

Review

Potential Diets to Improve Mitochondrial Activity in Amyotrophic Lateral Sclerosis

Sayuri Yoshikawa [†], Kurumi Taniguchi [†], Haruka Sawamura, Yuka Ikeda, Ai Tsuji and Satoru Matsuda ^{*†} 

Department of Food Science and Nutrition, Nara Women's University, Kita-Uoya Nishimachi, Nara 630-8506, Japan

^{*} Correspondence: smatsuda@cc.nara-wu.ac.jp; Tel./Fax: +81-742-20-3451[†] These authors contributed equally to the work.

Abstract: Amyotrophic lateral sclerosis (ALS) is an incurable neurodegenerative disease, the pathogenesis of which is based on alternations in the mitochondria of motor neurons, causing their progressive death. A growing body of evidence shows that more efficient mitophagy could prevent and/or treat this disorder by suppressing mitochondrial dysfunction-induced oxidative stress and inflammation. Mitophagy has been considered one of the main mechanisms responsible for mitochondrial quality control. Since ALS is characterized by enormous oxidative stress, several edible phytochemicals that can activate mitophagy to remove damaged mitochondria could be considered a promising option to treat ALS by providing neuroprotection. Therefore, it is of great significance to explore the mechanisms of mitophagy in ALS and to understand the effects and/or molecular mechanisms of phytochemical action, which could translate into a treatment for neurodegenerative diseases, including ALS.

Keywords: ALS; mitophagy; ROS; AMPK; mTOR; mTORC1; natural product

Citation: Yoshikawa, S.; Taniguchi, K.; Sawamura, H.; Ikeda, Y.; Tsuji, A.; Matsuda, S. Potential Diets to Improve Mitochondrial Activity in Amyotrophic Lateral Sclerosis. *Diseases* **2022**, *10*, 117. <https://doi.org/10.3390/diseases10040117>

Academic Editor: Alessandra Stacchiotti

Received: 2 September 2022

Accepted: 30 November 2022

Published: 1 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Mitochondrial activity and the generation of reactive oxygen species (ROS) are important for cell proliferation, survival, and/or differentiation [1]. In addition, many diseases, such as cardiac failure, cancer, and age-related pathological conditions, have been related to altered mitochondrial function [2]. For example, the energy deficiency resulting from local hypoxia during an ischemic heart attack leads to mitochondrial dysfunction, which could have arrhythmogenic consequences and lead to sudden cardiac death [3]. The significance of mitochondria has been emphasized in a variety of neurodegenerative diseases, including amyotrophic lateral sclerosis (ALS). ALS is an incurable neurodegenerative disease whose etiology is based on the progressive death of motor neurons [4]. At present, ALS has no effective therapy. A better understanding of the mitochondrial regulating pathways may raise several promising neuroprotection perspectives in the effective treatment of ALS. Furthermore, the development of new treatments would be useful not only for the mitochondrial disorders of ALS but also for the wide spectrum of age-related neurodegenerative diseases.

The mitochondrion is the main place for adenosine triphosphate (ATP) synthesis, fatty acid β -oxidation, and ROS production [5]. Mitochondrial function and ATP production are crucial for neuronal cell survival and excitability [6]. However, mitochondrial dysfunction leads to the overproduction of ROS and neuronal apoptosis, which is closely related to neurodegenerative diseases [6]. To slow down the progression of the pathology of ALS, the high oxidative stress and mitochondrial activity of neurons should at least be improved. Mitophagy, which is a specific kind of autophagy, can selectively degrade damaged mitochondria to reduce mitochondrial dysfunction and maintain mitochondrial function. Thus, mutations in the genes that encode factors essential for mitophagy may

result in the impairment of this process, which can lead to neurodegenerative conditions such as ALS [7]. For example, hypoxia is a common factor in several disease conditions, such as inflammation, which can lead to a depletion of oxygen and, eventually, through the production of ROS, directly to an alteration of intracellular proteins, lipids, or DNA [8]. Therefore, the control of the mitochondrial ROS level is of key relevance for maintaining cellular homeostasis [9]. The conserved pathway of mitophagy is required to prevent and/or counteract the pathogenic actions that may lead to neurodegeneration. Here, we outline the mechanisms leading to mitochondrial dysfunction in ALS and discuss the potential of targeting mitophagy with several phytochemicals for the possible treatment of ALS, with a view to providing a direction for finding certain phytochemicals that can target mitophagy to prevent and/or treat ALS.

2. Mitochondrial Dysfunction Involved in the Pathogenesis of ALS

Mitochondria are pretty vigorous organelles whose number and activity can be adjusted to the changes in cellular energy metabolism by regulating their biogenesis, fusion/fission events, and removing damaged ones [5]. The most common reasons for mitochondrial dysfunction are hypoxia and the overproduction of ROS. In particular, oxidative stress caused by numerous inflammations is related to the development of many neurological complications [10]. Excessive ROS also induce oxidative stress and apoptosis in cells [11]. Typically, in cells, ROS are generated in the mitochondrial respiratory chain. Accordingly, mitochondrial dysfunction could also lead to neuronal cell death and/or apoptosis [12], which is associated with neurological complications, including Alzheimer's disease, Parkinson's disease, and Huntington's disease [13]. These oxidative stresses that include oxygen radicals could be intercepted [14]. In various neurodegenerative diseases such as ALS, several defects in mitochondrial function leading to oxidative stresses have been identified in underlying relations [15]. Indeed, this emphasizes the importance of healthy mitochondria for the maintenance of healthy neuronal functions.

Under physiological conditions, ROS could work as regulators of the mechanism for maintaining cellular redox homeostasis [16]. During oxidative phosphorylation, mitochondria could produce a superoxide anion by-product, which may be further changed into ROS [17]. Mitochondrial ROS consist of superoxide, hydrogen peroxide, and hydroxyl, which can modify lipids, proteins, and DNA, resulting in mitochondrial dysfunction and/or neuronal cell death [18]. Accumulation of damaged mitochondria and the overproduction of ROS will strengthen each other, which may finally lead to severe mitochondrial dysfunction. Mitochondrial ROS are also regulators of a cellular redox environment linked to cellular metabolic balance [19]. Accordingly, hindering too much production of ROS is considered an effective way to prevent oxidative damage to cells, including the neuron [20]. It may be indispensable to explore the roles of mitochondrial ROS in ALS. Antioxidants are the first safety to clear ROS [21]. In addition, there are various systems to withstand ROS-induced oxidative stresses, based on superoxide dismutase, catalase, and/or glutathione peroxidases. Considering the crucial role of mitochondria in cellular homeostasis, monitoring the quality of mitochondria may be important for avoiding neurodegenerative diseases including ALS [22]. In fact, functional defects in and altered morphology of mitochondria have been found in the spinal motor neurons of ALS patients [23]. Likewise, mislocalization and aggregation of mitochondria have been detected in the motor neurons of ALS patients. These observations suggest that the dysfunction of mitochondria may be a regular feature of ALS [24]. Therefore, mitochondrial quality control in many forms of molecular and/or cellular levels should be performed to prevent neurodegenerative diseases, including ALS.

Mitophagy, a mitochondrial quality control mechanism, selectively removes dysfunctional mitochondria to preserve mitochondrial function and maintain cellular homeostasis. In this process, impaired mitochondria are trapped and surrounded by autophagic membranes and further delivered to lysosomes, where they are degraded. It is well-known that the clearance mechanism of damaged mitochondria can be a potent therapeutic strategy in

cases of increased oxidative stresses [25]. Some physiologic or chemical faults in mitochondria may trigger mitophagy by disrupting the mitochondrial inner membrane, possibly with the involvement of ROS. Mitophagy has been proven to be related to the development of various diseases, including ALS [26]. For example, treatment with progesterone can prolong the survival time in a mouse model of ALS, which might be associated with enhanced autophagy in the spinal cord [27]. On the contrary, autophagy induction could accelerate the progression of ALS, most likely through the excessive mitochondrial clearance in motor neurons [28]. The balance between synthesis and degradation of mitochondria may be essential for maintaining mitochondrial and/or cellular homeostasis, and the modulation of mitophagy represents a promising therapeutic intervention.

3. Mitochondrial Quality Control by Mitophagy in ALS

Mitochondrial dysfunction is considered an important cause in the pathogenesis of neurodegenerative diseases, which could be inhibited through mitophagy to retain the healthy functioning of the mitochondria. To put it simply, dysfunctional mitochondria could be separated from healthy ones and removed through mitophagy. In details of the molecular mechanisms, damaging mitochondria leads to the accumulation of PTEN-induced kinase 1 (PINK1) in the outer membrane of the mitochondria, where it recruits E3 ubiquitin ligase Parkin that can activate the removal of mitochondria by autophagosomes [29]. In general, PINK1 is located in the outer mitochondrial membrane, but PINK1 cannot be detected in healthy mitochondria (Figure 1). Consequently, PINK1 and Parkin are recruited to the outer mitochondrial membrane for the removal of impaired mitochondria [30], which in turn ubiquitinates mitochondrial surface proteins [31]. Therefore, PINK1 and Parkin are two critical mediators regulating mitophagy in mammalian cells [32] (Figure 1).

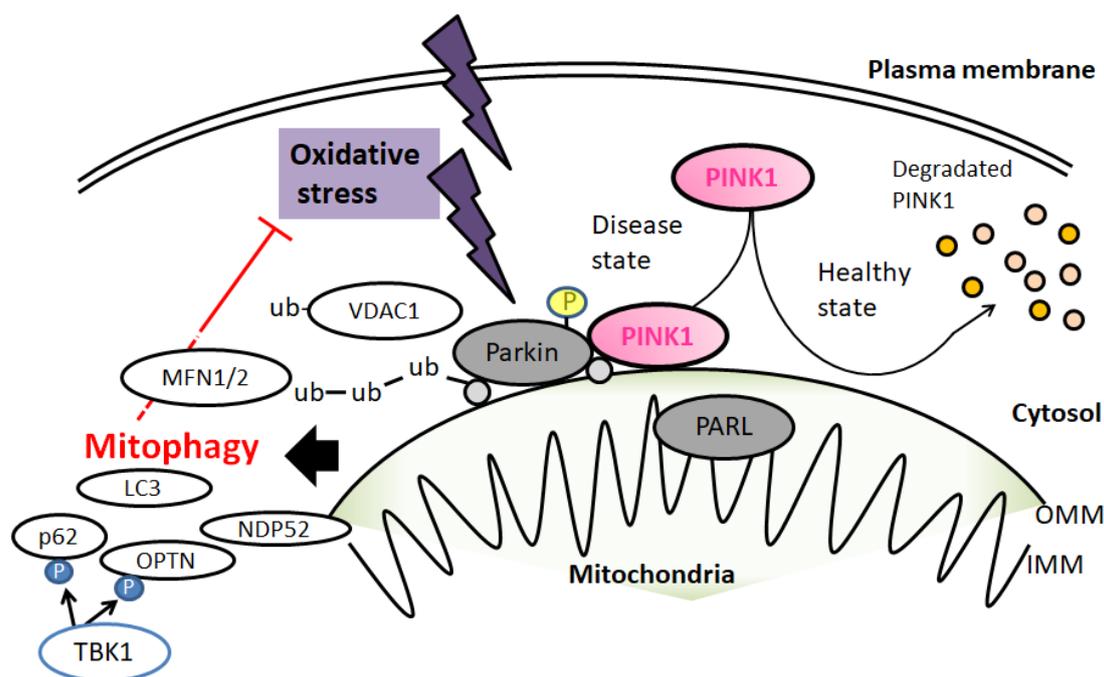


Figure 1. A hypothetical schematic representation and overview of PINK1, Parkin, and related molecules in a regulatory pathway for mitophagy. Under a healthy and steady state of cells, PINK1 is degraded within the surface of mitochondria. This may be inhibited by mitochondrial damage due to various oxidative stresses, resulting in PINK1 and Parkin accumulation in the outer membrane of mitochondria (OMM). Then, PINK1 phosphorylates ubiquitin to activate Parkin ubiquitin ligase activity. In addition, Parkin is also assumed to be phosphorylated and ubiquitinated, resulting in the induction of mitophagy. Note that some critical pathways have been omitted for clarity. OMM: outer mitochondrial membrane; IMM: inner mitochondrial membrane.

There are two major mechanisms involved in Parkin-dependent mitophagy. Parkin can promote the ubiquitination of the mitochondrial fusion proteins mitofusin 1 (MFN1) and mitofusin 2 (MFN2), resulting in their degradation, which leads to mitochondrial fission. This contributes to the separation of dysfunctional mitochondria from the healthy system and allows dysfunctional mitochondria to be surrounded by autophagosome membranes and degraded in lysosomes [33]. Also, Parkin-mediated ubiquitination of mitochondrial outer membrane protein voltage-dependent anion channel 1 (VDAC1) promotes the recognition of VDAC1 using autophagy receptors such as histone deacetylase 6 (HDAC6) [34,35]. Subsequently, PINK1, located in the outer mitochondrial membrane, recruits autophagy receptors, including p62, nuclear dot protein 52 (NDP52), and optineurin (OPTN), which can bind to an autophagosome-membrane-inserted LC3 protein to combine the dysfunctional mitochondria and autophagosomes. Finally, dysfunctional mitochondria are degraded in the lysosome [34] (Figure 1). To further activate mitophagy, PINK1 phosphorylates the ubiquitin of ubiquitin chains attached to mitochondrial proteins and the ubiquitin-like domain of Parkin to facilitate Parkin localization from the cytosol to the outer mitochondrial membrane of the impaired mitochondria [35,36]. Some ALS-associated gene-encoding proteins may play crucial roles in mitophagy. For example, mutant p62 shows a lower affinity for LC3, reducing the efficiency of autophagy [37]. It has been shown that mutations in gene-encoding TANK-binding kinase 1 (TBK1), which phosphorylates and regulates proteins like p62 and OPTN, result in impaired autophagy and contribute to ALS's pathology [38]. Lysosomal dysfunction has been found in cells with ALS, while lysosomal deficits accompanied by impaired autophagy may occur gradually in a mouse model of ALS [39].

Rapamycin, a commonly used immunosuppressive drug, has been shown to increase the survivability of ALS-affected motor neurons [40]. Rapamycin can induce autophagy by inhibiting the mechanistic/mammalian target of the rapamycin (mTOR) pathway [41]. In general, the mTOR pathway plays a key role in the proliferation, survival, and differentiation of various cells. Interestingly, the downmodulation of mTOR activity could increase longevity [42]. Metformin can downregulate the phosphorylation level of S6 kinase, which is a key signaling kinase downstream of the mTOR (Figure 2). Also, metformin can be used to treat a variety of aging-related metabolic disorders, like cardiovascular disease or cognitive decline [43]. The mTOR is involved in two types of protein complexes, mTOR complex 1 (mTORC1) and mTOR complex 2 (mTORC2), albeit with diverse functions [44]. The mTORC1 regularly obstructs autophagy by integrating the upstream signals from PI3 K and/or AKT. The other mTORC2 might not be closed with the regulation of autophagy [45]. However, mTOR could form two complexes, mTORC1, and mTORC2, probably with crosstalk between them. The activated PI3 K could trigger the AKT with 3-phosphoinositide-dependent protein kinase-1 (PDK1) [46]. Then the activated AKT further phosphorylates the tuberous sclerosis protein 2 (TSC2) and obstructs its interplay with TSC1, ultimately resulting in mTORC1 activation [47]. Adenosine monophosphate (AMP)-activated protein kinase (AMPK) also regulates the mTORC1 and serves as an energy sensor [48]. AMPK is stimulated by a decline in ATP concentration during ischemia that raises the ratio of AMP to ATP [49]. Interestingly, the longevity-enhancing effects of metformin mimic the effect of the stimulation of AMPK [50]. Therefore, AMPK-mTOR signaling pathways specifically regulate cellular homeostasis, including autophagy, cell proliferation, differentiation, and energy metabolism [51]. Modulation of autophagy has been shown to be a potential therapeutic target in neurodegeneration [52]. For example, treatment with n-butylidenephthalide could prolong the survival of ALS mice by abrogating autophagy [53]. Accordingly, there is a persistent reason to evaluate whether other agents can stimulate or inhibit autophagy and have a beneficial effect by reducing ALS pathogenesis.

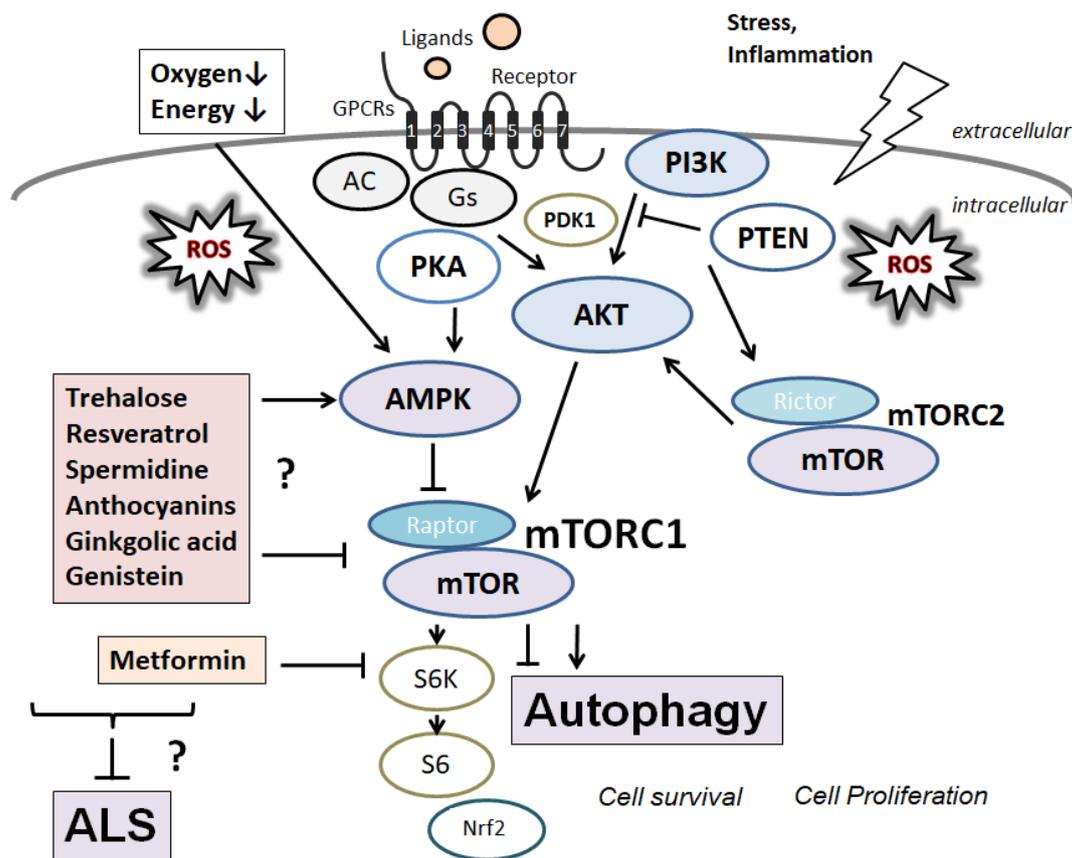


Figure 2. Selected signaling pathways involved in the induction of autophagy with indication of the sites of action of several natural compounds affecting them. Several modulator molecules linked to the PI3 K/AKT/mTOR/mTORC1 signaling pathway are demonstrated. Example compounds from natural sources known to act on the AMPK/mTOR and/or autophagy signaling are also shown. An arrow-head means stimulation, whereas a hammerhead represents inhibition. Note that some critical events, such as immune activation and/or antioxidant feedback, have been omitted for clarity. Abbreviations: AMPK, adenosine monophosphate-activated protein kinase; mTOR, mammalian/mechanistic target of rapamycin; PI3 K, phosphoinositide-3 kinase; PKA, protein kinase A; PTEN, phosphatase and tensin homolog deleted on chromosome 10; mTORC, mechanistic/mammalian target of rapamycin complex; ALS, Amyotrophic Lateral Sclerosis.

4. Beneficial Components from Natural Products for the Treatment of ALS

Increasing evidence emphasizes the beneficial role of naturally derived compounds from different natural sources for the prevention and/or treatment of human diseases, including ALS [54]. Currently, several compounds with antioxidant activity and neuroprotective effects are potential alternative therapies for neurological complications due to their unique therapeutic properties and considerable safety [55]. In addition, there is a variety of natural compounds known for autophagy-modulating properties as well as for controlling oxidative stress and/or active oxygen [56,57].

Trehalose is a natural saccharide that can modify autophagy [58], triggering transient lysosomal damage [59]. Induction of autophagy by trehalose may result in a significant reduction in the number of lysosomes in peripheral blood cells of sporadic ALS, suggesting that trehalose could represent an energetic treatment for ALS patients [60]. Trehalose's mechanisms of action may involve the inhibition of glucose transporters leading to AMPK activation, which could affect autophagy [61]. Consequently, trehalose could act as a weak inhibitor of the lysosome via the inactivation of mTORC1 [62]. Resveratrol, a natural polyphenolic compound, may indicate many beneficial effects. For example, resveratrol has neuroprotective properties against various neurological disorders [63] through the acti-

vation of the AMPK autophagy signaling pathway [64]. Resveratrol could reduce mTORC1 signaling for the activation of autophagy [65]. In addition, resveratrol can induce autophagy in embryonic stem cells by activating the AMPK pathway and the concomitant suppression of the mTORC1 signaling cascade [66]. Spermidine, which has a neuroprotective effect in patients with cerebral ischemia [67,68], can be found in a variety of foods [67,68]. Spermidine may also exert its beneficial effects with enhanced autophagy through the AMPK-mTOR pathway [69] and, thus, autophagy induction [70]. Spermidine could activate autophagy via the inhibition of mTORC1 [71]. Urolithin A is a natural polyphenol made by gut microbiota [72], which could induce autophagy for cell protection against kidney injury [73]. In addition, Urolithin A also protects against Parkinson's disease by enhancement of neuronal survival [74]. Urolithin A could induce mitophagy, which may be required for its neuroprotective effect [75]. Urolithin A could increase the phosphorylation level of AKT and mTOR [76]. Anthocyanins, common plant pigments, can activate autophagy and protect neuronal cells [77], which could improve the memory and motor performance of patients with neurodegeneration [78]. Anthocyanins could induce autophagy via the AMPK-mTOR signaling pathways [79]. Diallyl trisulfide is found in garlic oil, which has a neuroprotective effect against ALS model mice [80]. Diallyl trisulfide could decrease the PI3 K/mTOR signaling pathway and increase the expression of AMPK/TSC2 in Hep G2 cells [81], which might result in inducing autophagy and/or suppressing the levels of ROS [82]. Ginkgolic acid has an anti-cancer effect on colon cancer, triggering apoptosis and autophagy by controlling ROS production, in which mTORC1, p-mTOR, and p-S6 kinase could be dose-dependently reduced by the ginkgolic acid [83]. In addition, autophagy inhibitors could block ginkgolic acid-dependent clearance of α -synuclein in Parkinson's disease [84]. Genistein, the primary isoflavone from soy products, enhances antioxidant enzyme activities and could activate AMPK signaling through the downregulation of mTOR [85]. The SIRT1 (Sirtuin1)/AMPK pathway, combined with inhibiting mTOR signaling, has also been involved in accelerating autophagy by genistein [86], suggesting that genistein-dependent autophagy can diminish cellular senescence (Figure 2).

In view of the fact that more and more various phytochemicals could be applied to the treatment of ALS, it is necessary to have a more comprehensive understanding of the effects and/or potential mechanisms of the phytochemicals on autophagy and/or ALS. In addition, more research should focus on the regulatory mechanisms of mitophagy in ALS and further explore the potential of targeting mitophagy with certain phytochemicals for the prevention and/or treatment of ALS. The use of such compounds could be an opening point for the new therapy of ALS.

5. Current Therapeutic Possibilities and Limitations for ALS

With no current cure for the disease, ALS therapeutics seem to have been placed around attempts to slow the progression of the disease and provide symptomatic treatments to maintain patient quality of life (QOL). Therapeutic exercise and/or rehabilitation are also recommended for patients to slow symptomatic progression [87]. Furthermore, multidisciplinary therapy teams are known to improve patient QOL and have the potential to prolong patient survival [88]. However, there is still no cure for ALS that could reverse the progression of the disorder. At present, riluzole and edaravone may be two major disease-modifying drugs for the treatment of ALS [89,90]. The most widely-used drug showing a slight beneficial effect on patient survival [91], riluzole, might have a complex mechanism of biochemical action [92]. Riluzole may increase the survival of ALS patients by up to 18 months [93]. In the experimental study, enhanced mTOR levels and/or attenuated autophagic activity could increase the survival of motor neurons in a dose-dependent manner, suggesting that down-regulation of autophagy might be proffered as a therapeutic procedure for the treatment of ALS [53,94]. Riluzole could exhibit antioxidant capabilities against oxidative, but not nitrosative, stress [95]. Another drug, edaravone, is also an antioxidant compound anticipated to reduce oxidative stress and eliminate lipid peroxidation [96]. Edaravone has been described as having a therapeutic effect in ALS patients,

exhibiting reduced functional loss in several neurons [97]. Edaravone has been shown to remove peroxide and/or hydroxyl radicals protecting neurons in ALS [98]. Edaravone could also reduce excessive ROS, as a free radical scavenger, to prevent brain damage [99]. A somewhat unsatisfying efficacy and inconsistency in the potential mechanisms of these conventional drugs may indicate that new strategies are immediately needed to realize therapeutic development for the treatment of ALS. Since emerging evidence has supported the notion that dysregulation of autophagy is critical for the pathogenesis of ALS, the autophagic signal pathway may be a potential and/or crucial therapeutic target [100]. New therapeutic strategies for the ALS community are immediately necessary to combat the exponentially rising epidemiology of this disease [101].

6. Next Perspectives

With a complex etiology and no current cure for ALS, broadening the understanding of disease pathology is required to progress with patient care [102]. In general, mitophagy selectively degrades damaged mitochondria to suppress damaged mitochondria-derived ROS that would damage healthy mitochondria and ultimately result in mitochondrial dysfunction. As a major mechanism of mitochondrial quality control, mitophagy could degrade dysfunctional mitochondria to maintain mitochondrial integrity and function. Therefore, activated and/or appropriate mitophagy would prevent ROS from triggering oxidative stresses and inflammatory responses. In particular, counteraction of the process of oxidative stresses may be promising for prolonging life with ALS [103]. Looking for natural compounds with mitophagic actions would provide new insights into the therapeutic intervention for mitochondrial dysfunction-related diseases, including ALS. However, the therapeutic potential of autophagy modulation has not been fully exploited.

Additional efforts are being conducted to develop novel compounds with improved specificity and potency, in which a higher susceptibility of ALS lysosomes compared to healthy control could be suggested [104]. Additional studies and detailed mechanistic insights are also required to fully elucidate and decipher the function of autophagy in the pathogenesis of ALS. Another relevant and promising field of application may be various neurodegenerative diseases or aging-related disorders. The widespread inclusion of natural molecules in functional foods deserves consideration, as it might contribute significantly to the prevention and treatment of ALS, as well as to the improvement of public health. Furthermore, no curative therapeutic approach has been found to date, so further studies should be required to cure these lethal diseases one day.

Author Contributions: Conceptualization, S.Y. and S.M.; original draft preparation and editing, S.Y., K.T., H.S., Y.I., A.T. and S.M.; visualization, S.Y. and S.M.; supervision, S.M. Each author (S.Y., K.T., H.S., Y.I., A.T., S.M.) has participated sufficiently in this work of drafting the article and/or revising the article for the important rational content. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

ALS: amyotrophic lateral sclerosis
ATP: adenosine triphosphate
DNA: deoxyribonucleic acid
HDAC6; histone deacetylase 6
mTOR: mechanistic/mammalian target of rapamycin

MFN1: mitofusin 1	
mTORC: mechanistic/mammalian target of rapamycin complex	
NDP52: nuclear dot protein 52	
PARL: presenilin like	associated rhomboid
OPTN: optineurin	
PDK1: 3	phosphoinositide-dependent protein kinase
1	
PINK1: PTEN	induced kinase 1
PTEN: phosphatase and tensin homolog	
QOL: quality of life	
ROS: reactive oxygen species	
SIRT1: Sirtuin 1	
TBK1: TANK	binding kinase 1
TSC2: tuberous sclerosis protein 2	
VDAC1: voltage	dependent anion channel 1
Ub: ubiquitin	

References

- Harms, M.J.; Li, Q.; Lee, S.; Zhang, C.; Kull, B.; Hallen, S.; Thorell, A.; Alexandersson, I.; Hagberg, C.E.; Peng, X.R.; et al. Mature Human White Adipocytes Cultured under Membranes Maintain Identity, Function, and Can Transdifferentiate into Brown-like Adipocytes. *Cell Rep.* **2019**, *27*, 213–225.e5. [[CrossRef](#)] [[PubMed](#)]
- Sorrentino, V.; Menzies, K.J.; Auwerx, J. Repairing Mitochondrial Dysfunction in Disease. *Annu. Rev. Pharmacol. Toxicol.* **2018**, *58*, 353–389. [[CrossRef](#)] [[PubMed](#)]
- Osbakken, M.; Ito, K.; Zhang, D.; Ponomarenko, I.; Ivanics, T.; Jahngen, E.G.; Cohn, M. Creatine and cyclocreatine effects on ischemic myocardium: 31P nuclear magnetic resonance evaluation of intact heart. *Cardiology* **1992**, *80*, 184–195. [[CrossRef](#)] [[PubMed](#)]
- Lambert-Smith, I.A.; Saunders, D.N.; Yerbury, J.J. Proteostasis impairment and ALS. *Prog. Biophys. Mol. Biol.* **2022**, *174*, 3–27. [[CrossRef](#)]
- Kirtonia, A.; Gala, K.; Fernandes, S.G.; Pandya, G.; Pandey, A.K.; Sethi, G.; Khattar, E.; Garg, M. Repurposing of drugs: An attractive pharmacological strategy for cancer therapeutics. *Semin. Cancer Biol.* **2021**, *68*, 258–278. [[CrossRef](#)]
- Russo, E.; Nguyen, H.; Lippert, T.; Tuazon, J.; Borlongan, C.V.; Napoli, E. Mitochondrial targeting as a novel therapy for stroke. *Brain Circ.* **2018**, *4*, 84–94.
- Chua, J.P.; De Calbiac, H.; Kabashi, E.; Barmada, S.J. Autophagy and ALS: Mechanistic insights and therapeutic implications. *Autophagy* **2022**, *18*, 254–282. [[CrossRef](#)]
- Prakash, Y.S.; Pabelick, C.M.; Sieck, G.C. Mitochondrial Dysfunction in Airway Disease. *Chest* **2017**, *152*, 618–626. [[CrossRef](#)]
- Kirtonia, A.; Sethi, G.; Garg, M. The multifaceted role of reactive oxygen species in tumorigenesis. *Cell. Mol. Life Sci.* **2020**, *77*, 4459–4483. [[CrossRef](#)]
- Perrelli, A.; Retta, S.F. Polymorphisms in genes related to oxidative stress and inflammation: Emerging links with the pathogenesis and severity of Cerebral Cavernous Malformation disease. *Free Radic. Biol. Med.* **2021**, *172*, 403–417. [[CrossRef](#)]
- Chen, X.; Wang, X.; Yang, L.; Xu, H.; Wu, Y.; Wu, J.; Chen, L.; Xu, C. Magnesium isoglycyrrhizinate prevents cadmium-induced activation of JNK and apoptotic hepatocyte death by reversing ROS-inactivated PP2A. *J. Pharm. Pharmacol.* **2021**, *73*, 1663–1674. [[CrossRef](#)]
- Rabinowitz, A.R.; Li, X.; Levin, H.S. Sport and nonsport etiologies of mild traumatic brain injury: Similarities and differences. *Annu. Rev. Psychol.* **2014**, *65*, 301–331. [[CrossRef](#)]
- Pan, Q.; Ban, Y.; Xu, L. Silibinin-Albumin Nanoparticles: Characterization and Biological Evaluation Against Oxidative Stress-Stimulated Neurotoxicity Associated with Alzheimer's Disease. *J. Biomed. Nanotechnol.* **2021**, *17*, 1123–1130. [[CrossRef](#)]
- Rae, C.D.; Bröer, S. Creatine as a booster for human brain function. How might it work? *Neurochem. Int.* **2015**, *89*, 249–259. [[CrossRef](#)]
- Shah, S.I.; Paine, J.G.; Perez, C.; Ullah, G. Mitochondrial fragmentation and network architecture in degenerative diseases. *PLoS ONE* **2019**, *14*, e0223014. [[CrossRef](#)]
- Liu, Z.; Butow, R.A. Mitochondrial retrograde signaling. *Annu. Rev. Genet.* **2006**, *40*, 159–185. [[CrossRef](#)]
- Kalyanaraman, B.; Cheng, G.; Zielonka, J.; Bennett, B. Low-Temperature EPR Spectroscopy as a Probe-Free Technique for Monitoring Oxidants Formed in Tumor Cells and Tissues: Implications in Drug Resistance and OXPHOS-Targeted Therapies. *Cell Biochem. Biophys.* **2019**, *77*, 89–98. [[CrossRef](#)]
- Tan, Y.Q.; Zhang, X.; Zhang, S.; Zhu, T.; Garg, M.; Lobie, P.E.; Pandey, V. Mitochondria: The metabolic switch of cellular oncogenic transformation. *Biochim. Biophys. Acta Rev. Cancer* **2021**, *1876*, 188534. [[CrossRef](#)]

19. Bloch-Damti, A.; Bashan, N. Proposed mechanisms for the induction of insulin resistance by oxidative stress. *Antioxid. Redox Signal.* **2005**, *7*, 1553–1567. [[CrossRef](#)]
20. Mrakic-Sposta, S.; Vezzoli, A.; Rizzato, A.; Della Noce, C.; Malacrida, S.; Montorsi, M.; Paganini, M.; Cancellara, P.; Bosco, G. Oxidative stress assessment in breath-hold diving. *Eur. J. Appl. Physiol.* **2019**, *119*, 2449–2456. [[CrossRef](#)]
21. Kittimongkolsuk, P.; Pattarachotant, N.; Chuchawankul, S.; Wink, M.; Tencomnao, T. Neuroprotective Effects of Extracts from Tiger Milk Mushroom *Lignosus rhinocerus* Against Glutamate-Induced Toxicity in HT22 Hippocampal Neuronal Cells and Neurodegenerative Diseases in *Caenorhabditis elegans*. *Biology* **2021**, *10*, 30. [[CrossRef](#)] [[PubMed](#)]
22. Chan, D.C. Mitochondrial Dynamics and Its Involvement in Disease. *Annu. Rev. Pathol.* **2020**, *15*, 235–259. [[CrossRef](#)] [[PubMed](#)]
23. Boillée, S.; Vande, C.V.; Cleveland, D.W. ALS: A disease of motor neurons and their nonneuronal neighbors. *Neuron* **2006**, *52*, 39–59. [[CrossRef](#)] [[PubMed](#)]
24. Smith, E.F.; Shaw, P.J.; De Vos, K.J. The role of mitochondria in amyotrophic lateral sclerosis. *Neurosci. Lett.* **2019**, *710*, 132933. [[CrossRef](#)] [[PubMed](#)]
25. Jena, K.K.; Mehto, S.; Kolapalli, S.P.; Nath, P.; Sahu, R.; Chauhan, N.R.; Sahoo, P.K.; Dhar, K.; Das, S.K.; Chauhan, S.; et al. TRIM16 governs the biogenesis and disposal of stress-induced protein aggregates to evade cytotoxicity: Implication for neurodegeneration and cancer. *Autophagy* **2019**, *15*, 924–926. [[CrossRef](#)]
26. Su, Z.; Nie, Y.; Huang, X.; Zhu, Y.; Feng, B.; Tang, L.; Zheng, G. Mitophagy in Hepatic Insulin Resistance: Therapeutic Potential and Concerns. *Front. Pharmacol.* **2019**, *10*, 1193. [[CrossRef](#)]
27. Kim, J.; Kim, T.Y.; Cho, K.S.; Kim, H.N.; Koh, J.Y. Autophagy activation and neuroprotection by progesterone in the G93A-SOD1 transgenic mouse model of amyotrophic lateral sclerosis. *Neurobiol. Dis.* **2013**, *59*, 80–85. [[CrossRef](#)]
28. Perera, N.D.; Tomas, D.; Wanniarachchilage, N.; Cuic, B.; Luikinga, S.J.; Rytova, V.; Turner, B.J. Stimulation of mTOR-independent autophagy and mitophagy by rilmenidine exacerbates the phenotype of transgenic TDP-43 mice. *Neurobiol. Dis.* **2021**, *154*, 105359. [[CrossRef](#)]
29. Matsuda, S.; Kitagishi, Y.; Kobayashi, M. Function and characteristics of PINK1 in mitochondria. *Oxid. Med. Cell. Longev.* **2013**, *2013*, 601587. [[CrossRef](#)]
30. Matsuda, S.; Nakanishi, A.; Minami, A.; Wada, Y.; Kitagishi, Y. Functions and characteristics of PINK1 and Parkin in cancer. *Front. Biosci.* **2015**, *20*, 491–501. [[CrossRef](#)]
31. Rüb, C.; Wilkening, A.; Voos, W. Mitochondrial quality control by the Pink1/Parkin system. *Cell Tissue Res.* **2017**, *367*, 111–123. [[CrossRef](#)]
32. Narendra, D.; Tanaka, A.; Suen, D.F.; Youle, R.J. Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *J. Cell Biol.* **2008**, *183*, 795–803. [[CrossRef](#)]
33. Nardin, A.; Schrepfer, E.; Ziviani, E. Counteracting PINK/Parkin Deficiency in the Activation of Mitophagy: A Potential Therapeutic Intervention for Parkinson’s Disease. *Curr. Neuropharmacol.* **2016**, *14*, 250–259. [[CrossRef](#)]
34. Moreira, O.C.; Estébanez, B.; Martínez-Florez, S.; de Paz, J.A.; Cuevas, M.J.; González-Gallego, J. Mitochondrial Function and Mitophagy in the Elderly: Effects of Exercise. *Oxid. Med. Cell Longev.* **2017**, *2017*, 2012798. [[CrossRef](#)]
35. Wang, H.; Ni, H.M.; Chao, X.; Ma, X.; Rodriguez, Y.A.; Chavan, H.; Wang, S.; Krishnamurthy, P.; Dobrowsky, R.; Xu, D.X.; et al. Double deletion of PINK1 and Parkin impairs hepatic mitophagy and exacerbates acetaminophen-induced liver injury in mice. *Redox Biol.* **2019**, *22*, 101148. [[CrossRef](#)]
36. Eldeeb, M.A.; Ragheb, M.A. N-degron-mediated degradation and regulation of mitochondrial PINK1 kinase. *Curr. Genet.* **2020**, *66*, 693–701. [[CrossRef](#)]
37. Li, X.; Huang, L.; Lan, J.; Feng, X.; Li, P.; Wu, L.; Peng, Y. Molecular mechanisms of mitophagy and its roles in neurodegenerative diseases. *Pharmacol. Res.* **2021**, *163*, 105240. [[CrossRef](#)]
38. Oakes, J.A.; Davies, M.C.; Collins, M.O. TBK1: A new player in ALS linking autophagy and neuroinflammation. *Mol. Brain* **2017**, *10*, 5. [[CrossRef](#)]
39. Xie, Y.; Zhou, B.; Lin, M.Y.; Wang, S.; Foust, K.D.; Sheng, Z.H. Endolysosomal Deficits Augment Mitochondria Pathology in Spinal Motor Neurons of Asymptomatic fALS Mice. *Neuron* **2015**, *87*, 355–370. [[CrossRef](#)]
40. Chennampally, P.; Sayed-Zahid, A.; Soundararajan, P.; Sharp, J.; Cox, G.A.; Collins, S.D.; Smith, R.L. A microfluidic approach to rescue ALS motor neuron degeneration using rapamycin. *Sci. Rep.* **2021**, *11*, 18168. [[CrossRef](#)]
41. Mugume, Y.; Kazibwe, Z.; Bassham, D.C. Target of Rapamycin in Control of Autophagy: Puppet Master and Signal Integrator. *Int. J. Mol. Sci.* **2020**, *21*, 8259. [[CrossRef](#)] [[PubMed](#)]
42. Ejaz, A.; Mattesich, M.; Zwerschke, W. Silencing of the small GTPase DIRAS3 induces cellular senescence in human white adipose stromal/progenitor cells. *Aging* **2017**, *9*, 860–879. [[CrossRef](#)] [[PubMed](#)]
43. Barzilai, N.; Crandall, J.P.; Kritchevsky, S.B.; Espeland, M.A. Metformin as a Tool to Target Aging. *Cell Metab.* **2016**, *23*, 1060–1065. [[CrossRef](#)] [[PubMed](#)]
44. Ning, X.; He, J.; Shi, X.; Yang, G. Regulation of Adipogenesis by Quinine through the ERK/S6 Pathway. *Int. J. Mol. Sci.* **2016**, *17*, 504. [[CrossRef](#)] [[PubMed](#)]
45. Guertin, D.A.; Sabatini, D.M. The pharmacology of mTOR inhibition. *Sci. Signal.* **2009**, *2*, pe24. [[CrossRef](#)]
46. Stokoe, D.; Stephens, L.R.; Copeland, T.; Gaffney, P.R.; Reese, C.B.; Painter, G.F.; Holmes, A.B.; McCormick, F.; Hawkins, P.T. Dual role of phosphatidylinositol-3,4,5-trisphosphate in the activation of protein kinase B. *Science* **1997**, *277*, 567–570. [[CrossRef](#)]

47. Long, X.; Lin, Y.; Ortiz-Vega, S.; Yonezawa, K.; Avruch, J. Rheb binds and regulates the mTOR kinase. *Curr. Biol.* **2005**, *15*, 702–713. [[CrossRef](#)]
48. Egan, D.; Kim, J.; Shaw, R.J.; Guan, K.L. The autophagy initiating kinase ULK1 is regulated via opposing phosphorylation by AMPK and mTOR. *Autophagy* **2011**, *7*, 643–644. [[CrossRef](#)]
49. Høyer-Hansen, M.; Bastholm, L.; Szyniarowski, P.; Campanella, M.; Szabadkai, G.; Farkas, T.; Bianchi, K.; Fehrenbacher, N.; Ellingm, F.; Rizzuto, R.; et al. Control of macroautophagy by calcium, calmodulin-dependent kinase kinase-beta, and Bcl-2. *Mol. Cell* **2007**, *25*, 193–205. [[CrossRef](#)]
50. Fang, J.; Yang, J.; Wu, X.; Zhang, G.; Li, T.; Wang, X.; Zhang, H.; Wang, C.C.; Liu, G.H.; Wang, L. Metformin alleviates human cellular aging by upregulating the endoplasmic reticulum glutathione peroxidase 7. *Aging Cell* **2018**, *17*, e12765. [[CrossRef](#)]
51. Zhang, K.; Zhou, X.; Wang, J.; Zhou, Y.; Qi, W.; Chen, H.; Nie, S.; Xie, M. Dendrobium officinale polysaccharide triggers mitochondrial disorder to induce colon cancer cell death via ROS-AMPK-autophagy pathway. *Carbohydr. Polym.* **2021**, *264*, 118018. [[CrossRef](#)]
52. Williams, A.; Sarkar, S.; Cuddon, P.; Ttofi, E.K.; Saiki, S.; Siddiqi, F.H.; Jahreiss, L.; Fleming, A.; Pask, D.; Goldsmith, P.; et al. Novel targets for Huntington’s disease in an mTOR-independent autophagy pathway. *Nat. Chem. Biol.* **2008**, *4*, 295–305. [[CrossRef](#)]
53. Hsueh, K.W.; Chiou, T.W.; Chiang, S.F.; Yamashita, T.; Abe, K.; Borlongan, C.V.; Sanberg, P.R.; Huang, A.Y.; Lin, S.Z.; Harn, H.J. Autophagic down-regulation in motor neurons remarkably prolongs the survival of ALS mice. *Neuropharmacology*. **2016**, *108*, 152–160. [[CrossRef](#)]
54. Suntar, I.; Sureda, A.; Belwal, T.; Sanches Silva, A.; Vacca, R.A.; Tewari, D.; Sobarzo-Sánchez, E.; Nabavi, S.F.; Shirooie, S.; Dehpour, A.R.; et al. Natural products, PGC-1 α , and Duchenne muscular dystrophy. *Acta Pharm. Sin. B* **2020**, *10*, 734–745. [[CrossRef](#)]
55. Lilamand, M.; Mouton-Liger, F.; Di Valentin, E.; Sánchez, O.M.; Paquet, C. Efficacy and Safety of Ketone Supplementation or Ketogenic Diets for Alzheimer’s Disease: A Mini Review. *Front. Nutr.* **2022**, *8*, 807970. [[CrossRef](#)]
56. Chen, N.; Qi, Y.; Ma, X.; Xiao, X.; Liu, Q.; Xia, T.; Xiang, J.; Zeng, J.; Tang, J. Rediscovery of Traditional Plant Medicine: An Underestimated Anticancer Drug of Chelerythrine. *Front. Pharmacol.* **2022**, *13*, 906301. [[CrossRef](#)]
57. Hsu, C.M.; Yen, C.H.; Wang, S.C.; Liu, Y.C.; Huang, C.T.; Wang, M.H.; Chuang, T.M.; Ke, Y.L.; Yeh, T.J.; Gau, Y.C.; et al. Emodin Ameliorates the Efficacy of Carfilzomib in Multiple Myeloma Cells via Apoptosis and Autophagy. *Biomedicines* **2022**, *10*, 1638. [[CrossRef](#)]
58. Hosseinpour-Moghaddam, K.; Caraglia, M.; Sahebkar, A. Autophagy induction by trehalose: Molecular mechanisms and therapeutic impacts. *J. Cell. Physiol.* **2018**, *233*, 6524–6543. [[CrossRef](#)]
59. Rusmini, P.; Cortese, K.; Crippa, V.; Cristofani, R.; Cicardi, M.E.; Ferrari, V.; Vezzoli, G.; Tedesco, B.; Meroni, M.; Messi, E.; et al. Trehalose induces autophagy via lysosomal-mediated TFEB activation in models of motoneuron degeneration. *Autophagy* **2019**, *15*, 631–651. [[CrossRef](#)]
60. Bordoni, M.; Pansarasa, O.; Scarian, E.; Cristofani, R.; Leone, R.; Fantini, V.; Garofalo, M.; Diamanti, L.; Bernuzzi, S.; Gagliardi, S.; et al. Lysosomes Dysfunction Causes Mitophagy Impairment in PBMcs of Sporadic ALS Patients. *Cells* **2022**, *11*, 1272. [[CrossRef](#)]
61. DeBosch, B.J.; Heitmeier, M.R.; Mayer, A.L.; Higgins, C.B.; Crowley, J.R.; Kraft, T.E.; Chi, M.; Newberry, E.P.; Chen, Z.; Finck, B.N.; et al. Trehalose inhibits solute carrier 2A (SLC2A) proteins to induce autophagy and prevent hepatic steatosis. *Sci. Signal.* **2016**, *9*, ra21. [[CrossRef](#)] [[PubMed](#)]
62. Jeong, S.J.; Stitham, J.; Evans, T.D.; Zhang, X.; Rodriguez-Velez, A.; Yeh, Y.S.; Tao, J.; Takabatake, K.; Eelman, S.; Lodhi, I.J.; et al. Trehalose causes low-grade lysosomal stress to activate TFEB and the autophagy-lysosome biogenesis response. *Autophagy* **2021**, *17*, 3740–3752. [[CrossRef](#)] [[PubMed](#)]
63. Zhao, H.; Chen, S.; Gao, K.; Zhou, Z.; Wang, C.; Shen, Z.; Guo, Y.; Li, Z.; Wan, Z.; Liu, C.; et al. Resveratrol protects against spinal cord injury by activating autophagy and inhibiting apoptosis mediated by the SIRT1/AMPK signaling pathway. *Neuroscience* **2017**, *348*, 241–251. [[CrossRef](#)] [[PubMed](#)]
64. Maher, P.; Dargusch, R.; Bodai, L.; Gerard, P.E.; Purcell, J.M.; Marsh, J.L. ERK activation by the polyphenols fisetin and resveratrol provides neuroprotection in multiple models of Huntington’s disease. *Hum. Mol. Genet.* **2011**, *20*, 261–270. [[CrossRef](#)] [[PubMed](#)]
65. Hecht, J.T.; Coustry, F.; Veerisetty, A.C.; Hossain, M.G.; Posey, K.L. Resveratrol Reduces COMPopathy in Mice Through Activation of Autophagy. *JBMR Plus* **2021**, *5*, e10456. [[CrossRef](#)]
66. Suvorova, I.I.; Knyazeva, A.R.; Petukhov, A.V.; Aksenov, N.D.; Pospelov, V.A. Resveratrol enhances pluripotency of mouse embryonic stem cells by activating AMPK/Ulk1 pathway. *Cell Death Discov.* **2019**, *5*, 61. [[CrossRef](#)]
67. Atiya, A.M.; Poortvliet, E.; Strömberg, R.; Yngve, A. Polyamines in foods: Development of a food database. *Food Nutr. Res.* **2011**, *55*, 5572. [[CrossRef](#)]
68. Wirth, M.; Schwarz, C.; Benson, G.; Horn, N.; Buchert, R.; Lange, C.; Köbe, T.; Hetzer, S.; Maglione, M.; Michael, E.; et al. Effects of spermidine supplementation on cognition and biomarkers in older adults with subjective cognitive decline (SmartAge)-study protocol for a randomized controlled trial. *Alzheimers Res. Ther.* **2019**, *11*, 36. [[CrossRef](#)]
69. Yan, J.; Yan, J.Y.; Wang, Y.X.; Ling, Y.N.; Song, X.D.; Wang, S.Y.; Liu, H.Q.; Liu, Q.C.; Zhang, Y.; Yang, P.Z.; et al. Spermidine-enhanced autophagic flux improves cardiac dysfunction following myocardial infarction by targeting the AMPK/mTOR signalling pathway. *Br. J. Pharmacol.* **2019**, *176*, 3126–3142. [[CrossRef](#)]
70. Eisenberg, T.; Abdellatif, M.; Schroeder, S.; Primessnig, U.; Stekovic, S.; Pendl, T.; Harger, A.; Schipke, J.; Zimmermann, A.; Schmidt, A.; et al. Cardioprotection and lifespan extension by the natural polyamine spermidine. *Nat. Med.* **2016**, *22*, 1428–1438. [[CrossRef](#)]

71. Pietrocola, F.; Lachkar, S.; Enot, D.P.; Niso-Santano, M.; Bravo-San Pedro, J.M.; Sica, V.; Izzo, V.; Maiuri, M.C.; Madeo, F.; Mariño, G.; et al. Spermidine induces autophagy by inhibiting the acetyltransferase EP300. *Cell Death Differ.* **2015**, *22*, 509–516. [[CrossRef](#)]
72. Cerdá, B.; Periago, P.; Espín, J.C.; Tomás-Barberán, F.A. Identification of urolithin A as a metabolite produced by human colon microflora from ellagic acid and related compounds. *J. Agric. Food Chem.* **2005**, *53*, 5571–5576. [[CrossRef](#)]
73. Wang, Y.; Huang, H.; Jin, Y.; Shen, K.; Chen, X.; Xu, Z.; Jin, B.; Pan, H. Role of TFEB in autophagic modulation of ischemia reperfusion injury in mice kidney and protection by urolithin A. *Food Chem. Toxicol.* **2019**, *131*, 110591. [[CrossRef](#)]
74. Kujawska, M.; Jourdes, M.; Kurpik, M.; Szulc, M.; Szafer, H.; Chmielarz, P.; Kreiner, G.; Krajka-Kuźniak, V.; Mikołajczak, P.L.; Teissedre, P.L.; et al. Neuroprotective Effects of Pomegranate Juice against Parkinson's Disease and Presence of Ellagitannin-Derived Metabolite-Urolithin A-In the Brain. *Int. J. Mol. Sci.* **2019**, *21*, 202. [[CrossRef](#)]
75. Ryu, D.; Mouchiroud, L.; Andreux, P.A.; Katsyuba, E.; Moullan, N.; Nicolet-Dit-Félix, A.A.; Williams, E.G.; Jha, P.; Lo Sasso, G.; Huzard, D.; et al. Urolithin A induces mitophagy and prolongs lifespan in *C. elegans* and increases muscle function in rodents. *Nat. Med.* **2016**, *22*, 879–888. [[CrossRef](#)]
76. Tuohetaerbaik, B.; Zhang, Y.; Tian, Y.; Zhang, N.N.; Kang, J.; Mao, X.; Zhang, Y.; Li, X. Pancreas protective effects of Urolithin A on type 2 diabetic mice induced by high fat and streptozotocin via regulating autophagy and AKT/mTOR signaling pathway. *J. Ethnopharmacol.* **2020**, *250*, 112479. [[CrossRef](#)]
77. Kim, Y.K.; Yoon, H.H.; Lee, Y.D.; Youn, D.Y.; Ha, T.J.; Kim, H.S.; Lee, J.H. Anthocyanin Extracts from Black Soybean (*Glycine max* L.) Protect Human Glial Cells Against Oxygen-Glucose Deprivation by Promoting Autophagy. *Biomol. Ther.* **2012**, *20*, 68–74. [[CrossRef](#)]
78. Youdim, K.A.; Shukitt-Hale, B.; Joseph, J.A. Flavonoids and the brain: Interactions at the blood-brain barrier and their physiological effects on the central nervous system. *Free Radic. Biol. Med.* **2004**, *37*, 1683–1693. [[CrossRef](#)]
79. Li, J.; Zhao, R.; Zhao, H.; Chen, G.; Jiang, Y.; Lyu, X.; Wu, T. Reduction of Aging-Induced Oxidative Stress and Activation of Autophagy by Bilberry Anthocyanin Supplementation via the AMPK-mTOR Signaling Pathway in Aged Female Rats. *J. Agric. Food Chem.* **2019**, *67*, 7832–7843. [[CrossRef](#)]
80. Wu, Y.; Hu, Y.; Zhou, H.; Zhu, J.; Tong, Z.; Qin, S.; Liu, D. Organosulfur compounds induce cytoprotective autophagy against apoptosis by inhibiting mTOR phosphorylation activity in macrophages. *Acta Biochim. Biophys. Sin.* **2018**, *50*, 1085–1093. [[CrossRef](#)]
81. Chu, Y.L.; Ho, C.T.; Chung, J.G.; Raghu, R.; Lo, Y.C.; Sheen, L.Y. Allicin induces anti-human liver cancer cells through the p53 gene modulating apoptosis and autophagy. *J. Agric. Food Chem.* **2013**, *61*, 9839–9848. [[CrossRef](#)] [[PubMed](#)]
82. Liu, C.; Leng, B.; Li, Y.; Jiang, H.; Duan, W.; Guo, Y.; Li, C.; Hong, K. Diallyl Trisulfide Protects Motor Neurons from the Neurotoxic Protein TDP-43 via Activating Lysosomal Degradation and the Antioxidant Response. *Neurochem. Res.* **2018**, *43*, 2304–2312. [[CrossRef](#)] [[PubMed](#)]
83. Liu, Y.; Yang, B.; Zhang, L.; Cong, X.; Liu, Z.; Hu, Y.; Zhang, J.; Hu, H. Ginkgolic acid induces interplay between apoptosis and autophagy regulated by ROS generation in colon cancer. *Biochem. Biophys. Res. Commun.* **2018**, *498*, 246–253. [[CrossRef](#)] [[PubMed](#)]
84. Vijayakumar, S.; Nakamura, Y.; Henley, J.M.; Pountney, D.L. Ginkgolic acid promotes autophagy-dependent clearance of intracellular alpha-synuclein aggregates. *Mol. Cell. Neurosci.* **2019**, *101*, 103416. [[CrossRef](#)] [[PubMed](#)]
85. Zhang, H.; Yang, X.; Pang, X.; Zhao, Z.; Yu, H.; Zhou, H. Genistein protects against ox-LDL-induced senescence through enhancing SIRT1/LKB1/AMPK-mediated autophagy flux in HUVECs. *Mol. Cell. Biochem.* **2019**, *455*, 127–134. [[CrossRef](#)]
86. Wang, Y.; Li, Y.; Zhang, T.; Chi, Y.; Liu, M.; Liu, Y. Genistein and Myd88 Activate Autophagy in High Glucose-Induced Renal Podocytes In Vitro. *Med. Sci. Monit.* **2018**, *24*, 4823–4831. [[CrossRef](#)]
87. Jopowicz, A.; Wiśniowska, J.; Tarnacka, B. Cognitive and Physical Intervention in Metals' Dysfunction and Neurodegeneration. *Brain Sci.* **2022**, *12*, 345. [[CrossRef](#)]
88. Aridegbe, T.; Kandler, R.; Walters, S.J.; Walsh, T.; Shaw, P.J.; McDermott, C.J. The natural history of motor neuron disease: Assessing the impact of specialist care. *Amyotroph. Lateral Scler. Front. Degener.* **2013**, *14*, 13–19. [[CrossRef](#)]
89. Van Es, M.A.; Hardiman, O.; Chio, A.; Al-Chalabi, A.; Pasterkamp, R.J.; Veldink, J.H.; van den Berg, L.H. Amyotrophic lateral sclerosis. *Lancet* **2017**, *390*, 2084–2098. [[CrossRef](#)]
90. Scott, A. Drug therapy: On the treatment trail for ALS. *Nature* **2017**, *550*, S120–S121. [[CrossRef](#)]
91. Wobst, H.J.; Mack, K.L.; Brown, D.G.; Brandon, N.J.; Shorter, J. The clinical trial landscape in amyotrophic lateral sclerosis—Past, present, and future. *Med. Res. Rev.* **2020**, *40*, 1352–1384. [[CrossRef](#)]
92. Bellingham, M.C. A review of the neural mechanisms of action and clinical efficiency of riluzole in treating amyotrophic lateral sclerosis: What have we learned in the last decade? *CNS Neurosci. Ther.* **2011**, *17*, 4–31. [[CrossRef](#)]
93. Andrews, J.A.; Jackson, C.E.; Heiman-Patterson, T.D.; Bettica, P.; Brooks, B.R.; Pioro, E.P. Real-world evidence of riluzole effectiveness in treating amyotrophic lateral sclerosis. *Amyotroph. Lateral Scler. Front. Degener.* **2020**, *21*, 509–518. [[CrossRef](#)]
94. West, R.J.H.; Ugbo, C.; Fort-Aznar, L.; Sweeney, S.T. Neuroprotective activity of ursodeoxycholic acid in CHMP2B^{Intron5} models of frontotemporal dementia. *Neurobiol. Dis.* **2020**, *144*, 105047. [[CrossRef](#)]
95. Sala, G.; Arosio, A.; Conti, E.; Beretta, S.; Lunetta, C.; Riva, N.; Ferrarese, C.; Tremolizzo, L. Riluzole Selective Antioxidant Effects in Cell Models Expressing Amyotrophic Lateral Sclerosis Endophenotypes. *Clin. Psychopharmacol. Neurosci.* **2019**, *17*, 438–442. [[CrossRef](#)]
96. Rothstein, J.D. Edaravone: A new drug approved for ALS. *Cell* **2017**, *171*, 725. [[CrossRef](#)]

97. Sawada, H. Clinical efficacy of edaravone for the treatment of amyotrophic lateral sclerosis. *Expert Opin. Pharmacother.* **2017**, *18*, 735–738. [[CrossRef](#)]
98. Guo, Z.; Wu, H.T.; Li, X.X.; Yu, Y.; Gu, R.Z.; Lan, R.; Qin, X.Y. Edaravone protects rat astrocytes from oxidative or neurotoxic inflammatory insults by restoring Akt/Bcl-2/Caspase-3 signaling axis. *IBRO Rep.* **2020**, *8*, 122–128. [[CrossRef](#)]
99. Matsumoto, S.; Murozono, M.; Kanazawa, M.; Nara, T.; Ozawa, T.; Watanabe, Y. Edaravone and cyclosporine A as neuroprotective agents for acute ischemic stroke. *Acute Med. Surg.* **2018**, *5*, 213–221. [[CrossRef](#)]
100. Xu, X.; Shen, D.; Gao, Y.; Zhou, Q.; Ni, Y.; Meng, H.; Shi, H.; Le, W.; Chen, S.; Chen, S. A perspective on therapies for amyotrophic lateral sclerosis: Can disease progression be curbed? *Transl. Neurodegener.* **2021**, *10*, 29. [[CrossRef](#)]
101. Arthur, K.C.; Calvo, A.; Price, T.R.; Geiger, J.T.; Chiò, A.; Traynor, B.J. Projected increase in amyotrophic lateral sclerosis from 2015 to 2040. *Nat. Commun.* **2016**, *7*, 12408. [[CrossRef](#)] [[PubMed](#)]
102. Roberts, B.; Theunissen, F.; Mastaglia, F.L.; Akkari, P.A.; Flynn, L.L. Synucleinopathy in Amyotrophic Lateral Sclerosis: A Potential Avenue for Antisense Therapeutics? *Int. J. Mol. Sci.* **2022**, *23*, 9364. [[CrossRef](#)] [[PubMed](#)]
103. Gordon, P.H. Amyotrophic Lateral Sclerosis: An update for 2013 Clinical Features, Pathophysiology, Management and Therapeutic Trials. *Aging Dis.* **2013**, *4*, 295–310. [[CrossRef](#)] [[PubMed](#)]
104. Amin, A.; Perera, N.D.; Beart, P.M.; Turner, B.J.; Shabanpoor, F. Amyotrophic Lateral Sclerosis and Autophagy: Dysfunction and Therapeutic Targeting. *Cells* **2020**, *9*, 2413. [[CrossRef](#)]