

Figure S1: Expression of neural progenitor markers in NPCs and neuronal markers in differentiated neurons. Representative micrographs confirm the presence of neural progenitor markers (**A**) Nestin and SOX1, (**B**) PAX6 and SOX2 in Mut1, GC1, Mut2, and GC2. Scale bar = 100 μ m. (**C**) Representative micrographs confirm the presence of pan-neuronal markers MAP2 and TUBB3 in neurons differentiated from Mut1, GC1, Mut2, and GC2. Scale bar = 25 μ m.

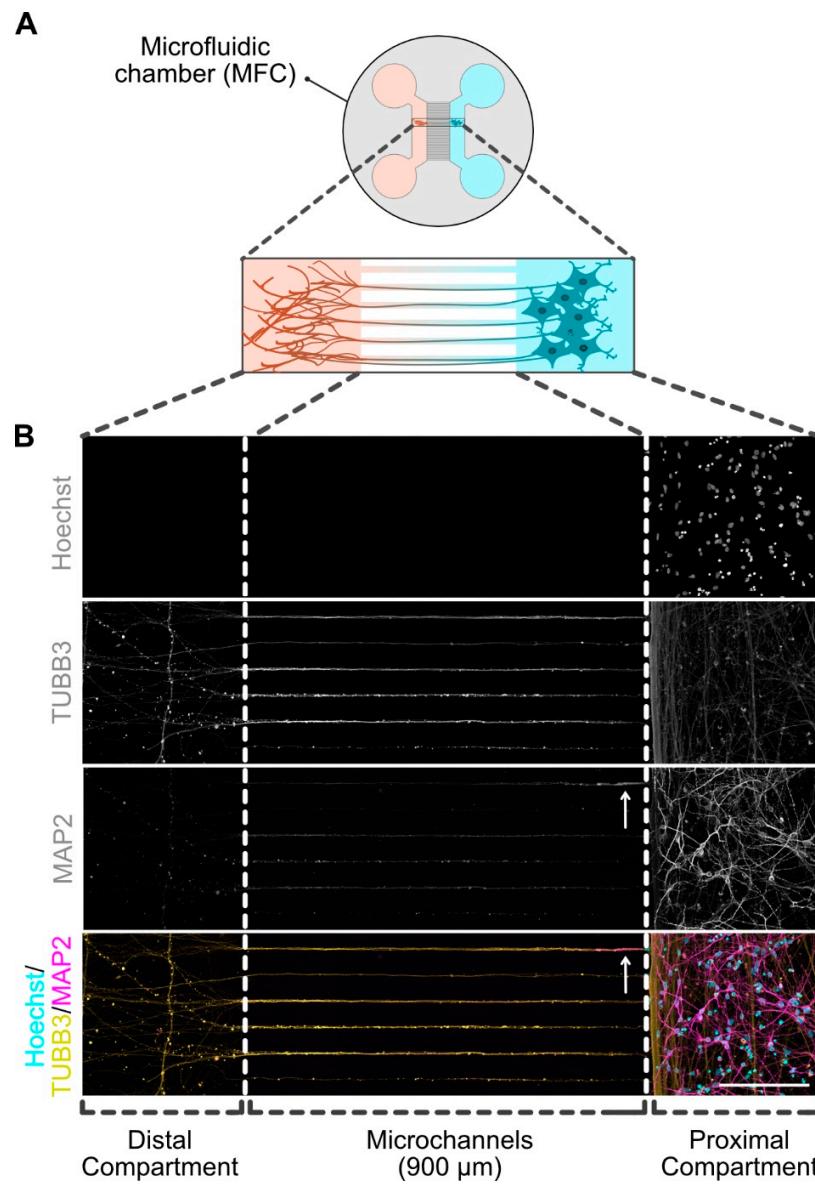


Figure S2: Spatial isolation of axons from soma and dendrites using microfluidic chambers.

(**A**) Schematic representation of MFCs, with soma in the proximal compartment and axonal outgrowth through microchannels (900 μ m) to the distal compartment. (**B**) Representative micrographs confirm TUBB3-positive axons in the distal compartment, while nuclei (Hoechst) and dendrites (MAP2) remain primarily in the proximal compartment. A white arrow marks the initial section of a microchannel penetrated by dendrites. Scale bar = 200 μ m.

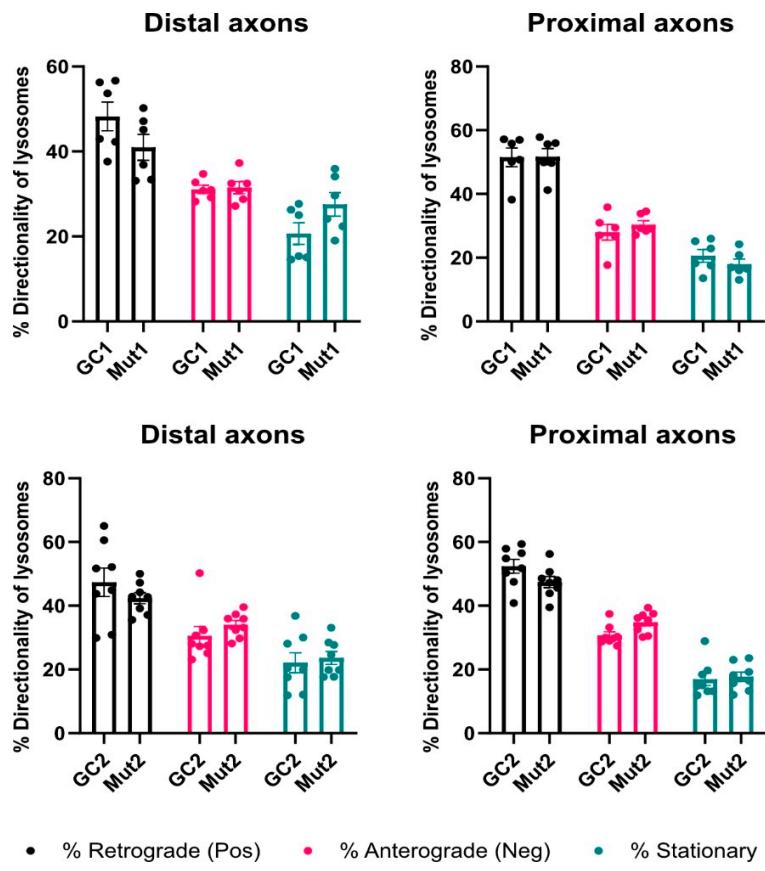


Figure S3: Directionality of lysosomes.

The relative proportion of retrograde, anterograde, and stationary lysosomes in distal and proximal axons of GC1-Mut1 (top) and GC2-Mut2 (bottom). There is no significant difference between GC and Mut in any direction, according to 2-way ANOVA.

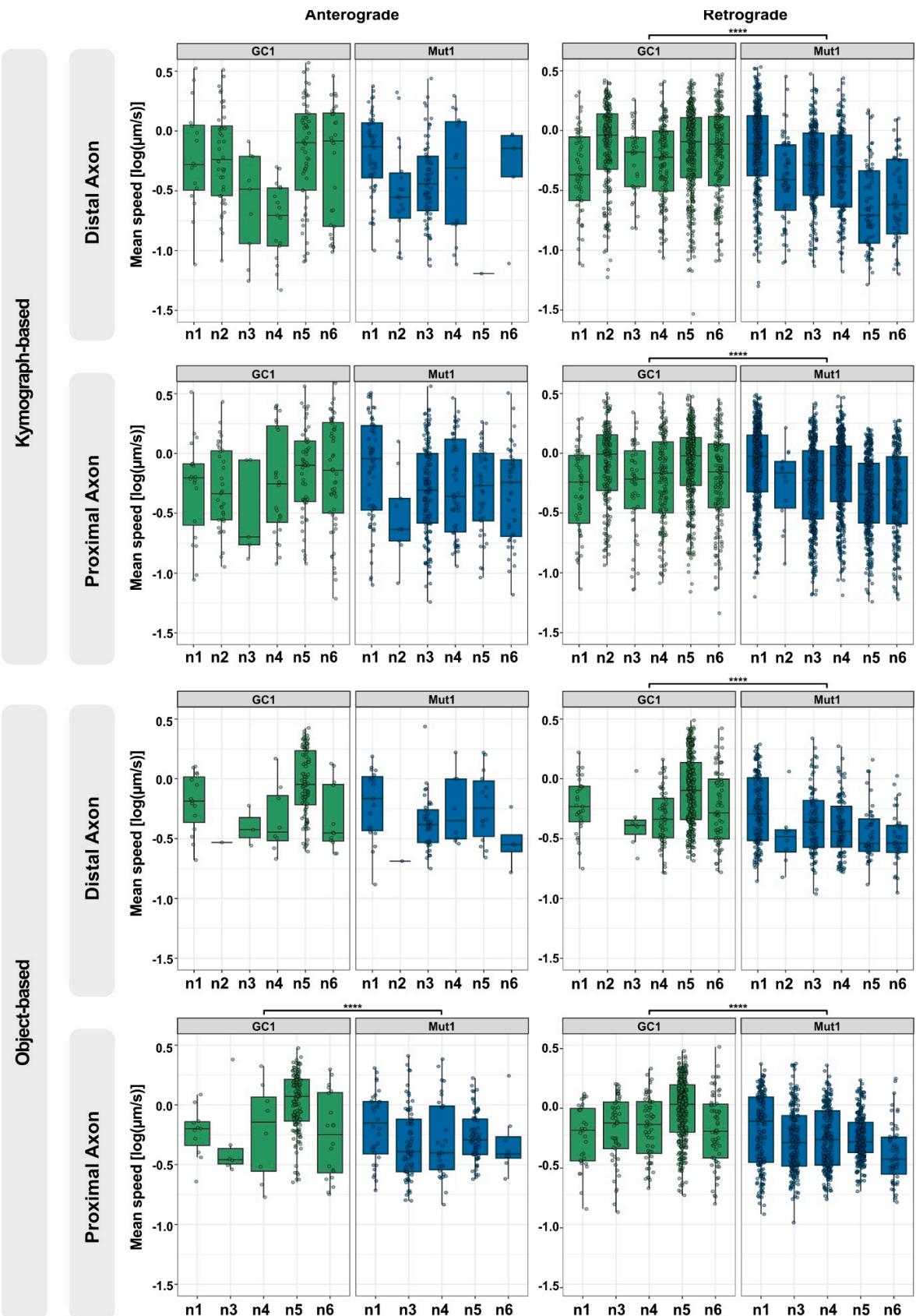


Figure S4. Quantification of mean speed of lysosomal trafficking in GC1 and Mut1 neurons.

Modified graphical representation of Figure 2A, with box-plots of individual experimental replicates (n), quantifying the mean speed of anterograde and retrograde lysosomes in distal and proximal axons of GC1 and Mut1 neurons by kymograph-based analysis (top) and object-based analysis (bottom). Statistics were calculated using 2-way ANOVA with post-hoc Tukey test to compare GC and Mut, considering any variation between the experimental replicates. **** corresponds to $p\text{-adj} \leq 0.0001$. Comparisons without a marked * did not report any significant differences. The exact mean difference and p-values can be found in Table S1.

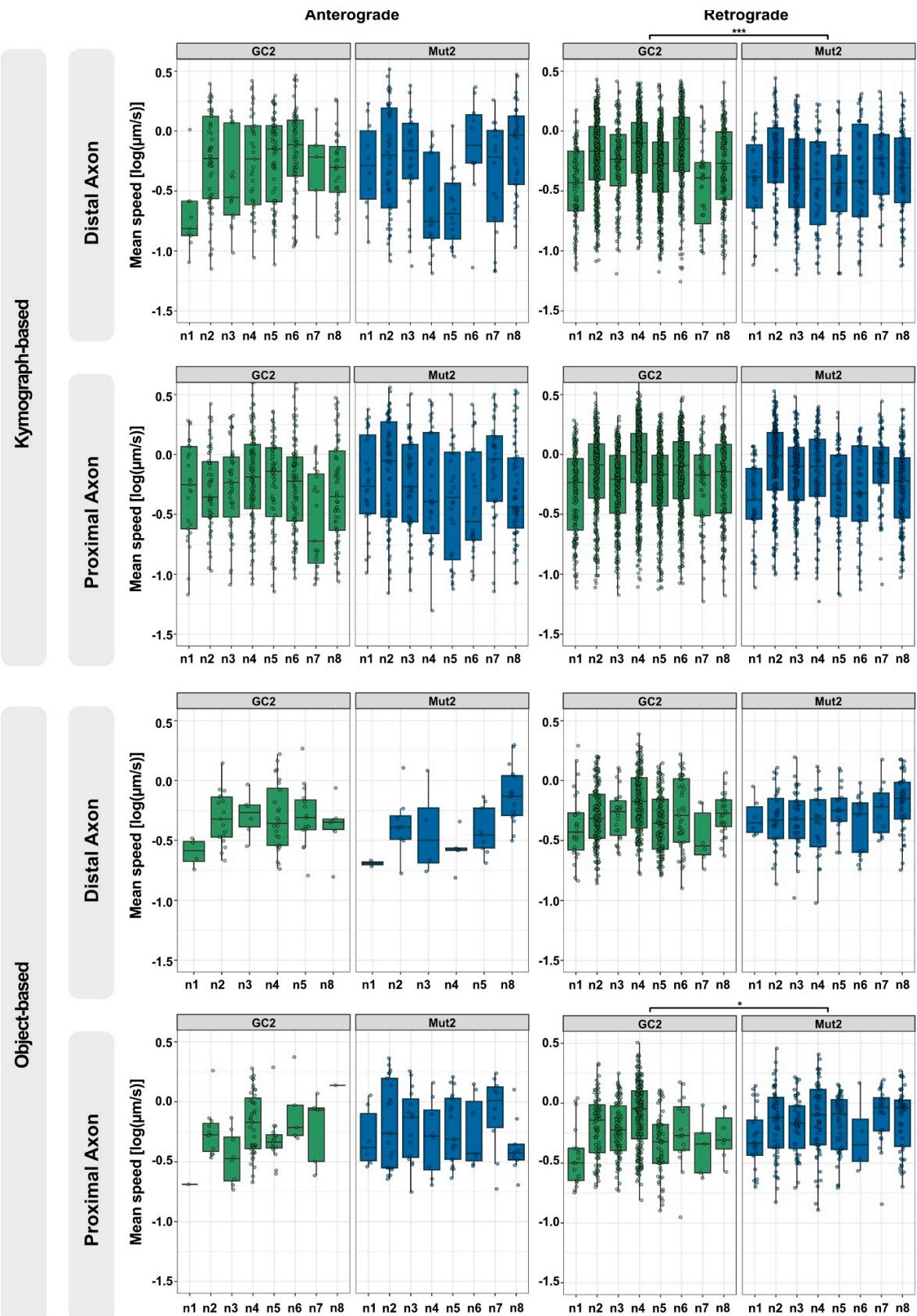


Figure S5. Quantification of mean speed of lysosomal trafficking in GC2 and Mut2 neurons.

Modified graphical representation of Figure 2B, with box-plots of individual experimental replicates (n), quantifying the mean speed of anterograde and retrograde lysosomes in distal and proximal axons of GC1 and Mut1 neurons by kymograph-based analysis (top) and object-based analysis (bottom). Statistics were calculated using 2-way ANOVA with post-hoc Tukey test to compare GC and Mut, considering any variation between the experimental replicates. *** corresponds to $p\text{-adj} \leq 0.001$, and * to $p\text{-adj} \leq 0.05$. Comparisons without a marked * did not report any significant differences. The exact mean difference and p-values can be found in Table S1.

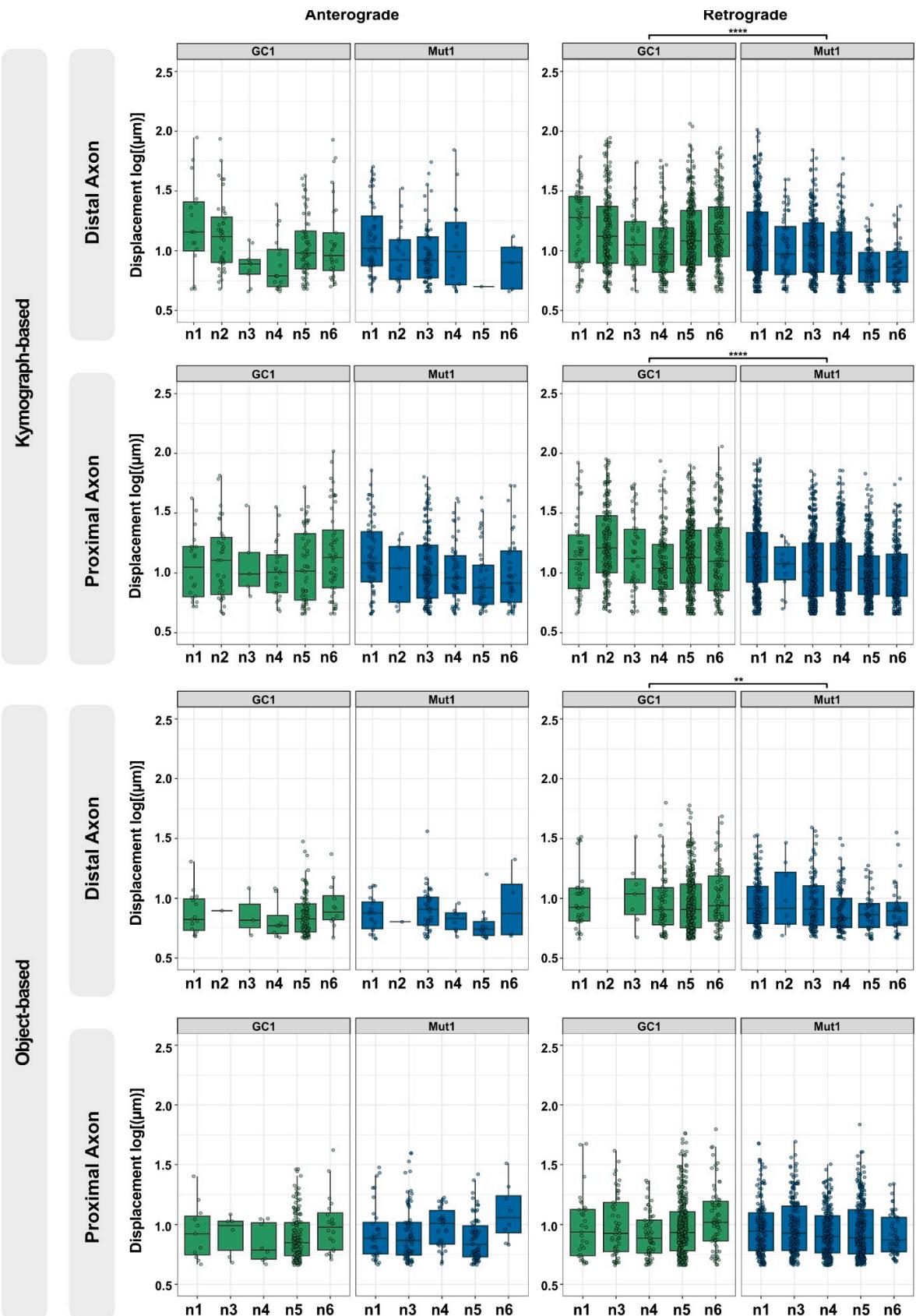


Figure S6: LRRK2 G2019S is associated with a small reduction in the displacement of retrograde lysosomal trafficking in Mut1 neurons.

Quantification of displacement of anterograde and retrograde lysosomes in the distal and proximal axons of GC1 and Mut1 neurons by kymograph-based analysis (top) and object-based analysis (bottom). Statistics were calculated using 2-way ANOVA with post-hoc Tukey test to compare GC and Mut, taking into consideration any variation between the experimental replicates (n). **** corresponds to p-adj <= 0.0001, ** to p-adj <= 0.01. Comparisons without a marked * did not report any significant differences. The exact mean difference and p-values can be found in Table S1.

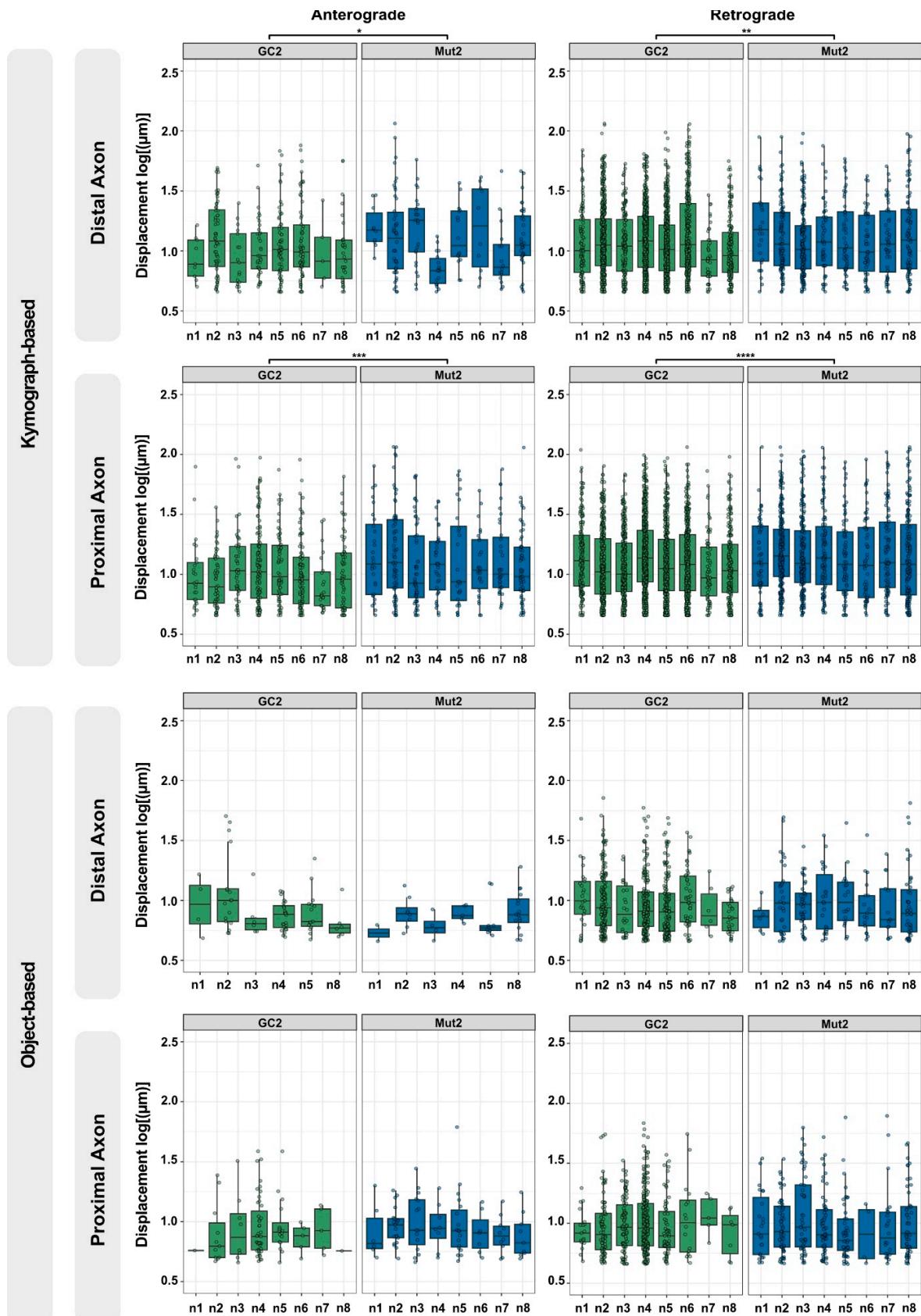


Figure S7: LRRK2 G2019S is not associated with a consistent change in the displacement of lysosomal trafficking in Mut2 neurons.

Quantification of displacement of anterograde and retrograde lysosomes in the distal and proximal axons of GC2 and Mut2 neurons by kymograph-based analysis (top) and object-based analysis (bottom). Statistics were calculated using 2-way ANOVA with post-hoc Tukey test to compare GC and Mut, taking into consideration any variation between the experimental replicates (n). **** corresponds to p-adj <= 0.0001, *** to p-adj <= 0.001, ** to p-adj <= 0.01, and * to p-adj <= 0.05. Comparisons without a marked * did not report any significant differences. The exact mean difference and p-values can be found in Table S1.

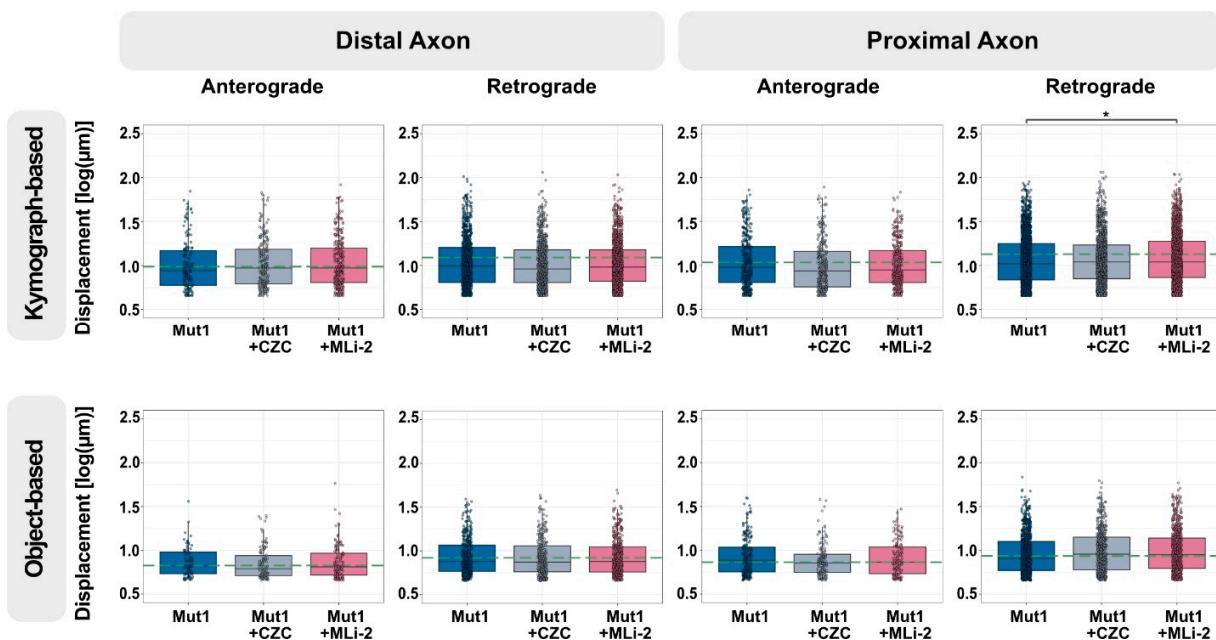


Figure S8: LRRK2 kinase inhibitors do not significantly affect the displacement of retrograde lysosomes in the distal or proximal axon.

Quantification of displacement of anterograde and retrograde lysosomes in the distal and proximal axons of Mut1 neurons without and with 2 μ M LRRK2 kinase inhibitors CZC-25146 or MLi-2 for 48 hours by kymograph-based analysis (top) and object-based analysis (bottom). The green dashed line marks the median lysosomal displacement of GC1 in the specific direction and position. For all graphs: Pooled individual measurements from $N =$ at least 6 independent experimental replicates. Statistics were calculated using 2-way ANOVA with post-hoc Tukey test to compare the effect of compound treatment, taking into consideration any variation between the experimental replicates. * corresponds to $p\text{-adj} \leq 0.05$. Comparisons without a marked * were not significantly different than Mut1. The exact mean difference and p -values can be found in Table S1.

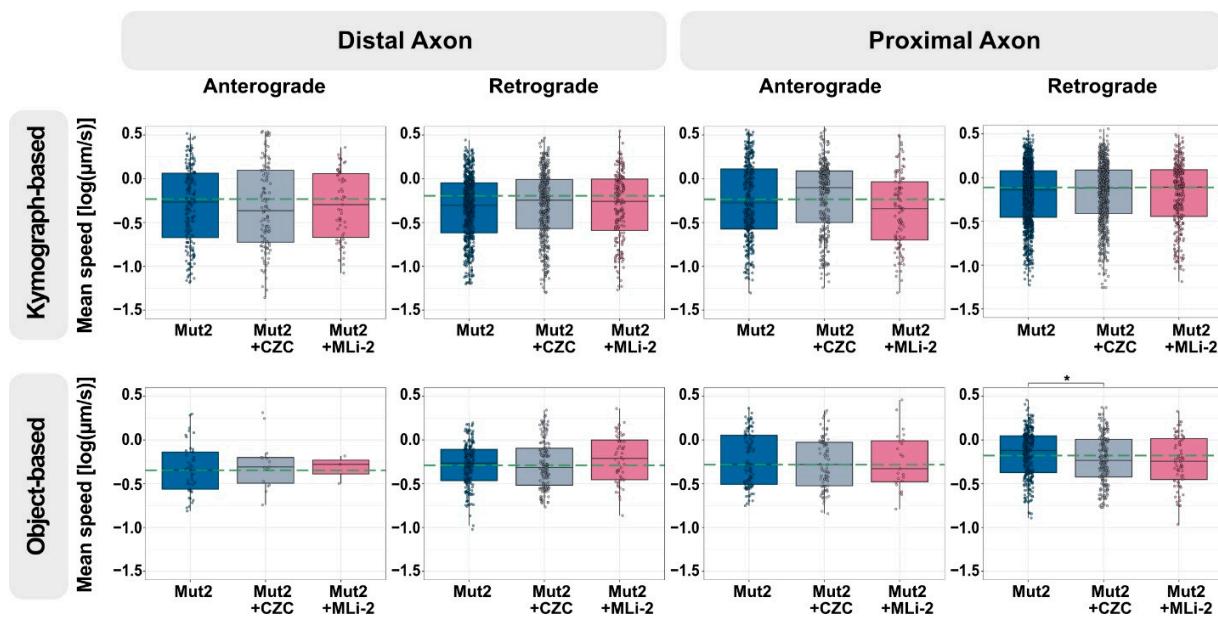


Figure S9: LRRK2 inhibitor treatment does not affect the speed of Mut2 axonal lysosomes.

Quantification of the mean speed of anterograde and retrograde lysosomes in the distal and proximal axons of Mut2 neurons without and with 2 μ M LRRK2 kinase inhibitors Czc-25146 or MLi-2 for 48 hours by kymograph-based analysis (top) and object-based analysis (bottom). The green dashed line marks the median lysosomal speed of GC2 in the specific direction and position. For all graphs: Pooled individual measurements from $N =$ at least 6 independent experimental replicates. Statistics were calculated using 2-way ANOVA with post-hoc Tukey test to compute the effect of compound treatment on comparison with no treatment, taking into consideration any variation between the experimental replicates. * corresponds to $p\text{-adj} \leq 0.05$. Comparisons without a marked * were not significantly different than Mut2. The exact mean difference and p-values can be found in Table S1.

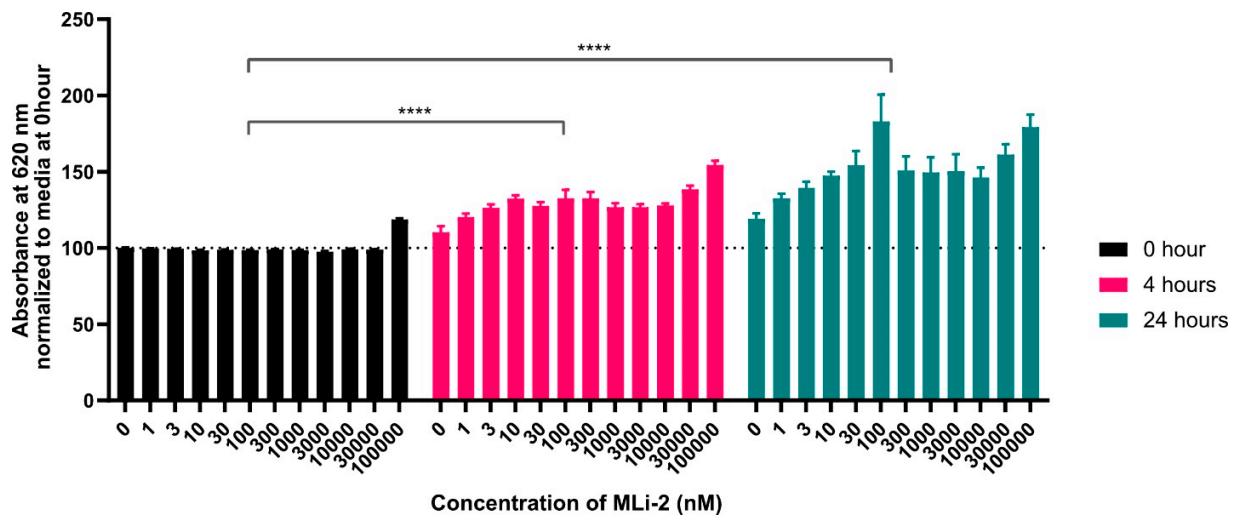


Figure S10: Precipitation of MLi-2 in media incubated at 37°C over time.

MLi-2 diluted in maturation medium over a range of concentrations was incubated at 37°C for 24 hours. Turbidimetric solubility assay measuring the absorbance at 620 nm at 0, 4, and 24 hours revealed the onset of precipitation and lack of solubility of MLi-2 over time based on the increase in absorbance value with MLi-2 relative to that of only media control at the start of the experiment. Statistics were calculated using 2-way ANOVA with post-hoc Tukey's multiple comparisons test. **** corresponds to p-adj<= 0.0001.

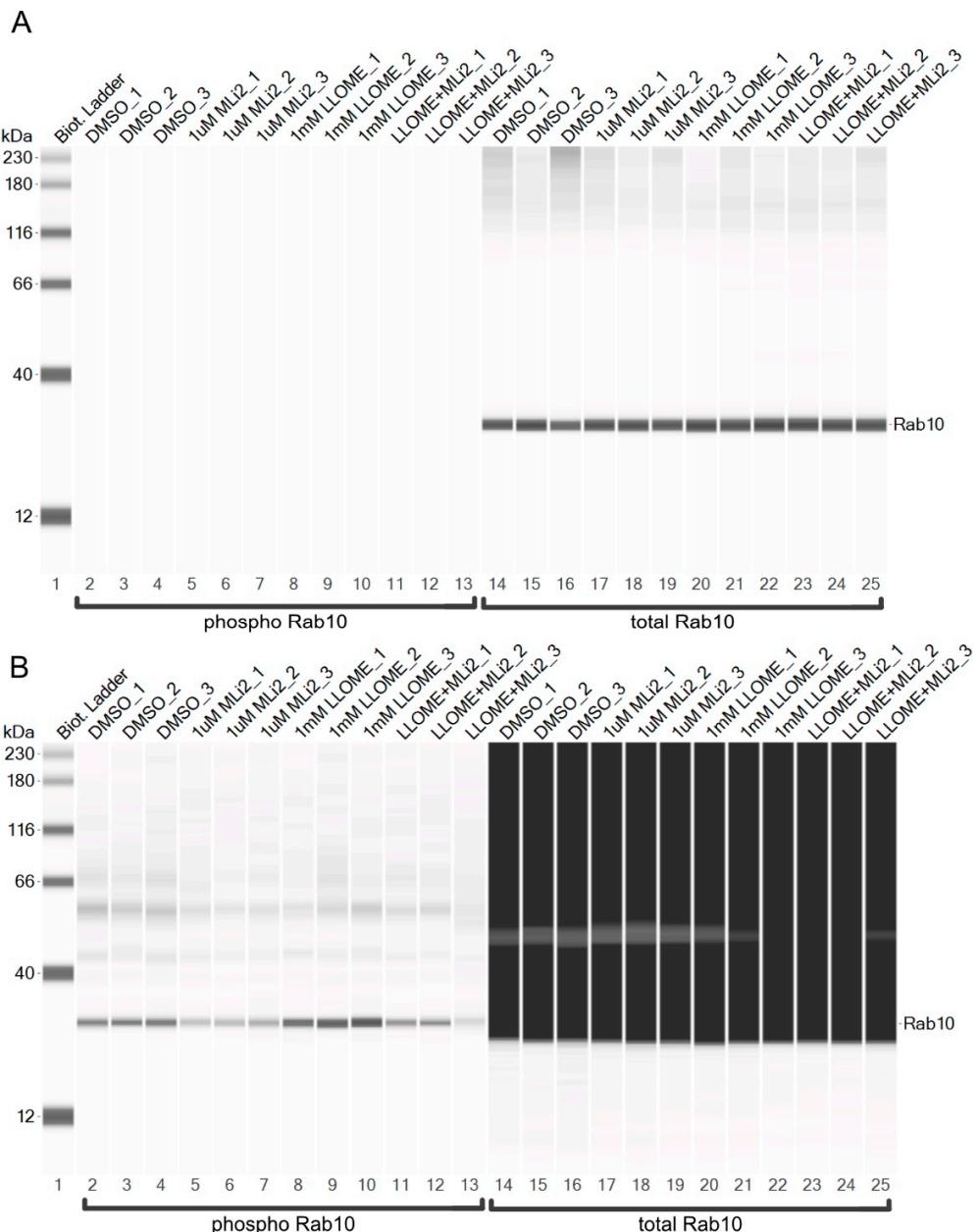


Figure S11: Uncropped WES blots of phospho-Rab10 and total Rab10 levels in LLOME and MLi-2 treated Mut1 neurons.

Uncropped Wes lane view (from Figure 5) measuring levels of phospho Rab10 and total Rab10 from whole cell lysates of DMSO, MLi-2, LLOME and MLi-2+LLOME treated Mut1 neurons (3 technical replicates for each condition). **(A)** Exposed Lanes 14-25 detecting the 30kDa total Rab10. **(B)** Adjusted contrast to detect phospho-Rab10 in Lanes 2-13.

| Figure No. | Position | Direction | Parameter | Analysis method | Condition 1 | | Condition 2 | Mean difference | p-adj-value | 95% confidence interval | |
|------------|----------|-------------|------------------------|-----------------|-------------|------------|--------------|-----------------|-------------|-------------------------|-------------|
| | | | | | Mut1 | Mut1 | | | | Lower bound | Upper bound |
| 2A | Distal | Retrograde | Mean speed [log(um/s)] | Kymograph-based | GC1 | Mut1 | 0.118901234 | 1.4E-11 | 0.085709452 | 0.152093017 | |
| 2A | Distal | Retrograde | Mean speed [log(um/s)] | Object-based | GC1 | Mut1 | 0.107607599 | 1.69E-08 | 0.070534005 | 0.144681133 | |
| 2A | Proximal | Anterograde | Mean speed [log(um/s)] | Object-based | GC1 | Mut1 | 0.141639852 | 3.25E-07 | 0.0880848 | 0.195194904 | |
| 2A | Proximal | Retrograde | Mean speed [log(um/s)] | Kymograph-based | GC1 | Mut1 | 0.072461235 | 1.45E-07 | 0.045506107 | 0.099416362 | |
| 2A | Proximal | Retrograde | Mean speed [log(um/s)] | Object-based | GC1 | Mut1 | 0.138734883 | 0 | 0.110857264 | 0.166612503 | |
| 2B | Distal | Retrograde | Mean speed [log(um/s)] | Kymograph-based | GC2 | Mut2 | 0.074955852 | 1.18692E-06 | 0.044754377 | 0.105117387 | |
| 2B | Proximal | Retrograde | Mean speed [log(um/s)] | Object-based | GC2 | Mut2 | -0.046588629 | 0.014573529 | -0.08394361 | -0.00923365 | |
| S4A | Distal | Retrograde | Displacement [log(um)] | Kymograph-based | GC1 | Mut1 | 0.088654611 | 4.38E-11 | 0.062606388 | 0.114702834 | |
| S4A | Distal | Retrograde | Displacement [log(um)] | Object-based | GC1 | Mut1 | 0.040756978 | 0.009466613 | 0.009997132 | 0.071516823 | |
| S4A | Proximal | Retrograde | Displacement [log(um)] | Kymograph-based | GC1 | Mut1 | 0.074596169 | 0 | 0.052226137 | 0.096966201 | |
| S4B | Distal | Anterograde | Displacement [log(um)] | Kymograph-based | GC2 | Mut2 | -0.056455741 | 0.037228959 | -0.10957542 | -0.00335606 | |
| S4B | Distal | Retrograde | Displacement [log(um)] | Kymograph-based | GC2 | Mut2 | -0.035949562 | 0.007478874 | -0.06228446 | -0.00961467 | |
| S4B | Proximal | Anterograde | Displacement [log(um)] | Kymograph-based | GC2 | Mut2 | -0.083751062 | 0.000151062 | -0.12691869 | -0.04057164 | |
| S4B | Proximal | Retrograde | Displacement [log(um)] | Kymograph-based | GC2 | Mut2 | -0.065969888 | 1.93E-08 | -0.08891582 | -0.04302396 | |
| 3 | Distal | Retrograde | Mean speed [log(um/s)] | Kymograph-based | Mut1 | Mut1+MLL-2 | -0.039556723 | 0.032508707 | -0.07651532 | -0.00259813 | |
| 3 | Proximal | Retrograde | Mean speed [log(um/s)] | Kymograph-based | Mut1 | Mut1+MLL-2 | -0.046533139 | 0.000138643 | -0.07330616 | -0.01976012 | |
| 3 | Proximal | Retrograde | Mean speed [log(um/s)] | Object-based | Mut1 | Mut1+MLL-2 | -0.073381438 | 5.51E-08 | -0.10384618 | -0.0429167 | |
| S5 | Proximal | Retrograde | Displacement [log(um)] | Kymograph-based | Mut1 | Mut1+MLL-2 | -0.023244226 | 0.036452445 | -0.04533914 | -0.00114931 | |
| S6 | Proximal | Retrograde | Mean speed [log(um/s)] | Object-based | Mut2 | Mut2+ZC | 0.066739437 | 0.022940438 | 0.00740384 | 0.126075033 | |
| 3 | Proximal | Retrograde | Mean speed [log(um/s)] | Kymograph-based | GC1 | Mut1+ZC | 0.080833407 | 2.71E-07 | 0.042904559 | 0.118763581 | |
| 3 | Proximal | Retrograde | Mean speed [log(um/s)] | Object-based | GC1 | Mut1+ZC | 0.122714657 | 0 | 0.082222893 | 0.163206421 | |
| 3 | Proximal | Retrograde | Mean speed [log(um/s)] | Kymograph-based | GC1 | Mut1+MLL-2 | 0.033603105 | 0.070854053 | -0.0018631 | 0.069074517 | |
| 3 | Proximal | Retrograde | Mean speed [log(um/s)] | Object-based | GC1 | Mut1+MLL-2 | 0.085325138 | 4.28E-08 | 0.047540621 | 0.123109655 | |

Table S1: Summary statistics and p-adj-values for axonal trafficking data

Mean difference, adjusted p-values, lower and upper bound of the confidence interval for the mean, resulting from 2-way ANOVA and Tukey's multiple comparisons test. Only comparisons reported as statistically significant in each of the listed conditions are specified.

| Pat ID | APOE Haplotype | MAPT Haplotype (H2=protective, H1=risk) | SNCA rs356220 or proxy rs356219 (C=protective, T=risk) | Other genes |
|---------------|-----------------------|--|---|------------------------------|
| Patient1 | E3/E3 | H1/H2 | C C | GBA, PRKN, PINK1 negative |
| Patient2 | E2/E3 | H1/H1 | T T | GBA, PRKN, PINK1 negative |

Table S2: Genetic information of the two LRRK2 G2019S-PD patient donors.

Genetic characteristics of the two LRRK2 G2019S-PD patients for Alzheimer APOE haplotype, and haplotype of frequent single nucleotide polymorphisms in PD risk factors MAPT and SNCA.

| Pat ID | Gender | Birth year | Age at onset | Age at examination | Disease duration | LEDD | Hoechst & Yahr Stadiu m | UPDRS III Score | MoCA A-Total value | CSF Abeta4 2 pg/ml | CSF h-Tau pg/ml | CSF p-Tau 181 pg/ml | CSF NFL pg/ml | CSF alpha synucle in total pg/ml | Number of positive seedings in 30h | RT-QuIC seeding result |
|----------|--------|------------|--------------|--------------------|------------------|------|-------------------------|-----------------|--------------------|--------------------|-----------------|---------------------|---------------|----------------------------------|------------------------------------|------------------------|
| Patient1 | F | 1931 | 70 | 75 | 5 | 2.5 | | | | | | | | | | |
| Patient1 | F | 1931 | 70 | 76 | 6 | 3 | | | | | | | | | | |
| Patient1 | F | 1931 | 70 | 77 | 7 | 120 | 2.5 | 41 | 547 | 421 | 53 | 1935 | | | | |
| Patient1 | F | 1931 | 70 | 78 | 8 | 380 | 3 | 60 | | | | | | | | |
| Patient1 | F | 1931 | 70 | 80 | 10 | 580 | 4 | 44 | 19 | 404 | 542 | 67 | | | | |
| Patient1 | F | 1931 | 70 | 80 | 10 | 760 | 3 | 56 | 24 | 450 | 556 | 81 | | | | |
| Patient1 | F | 1931 | 70 | 81 | 11 | | 3 | | | | | | | | | |
| Patient1 | F | 1931 | 70 | 83 | 13 | 675 | 3 | 63 | | | | | | | | |
| Patient1 | F | 1931 | 70 | 83 | 13 | | 3 | | 12 | | | | | | | |
| Patient2 | F | 1958 | 40 | 50 | 10 | 940 | 2 | 13 | 1003 | 144 | 31 | | | | | |
| Patient2 | F | 1958 | 40 | 52 | 12 | 1148 | | 25 | 28 | 819 | 148 | 26 | | | | |
| Patient2 | F | 1958 | 40 | 53 | 13 | 1148 | 2 | 21 | 27 | 923 | 237 | 35 | 794 | 444.25 | 3 | |
| Patient2 | F | 1958 | 40 | 54 | 14 | 894 | 2 | 24 | 26 | 794 | 200 | 32 | 830 | 267.15 | 4 | |
| Patient2 | F | 1958 | 40 | 54 | 14 | | 2 | | | | | | | | | |
| Patient2 | F | 1958 | 40 | 55 | 15 | 1290 | 2 | 11 | 26 | | | | | | | |
| Patient2 | F | 1958 | 40 | 64 | 24 | 1237 | 2 | 11 | 28 | | | | | | | |

Table S3: Demographic, clinical, and CSF biomarker information of the two LRRK2 G2019S-PD patient donors.

Characteristics of the two LRRK2 G2019S-PD patients for demographic, clinical, and CSF data. LEDD = Levodopa Equivalent Dose, UPDRS = Unified Parkinson's Disease Rating Scale, MoCA = Montreal Cognitive Assessment, NFL = neurofilament, RT-QuIC = real-time quaking-induced conversion.