

A Zebrafish Acromegaly Model Elevates DNA Damage and Impairs DNA Repair Pathways

Abdalla Elbially ¹, Shuichi Asakawa ¹, Shugo Watabe ² and Shigeharu Kinoshita ^{1,*}

Supplementary Tables

Table S1. Primers used for PCR.

Primer	Sequence	Reference
GH. F	ATCAGAACCACCGACTCACATCATAATC	This study
GH. R	CAATGCAACACATTTATTTACAGAATAACATTATTCC	This study

Table S2. Sequences of qPCR primers for selected genes.

Primer	Forward Primer (5 to 3)	Reverse Primer (5 to 3)	Reference
Insulin-like growth factor 1 (IGF1a)	ACACAGGGGGCAGAACTAT	AAGATGGGGCTTAAACGTCC	[42]
Insulin-like growth factor 3 (IGF1b)	CGTGATGTCCCTGCATCTGT	TTCTGGTATCGCCGCTGAAA	This study
Zebrafish growth hormone (GH _z)	AGGTCTTATGCCTGAGGAACG	AAGGTCTGGCTGGGAAACTC	[42]
Elongation factor 1 alpha (EF1)	TGTCCTCAAGCCTGGTATGG	TGGGTCGTTCTTGCTGTCTC	[42]
Tumor suppressor p53 (p53)	ATCATCTGAGCCCAAACAGG	AAATGACCCCTGTGACAAGC	[17]

Table S3. DNA damage gene set used for GSEA.

Zebrafish Gene Name	Gene Symbol
X-ray repair complementing defective repair in Chinese hamster cells 2	xrcc2
UDP glucuronosyltransferase 1 family a, b	utg1ab
uracil-DNA glycosylase a	unga
RAD50 homolog	rad50
RAD23 homolog Aa	rad23aa
excision repair cross-complementing rodent repair deficiency, complementation group 1	ercc1
excision repair cross-complementing rodent repair deficiency, complementation group 3	ercc3
damage specific DNA binding protein 1	ddb1
protein kinase Chk2	chk2
ataxia telangiectasia mutated	atm