] alignment of *CT* Scc1 and *Yersinia pestis* SycN from PDB 1XKP, chain B [4]. Details for interpreting the alignment are available on the Phyre2 web page [5].

3	c1xkpA_ Probab=	100.00	0 E-value=9.4e-40	Score=268.46	Aligned_columns=212	Identit	ies=	=14%
Q	ss pred	(ссссссссссннннннн	нннннннннн	нннннннннннннн	нсссснн		
Q	ss conf	3	35567630006365420135	66788767557788	88855444568988887643	1022136		
Q	CtCopN-intensi	37 (2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	ATRIKKKEEKFESI	LEARRKPTADKAEKKSESTE	EKGDTPL	96	(421)
Q	Consensus	37 4	- -~~s~ls~~~~~el~~~Aa	a~~r~~~eek~~~~	raE	~k~~~dl	96	(421)
•		-	++ .+++++ +	+++	+ +++	+		. ,
Т	Consensus	2 -	Ge-vs-ladaa	EEmS~1~~~~	r	~~~~	46	(217)
Т	c1xkpA_	2	RGESVQIVSGTLQSIADMA	EEVTELSLDR	LSDSQARVSD	VEEQV	46	(217)
Т	ss_dssp	I	BTTBCCCCCCSSCCCHHHHH	HHHCCCCCCC ·	CCCHHHHHHH	HHHHH		
Т	ss_pred	(CCCCEEECCCCCCHHHHH	 		HHHHH		
Т	ss_conf	-	76870055167566287699	99999999998	7876643335	66799		
Q	ss_pred	H	ннннсссссссннннннн	нннсссссннннн	нннннссснннннннннн	HHHCCCC		
Q	ss_conf		78730111111158899999	74067999899999	99998748812678999999	9622463		
Q	CtCopN-intensi	97 I	DRFTEDLSEVSGEDFRGLK	NSFDDDSSSDEIL	DALTSKFSDPTIKDLALDYL	IQIAPSD	156	(421)
Q	Consensus	97 <i>·</i>	-dr~~~~~e~~~q~L~~~F	≀~al~~d~S~~~il~	~~~~e~f~DpS~q~~AL~~1	~q~~~~	156	(421)
			+++.+.+- ++++-	• • • • • • • • • • • • • • • • • • • •	+.++++ + ++ ++ .++	.++		
Т	Consensus	47 <i>·</i>	k1-		~~a~q~FpD~Sd~~laL~~~	~~~~~~	104	(217)
Т	c1xkpA_	47 I	VQYLSVPELEQQNVSELL	.SLLSNSPNISLSQI	LAYLEGSEEPSEQFMLCGLR	DALGRPE	104	(217)
Т	ss_dssp	I	HHHCCCCCCC CHHHHH	HHHSSCSSCCTTT	сстттссссннннсснннн	HHCCCGG		
Т	ss_pred	I	ННННННННН НННННН	нннсссссннннн	нннннссссснннннннн	HHHCCCC		
Т	ss_conf	9	999742346899999999	999424689599999	999975899028999999999	9704689		
Q	ss_pred	ł	-ннннннннннннннссо	СННННННННННН	ННННННСССННННННННН	HHCCCCH		
Q	ss_conf		2-58999999999999972065	577798888999999	999865048989999999999	8638622		
Q	CtCopN-intensi	157 (G-KLKSALIQAKHQLMSQNF	QAIVGGRNVLLAS	ETFASRANTSPSSLRSLYFQ	VTSSPSN	215	(421)
Q	Consensus	157	g-el~~~l~~a~~el~~~~e , ++++.+++++++++	g∼aIrAGiNvAl~A~ ++ + + ++	~~Fa~~l~~sp~~LRdlYR~ +. ++++++ +. ++ +	~v~~s~ 	215	(421)
Т	Consensus	105 ·	lial	~~i~AGiNvAl~A	~~Fs~~~~~~LR~1YR~	~V~~~~	164	(217)
Т	c1xkpA	105 I	AHLSHLVEQALVSMAEEQO	GETIVLGARITPEA	YRESQSGVNPLQPLRDTYRD	avmgyqg	164	(217)
Т	ss_dssp	(<u> ЭННННННННННННННННН</u>	нннннннннн	нннттѕѕсснннннннн	нннсссс		
Т	ss_pred	I	ннннннннннннннссн	нннннннннн	нннннссссннннннннн	HHCCCCC		
Т	ss_conf		7889999999999999982073	877798651468999	999874168989999999999	9617654		
_								
Q	ss_pred	I	ннннннннсснннннн	нннннннннннн	СССССННННННННННН			
Q	ss_cont		/9999999846904599999	999999998555421	18888999999999987998			
Q	CtCopN-intensi	216 (ANLHQMLASYSPSEKTAV	1EFLVNGMVADLKS	GPSIPPAKLQVYMTELSN	267 (421)	
Q	Consensus	216 -	~d~y~s~L~rYg~~~~~vj ++++ ++ +++ +	L~FL~~aL~~DL~S^ -+ + ++ + +	~~PS~~~~L~~llt~l~~ ++ + + +++ ++ .+	267 (421)	
Т	Consensus	165 <i>·</i>	v~~~~i~~fg~~~~~v]	l~fl-~aL~~Dl~S∽	~~PS~~~-el~~1m~~l~k	214 (217)	
Т	c1xkpA_	165 3	[YAIWSDLQRFPNGDIDSV]	LFL-QALSADLQS	QQSGSGR-ELGIVISDLQL	214 (217)	
Т	ss_dssp	I	ННННННСССТТЅСНННН	ННН-ССННННННК	CCCSTTT-CCHHHHHHHCC			
Т	ss_pred	I	ннннннннсснннннн	ННН-НННННННК	ССССНН-НННННННННН			
Т	ss_conf	1	399999999823925599999	9999-9999766423	3899736-99999999866			

Figure S12. Phyre2 [3] alignment of *CT* CopN and *Yersinia pestis* YopN from PDB 1XKP, chain A [4]. Details for interpreting the alignment are available on the Phyre2 web page [5].



Figure S13. Scc4 homology model with Scc1 interface and overlapping resonances. Scc4 residues less than 5 Å from the homology model-predicted Scc1 interface are shown in blue. Scc4 residues with ¹H,¹⁵N-HSQC peaks that overlap peaks from the ¹H,¹⁵N-HSQC spectrum of ¹⁵N-Scc4:His₆-Scc1 are shown in purple.

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