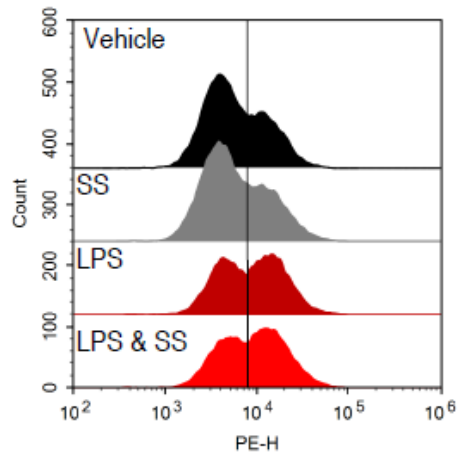
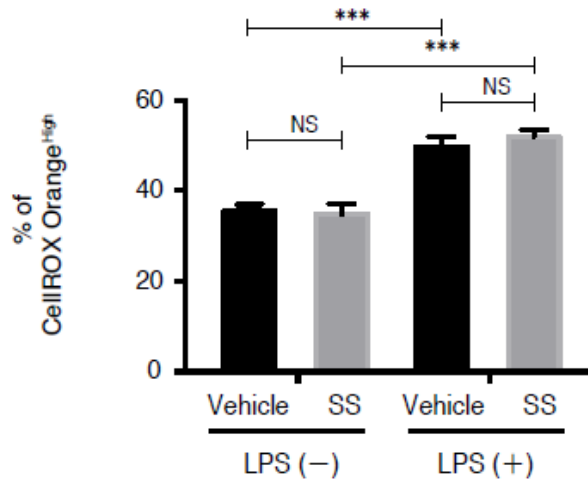
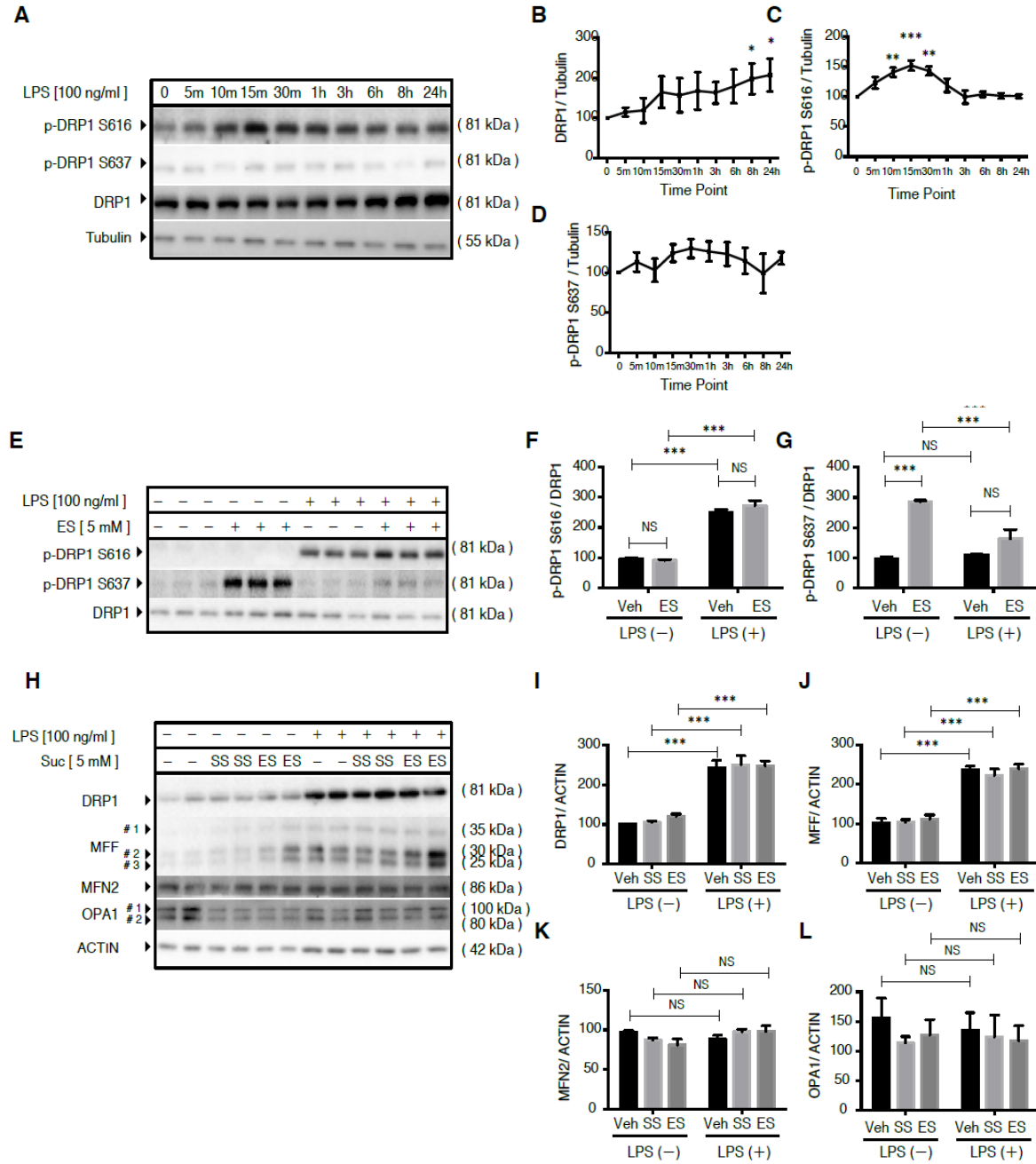


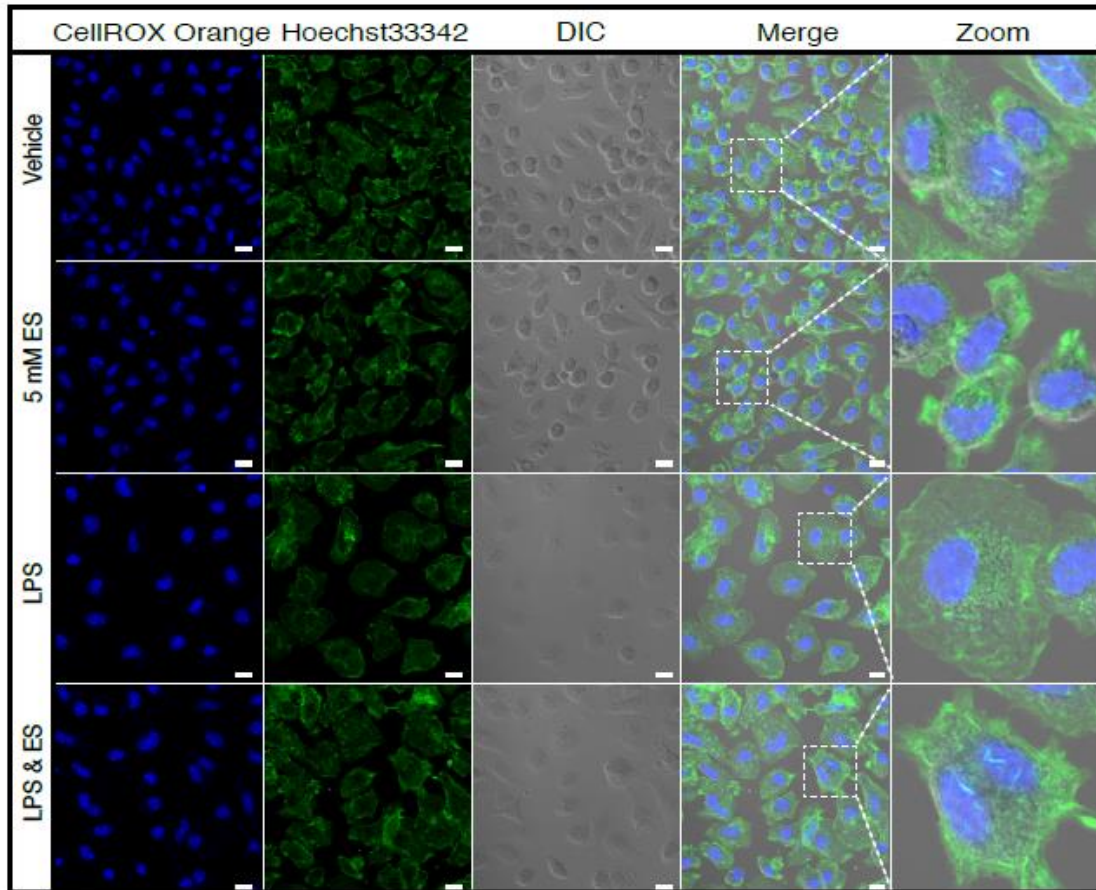
**Figure S1.** Disodium succinate did not alter M1 population in primary microglial cells. Primary microglial cells were pretreated without or with disodium succinate (ES: 5 mM) for 3 h before being stimulated with vehicle (PBS) or LPS (100 ng/mL) for 24 h. Cells were washed and analyzed for flow cytometry. (A,B) Representative overlay image and quantification analysis of M1 surface marker. (C,D) Representative overlay image and quantification analysis of M2 surface marker ( $n = 3$ ). Data represent mean  $\pm$  se. \*\*\*  $p < 0.001$ .

**A****B**

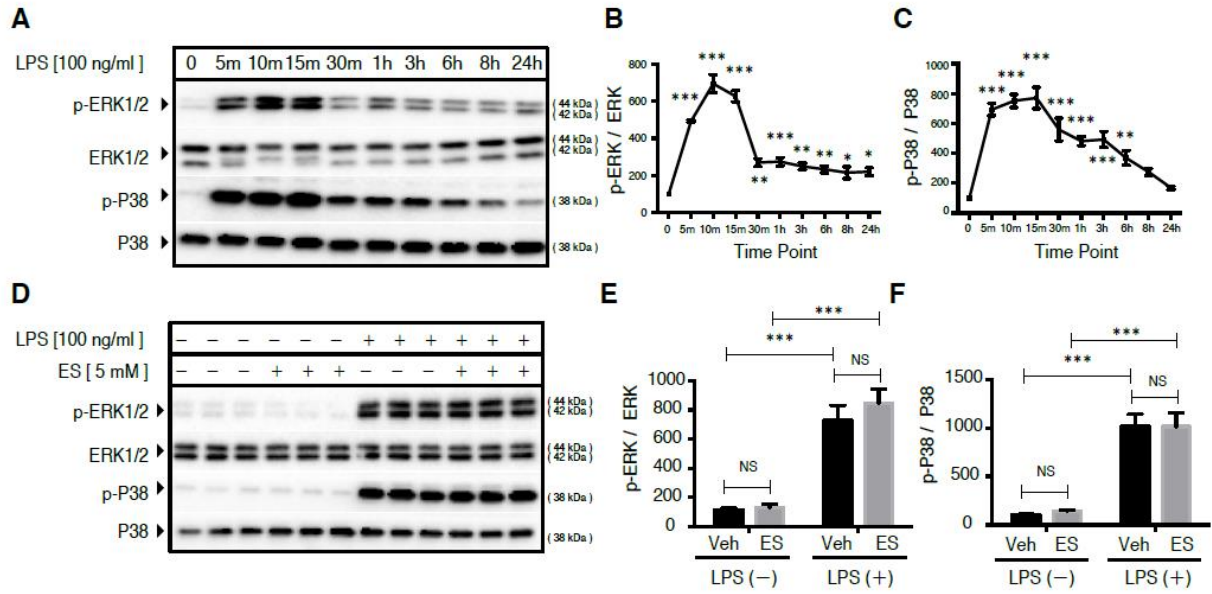
**Figure S2.** Disodium succinate did not reduce ROS production in primary microglial cells. Primary microglial cells were pretreated without or with disodium succinate (ES: 5 mM) for 3 h before being stimulated with vehicle (PBS) or LPS (100 ng/mL) for 1 h. The cells were incubated with a cellular ROS indicator, CellROX Orange, for 30 min at 37 °C. Cells were washed and analyzed by flow cytometry. **(A)** Representative overlay image of cellular ROS. **(B)** Quantification analysis of cellular ROS by flow cytometry ( $n = 3$ ). Data represent mean  $\pm$  se. \*\*\*  $p < 0.001$ .



**Figure S3.** Diethyl succinate promoted phosphorylation of DRP1 at Ser-637 in primary microglial cells. (A–D) Primary microglial cells were treated with LPS (100 ng/mL) at in-dicated time point. The protein levels of p-DRP1 S616, p-DRP1 S637, DRP1 and tubulin were determined by western blot analysis. (E–G) Primary microglial cells were pretreated without or with diethyl succinate (ES: 5 mM) for 3 h before being stimulated with vehicle (PBS) or LPS (100 ng/mL) for 10 min. The protein levels of p-DRP1 S616, p-DRP1 S637 and DRP1 were determined by western blot analysis. (H–L) Primary microglial cells were pretreated without or with disodium succinate (SS: 5 mM) or diethyl succinate (ES: 5 mM) for 3 h before being stimulated with LPS (100 ng/mL) for 24 h. The protein levels of DRP1, MFF, MFN2, OPA1 and ACTIN were determined by western blot analysis. ( $n = 3$ ) Data represent mean  $\pm$  se. \*  $p < 0.05$ . \*\*  $p < 0.01$ . \*\*\*  $p < 0.001$ .



**Figure S4.** Diethyl succinate prevented morphological transformation of *Sucnr1*<sup>-/-</sup> primary microglial cells associated with M1 activation. (A) Primary *Sucnr1*<sup>-/-</sup> microglial cells were pre-treated without or with diethyl succinate (ES: 5 mM) for 3 h before being stimulated with vehicle (PBS) or LPS (100 ng/mL) for 24 h. The cells were fixed and then immunocytochemically stained for ActinGreen 488, a marker for actin filaments. Nuclei were stained with hoechst33342, and images were captured with an LSM880 confocal microscope. Scale bar = 10  $\mu$ m.



**Figure S5.** Diethyl succinate did not affect lipopolysaccharide-induced MAPK activation in primary microglial cells. (A–C) Primary microglial cells were treated with LPS (100 ng/mL) at indicated time point. The protein levels of p-ERK1/2, ERK1/2, p-P38 and P38 were determined by western blot analysis. (D–F) Primary microglial cells were pretreated without or with diethyl succinate (ES: 5 mM) for 3 h before being stimulated with vehicle (PBS) or LPS (100 ng/mL) for 10 min. The protein levels of p-ERK1/2, ERK1/2, p-P38 and P38 were determined by western blot analysis ( $n = 3$ ). Data represent mean  $\pm$  se. \*\*  $p < 0.01$ . \*\*\*  $p < 0.001$ .

**Table S1. Antibodies used in this study.**

|   |
|---|
| Antibodies used for Flow Cytometry  |
| PE anti-Mouse/Human CD11b ( Cat # 101208; 1:100 dilution ; BioLegend, San Diego, CA, USA)                                 |
| APC anti-Mouse CD86 ( Cat # 105012; 1:100 dilution ; BioLegend, San Diego, CA, USA)                                       |
| Brilliant Violet 421™ anti-mouse CD206 ( Cat # 141717; 1:100 dilution ; BioLegend, San Diego, CA, USA)                    |
| Antibodies used for Western Blot  |
| Phospho-D RP1 (Ser637) Antibody (Cat # 4867s; 1:1000 dilution; Cell signaling, Danvers, MA, USA)                          |
| Phospho-D RP1 (Ser616) Antibody (Cat # 3455s; 1:1000 dilution; Cell signaling, Danvers, MA, USA)                          |
| Purified Mouse Anti-DLP1 ( Cat # 611113; 1:1000 dilution ; BD Biosciences, Sparks, MD, USA)                               |
| Mouse monoclonal anti-OPA1 ( Cat # 612606; 1:1000 dilution ; BD Biosciences, Sparks, MD, USA)                             |
| MFN2 monoclonal antibody ( Cat # H00009927-M03; 1:1000 dilution ; Abnova, Taipei, Taiwan)                                 |
| Mff antibody ( Cat # 17090-1-AP; 1:1000 dilution ; Proteintech, Rosemont, IL, USA)  |
| Rabbit polyclonal anti-phospho-p38 MAPK (Thr180/Tyr182 ) (Cat # 4511s; 1:1000 dilution; Cell signaling, Danvers, MA, USA) |
| Rabbit polyclonal anti-p38 MAPK (Cat # 8690; 1:1000 dilution; Cell signaling, Danvers, MA, USA)                           |
| Rabbit monoclonal anti-phospho-ERK1/2 (Thr202/Tyr204) (Cat # 4370s; 1:1000 dilution; Cell signaling, Danvers, MA, USA)    |
| Rabbit monoclonal anti-ERK1/2(137F5) (Cat # 4695s; 1:1000 dilution; Cell signaling, Danvers, MA, USA)                     |
| β-Actin (13E5) Rabbit mAb (Cat # 4970s; 1:1000 dilution; Cell signaling, Danvers, MA, USA)                                |
| α-Tubulin (11H10) Rabbit mAb (Cat # 2125s; 1:1000 dilution; Cell signaling, Danvers, MA, USA)                             |
| Horse anti-mouse IgG, HRP-linked antibody (1:5000 dilution; Cell signaling, Danvers, MA, USA)                             |
| Goat anti-rabbit IgG, HRP-linked antibody (1:5000 dilution; Cell signaling, Danvers, MA, USA)                             |

**Table S2. Real-time PCR primers used in this study.**

| Gene         | Forward primer (5' – 3') | Reverse primer (5' – 3') |
|--------------|--------------------------|--------------------------|
| <i>Cd86</i>  | AGCACGGACTTGAACAACCAGAC  | TTGTAAATGGGCACGGCAGATA   |
| <i>Cd206</i> | CTGATGCAAACCACACATGCAC   | TTCAGGCCTCAATCCAACCAA    |
| <i>Tnfa</i>  | ATCCGCGACGTGGAAGT        | ACCGCCTGGAGTTCTGGAA      |
| <i>Il-1b</i> | GGAGAACCAAGCAACGACAAAATA | TGGGGAAGTCTGCAGACTCAAAC  |
| <i>Actin</i> | TCATCACTATTGGCAACGACG    | AACAGTCCGCCTAGAAGCAC     |