

# SUPPLEMENTARY FILE

## **Urinary metabolite profiling to non-invasively monitor the omega-3 index: An exploratory secondary analysis of a randomized clinical trial in young adults.**

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**Table S1:** Full annotation of the 17 urinary metabolites significantly associated with n3-LCPUFA supplementation or the O3I reported in Table 3.

Metabolite/Identifier <sup>1</sup>	m/z:RMT:mode <sup>2</sup>	Molecular Formula <sup>2</sup>	Mass Error <sup>2</sup> (ppm)
Choline	104.108:0.333:p	C <sub>5</sub> H <sub>14</sub> NO	2.9
Tiglylglycine (HMDB0000959)	156.066:0.843:n	C <sub>7</sub> H <sub>11</sub> NO <sub>3</sub>	4.5
S-Carboxypropylcysteamine (HMDB0002169)	164.074:0.613:p	C <sub>6</sub> H <sub>13</sub> NO <sub>2</sub> S	1.8
N1-Acetylspermidine (HMDB0001276)	188.175:0.259:p	C <sub>9</sub> H <sub>21</sub> N <sub>3</sub> O	6.4
Quinic acid (HMDB0003072)	191.056:0.798:n	C <sub>7</sub> H <sub>12</sub> O <sub>6</sub>	4.2
Glucuronic acid (HMDB0000127)	193.035:0.772:n	C <sub>6</sub> H <sub>10</sub> O <sub>7</sub>	2.6
Tryptophan	205.098:0.899:p	C <sub>11</sub> H <sub>12</sub> N <sub>2</sub> O <sub>2</sub>	2.9
Unknown dianion [M-H <sub>2</sub> ] <sup>2-</sup>	221.075:0.927:n	C <sub>22</sub> H <sub>28</sub> N <sub>4</sub> O <sub>2</sub> S <sub>2</sub>	--
Carboxybutylhomocysteine	222.080:0.762:p	C <sub>8</sub> H <sub>15</sub> NO <sub>4</sub> S	2.7
O-Butyrylcarnitine (HMDB0002013)	232.155:0.718:p	C <sub>8</sub> H <sub>10</sub> NO <sub>4</sub>	4.7
Pyroglutamylisoleucine (HMDB0341381)	241.120:0.653:n	C <sub>11</sub> H <sub>18</sub> N <sub>2</sub> O <sub>4</sub>	6.7
Pyroglutamylleucine (HMDB0341382)	241.120:0.662:n	C <sub>11</sub> H <sub>18</sub> N <sub>2</sub> O <sub>4</sub>	3.0
Unknown cation isobar#2	258.110:0.788:p	C <sub>11</sub> H <sub>19</sub> N <sub>3</sub> S <sub>2</sub>	--
Unknown cation	300.215:0.841:p	C <sub>17</sub> H <sub>25</sub> N <sub>5</sub>	--
Tetrahydroaldosterone glucuronide (HMDB0010357)	539.249:0.472:n	C <sub>27</sub> H <sub>40</sub> O <sub>11</sub>	0.6
Unknown dianion [M-H <sub>2</sub> ] <sup>2-</sup>	88.004:1.622:n	C <sub>4</sub> H <sub>6</sub> N <sub>2</sub> O <sub>6</sub>	5.7
Symmetric dimethylarginine	203.151:0.478:p	C <sub>8</sub> H <sub>18</sub> N <sub>4</sub> O <sub>2</sub>	3.4

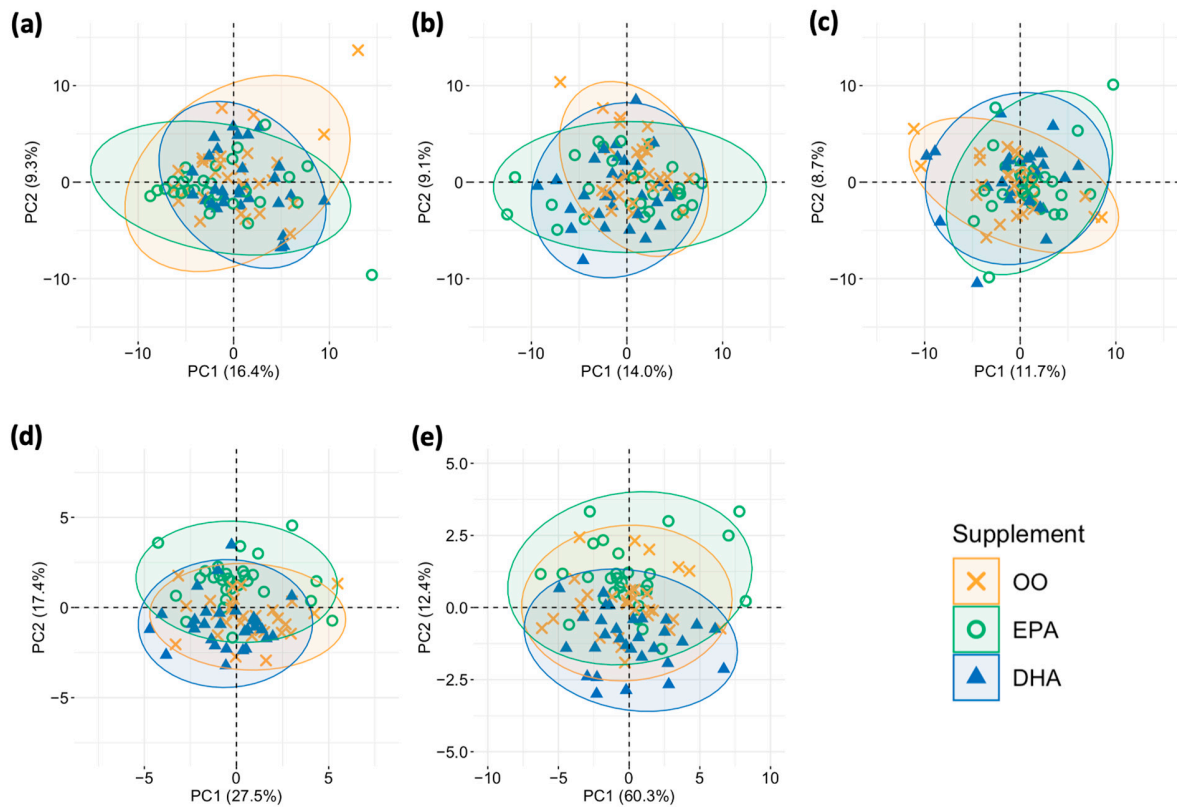
<sup>1</sup>Putative urinary metabolite identified together with their corresponding HMDB ID# if reported.

<sup>2</sup>All urinary metabolites were annotated based on their accurate mass, relative migration time and ionization mode (m/z:RMT:mode), most likely molecular formulae, and mass error (if structure known).

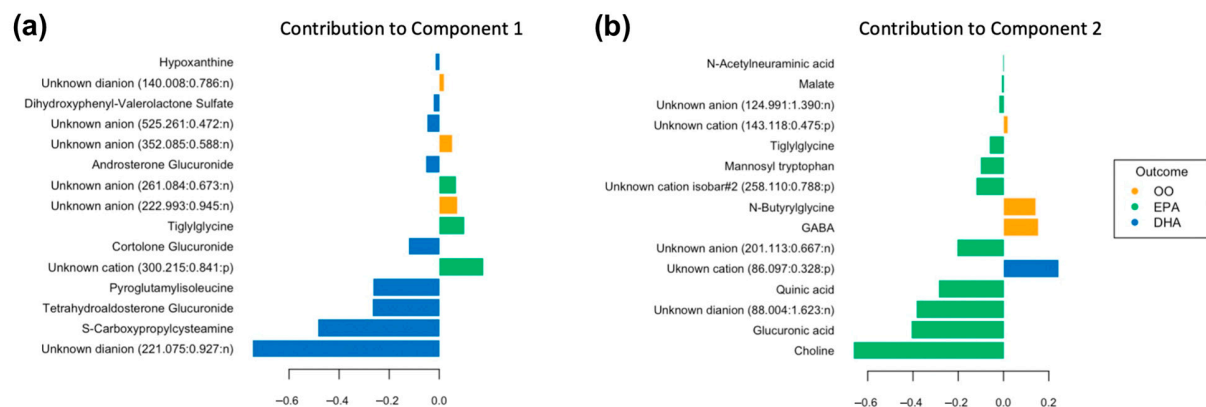
**Table S2:** glmmlasso selected urinary metabolites on the Omega-3 Index (O3I) and EPA+DHA (ng/mg Hb) integrated datasets.

Metabolite <sup>1</sup>	m/z:RMT:mode	O3I Outcome		EPA+DHA (ng/mg Hb) Outcome	
		Estimate	P value	Estimate	P value
Unknown dianion [M-H <sub>2</sub> ] <sup>2-</sup>	221.068:0.927:n	1.079	< 0.001	0.832	< 0.001
S-Carboxypropylcysteamine	164.074:0.613:p	0.501	< 0.001	0.535	< 0.001
Tetrahydroaldosterone glucuronide	539.249:0.472:n	0.365	< 0.001	0.264	< 0.001
N1-Acetylspermidine	188.175:0.259:p	-0.538	< 0.001	-0.858	< 0.001
Symmetric dimethylarginine	203.151:0.478:p	-	-	0.551	< 0.001
Tryptophan	205.098:0.899:p	-	-	0.166	< 0.001
Carboxybutylhomocysteine	222.080:0.762:p	-	-	-0.234	< 0.001

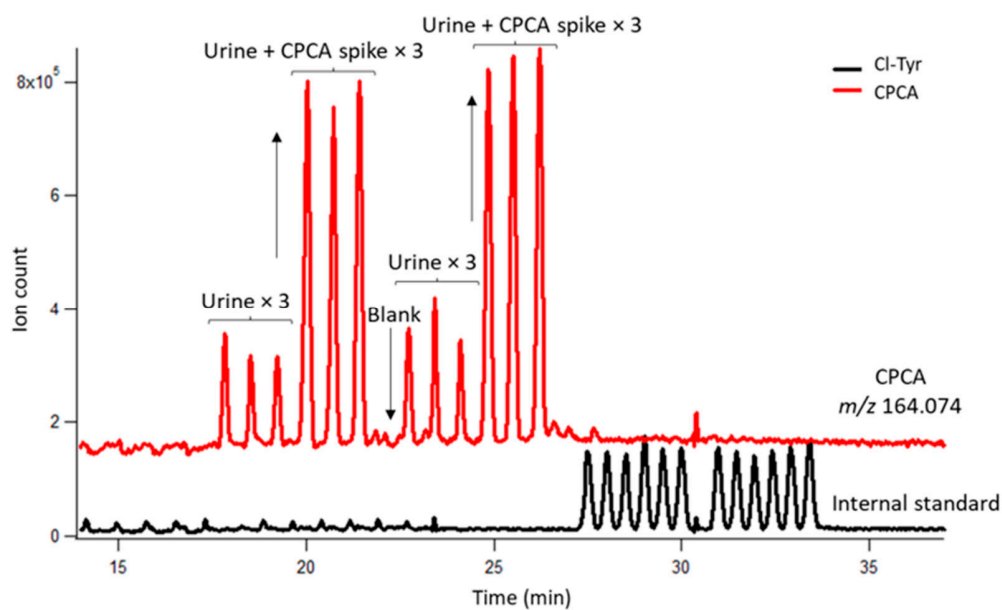
<sup>1</sup>Putative urinary metabolite as annotated by its accurate mass, relative migration time and ionization mode (m/z:RMT:mode). Most likely molecular formulae are provided for unknown chemical structures.



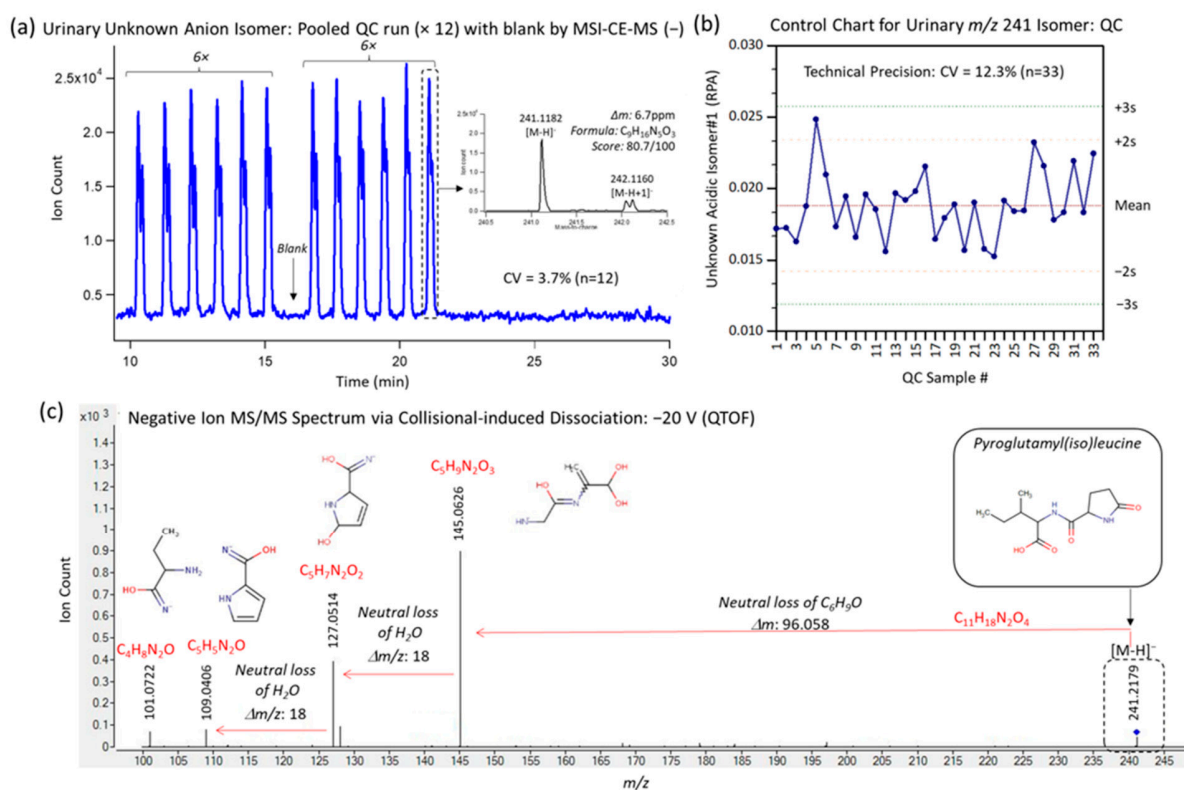
**Figure S1.** PCA plots of (a) baseline and (b) final (12 weeks) of 124 creatinine-normalized urinary metabolites for control (OO), EPA, and DHA groups (OO = orange, EPA = green, DHA = blue). Total variation explained in PC1 and PC2 plots was 25.7% and 23.1% respectively for each time point (a, b). (c) PCA plots for 88 participant delta values of 124 urinary metabolites using RPA; (d) 15 qualitative FA measurements (relative % FA), and (e) 15 quantitative FA measurements (ng FA/mg Hb). Total variation explained in PC1 and PC2 plots was 20.4%, 44.9%, 72.7% respectively for each dataset (c-e).



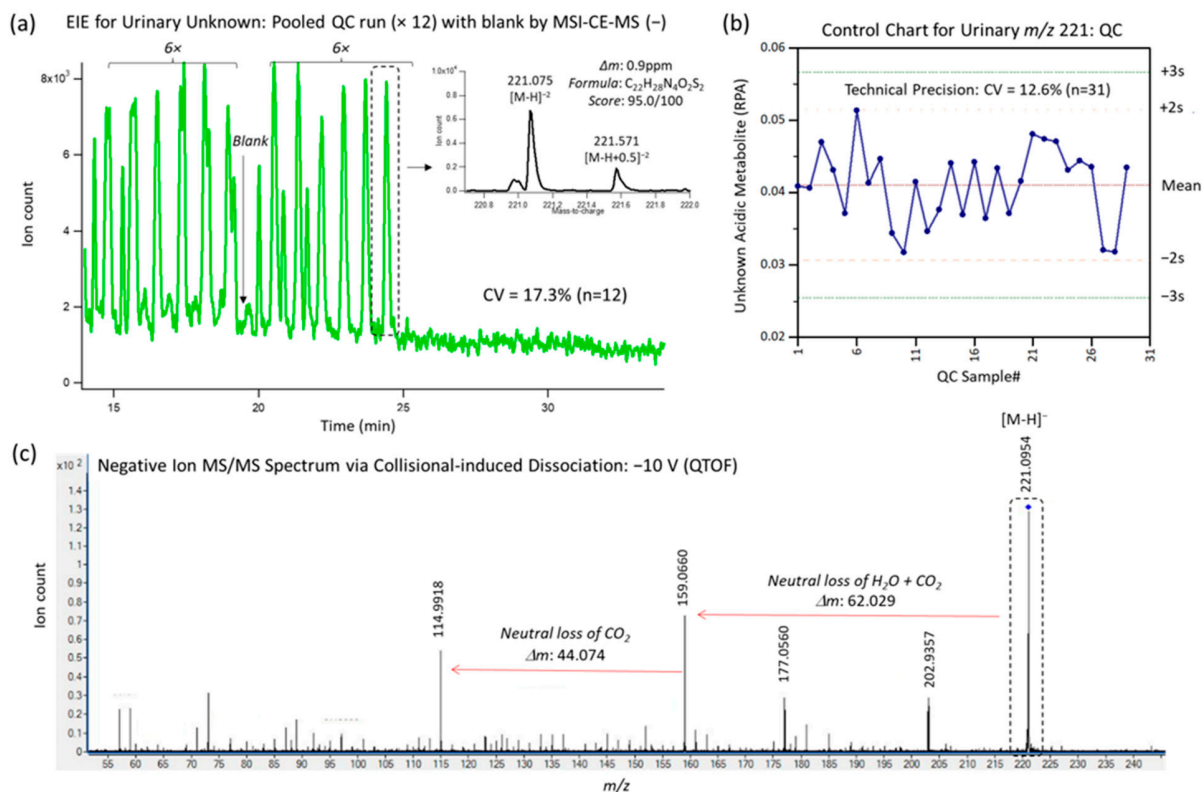
**Figure S2.** sPLS-DA loading weights reflecting the contribution for 15 urinary metabolites on (a) PC1 and (b) PC2 coloured by group (OO = orange, EPA = green, DHA = blue). All unknown metabolites are annotated based on their characteristic  $m/z$ :RMT:mode.



**Figure S3.** Tentative identification of CPCA based on spiking into pooled urine when using MSI-CE-MS with a 13 serial sample injection format, which confirmed its co-migration with the unknown sulfur-containing metabolite ( $m/z$  164.074) under positive ion mode conditions.

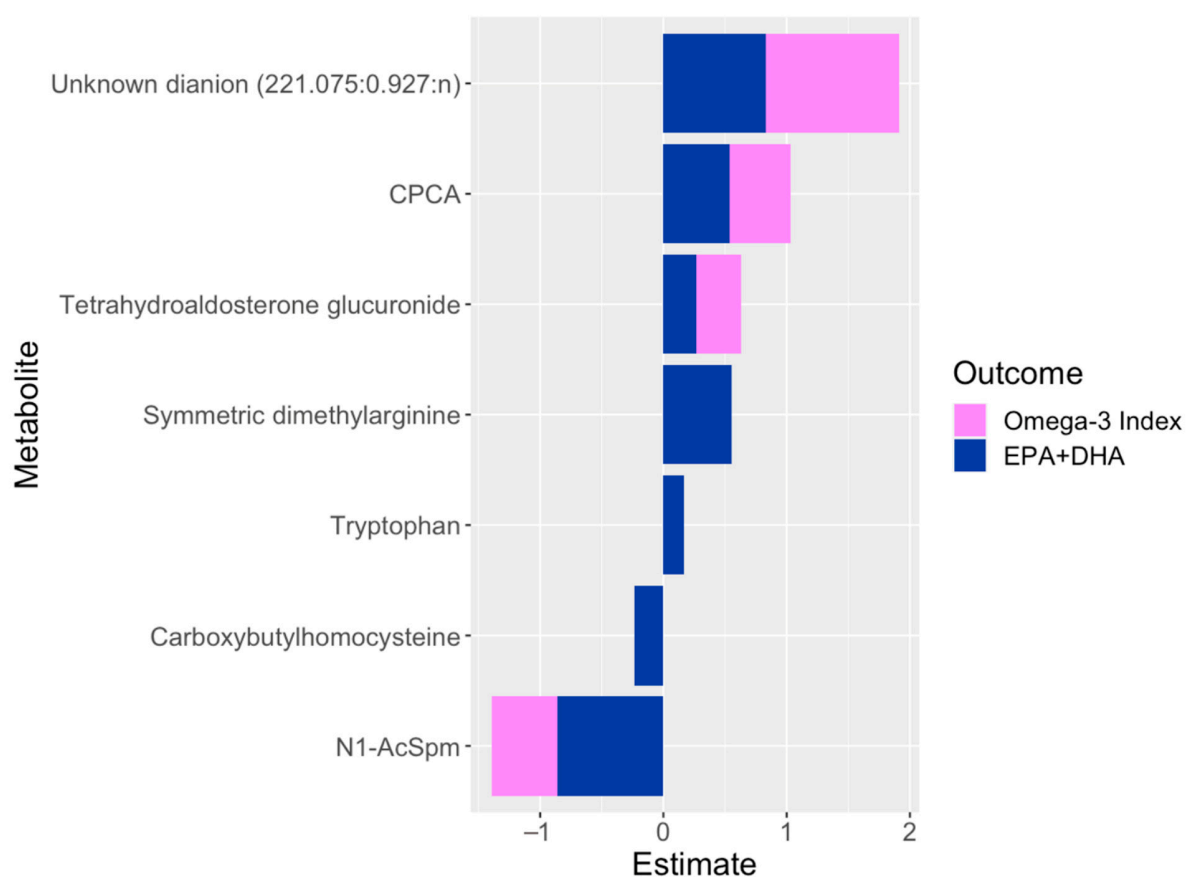


**Figure S4.** Tentative identification of two unknown anionic urinary metabolites that migrate as partially resolved isomers by MSI-CE-MS under negative ion mode conditions. **(a)** Representative extracted ion electropherogram (EIE) when using MSI-CE-MS under negative ion mode for the unknown anion isomer highlighting a serial injection of 12 urine samples and a blank in a single run. The first migrating isomer (241.120:0.653:n) was determined as the third most significant urinary metabolite associated with the O3I notably following DHA ingestion. **(b)** Technical precision of first migrating unknown anion isomer in QC samples (CV = 12.3%, n = 33). **(c)** Annotation of the first migrating unknown anion isomer MS/MS at an optimal collision energy, tentatively identified as pyroglutamylisoleucine (pGlu-Ile), whereas its slower migrating isomer is likely pyroglutamylleucine (pGlu-Leu).

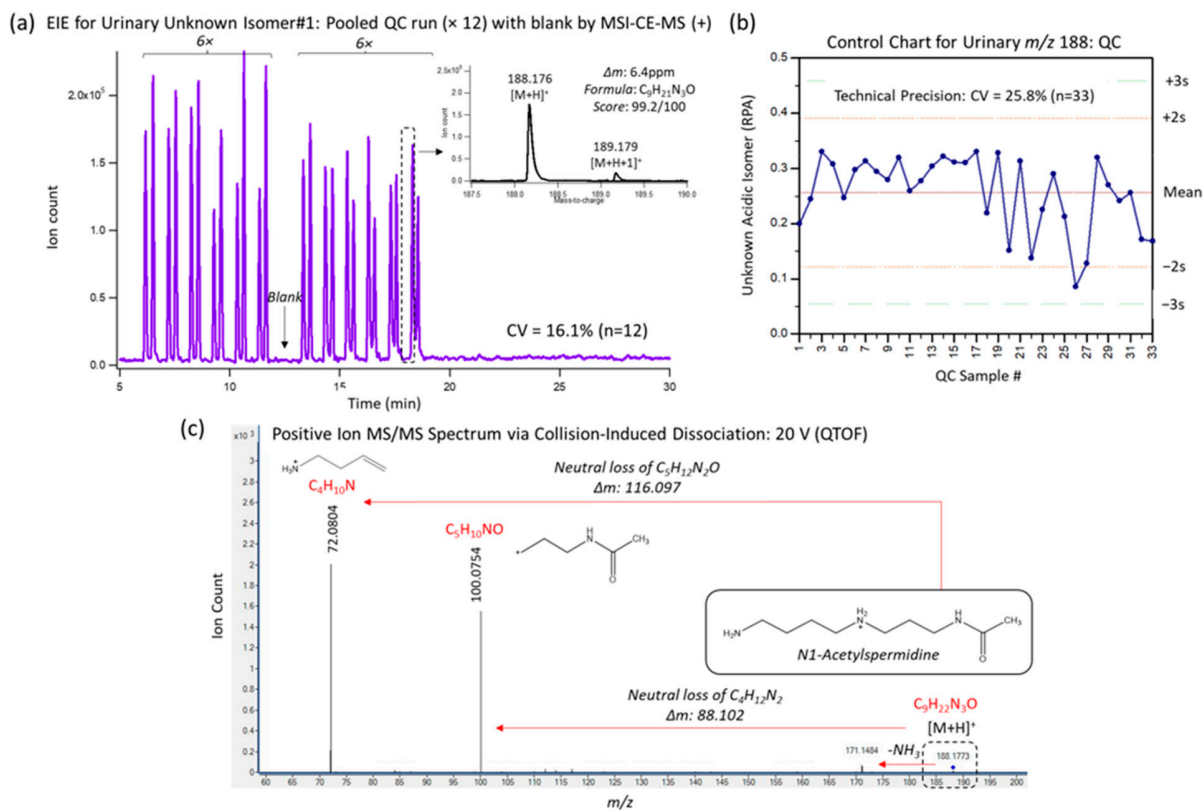


**Figure S5.** Annotation of an unknown urinary dianion ( $221.075:0.927:n$ ;  $[M-2H]^{2-}$ ) based on its (a) extracted ion electropherogram (EIE) when using MSI-CE-MS under negative ion mode demonstrating a charge state (i.e., isotopic pattern) that was associated with DHA supplementation. Although this acidic metabolite was measured with (b) adequate technical precision in QC samples ( $CV < 15\%$ ,  $n = 31$ ), its low abundance prevented acquisition of a good quality MS/MS spectra under negative ion mode conditions. (c) Collisional-induced dissociation experiments confirmed the likely presence of two acidic functional moieties ( $-COOH$ ) based on corresponding neutral mass losses.

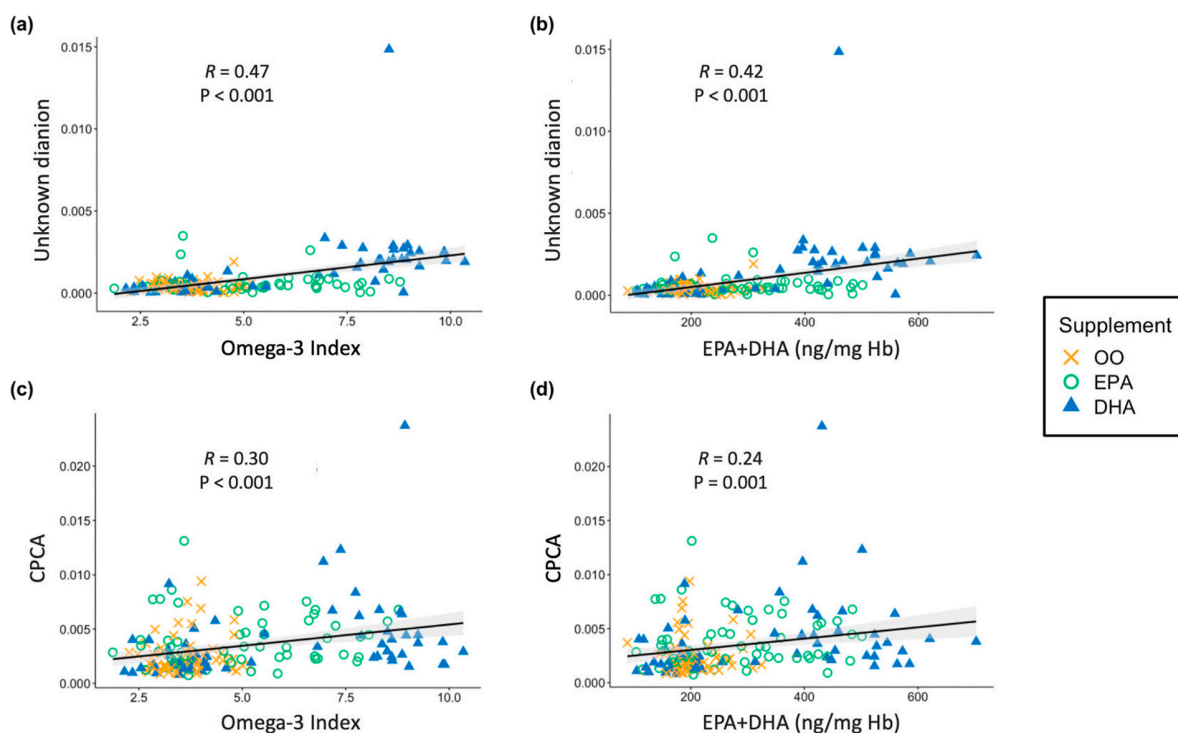




**Figure S6.** Estimate values for glmLasso urinary metabolites from two integrated datasets with scaled from 0-10 or 80-700 urine metabolites as covariates and the O3I or EPA+DHA (ng/mg Hb), respectively, as outcome variables. Urinary metabolites with a positive estimate result indicate a positive association with the outcome variable and metabolites with a negative estimate result indicate a negative association with the outcome variable. Estimates from values from the O3I (pink) and EPA+DHA (ng/mg Hb) (navy) are depicted in stacked bars. Unknown metabolites were annotated based on their characteristic m/z:RMT:mode. CPCA = 2-Carboxypropylcysteamine, N1-AcSpm = N1-Acetylspermidine.



**Figure S7.** Tentative identification of two unknown cationic urinary metabolites that (a) migrate as partially resolved isomers by MSI-CE-MS under positive ion mode conditions. However, only the first migrating isomer (188.175:0.259:p) was associated with the O3I. (b) This unknown isomer was measurable with acceptable technical precision in QC samples. (c) Annotation of this first migrating isomer MS/MS at an optimal collision energy, tentatively identified as *N1*-acetylspermidine (*N1*-AcSpm), whereas its slower migrating isomer is likely *N8*-acetylspermidine (*N8*-AcSpm).



**Figure S8.** Correlation plots for the (a) unknown dianion (221.075:0.927:n) and the O3I ( $R = 0.47$ ,  $P < 0.001$ ); (b) unknown dianion (221.075:0.927:n) and the sum of EPA+DHA (ng/mg Hb) ( $R = 0.42$ ,  $P < 0.001$ ); (c) S-carboxypropylcysteamine (CPCA; 164.074:0.613:p) and the O3I ( $R = 0.30$ ,  $P < 0.001$ ); and (d) CPCA and the sum of EPA+DHA (ng/mg Hb) ( $R = 0.24$ ,  $P = 0.001$ ). OO = yellow, EPA = green, DHA = blue.