Supplementary Material





Figure S1. A23187 induces bovine NETs release. 200.000 bovine PMN were incubated with 5 or 25 μ M of A23187 for 3h. Fixed in 4% PFA and treated with specific antibodies to detect Neutrophil elastase. Samples were mounted in DAPI-containing mounting media and the cells releasing NETs were counted. Representative images are shown in (A). Data is represented as box and whiskers plot (B). Statistical significance determined by Kruskal-Wallis and Dunn's post-test comparing against control condition (PMN + mock).



Figure S2. PMN (n = 3) were incubated with *B. besnoiti* tachyzoites, PMA 100 nM or plain media as described in the material and methods section to induce NETs and the % of cells releasing NETs was determined following the methodology proposed by Gonzalez et al. (2014) using Sytox Orange to stain DNA. As suggested, nuclear are expansion over a threshold of 80 μ m² was considered to

determine a PMN as positive for releasing NETs. Gonzalez, A.S., Bardoel, B.W., Harbort, C.J., Zychlinsky, A., 2014. Induction and quantification of neutrophil extracellular traps. Methods Mol. Biol. 1124, 307–318. https://doi.org/10.1007/978-1-62703-845-4_20



Figure S3. Representative figures obtained in under flow experiments and posterior staining with DAPI (Blue), Isolectin IB-4 coupled with Alexa Fluor 594 (Red). Control and infected conditions are shown. In both cases PMN were perfused at physiological flow (1 dyn/cm²) before fixing the samples.



Figure S4. *B. besnoiti* tachyzoites induce NETs. 2×10^5 PMN were confronted with 8×10^5 *B. besnoiti* tachyzoites (1:4 ratio) for 3 h at 37 °C and 5% CO₂. Samples were fixed and stained for DNA (Sytox Orange). Nuclear decondensation and extracellular DNA entangling *B. besnoiti* tachyzoites were observed under confocal microscopy.



