







## Review

# Plant–Fungi Interactions: Where It Goes?

A. K. Hasith Priyashantha <sup>1,†</sup> , Dong-Qin Dai <sup>1,†</sup> , Darbhe J. Bhat <sup>2,3</sup>, Steven L. Stephenson <sup>4</sup>,  
Itthayakorn Promputtha <sup>5</sup> , Prashant Kaushik <sup>6</sup> , Saowaluck Tibpromma <sup>1,\*</sup>   
and Samantha C. Karunarathna <sup>1,7,\*</sup> 

- <sup>1</sup> Center for Yunnan Plateau Biological Resources Protection and Utilization, College of Biological Resource and Food Engineering, Qujing Normal University, Qujing 655011, China; priyashanthahasith@gmail.com (A.K.H.P.); cidaidongqin@gmail.com (D.-Q.D.)
  - <sup>2</sup> Department of Botany and Microbiology, College of Science, King Saud University, P.O. Box 2455, Riyadh 11451, Saudi Arabia; bhatdj@gmail.com
  - <sup>3</sup> Biology Division, Vishnugupta Vishwavidyapeetam, Gokarna 581326, India
  - <sup>4</sup> Department of Biological Sciences, University of Arkansas, Fayetteville, AR 72701, USA; slsteph@uark.edu
  - <sup>5</sup> Department of Biology, Faculty of Science, Chiang Mai University, Chiang Mai 50200, Thailand; itthayakorn.p@cmu.ac.th
  - <sup>6</sup> Instituto de Conservación y Mejora de la Agrodiversidad Valenciana, Universitat Politècnica de València, 46022 Valencia, Spain; prakau@alumni.upv.es
  - <sup>7</sup> National Institute of Fundamental Studies (NIFS), Hantana Road, Kandy 20000, Sri Lanka
- \* Correspondence: saowaluckfai@gmail.com (S.T.); samanthakarunarathna@gmail.com (S.C.K.)  
† These authors contributed equally to this work.

**Simple Summary:** Fungi can form several different types of associations such as pathogenic and symbiotic with plants. Pathogenic fungi subject plants to a great deal of pressure by causing various diseases. Unlike animals, plants cannot escape from biotic and abiotic threats by moving from one place to another, but they survive such alarming conditions in some wonderful ways. In turn, plants enjoy the interactions with symbiotic fungi as they offer many benefits to the plants. Changes in micro- and macro-climates lead to modifying the interactions between plants and fungi, either positively or negatively. In this paper, we discuss all of those interactions and their relevance to better agricultural practices.



**Citation:** Priyashantha, A.K.H.; Dai, D.-Q.; Bhat, D.J.; Stephenson, S.L.; Promputtha, I.; Kaushik, P.; Tibpromma, S.; Karunarathna, S.C. Plant–Fungi Interactions: Where It Goes? *Biology* **2023**, *12*, 809. <https://doi.org/10.3390/biology12060809>

Academic Editor: James White

Received: 7 April 2023

Revised: 29 May 2023

Accepted: 30 May 2023

Published: 2 June 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

**Abstract:** Fungi live different lifestyles—including pathogenic and symbiotic—by interacting with living plants. Recently, there has been a substantial increase in the study of phytopathogenic fungi and their interactions with plants. Symbiotic relationships with plants appear to be lagging behind, although progressive. Phytopathogenic fungi cause diseases in plants and put pressure on survival. Plants fight back against such pathogens through complicated self-defense mechanisms. However, phytopathogenic fungi develop virulent responses to overcome plant defense reactions, thus continuing their deteriorative impacts. Symbiotic relationships positively influence both plants and fungi. More interestingly, they also help plants protect themselves from pathogens. In light of the nonstop discovery of novel fungi and their strains, it is imperative to pay more attention to plant–fungi interactions. Both plants and fungi are responsive to environmental changes, therefore construction of their interaction effects has emerged as a new field of study. In this review, we first attempt to highlight the evolutionary aspect of plant–fungi interactions, then the mechanism of plants to avoid the negative impact of pathogenic fungi, and fungal strategies to overcome the plant defensive responses once they have been invaded, and finally the changes of such interactions under the different environmental conditions.

**Keywords:** ascomycota; phytopathogenic fungi; plant defense mechanism; symbiotic fungi

## 1. Introduction

Multiple and intricate interactions between plants and fungi do exist in nature [1–3]. Plants are always prone to interact with various microbes in different ways which include phytopathogenic and symbiotic associations. In phytopathogenic associations fungi interact with different lifestyles, namely necrotrophic (e.g., *Alternaria alternata*, *A. solani*, *A. brassicae*, *Aspergillus flavus*, *Bipolaris sorokiniana*, *Botrytis cinerea*, *Claviceps gigantea*, *Colletotrichum beeveri*, *C. gloeosporioides*, *C. graminicola*, *C. musae*, *Sclerotinia sclerotiorum*, *Stenocarpella maydis*, *Zymoseptoria tritici*) [4–16]; biotrophic (e.g., *Blumeria graminis*, *Cladosporium fulvum*, *Hemileia vastatrix*, *Melampsora lini*, *Phakopsora pachyrhizi*, *Puccinia arachidis*, *Puccinia graminis*, *Puccinia kuehnii*, *Puccinia striiformis*, *Sporisorium scitamineum*, *Ustilago maydis*) [17–28]; and hemibiotrophic (e.g., *Colletotrichum higginsianum*, *C. trifolii*, *Fusarium equiseti*, *F. oxysporum*, *F. sacchari*, *Ganoderma boninense*, *Magnaporthe oryzae*, *Phomopsis longicolla*) [29–36]. A plethora of fungi also live as symbiotic, e.g., *Funnelformis mosseae*, *Glomus albidum*, *G. etunicatum*, *G. mosseae*, *G. fasciculatum*, *Glomus albidum*, *G. etunicatum*, *G. mosseae*, *G. fasciculatum*, *Glomus mosseae*, *Trichoderma virens* [37–43].

These interactions may significantly impact agriculture, the environment and, ultimately, the economy [44]. Interactions between plants and fungi can be highly effective in shaping plant community composition and diversity because they can affect the plants directly as well as indirectly by affecting competition and facilitation [45].

### 1.1. Overview of Phytopathogenic Fungi

The majority of fungal phytopathogens belong to the phyla Ascomycota and Basidiomycota. Plant pathogens are classified in a number of classes among Ascomycetes, such as the Dothideomycetes (including *Cladosporium* spp.), Sordariomycetes (including *Magnaporthe* spp.), or the Leotiomycetes (e.g., *Botrytis* spp.). Rusts (Pucciniomycetes) and smuts (spread among the subphylum of Ustilaginomycotina), the two major plant pathogen groups, are members of the Basidiomycetes [46].

Pathogenic fungi interact negatively with plants, can infect all parts of the plant and even kill them, and are responsible for causing ecologically and commercially significant plant diseases [5,6]. One of the most striking examples is *Botrytis cinerea*, which can infect and cause disease in more than 1000 species of plants, including many fruits, flowers, and leafy vegetable crops [10]. It has also been reported that the economic losses due to *B. cinerea* exceed USD 10 billion worldwide annually [9]. Further, in Sichuan Province, China disease caused by pathogens (e.g., *Blumeria graminis*) has resulted in wheat yield losses of 5 to 8% in general, and in an even more severe stage it goes up to 100% [19]. As aforementioned, plant pathogens are frequently grouped according to their lifestyle with their host plants. Necrotrophs infect living plants and destroy infected tissues as soon after they invade. The necrotrophs obtain nutrition from hosts, leading to the death of the affected organ or the entire plant [47]. Biotrophs infect living plants by weakening the plant's immune system and obtaining nutrients from the living cells. Hemibiotrophs first establish infection in living plant tissues, similar to biotrophs, and then uptake the nutrients from a combination of feeding from living and killed host cells [47,48]. Plants defend against those devastating pathogens through various mechanisms. Passive defense mechanisms are radially available against the pathogens, even before they contact the host. In these cases, physical barriers (e.g., cuticle, cell wall, stomatal aperture, and lenticels) and chemical barriers (e.g., pH, nutrient deprivation, and phytoanticipins) commonly work against the phytopathogenic fungi. Another type, the active defense mechanism (which we discuss in this article), is activated only after pathogen recognition and is mainly classified into rapid active defense reactions and delayed active defense reactions. Rapid active defense responses include the changes in membrane permeability (e.g., generation of reactive oxygen species or ROS, hydrogen peroxide or H<sub>2</sub>O<sub>2</sub>); hypersensitive cell death (HR); a fast, localized kind of programmed cell death; and cell wall fortification (e.g., papilla/callose deposition). Delayed active defense responses include wound repair (e.g., layers of cork cell formation),

expression of pathogenesis-related (PR) proteins (e.g., accumulation of chitinase), and systemic acquired resistance (e.g., salicylic acid (SA) biosynthesis) [49].

Compatible interactions occur when a plant fails to coordinate effective defense responses, enabling the disease to establish and spread throughout the host. On the other hand, an incompatible interaction occurs when the host's active defense mechanisms effectively stop the spread of the pathogen within the tissues of the host [50]. However, pathogens do not always create an interaction with the host. In fact, germination of a fungal spore can occur even in the absence of a host. The hypha may not penetrate the host tissues due to the passive defense mechanism of the host and ultimately die off. Thus, the fungus fails to establish a pathogenic relationship [51,52].

### 1.2. Overview of Symbiotic Fungi

In general, symbiosis refers to any type of close and long-term interaction between different organisms. Fungi live symbiotically, in general. They may interact with the plant, by either commensalism (one organism benefits from it and the other one shows no apparent beneficial or harmful effect) or mutualism (both gain reciprocal benefits and are usually mutual). In addition, some symbiotic fungi also can create harmful effects on the plant under certain conditions at some stage of their lifecycle, thus they are called pathogenic [3].

Among the symbionts, mycorrhizal glomeralean fungi are one of the most studied groups. They can be divided into several different relationship types at the coarsest level, including arbuscular mycorrhizal (AM), ectomycorrhizal (EcM), and ericoid mycorrhizal (ErM) [53]. Amongst them, AM fungi, belong to Glomeromycota and are the most prevalent endomycorrhizal (e.g., intracellular) mutualists with most vascular plants. They are also the earliest endomycorrhizal fungi to date and are found to be associated with about 85% of living plant species [54]. At the same time, EcM fungi are a huge group with a wide distribution, although they are only connected with 3–4% of the vascular plant families. Chiefly, they are members of the phyla Ascomycota and Basidiomycota [55]. However, ErM fungi are associated with a restricted diversity of plant species in the Ericaceae, Epacridaceae, and Empetraceae. In addition, they belong to a restricted group of fungi of the Ascomycetes [56]. In contrast to phytopathogens, symbiont interaction gives significant benefits to plants. Symbiont fungi enhance the structure and aggregation of the soil, which in turn influences the organization of plant communities and production.

Symbiotic fungi colonize the rhizosphere region in soil or the plant's internal tissues, usually take carbon (C) from the host plant and return to the plants with essential soil elements, and also improve the water and nutrition uptake by the plant [57,58]. Especially, some ErM and EcM secrete protease and phosphatases that access organic nitrogen (N) and phosphorus (P) to plants. In addition, some of them can also produce plant cell wall degrading enzymes, facilitating access to more organic N and P [59]. Moreover, they improve the N fixation, P solubilization, sulfur (S) oxidization, plant hormone production, or decomposition of organic compounds and further act as biofertilizers (e.g., *Alternaria* spp., *Aspergillus* spp., *Chaetomium* spp., *Fusarium* spp., *Penicillium* spp., *Serendipita* spp., *Phoma* spp., and *Trichoderma* spp.) [2,58]. In addition to those, symbiosis helps plants to retaliate against the biotic (e.g., parasites) and abiotic (e.g., drought, salinity, toxic compounds, and flooding) [54,57] stresses. Noteworthy that in addition to those symbionts, endophytic fungi also offer the aforementioned same benefits, while entomopathogenic fungi protect plants from insect pest attacks and also stimulate the plant defense responses [60–62]. However, unfortunately, in some cases, symbionts can also increase the activity of other plant pathogens, such as viruses (e.g., Potato virus), thus increasing disease susceptibility and reducing plant height and root development [41].

Accounting all, it is clear that their significance particularly in the agriculture sector, is not only because of their influence on improving farm productivity [63], but also because of their ability to minimize the application of agrochemicals. This leads to multi-dimensional

effects, including the degradation of the environment, health complications, and development of pest-resistance [64,65], therefore direct to sustainable agro farming [66–68].

In this article, we attempted to give an overall picture of the plant–fungi interaction, particularly addressing the phytopathogenic and symbiotic fungi. In order to acquire the relevant information, an extensive literature survey was also conducted by referring to the most recent studies wherever possible. Thus, the paper is worthy of up-to-date information. First, we provided the background data (other than in the introduction) that support the main content of this article, especially by sharing the evolutionary aspect of plant–fungi interactions. Second, we elaborated on how plants defend against pathogenic fungi and how fungi try to overcome those defense responses to create beneficial interactions.

According to the literature, the alteration of plant–fungi interaction under the changing environmental condition appears to be an emerging work of study. In addition, not enough comprehensive analyses have been completed in this regard. Therefore, this paper, most importantly, discusses how the broad group of fungi may change their interaction with plants due to the variation of environmental conditions, also bringing the most recent advancement in the field.

## 2. The Evolutionary Aspect of Plant-Fungal Interaction

### 2.1. Phytopathogenic Fungi

It is thought that fungal–plant association evolved at least 450 or 460 million years ago, most probably with the symbiotic fungi [69,70]. Unlike symbiotic fungi, diversion of phytopathogenic fungi occurred (relatively) recently [71]. It is observed that phytopathogenic fungi are unevenly distributed (phylogenetically) throughout the fungal kingdom [72]. The antagonistic interaction between plants and their diseases produces co-evolutionary dynamics in which plants respond to recognize the pathogens, and pathogens evolve to escape plant defense mechanisms in natural environments. Phytopathogenic fungi show high rates of molecular evolution, and their capacity to cope and adapt to the new environment ensures their survival throughout history [73]. Van Valen [74] presented the ‘Red Queen hypothesis’ (RQH) emphasizing the primacy of biotic interactions over abiotic forces in driving evolution. According to the RQH, any adaptation made by one species is countered by adaptations made by another interacting species, so constant evolutionary change is required for survival. Thereafter, several additional explanations were made by the scientists to the RQH. Today, collective theories provide a more conceptual framework for the evolution of a suite of characteristics such as mating systems, pathogen virulence, host resistance, and the maintenance of population genetic diversity [75–77]. For instance, in mating systems, the majority of phytopathogenic fungi can reproduce sexually or asexually, exclusive sexual and asexual species are significantly minor. In the case of asexuality, it has been hypothesized to arise frequently from sexual fungal species, during the evolution time frame [78]. In the sexual reproduction system, one of the most important benefits is that it generates genetic variation among the progenies, which may allow the population to adapt faster to novel and/or stressful environments through rapid adaptation to genetic variation. In spite of this, adaptation to stressful environments or new environments can still be accomplished without the involvement of sex through mutations [79]. Moreover, beneficial natural mutations can be detached from the deleterious mutations in sexual populations and can be evolved as separate lineages, thus taking evolutionary advantage [80]. A recent study conducted by Meng et al. [4] indicated that *Alternaria alternata*, combining many cycles of asexual propagation with fewer cycles of sexual reproduction, thereby enabling it to adapt to changing environments. In turn, species such as *Zymoseptoria tritici* which have limited asexuality showcase a number of mutations at a single nucleotide position leading to a higher level of genetic diversity. Moreover, new phenotypes evolve in a highly selective environment (e.g., monoculture of host plant) where the trait is desirable. In that case, the proportions of new phenotypes in the population can rapidly increase [18].

## 2.2. Symbiotic Fungi

Symbiotic interaction between plant fungi is unavoidable as same as connecting with pathogens. It is also worth noting that, unlike mycorrhizae, most fungal endophytes are considered commensalistic and can have either positive (mutualists) or rarely negative (pathogens) interactions with host plants. According to fossil records and recent genetic analysis, it is clear that in both mycorrhizal and endophytes symbiosis likely played a role in the early colonization of the land by plants, serving as the evolutionary cornerstone of the current flora of land plants [54,59]. Therefore, the historical bond of symbiotic interaction of plants with mycorrhizal and endophytic fungi runs over 460 [69] and 400 million [81] years back, respectively.

In an evolutionary scenario for mycorrhizal symbiosis, members of Zygomycota had a significant role in the early phases of land plant diversification. Later on, Mucoromycotina was replaced with Glomeromycotina in most plant lineages. It is believed that Glomeromycotina has transitioned into various Ascomycota and Basidiomycota lineages, leading to the emergence of several new mycorrhizal syndromes, e.g., orchids [82]. More precisely, among the mycorrhizal groups, AM and EcM fungi show clear differences in their evolutionary origins. AM symbiosis in plants and fungi has a single origin, with subsequent losses and sporadic reversions back to AM in the seed plants. EcM symbiosis shows multiple, independent evolutionary origins in both plants and fungi [83]. Furthermore, accounting for the AM, owing to their relatively simple spores, lack of sexual reproduction, and association with a wide range of plants, they are considered primitive. In addition, they are mostly unable to colonize without plants [84]. According to fossil records, the origin of glomeralean fungi runs 55 to 60 million years ago. Surprisingly, those evolved before the vascular plants arose and association with non-vascular plants such as Bryophyta (e.g., Hornworts) was found in early ages. However, they might have evolved into saprobes or developed Geosiphon (a non-mycorrhizal ancestral member of the Glomales, that formed an endosymbiosis with cyanobacteria)-type symbioses [69]. This ancient AM relationship is still present in 80% of plant species, proving the value of this mutualism to both parties [85]. In brief, endophytic fungi have been divided into two major groups namely, clavicipitaceous and non-clavicipitaceous endophytes. Clavicipitaceous endophytes infect some grasses limited to cool regions, while non-clavicipitaceous are found to be associated with non-vascular plants, ferns and allies, conifers, and angiosperms. Endophytes are also restricted to the division of Ascomycota and Basidiomycota [86].

It is also important to know what the possible reasons for such an association could be. It is accepted that early plants having poor development of roots or lack of true roots make it difficult to absorb the necessary nutrition, thus putting pressure on their survival. Therefore, it is evident that the evolution of plants with symbiosis was primarily aimed at large-scale colonization of the land [59]. In turn, EcM fungi have evolved from saprophytic fungi. This understanding is supported due to their production of enzymes that have the potential to digest plant cell walls, though these generally occur at considerably lower levels than in saprophytic fungi [84]. It has also been recognized that these symbionts have evolved to actively suppress the host's defense response [87].

## 3. Plant–Fungal Interactions: Heaven or Hell

### 3.1. Plant Defense Mechanism in Plant–Fungi Interaction

In response to the pathogen attack, plants impose defense mechanisms to protect themselves. Switching to such a defense mechanism also has negative consequences for the plants as it can negatively influence plants' growth. Nonetheless, plants may survive as it is the primary need of such a mechanism. Gene-for-gene (GFG) identification of the pathogen frequently marks the start of the responses towards immunological responses, which are triggered by the identification of pathogen-associated molecular patterns (PAMPs) or microbe-associated molecular patterns (MAMPs), by plant pattern recognition receptors (PRRs) localized in the plasma membrane and are mainly found in the forms of receptor-like protein kinases and receptor-like proteins [88–90]. MAMPs molecules are crucial for the

fitness and survival of microbes and are conserved across species, giving plants an effective way to detect the contact of pathogens. In the case of fungal contact, plants secrete chitin-producing enzymes called chitinases to release chitin fragments (chitin oligomers) from the cell wall. Then, it serves as an elicitor in MAMP [91]. For example, Hawkins et al. [92] identified seven chitinase genes in maize (*Zea mays*) that had alleles associated with increased resistance against the *Aspergillus flavus* infection. PRRs trigger the immune response and are called PAMP-triggered immunity (PTI), which provides a shield against non-host pathogens and minimizes the disease caused by virulent pathogens [89]. Further addressing, activation of PRRs signaling causes accumulation of reactive oxygen intermediates; activation of ion channels; activation of specific, defense-related mitogen-activated protein kinase cascades; and extensive transcriptional reprogramming of the host [93]—which altogether leads to accumulation of antifungal compounds such as phenols and phenolic glycosides, unsaturated lactones, sulfur compounds, saponins, cyanogenic glycosides, and glucosinolates [94]. R proteins have two conserved features, nucleotide-binding (NB) and leucine-rich repeat (LRR) domains, collectively referred to as NLRs. A wide range of NLRs is used by host plants to rapidly detect fungal effectors, which suppress the plant immunity responses' interaction with host proteins to modulate plant metabolism, either producing harmful secondary metabolites or proteins that kill the host plant during pathogen invasion. NLRs either directly or indirectly recognize the effectors, and this recognition frequently triggers an HR [90,93,95]. Moreover, HR often results through the activation of complex signal transduction pathways, case in point, ERK-like (extracellular-signal-regulated kinase) mitogen-activated protein kinases (MAPKs) [14]. It is widely recognized that the immune responses elicited by PRRs and NLRs are similar. Nevertheless, differences exist in the duration and amplitude of ETI responses, which are generally considered more significant than those of PTI (PAMP-triggered immunity) [90]. PTI and ETI extensively overlap, also signaling likely interactions [96].

The activation of plant defensive responses mediated by molecular mechanisms is enormously complex. In species, numerous biotrophic pathogens interact with their hosts in a framework of GFG that is traditionally described in which plant disease resistance (*R*) genes recognize the products of specific avirulent (*Avr*) genes in pathogens, resulting in disease resistance. For example, the first fungal avirulence gene—*Avr9* in *Cladosporium fulvum*—causes disease in tomatoes [97]. In these interactions, the disease is caused by a lack of either the *R* gene or the matching *Avr* gene [98,99]. An oxidative burst, or the rapid generation of ROS, typically occurs in conjunction with *R* gene-mediated resistance. Other than the host-induced ROS, some pathogens (e.g., *Alternaria brassicae*) produce those ROS during the compatible interaction at the hyphal tips [6]. Note that this production of ROS is also needed for HR response. The expression of several pathogenesis-related (PR) proteins, which are hypothesized to contribute to resistance, is linked to the activation of a signaling pathway dependent on salicylic acid (SA) in *R* gene-mediated resistance [100]. Other plant defensive responses are regulated by jasmonates (JAs), salicylic acid (Sa)- and/or ethylene-dependent systems (ET) [101]. It also needs to be highlighted that JAs represent the jasmonic acid (JA) and its derivatives, including its methyl ester (MeJA) and amino acid isoleucine conjugate (JA-Ile). JA improves plant disease tolerance through the JA signaling pathways in general, and aforementioned PAMPs are also associated with the JA signaling pathways [102]. JA and ET are typically linked to the defense against necrotrophic development. In contrast, the activation of defense mechanisms against biotrophic and hemibiotrophic diseases and the development of systemic acquired resistance are all facilitated by SA, which plays a crucial role in the plant defense system. Despite the fact that the SA and JA/ET defense pathways are antagonistic to one another, evidence of synergistic interactions has been found [101]. In a study, Tamaoki, et al. [103] attempted to understand the activation of the common defense mechanism of rice via JA and SA. According to them, SA signaling mainly contributes to the basal defense system in normal conditions due to the high endogenous SA concentrations. Nevertheless, sharp drops of endogenous SA are reported once the JA signal is switched on, and SA signaling

is found to be suppressed. Then, instead of SA, JA turns on the common defensive system. In contrast, Pena-Cortés et al. [104] found that SA blocks the biosynthesis of JA in tomato leaves. Thus, it is clear that the production of SA and JA is antagonistically inhibited. In a comprehensive study, Riemann et al. [105] show the involvement of JA derivative in rice defense response against the blast fungus, *Magnaporthe oryzae*. They have highlighted the role of exogenous JA, inducing the production of phytoalexins, which are important antimicrobial secondary metabolites that are produced in response to pathogen infection. Further, Riemann et al. [105] found that the JA-independent pathway worked against the *M. oryzae* induced accumulation of phytocassanes and partially for the accumulation of momilactones. Thus, it is conceivable that phytoalexin buildup is mediated by JA in the defense against blast fungal infection.

### 3.2. Fungi Overcome Plant Defense Mechanism in Plant–Fungi Interactions

Recent studies have shown that interactions between necrotrophs and their host plants are much more nuanced and intricate than previously thought [106]. For instance, when compared to biotrophs, some necrotrophs express effector proteins that are internalized by host cells and interact with the host in a GFG manner to cause disease, by suppressing or avoiding host basal defense or PAMP-triggered immunity (PTI) [107].

Conversely, the initial pathogenesis phase does not differ among the hemibiotrophic and biotrophic fungi (obligate), though various mechanisms are applied to take up the nutrients from the host. It is well understood that compared to the biotrophic and hemibiotrophic fungi, necrotrophs have broader host ranges and overcome the natural physical barriers of the plants (e.g., cell wall) through the secretion of toxins and extracellular cell wall degrading enzymes, which degrade a wide range of complex and cross-linked polysaccharides and glycoproteins [108].

In particular, *Alternaria alternata* produces the most prominent determinant of their virulence. The host-selective toxin (HST) called ACT (9,10-epoxy-8-hydroxy-9-methyl-decatrienoic acid) induces rapid electrolyte leakage from citrus cells. Furthermore, the release of cutinase, cellulose, and pectate lyase enzymes degrade the cells and form necrotic lesions [109]. Unlike necrotrophs, biotrophic fungi predominantly take up the necessary nutrients by creating invading structures of melanized appressoria, penetration hyphae, and infection hyphae to make a closer association with the host and to also avoid extreme damage within active plant cells [110]. In order to enter into leaf epidermal cells, some fungi (e.g., *Magnaporthe oryzae*) generate tremendous turgor pressure in the appressorium, and this pressure is used to puncture the host surface with a thin penetration peg, leading to the further development of the fungi inside the tissue [35].

As aforementioned, hemibiotrophs follow a short biotrophic phase and absorb the nutrients similarly once keeping the host cells alive. Next, they switch into the destructive necrotrophic development characterized by extended secondary hyphae that grow both intracellularly and intercellularly and cause the death of tissues [111,112]. In some cases, hemibiotrophs such as *Colletotrichum lindemuthianum* mutant H433 retain biotrophically and will not be moved to the necrotrophic phase [111]. Nonetheless, the difference between the necrotrophs and biotrophs during the initial infection process is quite interesting. As aforesaid, cell death is obvious at the primary stage of the pathogenic infection in both pathogenic types; however, cell death has remarkably different roles in plant responses to necrotrophs and biotrophs. While cell death caused by the necrotrophs is a sign of successful infection, in biotrophs, HR-associated cell death or the death of cells at the place of infection is actually a plant defense response where it stopped the further spread of pathogen hyphae, thereby limiting the nutrient intake to the pathogen which restricts their further development. In other words, necrotrophs aggressively encourage cell death by utilizing a variety of virulence factors, whereas biotrophs actively inhibit HR cell death [113]. Intriguingly, HR is also associated with the hemibiotrophs such as *Phytophthora infestans*, even though they show necrotrophic action in a later stage [114].

In the interest of endophytic fungi, many studies conducted to evaluate how they counteract the defense response of plants leading to inter and intracellular colonization. Endophytes are initially perceived as potential invaders. Like pathogens, they also have evolved sophisticated strategies to avoid recognition and elude plant immune systems. More precisely, endophytic fungi need to avoid either eliciting PTI or adapting to it or suppressing it to establish a compatible interaction that leads to proliferation [61,87,93]. For example, as previously highlighted, in plants, chitinases are effective immune molecules, as they break down chitin and weaken fungal cell walls. This type of detection by the plant immune system has to be prevented by the fungi, and they do so in two ways: (1) fungi could mask the chitin in their cell wall by covering it with other polymers or deacetylating it into chitosan and (2) they modify the elicitor-active chitin oligomers produced by the chitinases involved in the plant's immune response. These chitin oligomers may be inactivated by being bound, degraded, or deacetylated. In both cases, deacetylation appears to be the most likely inactivator of chitin since it is known that fully deacetylated chitosan oligomers do not bind to plant receptors and therefore do not induce an immune response [1]. On the other hand, fungi also must either inactivate toxic metabolites or secrete effectors to accomplish further survival in or inside the plant host. The effectors can also release proteins that act as a barrier for the fungus, reducing the host's immunological response, or altering the physiology of the host cell [93]. Some pathogens—for instance, *Sclerotinia sclerotiorum*—kill host plants via secretion of small, toxin, effector-like protein identified as SsSSVP1 and are expected to form disulfide bonds intra-molecularly [115]. As those effectors are proteins, they also export in fungi encompassing the signal peptide-mediated transfer through the endomembrane system. They first enter into the endoplasmic reticulum (ER), undergo proper folding, translocate the effectors to the Golgi apparatus, pack into secretory vesicles, fuse with the cytoplasmic membrane, and finally release into the extracellular space. This conventional endoplasmic reticulum–Golgi apparatus route is the most opted pathway to secrete [116]. In general, to secrete the effectors, contact should be there between the fungi and plants. Haustoria and hyphae are both secretion effectors. Primarily, localized release of those effectors can be observed at the interface between fungal pathogen and host plant, also associated with the penetration pore. This is important for hemibiotrophs since this stage leads to biotrophic changes to a necrotrophic lifestyle [29]. Those effectors then pass through the extra-haustorial matrix, extra-haustorial membrane for effectors secreted from haustoria, and apoplast, plant cell wall, and plant plasma membrane for effectors secreted from hyphae, then translocated into the host cell cytoplasm, although the followed mechanisms are still unclear [117].

One of the widespread effectors called necrosis-and ethylene-inducing-like proteins (NLPs) contributes to pathogen virulence through phytotoxic activity [118]. In this regard, Santhanam and co-workers [119] demonstrated the cytotoxic activity of NLP family members of a tomato-pathogenic *Verticillium dahliae* strain and found that two of the seven NLP-induced plant cell death. In *V. dahliae*, the genes encoding these cytotoxic NLP are found to be induced upon colonization of tomato. In addition to their role in virulence, Santhanam et al. [119] have found that one of the NLP genes also contributes to vegetative growth and conidiospore generation (asexual reproduction). In a similar study, Kombrink et al. [120] studied Chitin-binding lysin motif (LysM) effectors that contribute to the virulence of *V. dahliae*, which are causal agents of foliar diseases on various plants including *Arabidopsis*, tomato, and *Nicotiana benthamiana*. Most intriguingly, Kombrink and co-workers [120] found that the LysM effector binds chitin, has the ability to block immunological responses, and shields fungal hyphae from degradation by hydrolytic plant enzymes.

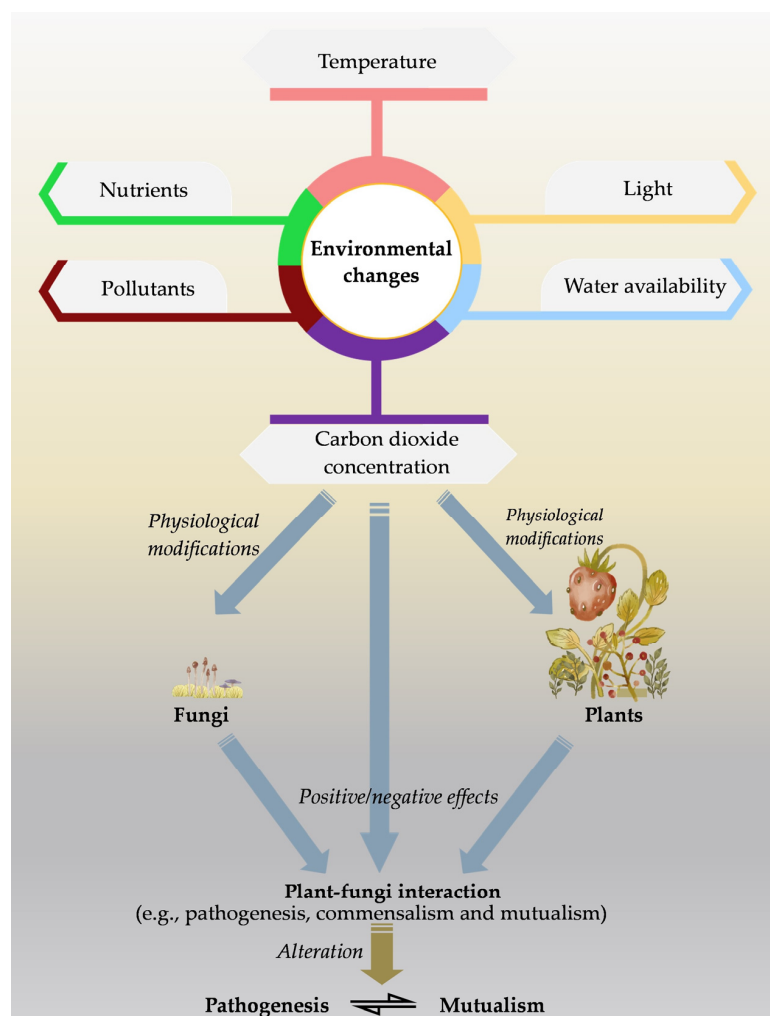
Another important consideration is that the fungi can also overcome the defense responses by inactivating previously mentioned endogenous JA through the biosynthesis of monooxygenase (Abm) to hydroxylate it, where such findings revealed studies along with *Magnaporthe oryzae*. More closely, Abm converts endogenous free JA into 12OH-JA. This 12OH-JA is released during pathogen-host penetration and helps to avoid a defense

response. Importantly, after the invasion, Abm itself is secreted and likely changes plant JA into 12OH-JA to aid host colonization [121].

#### 4. Plant–Fungal Interactions under the Changing Environmental Conditions

##### 4.1. Environmental Factors and Plant–Fungi Interaction

Under natural settings, many of the environmental conditions can change abruptly. Therefore, the response of fungi can also change, and subsequently could have significant ramifications for interaction between the plant fungi [122]. Changes in air temperature and moisture can affect fungal physiology and metabolism since they are most sensitive to these changes [123]. The majority of soil fungi are affected directly due to those environmental changes, although changes in plant physiology, morphology, immunological response, phenological traits, and root exudation impact the fungi indirectly (Figure 1) [124]. In other words, there is a direct impact of such conditions on the plant disease resistance mechanism as well as the virulence of the pathogens [125,126]. Beneficial microbes interact with plants, need to be functional under such unbefitting conditions and improve the plant tolerance to certain stress. However, there are possibilities for shifting those mutualistic interactions to non-convenient status [127,128]. There is also a possibility for the emergence of new pathogens as altering their virulence system, potentially leading to the downfall of *R* gene-mediated plant resistance. Such emergence has also been reported, e.g., *Puccinia striiformis* f.sp. *tritici* and *Fusarium graminearum* [124,129].



**Figure 1.** As a result of the changing environmental conditions, the physiology of plants and fungi can be affected, thereby altering the interaction between them.

#### 4.1.1. Temperature

Rising temperatures trigger a series of cellular processes and the production of heat shock proteins, which reduce plant cell damage. In contrast, heat stress worsens cell machinery and alters chromatin modifications in plants. It also increases membrane fluidity, which causes a reaction series to become uncoupled and disrupt metabolism, thus becoming more vulnerable to the pathogens [130]. On the other hand, there is a clear surfeit of consensus among researchers that higher and lower temperatures obviously affect fungal colonization and hyphal length. However, in most cases, the elevated temperatures would be beneficial to the fungi, particularly to mycorrhizal fungi. According to the literature, such temperature change has little positive effect towards the EcM fungi, but a strong positive effect on the AM fungi. One possible reason for this could be the faster plant C allocation to the rhizosphere where these fungi live [131,132].

More precisely, however, each plant–fungal interaction has a unique optimum temperature range. In this regard, a number of studies have assessed the mutualistic symbiosis between plants and the mycorrhizal fungal—phylum Glomeromycota. Of this, it was found that the fungal group is more prone to cold temperatures, and instead, a warming climate enhances its growth. For instance, Mathur et al. [133] observed that a mixture of *Glomus* species (e.g., *Rhizophagus irregularis*, *Funnelformis mosseae*) colonized about 75–80% in maize under the ambient condition, but reduced to 40–45% with higher temperature, which is about 43 °C. A comprehensive study conducted by Martin and Stutz [134] found that colonization of *Glomus intraradices* in pepper (*Capsicum annuum*) roots is higher in the 20.7–25.4 °C temperature range, while colonization is minimal with temperature ranges; 32.1–38 °C. In a supportive study, Liu and colleagues [135] revealed that lowering the temperature would lead to the failure of the colonization of *G. intraradices* wherein 15 °C would significantly reduce the mycelial development and further completely inhibit at 10 °C. In another study, de Vallavieille-Pope et al. [136] showed the highest infection ability of wheat pathogen *Puccinia striiformis* at 10 and 15 °C. However, inactivity occurred under the warmest (20 °C) and coldest (5 °C) temperatures. According to published research findings, even if the optimum temperature condition (thus reduced colonization) is not reached, the available fungi can give considerable benefits to the plants and allow them to tolerate critical environmental conditions [137–139].

Mathur et al. [133] experimented about the effect of high temperature ( $43 \pm 0.2$  °C) on maize and also the protective role of AM fungi, *Rhizophagus irregularis*, *Funnelformis mosseae*, and other *Glomus* species. This study's findings revealed that interactions with mycorrhizal fungi helped the plants to tolerate high temperatures while maintaining the stability of their photosynthetic apparatus—photosystem (PS) II and PSI. Furthermore, results showed that under normal conditions, mycorrhizal colonized plants had higher total chlorophyll ( $46 \pm 1$  SPAD Units) content than control plants ( $41 \pm 1$  SPAD Units). However, more elevated temperature stress plants and ultimately drop the chlorophyll contents ( $20 \pm 1$  SPAD Units), though mycorrhizal association helps the plant to recover from such conditions ( $37 \pm 2$  SPAD Units). In contrast, it is also worthwhile to mention that the heat stress turns endophytic *Botryosphaeria dothidea* into the pathogenic form, causing sudden disease severity conditions. In particular, this is common in Botryosphaeriaceae where the species of the family are recognized as stress-associated pathogens [140].

#### 4.1.2. Light

Ballhorn et al. [141] have inoculated the group microorganisms (*Glomus aggregatum*, *G. clarum*, *G. deserticola*, *G. etunicatum*, *G. monosporus*, *G. mosseae*, and *Gigaspora margarita*, *Paraglomus brasilianum*, *Rhizophagus irregularis*) with Lima bean (*Phaseolus lunatus*) in order to assess the impact of them under the full light and 50% shading conditions. After a 14-week trial, the researchers discovered that below-ground symbionts had boosting effects on growth and reproduction under full light. In contrast, infected plants under shaded conditions saw decreased plant growth and reproduction. This reduced plant growth is probably caused by the high C cost of the symbiosis in comparison to the available C and

the inability of plants to properly offset the fungal C demand in low light [122]. On the other hand, as mycorrhizal plants decline rapidly with decreasing light intensity, P uptake by roots becomes marginal, thus leading to poor growth and development of plants [122,142]. In a recent study, Garnica et al. [143] demonstrated the impact of light on the development of root endophyte *Serendipita herbamans* in knotweed (*Reynoutria* spp.). The researchers have noticed that approximately 20% light levels increase the colonization ability of pathogens, which is also significant; however, it resulted in lower biomass (decreased by 10%) of plants compared to the control treatment. Furthermore, results show that endophyte inoculation decreased chlorophyll content by 5% ( $p = 0.020$ ) under this condition.

In an interesting study, Hevia et al. [144] recognized the microbial biological clock-mediated plant-pathogenic fungi interaction for the first time. The researchers have used *Botrytis cinerea*, which infected *Arabidopsis thaliana* and disrupted the circadian oscillator of the fungi by providing light and dark cycling conditions. The fungal clock is the major factor influencing the outcome of the interaction between the *Arabidopsis* and *Botrytis* species. The researchers reported that the fungus may grow to its most virulent state even at daybreak, provided that its internal clock reads dusk time, thus fungal virulence potential can bypass a plant's natural defense mechanisms.

#### 4.1.3. Water Availability

Too little water (underwater deficit or osmotic stress) or too much water (during flooding) can greatly affect many aspects of plant and microbe biology. In general, lack of water (drought conditions) restricts mycelial growth and limits its ability to supply nutrients to plants. On the contrary, plants suppressed the symbiont fungi by limiting C flow to the roots, increasing saprotrophic fungi and slightly affecting the pathogenic fungi [145]. However, such a scenario depends on the type of host plant and the associated fungi, type of soil and the stage of the plant life cycle (e.g., seedling, young, matured) [146,147]. Augé [148] showed that under drought conditions, mycorrhizal fungi could give extra tolerance to plants, also because of the higher water uptake ability. Morte et al. [149] found that colonization of mycorrhizal fungi (associated with Aleppo pine—*Pinus halepensis*) was not affected by the lowering of the feasible water content. Moreover, they have recognized that mycorrhizal interactions help Aleppo pine to overcome water stress. In contrast, Boczoń et al. [150] further highlighted that the water shortage can shift the endophytic phase of *Cenangium ferruginosum* to phytopathogenic and saprotrophic lifestyles. This stressed environment activates *C. ferruginosum* and causes pine dieback disease as the plant reduces its resistance mechanism against the pathogen. Nevertheless, it is enthralling to know how mycorrhizal fungi take up the necessary water. According to Boczoń et al. [150] this could have happened because of the ability of mycorrhizal hyphae to explore small water pores in the soil, where plant roots are not accessible, thus improving plant water status under low water availability. Added to this explanation, Bennett and Classen [132] reported that mycorrhiza support the plant via improved apoplastic water flow facilitated water uptake through fungal water channels, increased stomatal conductance in host plants, and modified host gene expression of drought-related genes encoding plant aquaporins. Moreover, it has been recognized that symbiotic endophytic fungi help plants to survive this hostile condition by increasing sugar content in cells, which enhances the osmotic adjustment by limiting drought-induced damage to the host plant. In addition, it mitigates the buildup of drought stimulated destructive  $H_2O_2$  and saves plants from cellular damage [151].

In a separate study, Andreo-Jimenez et al. [152] found that under normal environmental conditions, instead of members of the Glomeromycota forming mycorrhizal associations, fungi belonging to the Zygomycota, Ascomycota and Basidiomycota were present in rice (*Oryza sativa*). However, along with drought, the composition of the endophytic fungal microbiota changes and increases the proportion of the Ascomycota and Basidiomycota. Moreover, Andreo-Jimenez et al. [152] examined the effect of the Ascomycota fungus—*Arthrinium phaeospermum* on rice growth and found a significant correlation between higher plant yield under drought conditions. Furthermore, the early work of Sten-

ström [153] opened a new window as he tested the impact of a water-logged environment on the fungal associations of five species with *Pinus sylvestris* seedlings. He found that *Suillus bovinus* and *S. flavidus* are highly sensitive to flooding and the reverse was true in *Hebeloma crustuliniforme*, *Laccaria laccata*, and *Thelephora terrestris*.

#### 4.1.4. CO<sub>2</sub> Concentration

The rise of CO<sub>2</sub> is a universal concern. The effect on fungi is uncertain since they are both for and against depending on the group of the microorganism—among them, symbionts are generally promoted by such conditions [154,155]. In the natural environment, the climbing of CO<sub>2</sub> (up to a certain level) enhances the photosynthesis of plants and improves C assimilation and allocation to roots. The root-associated microbiota favors this situation—AM fungi as they receive a higher amount of photosynthates before other soil microbes, thus higher the proliferation [156]. Nonetheless, in vitro assay conducted by Baazeem et al. [157] observed that an elevated CO<sub>2</sub> (also with little raised water supply) level speeds the colony growth of *Aspergillus flavus* as well as the metabolite activities. According to their findings, when exposed to CO<sub>2</sub> at 1000 ppm (37 °C), the fungi produced a substantial amount of the secondary metabolite—afatoxin B1. This made it clear that the microbe's metabolic activity had changed due to this unfamiliar condition, therefore the changing interaction between plants and fungi is obvious. A Meta-analysis conducted by Dong et al. [155] found increased mycorrhizal plant biomass (+26.20%), nutrient contents (N: +2.45%, P: 10.66%), and mycorrhizal fungal growth (extraradical hyphal length: +22.87%, mycorrhizal fungal biomass: +21.77%) due to the elevated CO<sub>2</sub>.

In a field experiment, Garcia et al. [158] attempted to understand the mycorrhizal dynamics under increased CO<sub>2</sub> (200 ppm) in a warm temperate (above 4 °C) condition. In more detail, they have shown the EcM root colonization increased significantly (by 14%) under increased CO<sub>2</sub>, whereas the length of the AM fungi hyphae and the stocks of its glomalin (an obstinate glycoprotein that persists in the soil even after the fungus has died) concentration did not significantly change in response to CO<sub>2</sub> enrichment, and the effects of CO<sub>2</sub> on AM fungi root colonization varied by date. For instance, compared to the ambient CO<sub>2</sub> treatment, glomalin concentrations in the elevated CO<sub>2</sub> treatment tended to be 6% higher in one month, while 29% lower in another. It is quite interesting to bring the possible effect of those to plants. In this regard, Matamala and Schlesinger [159] assessed the biomass of fine roots of pine (*Pinus taeda*) forests and found biomass enhancement by 87% under elevated CO<sub>2</sub>. More precisely, Brosi et al. [147] evaluated the effect of elevated CO<sub>2</sub> conditions on the symbiotic association between tall fescue (*Lolium arundinaceum*) and endophytic fungi *Neotyphodium coenophialum*. The researchers found that endophyte infection frequency changed under the elevated CO<sub>2</sub> condition, 81% (±3) during the ambient and 91% (±2) increased CO<sub>2</sub> level, thus promoting this grass–fungal symbiosis.

Contrarily, Váry et al. [160] conducted a study to evaluate wheat Fusarium head blight (FHB) and Septoria tritici blotch (STB) disease under the elevated CO<sub>2</sub>. The authors reported an increased level of pathogenicity of both the *Zymoseptoria tritici* and *Fusarium graminearum* elevated CO<sub>2</sub> (780 ppmv) condition compared to the optimum CO<sub>2</sub> conditions (390 ppmv). Further, pathogen and plant acclimation to elevated CO<sub>2</sub> leads to the rapid development of STB and FHB disease and its severity on the plant. It is also recognized that the overall pathogen acclimation to elevated CO<sub>2</sub> had a greater effect on FHB development than on STB disease.

#### 4.1.5. Pollutants

Changes in environmental health via pollutants become another threat to any sort of plant–fungal interaction, where many studies have been conducted to understand the effect of heavy metals. For instance, El-Shafey et al. [161] attempted to find out the response of Rose-scented geranium (*Pelargonium graveolens*) to Cadmium (Cd)-stress. Here, further, they evaluated the effect of three endophytic fungi namely *Talaromyces versatilis*, *Emericella nidulans*, and *Aspergillus niger* on geranium and also the potentiality of the endophytes to

alleviate in the changing environment. According to them, *T. versatilis* and *A. niger* had the most stimulating effects on fresh biomass of geranium leaves under normal conditions, while *E. nidulans* had the reverse impact. In this case, whereas *A. niger* only caused a non-significant increase of 14% in biomass, *T. versatilis* considerably increased biomass by 73% above the control. The biomass of non-inoculated geranium under Cd stress, however, significantly decreased to 58.4% of control. Because they considerably increased the biomass of geranium leaves to 312% and 182%, respectively, in comparison to the non-inoculated one during Cd-stress, *T. versatilis* and *A. niger*'s stimulatory impact was more evident under stressful circumstances. *E. nidulans*, on the other hand, inhibited leaf growth. Furthermore, El-Shafey et al. [161] highlighted the significant beneficial effect of the above fungi on alleviating the Cd-toxic effect of the plant by improving the resistance and enhancing the tissue quality.

Under this stressed environment, chiefly *T. versatilis* and *A. niger* stimulate the plant antioxidant enzymes, also upregulate the detoxification mechanisms of glutathione-S-transferase, phytochelatin, and metallothionein levels. In another study, Selim et al. [162] stressed the rye (*Secale cereale*) and sorghum (*Sorghum bicolor*) plants with Vanadium (V), and checked AM fungi (*Rhizophagus irregularis*) responses. Convincing data showed that interaction with *R. irregularis* places a beneficial effect by restricting the V intake. Further results by Selim et al. [162] noticed considerable changes in plants' minerals content in the roots and shoots compared to *R. irregularis* with and without V. Accumulation of V in shoots was reduced by mycorrhizal treatment to 29% and 58% in rye and sorghum, respectively. Roots of rye and sorghum show a similar, though the more pronounced reduction in vanadium accumulation (40% and 68% reduction, respectively). Moreover, in agreement with El-Shafey et al.'s [161] findings, Selim et al. [162] reported the mycorrhizal ability to restore the plant biomass that was reduced due to the V stress. For instance, the fresh weight of the shoots and roots of the sorghum plant is noticeably increased by 216% and 158%, respectively. In contrast, the researcher also noticed the growth of the shoots and roots in rye are insensitive to mycorrhizal availability.

#### 4.1.6. Nutrients

Deprivation of available soil nutrients also leads to a stressful environment for the plants; the priceless association with fungi may play a role in mitigating such effects on plants. In a study, Garnica and co-workers [143] recognized that under low-nutrient conditions, *Serendipita herbamans* strongly colonized knotweed and had positive effects on plant growth, where biomass increased by 15% and chlorophyll content increased by 13% ( $p = 0.006$ ). Mycorrhizae's role in promoting plant growth has been well-documented, markedly in conjunction with the increased uptake of P [163]. Patricia Guadarrama et al. [164] studied the effect of increasing P nutrition and mycorrhizal growth response of *Lotus corniculatus* and *L. glaber* in soil with little accessible P. Interestingly, the study revealed that there is no significant effect with the addition of P in plant shoot yield, as both mycorrhizal and non-mycorrhizal plants responded strongly to added P in soil. This shows that the interaction between fungi–plant may not be an advantage or disadvantage as a means of increased P availability. Similarly, Garcia et al. [158] noticed that N fertilization had no effect on the colonization of EcM roots though increased the colonization of AM fungi. More recent work by Garces et al. [165] found the influence of N enrichment on *Epichloë* colonization of the dune-building grass, *Ammophila breviligulata*. It is reported that the presence of *Epichloë* spp. in the host grass increases the species richness of root endophytes by 17%. The addition of N, however, exhibited no noticeable main or interaction influence on the richness of root endophytes. In contrast, N addition impacted the composition of the root endophyte population, primarily in areas where *Epichloë* spp. were prevalent.

#### 4.2. Environmental Factors and Lifestyle Switching of Fungi

Most of the fungi (possibly all) can express different lifestyles, e.g., *Colletotrichum magna* is pathogenic in a variety of cucurbits, a saprophyte in dead plant parts, and a

nonpathogenic endophyte in tomatoes. It is difficult to pinpoint the exact reason for this, but environmental changes could be one of the driving factors behind it [166]. The environmental factors on lifestyle switching are generally coupled and discussed with the symbiotic fungi, which often live with plants by creating mutualistic relationships; however, they may change their lifestyle into a parasitic one [167,168]. In a study, Delaye et al. [169] analyzed the lifestyle switching of endophytes, necrotrophs, and biotrophs. Accordingly, lifestyle switching of fungi is generally noticed as the conversion of endophyte to necrotroph and vice versa. Such changes are identified to happen at an equal frequency, however, endophytic to a pathogenic biotrophic lifestyle is rare. Further, once biotrophy has evolved, it is suggested that lifestyle change to another form is impossible to occur. There are clear findings of changing symbiotic endophytes into pathogens under various unfamiliar environmental conditions [170]. This can be explained as endophytes interacting with the plants via a balanced antagonism, in order to recognize the host and colonization they need to switch to the virulence mechanism, and as a result, it triggers the plant defense mechanism. Endophytes maintain equilibrium in plant interaction, while fungi survive with the nutrient exchange. However, as aforementioned, the hazardous nature of the environment affects the health of the plant, thus weakening the defense response. This makes an opportunity for the endophytes (not the true endophytes) to grow abruptly and change their life into a pathogenic form [171,172]. For instance, *Alternaria* species can become pathogenic when the plant becomes stressed and weakened [173].

Changes in the nutrient content are the most recognized environmental factor, which stimulates lifestyle transitions. Unlike nutrient deprivation, the higher nutrient condition is highly responsible for this [167]. Enriched nutrient conditions (e.g., heavy application of fertilizers in agro farming) generally improve plant growth and development. At the same time, lifestyle switching of mycorrhizal into pathogenic is noticeable as balanced mutualistic interaction becomes less balanced. Anyhow, it is difficult to conclude which nutrient (e.g., P or N) could lead to this condition, as various results appeared throughout the literature [174]. Other than the nutritional costs and benefits, Mandyam and Jump-ponen [175] illustrated that *Periconia macrospinoso*, a dark septate endophyte, changes its lifestyle symbiosis to pathogenic with the increasing lower light condition/shade. In support of this, Álvarez-Loayza et al. [176] depicted that *Diplodia mutila* favors low light and endosymbiotic continues with young palm seedlings (*Iriarteia deltoidea*), though high light triggers pathogenicity of the fungus owing to the enhanced produces of Melanin (correlate with increased production of ROS) and H<sub>2</sub>O<sub>2</sub>. Álvarez-Loayza et al. [176] suggested that higher light intensity increases fungus virulence by triggering the HR response in plants.

The reverse is also possible, as pathogens switch into symbiosis [167]. Of this, if the lifestyle characterization (molecular level) of the fungi is based on the capability to cause host disease resistance, stress tolerance (e.g., drought), or growth enhancement, then the parasitic lifestyle can convert into mutualistic when the right time comes. Such condition is recognized with the *Colletotrichum* species [177].

Additionally, the precise mechanism that causes this scenario to arise is yet to be resolved. However, several fragmented attempts made little understanding. For example, Hill et al. [178] worked with the lifestyle changes of *Fusarium* spp. and found that the copy number variation of gene *CSEP* and *CAZyme* is the main driving force as no significant difference in *CSEP*, *CAZyme*, or gene repertoires between phytopathogenic and endophytic strains were noticed. In a separate study, Muszewska et al. [179] highlighted that serine proteases play a crucial role in changing lifestyles.

## 5. Conclusions

Interactions between plants and fungi have become unavoidable and rather indispensable. Understanding plant–fungi interactions is essential in the current world, particularly for sustainable agriculture and food security and ultimately the well-being of humans and livestock. Plants and fungi have a complex and dynamic relationship that can be beneficial or detrimental for both partners. On one hand, some fungi are pathogenic to plants, causing

diseases that reduce plant reproduction, growth, development, and productivity, ultimately affecting their persistence. On the other hand, others form symbiotic associations with plants, such as AM and endophytic fungi, which enhance plant nutrient uptake, growth, and stress tolerance. Moreover, the outcome of plant–fungi interactions can be influenced by various environmental factors, including temperature, light, water, CO<sub>2</sub>, pollutants, and nutrient concentration in the soil, which can modulate the balance between mutualism and antagonism. The intricate molecular mechanisms that trigger the cascade of interactions between plants and their associated fungi under environmental changes, however, are still little understood. There are only a handful of papers documenting the specific non-molecular based mechanism employed by fungi in modifying the plant physiology and themselves to protect against the changing unwelcome environmental conditions. Overall, plant–fungi interactions are a double-edged sword for plant health and survival, depending on the type of fungi involved, the physiological state of the plant, and the abiotic conditions of the habitat.

**Author Contributions:** Conceptualization, A.K.H.P. and S.C.K.; methodology, A.K.H.P., S.T., D.J.B., D.-Q.D. and S.C.K.; validation, S.C.K., I.P. and S.T.; investigation, A.K.H.P. and P.K.; resources, A.K.H.P.; writing—original draft preparation, A.K.H.P.; writing—review and editing, A.K.H.P., S.L.S. and S.C.K.; visualization, A.K.H.P. and S.C.K.; supervision, S.C.K., S.T. and S.L.S.; project administration, S.C.K.; funding acquisition, S.C.K. and D.-Q.D. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was funded by the National Natural Science Foundation of China (No. NSFC 31760013 and 32260004) and the High-Level Talent Recruitment Plan of Yunnan Provinces (“Young Talent” Program).

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** Itthayakorn Promputtha thanks the Faculty of Science, Chiang Mai University for the support. D. J. Bhat gratefully acknowledges the financial support provided under the Distinguished Scientist Fellowship Programme (DSFP), at King Saud University, Riyadh, Saudi Arabia.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Cord-Landwehr, S.; Melcher, R.L.J.; Kolkenbrock, S.; Moerschbacher, B.M. A Chitin Deacetylase from the Endophytic Fungus *Pestalotiopsis* sp. Efficiently Inactivates the Elicitor Activity of Chitin Oligomers in Rice Cells. *Sci. Rep.* **2016**, *6*, 38018. [\[CrossRef\]](#)
2. Karunarathna, S.C.; Ashwath, N.; Jeewon, R. Editorial: The Potential of Fungi for Enhancing Crops and Forestry Systems. *Front. Microbiol.* **2021**, *12*, 813051. [\[CrossRef\]](#)
3. Redman, R.S.; Dunigan, D.D.; Rodriguez, R.J. Fungal Symbiosis from Mutualism to Parasitism: Who Controls the Outcome, Host or Invader? *New Phytol.* **2001**, *151*, 705–716. [\[CrossRef\]](#)
4. Meng, J.-W.; Zhu, W.; He, M.-H.; Wu, E.-J.; Duan, G.-H.; Xie, Y.-K.; Jin, Y.-J.; Yang, L.-N.; Shang, L.-P.; Zhan, J. Population Genetic Analysis Reveals Cryptic Sex in the Phytopathogenic Fungus *Alternaria alternata*. *Sci. Rep.* **2015**, *5*, 18250. [\[CrossRef\]](#)
5. Niu, Z.; Zheng, L.; Yang, P.; Wang, J.; Tian, M.; Pan, Y.; Zhao, D.; Yang, Z.; Zhu, J. Detection of *Alternaria solani* with High Accuracy and Sensitivity during the Latent Period of Potato Early Blight. *Front. Microbiol.* **2022**, *13*, 1016996. [\[CrossRef\]](#)
6. Mandal, S.; Rajarammohan, S.; Kaur, J. *Alternaria brassicae* Interactions with the Model Brassicaceae Member *Arabidopsis thaliana* Closely Resembles Those with Mustard (*Brassica juncea*). *Physiol. Mol. Biol. Plants* **2018**, *24*, 51–59. [\[CrossRef\]](#)
7. Liu, H.; Wu, H.; Wang, Y.; Wang, H.; Chen, S.; Yin, Z. Comparative Transcriptome Profiling and Co-Expression Network Analysis Uncover the Key Genes Associated With early-Stage Resistance to *Aspergillus flavus* in Maize. *BMC Plant Biol.* **2021**, *21*, 216. [\[CrossRef\]](#)
8. Manan, F.; Shi, G.; Gong, H.; Hou, H.; Khan, H.; Leng, Y.; Castell-Miller, C.; Ali, S.; Faris, J.D.; Zhong, S.; et al. Prevalence and Importance of the Necrotrophic Effector Gene *ToxA* in *Bipolaris sorokiniana* Populations Collected from Spring Wheat and Barley. *Plant Dis.* **2023**. [\[CrossRef\]](#)
9. Hua, L.; Yong, C.; Zhanquan, Z.; Boqiang, L.; Guozheng, Q.; Shiping, T. Pathogenic Mechanisms and Control Strategies of *Botrytis cinerea* Causing Post-Harvest Decay in Fruits and Vegetables. *Food Qual. Saf.* **2018**, *2*, 111–119. [\[CrossRef\]](#)

10. Petrasch, S.; Knapp, S.J.; van Kan, J.A.L.; Blanco-Ulate, B. Grey Mould of Strawberry, a Devastating Disease Caused by the Ubiquitous Necrotrophic Fungal Pathogen *Botrytis cinerea*. *Mol. Plant Pathol.* **2019**, *20*, 877–892. [\[CrossRef\]](#)
11. Moreno-Manzano, C.E.; De León-García de Alba, C.; Nava-Díaz, C.; Sánchez-Pale, J.R. Sclerotial Germination And Ascospore Formation of *Claviceps gigantea* Fuentes, De la Isla, Ullstrup y Rodríguez. *Rev. Mex. Fitopatol.* **2016**, *34*, 223–241. [\[CrossRef\]](#)
12. Cannon, P.F.; Damm, U.; Johnston, P.R.; Weir, B.S. *Colletotrichum*—Current Status and Future Directions. *Stud. Mycol.* **2012**, *73*, 181–213. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Jain, A.; Sarsaiya, S.; Wu, Q.; Lu, Y.; Shi, J. A Review of Plant Leaf Fungal Diseases and Its Environment Speciation. *Bioengineered* **2019**, *10*, 409–424. [\[CrossRef\]](#)
14. Hammond-Kosack, K.E.; Rudd, J.J. Plant Resistance Signalling Hijacked by a Necrotrophic Fungal Pathogen. *Plant Signal. Behav.* **2008**, *3*, 993–995. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Downie, R.C.; Lin, M.; Corsi, B.; Ficke, A.; Lillemo, M.; Oliver, R.P.; Phan, H.T.T.; Tan, K.-C.; Cockram, J. Septoria Nodorum Blotch of Wheat: Disease Management and Resistance Breeding in the Face of Shifting Disease Dynamics and a Changing Environment. *Phytopathology* **2021**, *111*, 906–920. [\[CrossRef\]](#) [\[PubMed\]](#)
16. Xia, S.; Xu, Y.; Hoy, R.; Zhang, J.; Qin, L.; Li, X. The Notorious Soilborne Pathogenic Fungus *Sclerotinia sclerotiorum*: An Update on Genes Studied with Mutant Analysis. *Pathogens* **2020**, *9*, 27. [\[CrossRef\]](#)
17. dos Santos, J.P.R.; Pires, L.P.M.; de Castro Vasconcellos, R.C.; Pereira, G.S.; Von Pinho, R.G.; Balestre, M. Genomic Selection to Resistance to *Stenocarpella maydis* in Maize Lines Using DArTseq Markers. *BMC Genet.* **2016**, *17*, 86. [\[CrossRef\]](#)
18. Hassine, M.; Siah, A.; Hellin, P.; Cadalen, T.; Halama, P.; Hilbert, J.-L.; Hamada, W.; Baraket, M.; Yahyaoui, A.; Legrève, A.; et al. Sexual Reproduction of *Zymoseptoria tritici* on Durum Wheat in Tunisia Revealed by Presence of Airborne Inoculum, Fruiting Bodies and High Levels of Genetic Diversity. *Fungal Biol.* **2019**, *123*, 763–772. [\[CrossRef\]](#)
19. Liu, N.; Liu, Z.L.; Gong, G.; Zhang, M.; Wang, X.; Zhou, Y.; Qi, X.; Chen, H.; Yang, J.; Luo, P.; et al. Virulence Structure of *Blumeriagraminis* f. sp. *tritici* and Its Genetic Diversity by ISSR and SRAP Profiling Analyses. *PLoS ONE* **2015**, *10*, e0130881. [\[CrossRef\]](#)
20. Thomma, B.P.H.J.; Van Esse, H.P.; Crous, P.W.; De Wit, P.J.G.M. *Cladosporium fulvum* (syn. *Passalora fulva*), a Highly Specialized Plant Pathogen as a Model for Functional Studies on Plant Pathogenic Mycosphaerellaceae. *Mol. Plant Pathol.* **2005**, *6*, 379–393. [\[CrossRef\]](#)
21. Silva, M.D.C.; Guerra-Guimarães, L.; Diniz, I.; Loureiro, A.; Azinheira, H.; Pereira, A.P.; Tavares, S.; Batista, D.; Várzea, V. An Overview of the Mechanisms Involved in Coffee-*Hemileia vastatrix* Interactions: Plant and Pathogen Perspectives. *Agronomy* **2022**, *12*, 326. [\[CrossRef\]](#)
22. Lawrence, G.J.; Dodds, P.N.; Ellis, J.G. Transformation of the Flax Rust Fungus, *Melampsora lini*: Selection via Silencing of an Avirulence Gene: Transformation of Flax Rust. *Plant J.* **2010**, *61*, 364–369. [\[CrossRef\]](#) [\[PubMed\]](#)
23. Mapuranga, J.; Zhang, N.; Zhang, L.; Chang, J.; Yang, W. Infection Strategies and Pathogenicity of Biotrophic Plant Fungal Pathogens. *Front. Microbiol.* **2022**, *13*, 799396. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Olivera, P.D.; Szabo, L.J.; Kokhmetova, A.; Morgounov, A.; Luster, D.G.; Jin, Y. *Puccinia graminis* f. sp. *tritici* Population Causing Recent Wheat Stem Rust Epidemics in Kazakhstan Is Highly Diverse and Includes Novel Virulence Pathotypes. *Phytopathology* **2022**, *112*, 2403–2415. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Cadavid, M.; Ángel, J.C.; Victoria, J.I. First Report of Orange Rust of Sugarcane Caused by *Puccinia kuehnii* in Colombia. *Plant Dis.* **2012**, *96*, 143. [\[CrossRef\]](#)
26. Wu, N.; Ozketen, A.C.; Cheng, Y.; Jiang, W.; Zhou, X.; Zhao, X.; Guan, Y.; Xiang, Z.; Akkaya, M.S. *Puccinia striiformis* f. sp. *tritici* Effectors in Wheat Immune Responses. *Front. Plant Sci.* **2022**, *13*, 1012216. [\[CrossRef\]](#)
27. Teixeira-Silva, N.S.; Schaker, P.D.C.; Rody, H.V.S.; Maia, T.; Garner, C.M.; Gassmann, W.; Monteiro-Vitorello, C.B. Leaping into the Unknown World of *Sporisorium scitamineum* Candidate Effectors. *J. Fungi* **2020**, *6*, 339. [\[CrossRef\]](#)
28. Ferris, A.C.; Walbot, V. Understanding *Ustilago maydis* Infection of Multiple Maize Organs. *J. Fungi* **2021**, *7*, 8. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Yan, Y.; Yuan, Q.; Tang, J.; Huang, J.; Hsiang, T.; Wei, Y.; Zheng, L. *Colletotrichum higginsianum* as a Model for Understanding Host–Pathogen Interactions: A Review. *Int. J. Mol. Sci.* **2018**, *19*, 2142. [\[CrossRef\]](#)
30. Samac, D.A.; Allen, S.; Witte, D.; Miller, D.; Peterson, J. First Report of Race 2 of *Colletotrichum trifolii* Causing Anthracnose on Alfalfa (*Medicago sativa*) in Wisconsin. *Plant Dis.* **2014**, *98*, 843. [\[CrossRef\]](#)
31. Hami, A.; Rasool, R.S.; Khan, N.A.; Mansoor, S.; Mir, M.A.; Ahmed, N.; Masoodi, K.Z. Morpho-Molecular Identification and First Report of *Fusarium equiseti* in Causing Chilli Wilt from Kashmir (Northern Himalayas). *Sci. Rep.* **2021**, *11*, 3610. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Lyons, R.; Stiller, J.; Powell, J.; Rusu, A.; Manners, J.M.; Kazan, K. *Fusarium oxysporum* Triggers Tissue-Specific Transcriptional Reprogramming in *Arabidopsis thaliana*. *PLoS ONE* **2015**, *10*, e0121902. [\[CrossRef\]](#)
33. Huang, Z.; Li, H.; Zhou, Y.; Bao, Y.; Duan, Z.; Wang, C.; Powell, C.A.; Chen, B.; Zhang, M.; Yao, W. Predication of the Effector Proteins Secreted by *Fusarium sacchari* Using Genomic Analysis and Heterogenous Expression. *J. Fungi* **2022**, *8*, 59. [\[CrossRef\]](#)
34. Bahari, M.N.A.; Sakeh, N.M.; Abdullah, S.N.A.; Ramli, R.R.; Kadkhodaei, S. Transcriptome Profiling at Early Infection of *Elaeis guineensis* by *Ganoderma boninense* Provides Novel Insights on Fungal Transition from Biotrophic to Necrotrophic Phase. *BMC Plant Biol.* **2018**, *18*, 377. [\[CrossRef\]](#)
35. Park, J.-Y.; Jin, J.; Lee, Y.-W.; Kang, S.; Lee, Y.-H. Rice Blast Fungus (*Magnaporthe oryzae*) Infects *Arabidopsis* via a Mechanism Distinct from That Required for the Infection of Rice. *Plant Physiol.* **2009**, *149*, 474–486. [\[CrossRef\]](#) [\[PubMed\]](#)

36. Zhao, X.; Zhang, Z.; Zheng, S.; Ye, W.; Zheng, X.; Wang, Y. Genome Sequence Resource of *Phomopsis longicolla* YC2-1, a Fungal Pathogen Causing Phomopsis Stem Blight in Soybean. *Mol. Plant-Microbe Interact.* **2021**, *34*, 842–844. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Maffei, G.; Miozzi, L.; Fiorilli, V.; Novero, M.; Lanfranco, L.; Accotto, G.P. The Arbuscular Mycorrhizal Symbiosis Attenuates Symptom Severity and Reduces Virus Concentration in Tomato Infected by Tomato Yellow Leaf Curl Sardinia Virus (TYLCSV). *Mycorrhiza* **2014**, *24*, 179–186. [\[CrossRef\]](#)
38. Gworgwor, N.A.; Weber, H.C. Arbuscular Mycorrhizal Fungi-Parasite-Host Interaction for the Control of *Striga hermonthica* (Del.) Benth. in Sorghum [*Sorghum bicolor* (L.) Moench]. *Mycorrhiza* **2003**, *13*, 277–281. [\[CrossRef\]](#)
39. Nemec, S. Virus-Glomus *etunicatum* Interactions in Citrus Rootstocks. *Plant Dis.* **1984**, *68*, 311. [\[CrossRef\]](#)
40. Shaul, O.; Galili, S.; Volpin, H.; Ginzberg, I.I.; Elad, Y.; Chet, I.I.; Kapulnik, Y. Mycorrhiza-Induced Changes in Disease Severity and PR Protein Expression in Tobacco Leaves. *Mol. Plant-Microbe Interact.* **1999**, *12*, 1000–1007. [\[CrossRef\]](#)
41. Sipahioglu, M.H.; Demir, S.; Usta, M.; Akkopru, A. Biological Relationship of Potato Virus Y and Arbuscular Mycorrhizal Fungus *Glomus intraradices* in Potato. *Pest Tec.* **2009**, *3*, 63–66.
42. Elsharkawy, M.M.; Shimizu, M.; Takahashi, H.; Hyakumachi, M. The Plant Growth-Promoting Fungus *Fusarium equiseti* and the Arbuscular Mycorrhizal Fungus *Glomus mosseae* Induce Systemic Resistance against Cucumber Mosaic Virus in Cucumber Plants. *Plant Soil* **2012**, *361*, 397–409. [\[CrossRef\]](#)
43. Harman, G.E. Multifunctional Fungal Plant Symbionts: New Tools to Enhance Plant Growth and Productivity: Commentary. *New Phytol.* **2011**, *189*, 647–649. [\[CrossRef\]](#)
44. Balestrini, R. Grand Challenges in Fungi-Plant Interactions. *Front. Fungal Biol.* **2021**, *2*, 750003. [\[CrossRef\]](#)
45. Bennett, J.A.; Cahill, J.F., Jr. Fungal Effects on Plant-Plant Interactions Contribute to Grassland Plant Abundances: Evidence from the Field. *J. Ecol.* **2016**, *104*, 755–764. [\[CrossRef\]](#)
46. Doehlemann, G.; Ökmen, B.; Zhu, W.; Sharon, A. Plant Pathogenic Fungi. *Microbiol. Spectr.* **2017**, *5*, 14. [\[CrossRef\]](#)
47. Shao, D.; Smith, D.L.; Kabbage, M.; Roth, M.G. Effectors of Plant Necrotrophic Fungi. *Front. Plant Sci.* **2021**, *12*, 687713. [\[CrossRef\]](#)
48. Fei, W.; Liu, Y. Biotrophic Fungal Pathogens: A Critical Overview. *Appl. Biochem. Biotechnol.* **2023**, *195*, 1–16. [\[CrossRef\]](#)
49. Chakraborty, N.; Chakraborty, P.; Bandopadhyay, R.; Basak, J. Deciphering the Molecular Mechanisms of Biotic Stress Tolerance Unravels the Mystery of Plant-Pathogen Interaction. In *Sustainable Agriculture Reviews 51*; Springer: Cham, Switzerland, 2021; pp. 295–316, ISBN 9783030688271.
50. Backer, R.; Naidoo, S.; van den Berg, N. The Nonexpressor of Pathogenesis-Related Genes 1 (NPR1) and Related Family: Mechanistic Insights in Plant Disease Resistance. *Front. Plant Sci.* **2019**, *10*, 102. [\[CrossRef\]](#)
51. Nesher, I.; Minz, A.; Kokkelink, L.; Tudzynski, P.; Sharon, A. Regulation of Pathogenic Spore Germination by CgRac1 in the Fungal Plant Pathogen *Colletotrichum gloeosporioides*. *Eukaryot. Cell* **2011**, *10*, 1122–1130. [\[CrossRef\]](#)
52. Balotf, S.; Tegg, R.S.; Nichols, D.S.; Wilson, C.R. Spore Germination of the Obligate Biotroph *Spongospora subterranea*: Transcriptome Analysis Reveals Germination Associated Genes. *Front. Microbiol.* **2021**, *12*, 691877. [\[CrossRef\]](#)
53. Vohník, M. Ericoid Mycorrhizal Symbiosis: Theoretical Background and Methods for Its Comprehensive Investigation. *Mycorrhiza* **2020**, *30*, 671–695. [\[CrossRef\]](#)
54. Strullu-Derrien, C.; Selosse, M.-A.; Kenrick, P.; Martin, F.M. The Origin and Evolution of Mycorrhizal Symbioses: From Palaeomycology to Phylogenomics. *New Phytol.* **2018**, *220*, 1012–1030. [\[CrossRef\]](#)
55. Huey, C.J.; Gopinath, S.C.B.; Uda, M.N.A.; Zulhaimi, H.I.; Jaafar, M.N.; Kasim, F.H.; Yaakub, A.R.W. Mycorrhiza: A Natural Resource Assists Plant Growth under Varied Soil Conditions. *3 Biotech* **2020**, *10*, 204. [\[CrossRef\]](#)
56. Dighton, J. *Encyclopedia of Microbiology*, 3rd ed.; Elsevier: Cambridge, MA, USA, 2009; Mycorrhizae; pp. 153–162, ISBN 9780123739445.
57. Berruti, A.; Lumini, E.; Balestrini, R.; Bianciotto, V. Arbuscular Mycorrhizal Fungi as Natural Biofertilizers: Let's Benefit from Past Successes. *Front. Microbiol.* **2016**, *6*, 1559. [\[CrossRef\]](#)
58. Pirttilä, A.M.; Mohammad Parast Tabas, H.; Baruah, N.; Koskimäki, J.J. Biofertilizers and Biocontrol Agents for Agriculture: How to Identify and Develop New Potent Microbial Strains and Traits. *Microorganisms* **2021**, *9*, 817. [\[CrossRef\]](#)
59. Cairney, J.W. Evolution of Mycorrhiza Systems. *Sci. Nat.* **2000**, *87*, 467–475. [\[CrossRef\]](#)
60. Jensen, R.E.; Cabral, C.; Enkegaard, A.; Steenberg, T. Influence of the Plant Interacting Entomopathogenic Fungus *Beauveria bassiana* on Parasitoid Host Choice-Behavior, Development, and Plant Defense Pathways. *PLoS ONE* **2020**, *15*, e0238943. [\[CrossRef\]](#)
61. Lu, H.; Wei, T.; Lou, H.; Shu, X.; Chen, Q. A Critical Review on Communication Mechanism within Plant-Endophytic Fungi Interactions to Cope with Biotic and Abiotic Stresses. *J. Fungi* **2021**, *7*, 719. [\[CrossRef\]](#)
62. Chen, X.-J.; Yin, Y.-Q.; Zhu, X.-M.; Xia, X.; Han, J.-J. High Ambient Temperature Regulated the Plant Systemic Response to the Beneficial Endophytic Fungus *Serendipita indica*. *Front. Plant Sci.* **2022**, *13*, 844572. [\[CrossRef\]](#)
63. He, W.-X.; Wu, Q.-S.; Hashem, A.; Abd\_Allah, E.F.; Muthuramalingam, P.; Al-Arjani, A.-B.F.; Zou, Y.-N. Effects of Symbiotic Fungi on Sugars and Soil Fertility and Structure-Mediated Changes in Plant Growth of *Vicia villosa*. *Agriculture* **2022**, *12*, 1523. [\[CrossRef\]](#)
64. Priyashantha, A.K.H.; Attanayake, R.N. Can Anaerobic Soil Disinfestation (ASD) Be a Game Changer in Tropical Agriculture? *Pathogens* **2021**, *10*, 133. [\[CrossRef\]](#)
65. Tudi, M.; Daniel Ruan, H.; Wang, L.; Lyu, J.; Sadler, R.; Connell, D.; Chu, C.; Phung, D.T. Agriculture Development, Pesticide Application and Its Impact on the Environment. *Int. J. Environ. Res. Public Health* **2021**, *18*, 1112. [\[CrossRef\]](#) [\[PubMed\]](#)

66. Rillig, M.C.; Aguilar-Trigueros, C.A.; Camenzind, T.; Cavagnaro, T.R.; Degruene, F.; Hohmann, P.; Lammel, D.R.; Mansour, I.; Roy, J.; van der Heijden, M.G.A.; et al. Why Farmers Should Manage the Arbuscular Mycorrhizal Symbiosis. *New Phytol.* **2019**, *222*, 1171–1175. [[CrossRef](#)]
67. Duley, G.; Boselli, E. Mutual Plant-Fungi Symbiosis Compromised by Fungicide Use. *Commun. Biol.* **2022**, *5*, 1069. [[CrossRef](#)] [[PubMed](#)]
68. Edlinger, A.; Garland, G.; Hartman, K.; Banerjee, S.; Degruene, F.; García-Palacios, P.; Hallin, S.; Valzano-Held, A.; Herzog, C.; Jansa, J.; et al. Agricultural Management and Pesticide Use Reduce the Functioning of Beneficial Plant Symbionts. *Nat. Ecol. Evol.* **2022**, *6*, 1145–1154. [[CrossRef](#)] [[PubMed](#)]
69. Redecker, D.; Kodner, R.; Graham, L.E. Glomalean Fungi from the Ordovician. *Science* **2000**, *289*, 1920–1921. [[CrossRef](#)] [[PubMed](#)]
70. Brundrett, M.C.; Tedersoo, L. Evolutionary History of Mycorrhizal Symbioses and Global Host Plant Diversity. *New Phytol.* **2018**, *220*, 1108–1115. [[CrossRef](#)]
71. Hiruma, K.; Kobae, Y.; Toju, H. Beneficial Associations between Brassicaceae Plants and Fungal Endophytes under Nutrient-Limiting Conditions: Evolutionary Origins and Host–Symbiont Molecular Mechanisms. *Curr. Opin. Plant Biol.* **2018**, *44*, 145–154. [[CrossRef](#)]
72. Bahram, M.; Netherway, T. Fungi as Mediators Linking Organisms and Ecosystems. *FEMS Microbiol. Rev.* **2022**, *46*, fuab058. [[CrossRef](#)]
73. Möller, M.; Stukenbrock, E.H. Evolution and Genome Architecture in Fungal Plant Pathogens. *Nat. Rev. Microbiol.* **2017**, *15*, 756–771. [[CrossRef](#)]
74. Van Valen, L. A new evolutionary law. *Evol. Theory* **1973**, *1*, 1–30.
75. Clay, K.; Kover, P.X. The Red Queen Hypothesis and Plant/Pathogen Interactions. *Annu. Rev. Phytopathol.* **1996**, *34*, 29–50. [[CrossRef](#)]
76. Delaye, L.; Ruiz-Ruiz, S.; Calderon, E.; Tarazona, S.; Conesa, A.; Moya, A. Evidence of the Red-Queen Hypothesis from Accelerated Rates of Evolution of Genes Involved in Biotic Interactions in *Pneumocystis*. *Genome Biol. Evol.* **2018**, *10*, 1596–1606. [[CrossRef](#)] [[PubMed](#)]
77. Solé, R. Revisiting Leigh van Valen’s “A New Evolutionary Law” (1973). *Biol. Theory* **2022**, *17*, 120–125. [[CrossRef](#)]
78. Sun, S.; Heitman, J. Is Sex Necessary? *BMC Biol.* **2011**, *9*, 56. [[CrossRef](#)]
79. Ashu, E.E.; Xu, J. The Roles of Sexual and Asexual Reproduction in the Origin and Dissemination of Strains Causing Fungal Infectious Disease Outbreaks. *Infect. Genet. Evol.* **2015**, *36*, 199–209. [[CrossRef](#)] [[PubMed](#)]
80. Newman, T.E.; Derbyshire, M.C. The Evolutionary and Molecular Features of Broad Host-Range Necrotrophy in Plant Pathogenic Fungi. *Front. Plant Sci.* **2020**, *11*, 591733. [[CrossRef](#)]
81. Krings, M.; Taylor, T.N.; Hass, H.; Kerp, H.; Dotzler, N.; Hermsen, E.J. Fungal Endophytes in a 400-Million-Yr-Old Land Plant: Infection Pathways, Spatial Distribution, and Host Responses. *New Phytol.* **2007**, *174*, 648–657. [[CrossRef](#)]
82. Feijen, F.A.A.; Vos, R.A.; Nuytinck, J.; Merckx, V.S.F.T. Evolutionary Dynamics of Mycorrhizal Symbiosis in Land Plant Diversification. *Sci. Rep.* **2018**, *8*, 10698. [[CrossRef](#)]
83. Hoeksema, J.D.; Bever, J.D.; Chakraborty, S.; Chaudhary, V.B.; Gardes, M.; Gehring, C.A.; Hart, M.M.; Housworth, E.A.; Kaonongbua, W.; Klironomos, J.N.; et al. Evolutionary History of Plant Hosts and Fungal Symbionts Predicts the Strength of Mycorrhizal Mutualism. *Commun. Biol.* **2018**, *1*, 116. [[CrossRef](#)]
84. Brundrett, M.C. Coevolution of Roots and Mycorrhizas of Land Plants. *New Phytol.* **2002**, *154*, 275–304. [[CrossRef](#)]
85. Cameron, D.D.; Neal, A.L.; van Wees, S.C.M.; Ton, J. Mycorrhiza-Induced Resistance: More than the Sum of Its Parts? *Trends Plant Sci.* **2013**, *18*, 539–545. [[CrossRef](#)] [[PubMed](#)]
86. Gouda, S.; Das, G.; Sen, S.K.; Shin, H.-S.; Patra, J.K. Endophytes: A Treasure House of Bioactive Compounds of Medicinal Importance. *Front. Microbiol.* **2016**, *7*, 1538. [[CrossRef](#)]
87. Zamioudis, C.; Pieterse, C.M.J. Modulation of Host Immunity by Beneficial Microbes. *Mol. Plant-Microbe Interact.* **2012**, *25*, 139–150. [[CrossRef](#)]
88. Giraldo, M.C.; Valent, B. Filamentous Plant Pathogen Effectors in Action. *Nat. Rev. Microbiol.* **2013**, *11*, 800–814. [[CrossRef](#)]
89. Ponce de León, I.; Montesano, M. Activation of Defense Mechanisms against Pathogens in Mosses and Flowering Plants. *Int. J. Mol. Sci.* **2013**, *14*, 3178–3200. [[CrossRef](#)]
90. Nguyen, Q.-M.; Iswanto, A.B.B.; Son, G.H.; Kim, S.H. Recent Advances in Effector-Triggered Immunity in Plants: New Pieces in the Puzzle Create a Different Paradigm. *Int. J. Mol. Sci.* **2021**, *22*, 4709. [[CrossRef](#)]
91. Wan, J.; Zhang, X.-C.; Stacey, G. Chitin Signaling and Plant Disease Resistance. *Plant Signal. Behav.* **2008**, *3*, 831–833. [[CrossRef](#)]
92. Hawkins, L.K.; Mylroie, J.E.; Oliveira, D.A.; Smith, J.S.; Ozkan, S.; Windham, G.L.; Williams, W.P.; Warburton, M.L. Characterization of the Maize Chitinase Genes and Their Effect on *Aspergillus flavus* and Aflatoxin Accumulation Resistance. *PLoS ONE* **2015**, *10*, e0126185. [[CrossRef](#)]
93. Lo Presti, L.; Lanver, D.; Schweizer, G.; Tanaka, S.; Liang, L.; Tollot, M.; Zuccaro, A.; Reissmann, S.; Kahmann, R. Fungal Effectors and Plant Susceptibility. *Annu. Rev. Plant Biol.* **2015**, *66*, 513–545. [[CrossRef](#)]
94. Osbourn, A.E. Preformed Antimicrobial Compounds and Plant Defense against Fungal Attack. *Plant Cell* **1996**, *8*, 1821–1831. [[CrossRef](#)]
95. Rajamuthiah, R.; Mylonakis, E. Effector Triggered Immunity: Activation of Innate Immunity in Metazoans by Bacterial Effectors. *Virulence* **2014**, *5*, 697–702. [[CrossRef](#)]

96. Hatsugai, N.; Igarashi, D.; Mase, K.; Lu, Y.; Tsuda, Y.; Chakravarthy, S.; Wei, H.-L.; Foley, J.W.; Collmer, A.; Glazebrook, J.; et al. A Plant Effector-Triggered Immunity Signaling Sector Is Inhibited by Pattern-Triggered Immunity. *EMBO J.* **2017**, *36*, 2758–2769. [\[CrossRef\]](#)
97. van Kan, J.A.; van den Ackerveken, G.F.; de Wit, P.J. Cloning and Characterization of cDNA of Avirulence Gene *Avr9* of the Fungal Pathogen *Cladosporium fulvum*, Causal Agent of Tomato Leaf Mold. *Mol. Plant-Microbe Interact.* **1991**, *4*, 52–59. [\[CrossRef\]](#)
98. de Wit, P.J.G.M. *Genes Involved in Plant Defense*; Springer Vienna: Vienna, Australia, 1992; Functional Models to Explain Gene-for-Gene Relationships in Plant-Pathogen Interactions; pp. 25–47, ISBN 9783709173800.
99. Frank, S.A. Coevolutionary Genetics of Plants and Pathogens. *Evol. Ecol.* **1993**, *7*, 45–75. [\[CrossRef\]](#)
100. Glazebrook, J. Contrasting Mechanisms of Defense against Biotrophic and Necrotrophic Pathogens. *Annu. Rev. Phytopathol.* **2005**, *43*, 205–227. [\[CrossRef\]](#)
101. Bari, R.; Jones, J.D.G. Role of Plant Hormones in Plant Defence Responses. *Plant Mol. Biol.* **2009**, *69*, 473–488. [\[CrossRef\]](#)
102. Wang, Y.; Mostafa, S.; Zeng, W.; Jin, B. Function and Mechanism of Jasmonic Acid in Plant Responses to Abiotic and Biotic Stresses. *Int. J. Mol. Sci.* **2021**, *22*, 8568. [\[CrossRef\]](#)
103. Tamaoki, D.; Seo, S.; Yamada, S.; Kano, A.; Miyamoto, A.; Shishido, H.; Miyoshi, S.; Taniguchi, S.; Akimitsu, K.; Gomi, K. Jasmonic Acid and Salicylic Acid Activate a Common Defense System in Rice. *Plant Signal. Behav.* **2013**, *8*, e24260. [\[CrossRef\]](#)
104. Pena-Cortés, H.; Albrecht, T.; Prat, S.; Weiler, E.; Willmitzer, L. Aspirin Prevents Wound-Induced Gene Expression in Tomato Leaves by Blocking Jasmonic Acid Biosynthesis. *Planta* **1993**, *191*, 123–128. [\[CrossRef\]](#)
105. Riemann, M.; Haga, K.; Shimizu, T.; Okada, K.; Ando, S.; Mochizuki, S.; Nishizawa, Y.; Yamanouchi, U.; Nick, P.; Yano, M.; et al. Identification of Rice *Allene Oxide Cyclase* Mutants and the Function of Jasmonate for Defence against *Magnaporthe oryzae*. *Plant J.* **2013**, *74*, 226–238. [\[CrossRef\]](#)
106. Williams, B.; Kabbage, M.; Kim, H.-J.; Britt, R.; Dickman, M.B. Tipping the Balance: *Sclerotinia sclerotiorum* Secreted Oxalic Acid Suppresses Host Defenses by Manipulating the Host Redox Environment. *PLoS Pathog.* **2011**, *7*, e1002107. [\[CrossRef\]](#)
107. Liang, X.; Rollins, J.A. Mechanisms of Broad Host Range Necrotrophic Pathogenesis in *Sclerotinia sclerotiorum*. *Phytopathology* **2018**, *108*, 1128–1140. [\[CrossRef\]](#)
108. Ito, K.; Tanaka, T.; Hatta, R.; Yamamoto, M.; Akimitsu, K.; Tsuge, T. Dissection of the Host Range of the Fungal Plant Pathogen *Alternaria alternata* by Modification of Secondary Metabolism: Host Specificity of *Alternaria alternata*. *Mol. Microbiol.* **2004**, *52*, 399–411. [\[CrossRef\]](#)
109. Ma, H.; Zhang, B.; Gai, Y.; Sun, X.; Chung, K.-R.; Li, H. Cell-Wall-Degrading Enzymes Required for Virulence in the Host Selective Toxin-Producing Necrotroph *Alternaria alternata* of Citrus. *Front. Microbiol.* **2019**, *10*, 2514. [\[CrossRef\]](#)
110. Mendgen, K.; Hahn, M. Plant Infection and the Establishment of Fungal Biotrophy. *Trends Plant Sci.* **2002**, *7*, 352–356. [\[CrossRef\]](#)
111. Dufresne, M.; Perfect, S.; Pellier, A.L.; Bailey, J.A.; Langin, T. A GAL4-like Protein Is Involved in the Switch between Biotrophic and Necrotrophic Phases of the Infection Process of *Colletotrichum lindemuthianum* on Common Bean. *Plant Cell* **2000**, *12*, 1579–1590. [\[CrossRef\]](#)
112. Münch, S.; Lingner, U.; Floss, D.S.; Ludwig, N.; Sauer, N.; Deising, H.B. The Hemibiotrophic Lifestyle of *Colletotrichum* Species. *J. Plant Physiol.* **2008**, *165*, 41–51. [\[CrossRef\]](#)
113. Mengiste, T. Plant Immunity to Necrotrophs. *Annu. Rev. Phytopathol.* **2012**, *50*, 267–294. [\[CrossRef\]](#)
114. Zimnoch-Guzowska, E.; Lebecka, R.; Kryszczuk, A.; Maciejewska, U.; Szczerbakowa, A.; Wielgat, B. Resistance to *Phytophthora infestans* in Somatic Hybrids of *Solanum nigrum* L. and Diploid Potato. *Züchter Genet. Breed. Res.* **2003**, *107*, 43–48. [\[CrossRef\]](#)
115. Lyu, X.; Shen, C.; Fu, Y.; Xie, J.; Jiang, D.; Li, G.; Cheng, J. A Small Secreted Virulence-Related Protein Is Essential for the Necrotrophic Interactions of *Sclerotinia sclerotiorum* with Its Host Plants. *PLoS Pathog.* **2016**, *12*, e1005435. [\[CrossRef\]](#)
116. Reindl, M.; Hänsch, S.; Weidtkamp-Peters, S.; Schipper, K. A Potential Lock-Type Mechanism for Unconventional Secretion in Fungi. *Int. J. Mol. Sci.* **2019**, *20*, 460. [\[CrossRef\]](#)
117. Petre, B.; Kamoun, S. How Do Filamentous Pathogens Deliver Effector Proteins into Plant Cells? *PLoS Biol.* **2014**, *12*, e1001801. [\[CrossRef\]](#)
118. Kombrink, A.; Thomma, B.P.H.J. LysM Effectors: Secreted Proteins Supporting Fungal Life. *PLoS Pathog.* **2013**, *9*, e1003769. [\[CrossRef\]](#)
119. Santhanam, P.; van Esse, H.P.; Albert, I.; Faino, L.; Nürnberger, T.; Thomma, B.P.H.J. Evidence for Functional Diversification within a Fungal NEP1-like Protein Family. *Mol. Plant-Microbe Interact.* **2013**, *26*, 278–286. [\[CrossRef\]](#)
120. Kombrink, A.; Rovenich, H.; Shi-Kunne, X.; Rojas-Padilla, E.; van den Berg, G.C.M.; Domazakis, E.; de Jonge, R.; Valkenburg, D.-J.; Sánchez-Vallet, A.; Seidl, M.F.; et al. *Verticillium dahlia* LysM Effectors Differentially Contribute to Virulence on Plant Hosts. *Mol. Plant Pathol.* **2017**, *18*, 596–608. [\[CrossRef\]](#)
121. Patkar, R.N.; Benke, P.I.; Qu, Z.; Chen, Y.Y.C.; Yang, F.; Swarup, S.; Naqvi, N.I. A Fungal Monooxygenase-Derived Jasmonate Attenuates Host Innate Immunity. *Nat. Chem. Biol.* **2015**, *11*, 733–740. [\[CrossRef\]](#)
122. Saha, H.; Kaloterakis, N.; Harvey, J.A.; Van der Putten, W.H.; Biere, A. Effects of Light Quality on Colonization of Tomato Roots by AMF and Implications for Growth and Defense. *Plants* **2022**, *11*, 861. [\[CrossRef\]](#)
123. Chanda, A.; Maghrawy, H.; Sayour, H.; Gummadidala, P.M.; Gomaa, O.M. Impact of Climate Change on Plant-Associated Fungi. In *Climate Change Impacts on Agriculture and Food Security in Egypt*; Springer: Cham, Switzerland, 2020; pp. 83–96, ISBN 9783030416287.

124. Trivedi, P.; Batista, B.D.; Bazany, K.E.; Singh, B.K. Plant-Microbiome Interactions under a Changing World: Responses, Consequences and Perspectives. *New Phytol.* **2022**, *234*, 1951–1959. [\[CrossRef\]](#)
125. Velásquez, A.C.; Castroverde, C.D.M.; He, S.Y. Plant–Pathogen Warfare under Changing Climate Conditions. *Curr. Biol.* **2018**, *28*, R619–R634. [\[CrossRef\]](#)
126. Perrone, G.; Ferrara, M.; Medina, A.; Pascale, M.; Magan, N. Toxigenic Fungi and Mycotoxins in a Climate Change Scenario: Ecology, Genomics, Distribution, Prediction and Prevention of the Risk. *Microorganisms* **2020**, *8*, 1496. [\[CrossRef\]](#)
127. Cohen, S.P.; Leach, J.E. High Temperature-Induced Plant Disease Susceptibility: More than the Sum of Its Parts. *Curr. Opin. Plant Biol.* **2020**, *56*, 235–241. [\[CrossRef\]](#)
128. Nieva, A.S.; Romero, F.M.; Erban, A.; Carrasco, P.; Ruiz, O.A.; Kopka, J. Metabolic Profiling and Metabolite Correlation Network Analysis Reveal That *Fusarium solani* Induces Differential Metabolic Responses in *Lotus japonicus* and *Lotus tenuis* against Severe Phosphate Starvation. *J. Fungi* **2021**, *7*, 765. [\[CrossRef\]](#)
129. Nnadi, N.E.; Carter, D.A. Climate Change and the Emergence of Fungal Pathogens. *PLoS Pathog.* **2021**, *17*, e1009503. [\[CrossRef\]](#)
130. Shaffique, S.; Khan, M.A.; Wani, S.H.; Pande, A.; Imran, M.; Kang, S.-M.; Rahim, W.; Khan, S.A.; Bhatta, D.; Kwon, E.-H.; et al. A Review on the Role of Endophytes and Plant Growth Promoting Rhizobacteria in Mitigating Heat Stress in Plants. *Microorganisms* **2022**, *10*, 1286. [\[CrossRef\]](#)
131. Compant, S.; van der Heijden, M.G.A.; Sessitsch, A. Climate Change Effects on Beneficial Plant-Microorganism Interactions: Climate Change and Beneficial Plant-Microorganism Interactions. *FEMS Microbiol. Ecol.* **2010**, *73*, 197–214. [\[CrossRef\]](#)
132. Bennett, A.E.; Classen, A.T. Climate Change Influences Mycorrhizal Fungal-Plant Interactions, but Conclusions Are Limited by Geographical Study Bias. *Ecology* **2020**, *101*, e02978. [\[CrossRef\]](#)
133. Mathur, S.; Agnihotri, R.; Sharma, M.P.; Reddy, V.R.; Jajoo, A. Effect of High-Temperature Stress on Plant Physiological Traits and Mycorrhizal Symbiosis in Maize Plants. *J. Fungi* **2021**, *7*, 867. [\[CrossRef\]](#)
134. Martin, C.A.; Stutz, J.C. Interactive Effects of Temperature and Arbuscular Mycorrhizal Fungi on Growth, P Uptake and Root Respiration of *Capsicum annuum* L. *Mycorrhiza* **2004**, *14*, 241–244. [\[CrossRef\]](#)
135. Liu, A.; Wang, B.; Hamel, C. Arbuscular Mycorrhiza Colonization and Development at Suboptimal Root Zone Temperature. *Mycorrhiza* **2004**, *14*, 93–101. [\[CrossRef\]](#) [\[PubMed\]](#)
136. de Vallavielle-Pope, C.; Bahri, B.; Leconte, M.; Zurfluh, O.; Belaid, Y.; Maghrebi, E.; Huard, F.; Huber, L.; Launay, M.; Bancal, M.O. Thermal Generalist Behaviour of Invasive *Puccinia striiformis* f. sp. *tritici* Strains under Current and Future Climate Conditions. *Plant Pathol.* **2018**, *67*, 1307–1320. [\[CrossRef\]](#)
137. Heinemeyer, A.; Fitter, A.H. Impact of Temperature on the Arbuscular Mycorrhizal (AM) Symbiosis: Growth Responses of the Host Plant and Its AM Fungal Partner. *J. Exp. Bot.* **2004**, *55*, 525–534. [\[CrossRef\]](#)
138. Jerbi, M.; Labidi, S.; Lounès-Hadj Sahraoui, A.; Chaar, H.; Ben Jeddi, F. Higher Temperatures and Lower Annual Rainfall Do Not Restrict, Directly or Indirectly, the Mycorrhizal Colonization of Barley (*Hordeum vulgare* L.) under Rainfed Conditions. *PLoS ONE* **2020**, *15*, e0241794. [\[CrossRef\]](#)
139. Carro-Huerga, G.; Mayo-Prieto, S.; Rodríguez-González, Á.; Álvarez-García, S.; Gutiérrez, S.; Casquero, P.A. The Influence of Temperature on the Growth, Sporulation, Colonization, and Survival of *Trichoderma* spp. in Grapevine Pruning Wounds. *Agronomy* **2021**, *11*, 1771. [\[CrossRef\]](#)
140. Marsberg, A.; Kemler, M.; Jami, F.; Nagel, J.H.; Postma-Smidt, A.; Naidoo, S.; Wingfield, M.J.; Crous, P.W.; Spatafora, J.W.; Hesse, C.N.; et al. *Botryosphaeria dothidea*: A Latent Pathogen of Global Importance to Woody Plant Health. *Mol. Plant Pathol.* **2017**, *18*, 477–488. [\[CrossRef\]](#)
141. Ballhorn, D.J.; Schädler, M.; Elias, J.D.; Millar, J.A.; Kautz, S. Friend or Foe—Light Availability Determines the Relationship between Mycorrhizal Fungi, Rhizobia and Lima Bean (*Phaseolus lunatus* L.). *PLoS ONE* **2016**, *11*, e0154116. [\[CrossRef\]](#) [\[PubMed\]](#)
142. Carstensen, A.; Herdean, A.; Schmidt, S.B.; Sharma, A.; Spetea, C.; Pribil, M.; Husted, S. The Impacts of Phosphorus Deficiency on the Photosynthetic Electron Transport Chain. *Plant Physiol.* **2018**, *177*, 271–284. [\[CrossRef\]](#)
143. Garnica, S.; Liao, Z.; Hamard, S.; Waller, F.; Parepa, M.; Bossdorf, O. Environmental Stress Determines the Colonization and Impact of an Endophytic Fungus on Invasive Knotweed. *Biol. Invasions* **2022**, *24*, 1785–1795. [\[CrossRef\]](#)
144. Hevia, M.A.; Canessa, P.; Müller-Esparza, H.; Larrondo, L.F. A Circadian Oscillator in the Fungus *Botrytis cinerea* Regulates Virulence When Infecting *Arabidopsis thaliana*. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 8744–8749. [\[CrossRef\]](#)
145. Lozano, Y.M.; Aguilar-Trigueros, C.A.; Roy, J.; Rillig, M.C. Drought Induces Shifts in Soil Fungal Communities That Can Be Linked to Root Traits across 24 Plant Species. *New Phytol.* **2021**, *232*, 1917–1929. [\[CrossRef\]](#)
146. Lamhamedi, M.S.; Bernier, P.Y.; André-Fortin, J. Hydraulic Conductance and Soil Water Potential at the Soil-Root Interface of *Pinus pinaster* Seedlings Inoculated with Different Dikaryons of *Pisolithus* sp. *Tree Physiol.* **1992**, *10*, 231–244. [\[CrossRef\]](#)
147. Brosi, G.B.; McCulley, R.L.; Bush, L.P.; Nelson, J.A.; Classen, A.T.; Norby, R.J. Effects of Multiple Climate Change Factors on the Tall Fescue-Fungal Endophyte Symbiosis: Infection Frequency and Tissue Chemistry. *New Phytol.* **2011**, *189*, 797–805. [\[CrossRef\]](#)
148. Augé, R.M. Water Relations, Drought and Vesicular-Arbuscular Mycorrhizal Symbiosis. *Mycorrhiza* **2001**, *11*, 3–42. [\[CrossRef\]](#)
149. Morte, A.; Díaz, G.; Rodríguez, P.; Alarcón, J.J.; Sánchez-Blanco, M.J. Growth and Water Relations in Mycorrhizal and Nonmycorrhizal *Pinus halepensis* Plants in Response to Drought. *Biol. Plant.* **2001**, *44*, 263–267. [\[CrossRef\]](#)
150. Boczoń, A.; Hilszczańska, D.; Wrzosek, M.; Szczepkowski, A.; Sierota, Z. Drought in the Forest Breaks Plant–Fungi Interactions. *Eur. J. For. Res.* **2021**, *140*, 1301–1321. [\[CrossRef\]](#)

151. Tufail, M.A.; Ayyub, M.; Irfan, M.; Shakoor, A.; Chibani, C.M.; Schmitz, R.A. Endophytic Bacteria Perform Better than Endophytic Fungi in Improving Plant Growth under Drought Stress: A Meta-Comparison Spanning 12 Years (2010–2021). *Physiol. Plant.* **2022**, *174*, e13806. [\[CrossRef\]](#)
152. Andreo-Jimenez, B.; Vandenkoornhuyse, P.; Lê Van, A.; Heutinck, A.; Duhamel, M.; Kadam, N.; Jagadish, K.; Ruyter-Spira, C.; Bouwmeester, H. Plant Host and Drought Shape the Root Associated Fungal Microbiota in Rice. *PeerJ* **2019**, *7*, e7463. [\[CrossRef\]](#) [\[PubMed\]](#)
153. Stenström, E. The Effects of Flooding on the Formation of Ectomycorrhizae in *Pinus sylvestris* Seedlings. *Plant Soil* **1991**, *131*, 247–250. [\[CrossRef\]](#)
154. Gavito, M.E.; Curtis, P.S.; Mikkelsen, T.N.; Jakobsen, I. Atmospheric CO<sub>2</sub> and Mycorrhiza Effects on Biomass Allocation and Nutrient Uptake of Nodulated Pea (*Pisum sativum* L.) Plants. *J. Exp. Bot.* **2000**, *51*, 1931–1938. [\[CrossRef\]](#) [\[PubMed\]](#)
155. Dong, Y.; Wang, Z.; Sun, H.; Yang, W.; Xu, H. The Response Patterns of Arbuscular Mycorrhizal and Ectomycorrhizal Symbionts under Elevated CO<sub>2</sub>: A Meta-Analysis. *Front. Microbiol.* **2018**, *9*, 1248. [\[CrossRef\]](#) [\[PubMed\]](#)
156. Habeeb, T.H.; Abdel-Mawgoud, M.; Yehia, R.S.; Khalil, A.M.A.; Saleh, A.M.; AbdElgawad, H. Interactive Impact of Arbuscular Mycorrhizal Fungi and Elevated CO<sub>2</sub> on Growth and Functional Food Value of *Thymus vulgare*. *J. Fungi* **2020**, *6*, 168. [\[CrossRef\]](#)
157. Baazeem, A.; Medina, A.; Magan, N. Effect of Acclimatization in Elevated CO<sub>2</sub> on Growth and Aflatoxin B<sub>1</sub> Production by *Aspergillus flavus* Strains on Pistachio Nuts. *Microorganisms* **2022**, *10*, 49. [\[CrossRef\]](#)
158. Garcia, M.O.; Ovasapyan, T.; Greas, M.; Treseder, K.K. Mycorrhizal Dynamics under Elevated CO<sub>2</sub> and Nitrogen Fertilization in a Warm Temperate Forest. *Plant Soil* **2008**, *303*, 301–310. [\[CrossRef\]](#)
159. Matamala, R.; Schlesinger, W.H. Effects of Elevated Atmospheric CO<sub>2</sub> on Fine Root Production and Activity in an Intact Temperate Forest Ecosystem: Root Production of Forest Growing at High CO<sub>2</sub>. *Glob. Chang. Biol.* **2000**, *6*, 967–979. [\[CrossRef\]](#)
160. Váry, Z.; Mullins, E.; McElwain, J.C.; Doohan, F.M. The Severity of Wheat Diseases Increases When Plants and Pathogens are Acclimatized to Elevated Carbon Dioxide. *Glob. Chang. Biol.* **2015**, *21*, 2661–2669. [\[CrossRef\]](#)
161. El-Shafey, N.M.; Marzouk, M.A.; Yasser, M.M.; Shaban, S.A.; Beemster, G.T.S.; AbdElgawad, H. Harnessing Endophytic Fungi for Enhancing Growth, Tolerance and Quality of Rose-Scented Geranium (*Pelargonium graveolens* (L'Hér) Thunb.) Plants under Cadmium Stress: A Biochemical Study. *J. Fungi* **2021**, *7*, 1039. [\[CrossRef\]](#)
162. Selim, S.; Abuelsoud, W.; Alsharari, S.S.; Alowaiesh, B.F.; Al-Sanea, M.M.; Al Jaouni, S.; Madany, M.M.Y.; AbdElgawad, H. Improved Mineral Acquisition, Sugars Metabolism and Redox Status after Mycorrhizal Inoculation are the Basis for Tolerance to Vanadium Stress in C3 and C4 Grasses. *J. Fungi* **2021**, *7*, 915. [\[CrossRef\]](#)
163. Bolan, N.S. A Critical Review on the Role of Mycorrhizal Fungi in the Uptake of Phosphorus by Plants. *Plant Soil* **1991**, *134*, 189–207. [\[CrossRef\]](#)
164. Guadarrama, P.; Álvarez-Sánchez, J.; Estrada-Torres, A. Phosphorus Dependence in Seedlings of a Tropical Pioneer Tree: The Role of Arbuscular Mycorrhizae. *J. Plant Nutr.* **2005**, *27*, 2159–2174. [\[CrossRef\]](#)
165. Garces, K.R.; Sage, H.E.; Christian, N.; Emery, S.M. *Epichloë* Increases Root Fungal Endophyte Richness and Alters Root Fungal Endophyte Composition in a Changing World. *J. Fungi* **2022**, *8*, 1142. [\[CrossRef\]](#) [\[PubMed\]](#)
166. Rodriguez, R.J.; Redman, R.S. *Fungal Life-Styles and Ecosystem Dynamics: Biological Aspects of Plant Pathogens, Plant Endophytes and Saprophytes*; Advances in Botanical Research; Elsevier: Cambridge, MA, USA, 1997; pp. 169–193, ISBN 9780120059249.
167. Rai, M.; Agarkar, G. Plant-Fungal Interactions: What Triggers the Fungi to Switch among Lifestyles? *Crit. Rev. Microbiol.* **2016**, *42*, 428–438. [\[CrossRef\]](#) [\[PubMed\]](#)
168. Eydoux, L.; Farrer, E.C. Does Salinity Affect Lifestyle Switching in the Plant Pathogen *Fusarium solani*? *Access Microbiol.* **2020**, *2*, acmi000114. [\[CrossRef\]](#)
169. Delaye, L.; García-Guzmán, G.; Heil, M. Endophytes versus Biotrophic and Necrotrophic Pathogens—Are Fungal Lifestyles Evolutionarily Stable Traits? *Fungal Divers.* **2013**, *60*, 125–135. [\[CrossRef\]](#)
170. Brader, G.; Compant, S.; Vescio, K.; Mitter, B.; Trognitz, F.; Ma, L.-J.; Sessitsch, A. Ecology and Genomic Insights into Plant-Pathogenic and Plant-Nonpathogenic Endophytes. *Annu. Rev. Phytopathol.* **2017**, *55*, 61–83. [\[CrossRef\]](#) [\[PubMed\]](#)
171. Baron, N.C.; Rigobelo, E.C. Endophytic Fungi: A Tool for Plant Growth Promotion and Sustainable Agriculture. *Mycology* **2022**, *13*, 39–55. [\[CrossRef\]](#) [\[PubMed\]](#)
172. Collinge, D.B.; Jensen, B.; Jørgensen, H.J. Fungal Endophytes in Plants and Their Relationship to Plant Disease. *Curr. Opin. Microbiol.* **2022**, *69*, 102177. [\[CrossRef\]](#)
173. Thomma, B.P.H.J. *Alternaria* spp.: From General Saprophyte to Specific Parasite. *Mol. Plant Pathol.* **2003**, *4*, 225–236. [\[CrossRef\]](#) [\[PubMed\]](#)
174. Johnson, N.C.; Graham, J.H.; Smith, F.A. Functioning of Mycorrhizal Associations along the Mutualism-Parasitism Continuum. *New Phytol.* **1997**, *135*, 575–585. [\[CrossRef\]](#)
175. Mandyam, K.G.; Jumpponen, A. Mutualism–parasitism paradigm synthesized from results of root-endophyte models. *Front. Microbiol.* **2014**, *5*, 776. [\[CrossRef\]](#)
176. Álvarez-Loayza, P.; White, J.F., Jr.; Torres, M.S.; Balslev, H.; Kristiansen, T.; Svenning, J.-C.; Gil, N. Light Converts Endosymbiotic Fungus to Pathogen, Influencing Seedling Survival and Niche-Space Filling of a Common Tropical Tree, *Iriarte deltoidea*. *PLoS ONE* **2011**, *6*, e16386. [\[CrossRef\]](#) [\[PubMed\]](#)

177. Redman, R.S.; Henson, J.M.; Rodriguez, R.J. Symbiotic Lifestyle Expression by Fungal Endophytes and the Adaptation of Plants to Stress: Unraveling the Complexities of Intimacy. In *Mycology*; CRC Press: Boca Raton, FL, USA, 2005; pp. 683–695, ISBN 9780824723552.
178. Hill, R.; Buggs, R.J.A.; Vu, D.T.; Gaya, E. Lifestyle Transitions in Fusarioid Fungi Are Frequent and Lack Clear Genomic Signatures. *Mol. Biol. Evol.* **2022**, *39*, msac085. [[CrossRef](#)] [[PubMed](#)]
179. Muszewska, A.; Stepniewska-Dziubinska, M.M.; Steczkiewicz, K.; Pawlowska, J.; Dziedzic, A.; Ginalski, K. Fungal Lifestyle Reflected in Serine Protease Repertoire. *Sci. Rep.* **2017**, *7*, 9147. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.