



Polyherbal and Multimodal Treatments: Kaempferol- and Quercetin-Rich Herbs Alleviate Symptoms of Alzheimer's Disease

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Simple Summary: Despite the well-documented pathophysiology of Alzheimer's Disease (AD), treatment options are limited in diversity and efficacy. Thus, the development of new treatments requires an extensive understanding of molecular pathways altered by drugs in development. In this review, we survey the literature regarding common herbal phytochemicals, kaempferol and quercetin, with a specific focus on their multiple mechanisms that alleviate the pathological underpinnings of AD. Here, we utilize the well-documented mechanisms of quercetin to propose a novel multimodal mechanism of kaempferol, and we discuss common herbal sources and the limitations of these potential treatments.

Abstract: Alzheimer's Disease (AD) is a progressive neurodegenerative disorder impairing cognition and memory in the elderly. This disorder has a complex etiology, including senile plaque and neurofibrillary tangle formation, neuroinflammation, oxidative stress, and damaged neuroplasticity. Current treatment options are limited, so alternative treatments such as herbal medicine could suppress symptoms while slowing cognitive decline. We followed PRISMA guidelines to identify potential herbal treatments, their associated medicinal phytochemicals, and the potential mechanisms of these treatments. Common herbs, including Ginkgo biloba, Camellia sinensis, Glycyrrhiza uralensis, *Cyperus rotundus*, and *Buplerum falcatum*, produced promising pre-clinical results. These herbs are rich in kaempferol and quercetin, flavonoids with a polyphenolic structure that facilitate multiple mechanisms of action. These mechanisms include the inhibition of A β plaque formation, a reduction in tau hyperphosphorylation, the suppression of oxidative stress, and the modulation of BDNF and PI3K/AKT pathways. Using pre-clinical findings from quercetin research and the comparatively limited data on kaempferol, we proposed that kaempferol ameliorates the neuroinflammatory state, maintains proper cellular function, and restores pro-neuroplastic signaling. In this review, we discuss the anti-AD mechanisms of quercetin and kaempferol and their limitations, and we suggest a potential alternative treatment for AD. Our findings lead us to conclude that a polyherbal kaempferol- and quercetin-rich cocktail could treat AD-related brain damage.

Keywords: Alzheimer's disease (AD); kaempferol; quercetin; flavonoids; traditional Chinese medicine; dementia

1. Introduction

Alzheimer's disease (AD) is a debilitating neurodegenerative disorder characterized by cognitive decline and memory impairment. AD could affect 152 million individuals by 2050 [1]. The progression of AD is influenced by multiple factors, including the accumulation of beta-amyloid plaques (A β) and the formation of neurofibrillary tangles (NFTs). The aggregation of A β plaques exacerbates the disease by impairing neuronal function and triggering neuroinflammation [2–5]. Oxidative stress and the presence of neurofibrillary



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). tangles (NFTs) also contribute to the aggregation of A β into senile plaques [6–14]. NFTs consist of hyperphosphorylated tau proteins that disrupt neuronal transport systems [15–21]. Neuroinflammation, in turn, exacerbates damage to neuronal integrity [22]. Symptoms of AD include memory loss, impaired learning, emotional changes, cognitive and speech deficits, shortened attention span, and impaired management of daily tasks [23–25].

Currently, the treatments available for AD are expensive and have minimal efficacy. Acetylcholinesterase inhibitors (AChEIs), including donepezil, and N-methyl-D-aspartate (NMDA) receptor antagonists, including memantine, are commonly prescribed for AD [26]. AChEIs inhibit the enzymatic degradation of ACh by inhibiting cholinesterase activity [27], while NMDA receptor antagonists limit calcium influx to prevent glutamate-induced cytotoxic cell death [28]. However, these drugs simply suppress symptoms and fail to halt disease progression [26], and only half of the population positively responds to these current treatments [29,30]. Herbal medicine boasts a well-documented history of safe and effective incorporation into traditional Asian diets [31,32]. Preclinical studies have demonstrated that these herbs can enhance cognitive and memory functions [33,34]. These herbs serve as dependable sources of phytochemicals, such as kaempferol and quercetin, that have limited side effects and could combat Alzheimer's disease [35–37]. Specifically, these flavonoids have anti-inflammatory, neuroprotective, and anti-degenerative effects [33,38–46].

The objective of this review is to elucidate the anti-AD mechanisms of kaempferol and quercetin. Here, we present a multimodal mechanism of action for kaempferol and quercetin in the treatment of Alzheimer's disease (AD). First, both flavonoids exert antioxidant effects, which stabilize cellular function and reduce neuroinflammation. Importantly, they also modulate PI3K/AKT signaling to limit A β and tau accumulation in toxic aggregates and enhance neuroplasticity by restoring BDNF signaling. These mechanisms ultimately improve memory and cognitive performance in AD patients. To our knowledge, this review represents the first comprehensive exploration of the literature that collectively shows kaempferol's potential to counteract both tau and A β via modulation of the PI3K/AKT/GSK-3 β pathway. Additionally, we propose that a polyherbal cocktail, incorporating sources rich in quercetin and kaempferol, could serve as an effective adjunctive or alternative treatment for AD. Finally, we explore the limitations of quercetin and kaempferol and discuss potential strategies for overcoming these challenges.

2. Materials and Methods

We collected data following the PRISMA guidelines for systematic review articles. The articles were sourced from PubMed, ScienceDirect, and Google Scholar, and data collection was conducted up until November 2023. We compiled a relevant list of articles to identify phytochemicals that have been studied for the treatment of Alzheimer's disease (AD) and their potential to induce therapeutic brain changes related to AD. Our search strategy initially yielded a total of 13,691 papers (13,688 from databases and an additional 3 from other sources). Of these, 2463 studies were screened based on their titles and abstracts, resulting in 378 articles that met the inclusion criteria (Figure 1). We included studies and reviews that explored the anti-AD mechanisms of phytochemicals and those that provided insights into the features of AD. The language was limited to English. Inclusion criteria required that articles discuss topics such as "Alzheimer's disease", "herbs", "kaempferol", "quercetin", "inflammation", "neuroprotection", "tau", and "A β ". The selected articles encompassed reviews, original research articles, and published clinical trials. Data extraction was carried out independently by a team of three investigators, considering factors such as the year of publication, article types, and the topic of herbs in relation to AD.



Figure 1. Flowchart depicting the article screening and selection process according to PRISMA guidelines.

3. Hallmarks of Alzheimer's Disease

Several features of AD, including A β plaque accumulation [47,48], tau hyperphosphorylation and neuroinflammation [49], and oxidative stress [50–53], have been identified as targets for drug development. Moreover, these deficits have been observed in studies with human patients [6,54–63]. This section will briefly explore the pathophysiology of AD, with a focus on the proposed molecular origins and outcomes of their aberrant activities.

While the origins are still debated, the literature greatly supports the roles of oxidative stress and neuroinflammation as critical drivers of neurodegeneration. Antioxidant deficits facilitate ROS production, driving oxidative stress via lipid peroxidation [57]. Consequently, mitochondrial energy production is impaired and pro-apoptotic signaling follows [57]. Glutamate-induced excitotoxicity could also facilitate oxidative stress [64–67]. Disrupted ROS clearance establishes the neuroinflammatory microglial and astrocytic hyperactivity [38,68–72] and favors neuronal signaling pathways that impair A β clearance [48,73,74]. Finally, proper mitochondrial function is required for A β clearance and can, in turn, maintain appropriate tau activity states [75].

Although normal A β levels can maintain regular neuronal function [76], failed A β clearance from the brain can expedite neurodegeneration by facilitating plaque accumulation and impairing neuronal communication [22,47,48,75,77–79]. Moreover, A β accumulation further promotes oxidative stress [80–83]. As A β plaques accumulate in the brain due to impaired clearance [84], overzealous astrocytic and microglial responses compound the neuroinflammatory environment by releasing pro-inflammatory factors, promoting neuronal apoptosis [6,49,85–89]. These findings were supported in postmortem tissue [6,54–56]. Finally, A β signaling significantly impairs LTP [90], facilitating neurodegeneration via low synaptic activity.

AD is one of the most common tauopathies [91]. A β plaque accumulation drives tau hyperphosphorylation [47,58,92–100], possibly by excess GSK-3 β signaling [101]. Likewise, tau hyperphosphorylation also compounds A β toxicity [102,103], which has been supported by PET imaging in humans with memory impairment and cognitive decline [60]. These studies demonstrate that A β toxicity is necessary for tau hyperphosphorylation [59,60]. Specifically, accumulating A β binds to NMDAR, generating excess calcium levels to acti-

vate calpain-mediated microtubule-associated protein cleavage [65,104,105]. These events impair mitochondrial function, invoking pro-apoptotic signaling [65,106]. Tau hyperphosphorylation dismantles axonal microtubules to degenerate the axon [15,107–110], impairing synaptic plasticity [102,103,111,112]. Hyperphosphorylated tau spreads throughout the hippocampus in AD models [113], and uptake may be mediated by clathrin-induced endocytosis [114]. Risk factors such as sleep apnea may potentiate the spread of tau in this manner [115]. Ultimately, these events result in neuronal death and compromise neuroplasticity, thereby driving neuroinflammation and impairing cognitive function.

4. Anti-AD Mechanisms of Quercetin and Kaempferol

Given the limited therapeutics available to AD patients, it is essential to explore alternative treatments, such as plant-derived phytochemicals. Flavonoids, including kaempferol and quercetin, belong to the class of polyphenols commonly found in various herbs. Notably, kaempferol and quercetin possess lipophilic properties [50], which facilitate their easy entry into cells. These phytochemicals are abundant, with an average daily consumption of approximately 23 mg of flavonoids in a typical diet [116,117]. Kaempferol and quercetin produce several beneficial properties, including anti-inflammatory, antioxidant, anti-A β , anti-tau, and pro-neuroplastic effects [37–39,57,74,118–128]. Moreover, they have demonstrated cognitive and memory-enhancing effects in animal studies [37]. Consequently, this section aims to delve into the commonly studied effects of these phytochemicals.

4.1. Quercetin

Quercetin, the most prevalent flavonoid, is found in several traditional medicinal herbs and is commonly found in fruits and vegetables, including berries, onions, and leeks [118,129–139]. Quercetin intake constitutes approximately 60–75% of total flavonols [140,141], and 25 mg of quercetin is found in the average diet [38]. Quercetin is commonly investigated for its potential anti-neurodegenerative efficacy and is considered safe [51,142]. Quercetin is a 15-carbon flavonoid with two benzene rings connected via a 3-carbon shape (Figure 2) [38,130,143].

Quercetin produces anti-inflammatory effects via multiple signaling pathways, including Nrf2, paraoxonase-2 (PON2), JNK, PKC, and NF-kB [51,118,128,144–147]. Quercetin dose-dependently protected HT22 hippocampal neurons from glutamate-induced apoptosis by limiting ROS production, impairing the calpain-mediated cleavage of cytoskeletal proteins, and preserving mitochondrial membrane potential [65]. Quercetin also inhibits NO release by inhibiting iNOS activity [33,38,148], which could reduce excess glutamate signaling and minimize the risk of glutamate-induced cytotoxicity in hippocampal neurons in a similar fashion to kaempferol and its derivatives [149]. Moreover, quercetin inhibits COX-2 and TLR4 activity to reduce inflammatory responses [6,39,148]. Interestingly, quercetin may have epigenetic mechanisms by inhibiting lysine acetyltransferase (KAT) activity [150,151] and increasing lysine deacetylase (KDAC) activity [152], suggesting that the flavonoid can bidirectionally regulate autophagy [153], neuroinflammation, and apoptosis [154]. Quercetin also inhibits acetylcholinesterase (AChE) [155], which can enhance alertness and cognitive function in AD patients.

The anti-A β effects of quercetin are well studied in AD and related models and have yielded promising therapeutic properties. The hydrophobic groups of quercetin can inhibit the formation of A β fibrils [120–123,156]. Chronic quercetin treatment also slowed A β aggregation by potentiating AMPK signaling and inhibiting mitochondrial ROS production, leading to improved memory and object recognition in APPswe/PS1dE9 [80,157]. Quercetin treatment also inhibits the BACE1-mediated cleavage of APP into A β by inhibiting NF-kB [74]. Consequently, mitochondrial membrane permeability is restored, and cellular survival is favored over oxidative stress [158]. This anti-neurodegenerative effect could be due to the free radical-quenching structure of the catechol group, reducing neuroinflammation, lipid peroxidation, mitochondrial stress, and DNA damage [38,51].

Elevated SOD, GPx, and Na⁺-K⁺ ATPase activity could also be due to quercetin's anti-A β effects [44,78].

In many studies, quercetin and its derivatives reduced tau hyperphosphorylation [23, 58,132,159]. In rodent HT22 hippocampal neurons, chronic quercetin treatment inhibited tau phosphorylation at four sites by reducing p-Cdk5 levels, limiting calpain activity, and dramatically reducing Ca^{2+} influx [58]. In 3xTgAD mice, chronic quercetin inhibited A β pathology, reduced NFT levels, and prevented astrocytic and microglial hyperactivity in the amygdala and hippocampus [132,160], showing that the anti-A β and anti-tau mechanisms of quercetin depend on its anti-inflammatory effects. Consequently, these mice demonstrated improved learning and memory and decreased anxiety [132], while combined exercise and quercetin treatment robustly improved spatial memory in AD rodents [161]. Studies also found that quercetin enhanced cell viability and morphology by reducing MDA and ROS levels and increasing antioxidant SOD and GSH activity [159,162], limiting NF- κ B signaling, restoring mitochondrial membrane potential to baseline, inhibiting tau hyperphosphorylation, and regulating Akt/PI3K/GSK-3β signaling pathway [159,163]. Taken together, these data show that quercetin has a multimodal mechanism of action in treating AD. Of note, the anti-tau and consequent pro-neuroplastic effect of quercetin is further explored in Section 5, but the primary anti-inflammatory, anti-A β , ant-tau, and pro-neuroplastic effects of this flavonoid are all dependent on each other.



Figure 2. The chemical structure of quercetin, deduced from PubChem [164].

4.2. Kaempferol

Kaempferol is a common 15-carbon polyphenol (Figure 3) that shares significant structural similarity with quercetin. It is one of the most common flavonoids and is found in a variety of common foods, including fruits and vegetables [129,130,165–170]. Multiple preclinical and clinical studies have supported the anti-AD activity of kaempferol [57,149,171–174]. Kaempferol has pro-neuroplastic, anti-A β , anti-tau, anti-inflammatory, and antioxidant properties [29,44,57,171,175–179]. Notably, kaempferol also inhibits AChE like quercetin [180], but this mechanism is beyond the scope of this review.

Like quercetin, kaempferol and its metabolites reduce inflammation and have potent antioxidant properties [181–184]. Kaempferol also directly modulates neuroinflammation by impairing microglial TLR4 and NF-kB signaling and inhibiting the release of NO, iNOS, PGE2, IL-1 β , TNF- α , and IFN- γ [167,185]. Kaempferol also reversed BBB damage [36,186,187]. Kaempferol can also modulate neuroinflammation by regulating epigenetic factors such as SIRT1, a subtype of KDAC [188–190]. Kaempferol also prevents cytotoxic damage to PC12 neurons by upregulating SIRT [191]. Other immune factors modulated by kaempferol include COX-2, lipoxygenases, prostacyclin, and leukotrienes [148,187,192–194]. Finally, kaempferol may reduce neuroinflammation via Nrf-2 signaling [185].

Like quercetin, kaempferol and its derivatives reverse Aβ-induced damage [29,120, 122,124,125,149,195]. Kaempferol-3-O-rhamnoside (K-3-Rh), a kaempferol derivative, lim-

ited total A β burden and toxicity by disrupting β -sheet formation and impairing A β plaque formation in human SH-SY5Y cells [195,196]. However, kaempferol antagonized fibrilization with lower potency compared to quercetin and morin [120,122]. In rodent neuroblastoma cells, kaempferol 3-O-(6''-acetyl)- β -glucopyranoside (KAG) robustly inhibited A β -mediated cytotoxic cell death and ROS generation [149]. KAG reversed A β -mediated oxidative stress and increased cell survival by regulating caspase-3, Bax, and Bcl-2 signaling [44,64,149,197–200]. Kaempferol dose-dependently and sex-dependently limited A β -induced mitochondrial toxicity in neurons, improving rodent memory in the Y-maze test [57,134,201]. Of note, studies regarding kaempferol's direct influence on tau are limited; thus, more research is necessary. However, due to its similar phenolic structure to quercetin [165,166], we hypothesize that kaempferol could also reduce tau hyperphosphorylation.



Figure 3. The chemical structure of kaempferol, deduced from PubChem [202].

5. Kaempferol, Quercetin, and Neuroplasticity

The aberrant brain changes described in Section 3 can impair memory and cognitive function by creating deficits in neuroplasticity. Thus, future AD treatments should also be designed to directly target signaling pathways that can counteract the etiologies of AD. Specifically, we identified the PI3K/AKT signaling pathway as a critical candidate to counteract neurodegeneration. Several studies have suggested that flavonoids can alleviate learning and memory deficits by targeting this signaling pathway [29,203–205]. However, other pathways, including the MAPK-ERK1/2 cascade [206], have also been proposed and outlined in a recent review [207]. In this section, we will first explore the impact of A β -and tau-mediated neuroinflammation on synaptic plasticity-related neuronal signaling. We will support the necessity of the PI3K/AKT/GSK-3 β pathway in AD treatments and investigate the potential roles of kaempferol and quercetin in improving memory and cognition through this pathway.

5.1. Neuroplasticity Deficits in AD

An ideal AD treatment should enhance the expression of plasticity-related genes such as BDNF, a neurotrophic factor that regulates neuronal plasticity and survival [208–214]. BDNF signaling begins with its binding to the receptor, Trk β , activating signaling via a variety of pathways like PI3K/AKT [211,215]. Then, AKT or protein kinase B (PKB) [216] can activate the CREB-mediated transcription of BDNF [217,218]. Since Trk β receptors mediate the pro-neuroplastic effects of BDNF [219], AD drugs must produce a direct or indirect effect on the receptor.

BDNF deficits increase the risk of AD development [220], and BDNF dysfunction due to impaired PI3K and AKT signaling can expedite neurodegeneration [7,41,221–223]. The PI3K/AKT signaling pathway has multiple functions, including regulating synaptic plasticity, glucose processing, cell cycle progression, cell proliferation, survival, and apoptosis [167,175,224–226]. Moreover, this pathway may protect neurons from A β toxic-

ity [224], oxidative stress [227], and neuroinflammation [217]. GSK-3 β is downstream of PI3K/AKT, and A β can specifically lead to its hyperactivity [7]. However, BDNF and CREB are also vulnerable to A β signaling [228] as CREB is regulated by the PI3K/AKT/GSK-3 β pathway [211,212,215,229–231].

Thus, the Aβ-mediated signaling cascade that degenerates the neuron is as follows (Figure 4A): Aβ binding to NMDAR inhibits PI3K/AKT signaling by activating GSK-3β-mediated tau hyperphosphorylation and CREB downregulation [93,97,210,211,223,229, 230,232–237]. Consequently, the impaired CREB-mediated transcription of BDNF genes decreases plasticity and facilitates plaque accumulation, as demonstrated in postmortem tissues from humans and human neuronal cells [209,210,229,232]. The absence of protective BDNF and PI3K/AKT activity facilitates the caspase-mediated pro-apoptotic signaling cascade [6,224], degenerating the neuronal circuitry, while tau dissociation from microtubules breaks down the neuronal cytoskeleton [7,233,238–242].

However, future AD treatments could reverse this toxic signaling via the following mechanism: A drug must either directly activate Trk β or should do so indirectly by enhancing BDNF transcription [210]. The drug can either directly activate PI3K and/or AKT, which would ultimately inhibit GSK-3 β via the phosphorylation of its Ser9 residue [224]. In turn, AKT can also inhibit caspase-9 and Bcl-3 to inhibit pro-apoptotic signaling [243–246]. One study showed that the GSK-3 β inhibitor, AR-A014418 (ARA), inhibited BACE1-mediated APP cleavage into A β proteins in rodents [48], supporting the necessity of a GSK-3 β -inhibiting drug for the treatment of AD. Finally, GSK-3 β inhibition also reversed oxidative stress [93,247]. In short, the PI3K/AKT pathway can not only reverse neuroinflammation but can also counteract A β -mediated tau hyperphosphorylation by inhibiting GSK-3 β .

5.2. Quercetin and Kaempferol Resolve AD-Related Plasticity Deficits

The multimodal mechanisms of kaempferol and quercetin collectively slow neurodegeneration by combating the impairments that are illustrated in Figure 4A and are described in Table 1. Specifically, the restoration of proper PI3K/AKT signaling will greatly improve synaptic plasticity deficits in AD [7]. While quercetin's interaction with each component of this signaling pathway has already been documented [7], kaempferol's mechanisms are still unclear. However, since kaempferol's structure is similar to that of quercetin [165], we propose that kaempferol has a nearly identical mechanism with respect to the signaling pathway in this subsection. Finally, we will propose the potential outcomes of these molecular interactions.

Table 1. Kaempferol and Quercetin and molecular interactions with select molecules relevant to neuroplasticity in AD. These affinity or potency values are deduced from molecular docking studies (affinity) and competition assays (IC50; potency) or were indirect interactions evidenced in the literature. Docking scores (DS) of 5 or higher indicate the high affinity of a compound for the protein of interest [248,249]. Or, affinity from docking studies may be expressed as binding energies (BE) in -kcal/mol. The more negative the value, the higher the binding affinity. If studies have not supported direct binding to a certain target, the affinity column is noted as "Indirect".

Molecular Target	Phytochemical	Mechanism	Affinity (DS, BE, or IC50)	References
GSK-3β	Kaempferol	Inhibit	4.6 (DS, mice); −7.9 kcal/mol (human brain docking) −9.2 kcal/mol (zebrafish)	[243,250,251]
	Quercetin	Inhibit	5.64 (DS); –8.8 kcal/mol (human brain docking) –9.0 kcal/mol (zebrafish)	[243,250,251]
A 0	Kaempferol	Inhibit	Indirect	[171]
Ар	Quercetin	Inhibit	Indirect	[252]
BACE1	Kaempferol	Inhibit	$IC50 = 14.7 \ \mu M$	[253,254]
	Quercetin	Inhibit	$IC50 = 5.4 \ \mu M$	[253,254]

Molecular Target	Phytochemical	Mechanism	Affinity (DS, BE, or IC50)	References
Tau	Kaempferol	Inhibit hyperactivation	Indirect	[47]
	Quercetin	Inhibit hyperactivation	Indirect	[255]
DIOK	Kaempferol	Activate	5.19 (DS, neurons)	[256]
РІЗК	Quercetin	Activate	7.04 (MD, neurons)	[256]
AKT1	Kaempferol	Activate	5.13 (MD, neurons); -9.3 kcal/mol	[256,257]
	Quercetin	Activate	5.03 (MD, neurons), —9.4 kcal/mol; —7.96 kcal/mol	[213,256,257]
BDNF Kaempferol Upregulate Quercetin Upregulate	Kaempferol	Upregulate	Indirect	[258]
	Upregulate	Indirect	[252]	
CRER	Kaempferol	Activate	Indirect	[211]
CKEB	Quercetin	etin Activate	Indirect	[252]
NMDAR -	Kaempferol	Reverse Aβ binding	-10.84 kcal/mol	[259]
	Quercetin	Reverse Aβ binding	Indirect	[255,260]
	Kaempferol	Activate	Not Found	[188,189]
HDAC	Quercetin	Activate	$IC50 = 105.1 \ \mu M$	[261]
AChE	Kaempferol	Inhibit	-10.26 kcal/mol; between -8.6 and -9.22 kcal/mol	[259,262,263]
	Quercetin	Inhibit	$\frac{-7.9 \text{ kcal/mol;}}{\text{IC50} = 4.59 \pm 0.27 \mu\text{M}}$	[155,263]

Table 1. Cont.

Molecular docking studies suggested that quercetin can bind PI3K, AKT, and GSK3β [213,250,255–257,260,264,265]. Specifically, quercetin can bind to PI3K [256], consequently activating AKT signaling [265], or quercetin can directly bind to AKT [257]. In preclinical studies, quercetin reduced GSK-3 β activity, which decreased tau hyperphosphorylation and reduced pro-apoptotic signaling [7,38,159]. Quercetin treatment in rodents also increased BDNF, Trkβ, PI3K, and AKT expression [243,266]. Consequently, quercetin enhanced neurite outgrowth in hippocampal neurons [36] and ameliorated the stress-induced downregulation of CREB and BDNF [40], suggesting that quercetin could potently replenish neuroplasticity in the AD brain. Moreover, quercetin inhibited A β by restoring Trk β signaling and CREB-mediated BDNF transcription, increasing the viability of SH-SY5Y cells [252]. Finally, quercetin's dual pro-neuroplastic and anti-inflammatory effects may also be related to the quercetin-mediated downregulation of BACE1 expression via the inhibition of NF-kB [253,254,264,267]. Taken together, these data suggest that quercetin antagonizes A β -induced GSK-3 β signaling relative to tau by activating the PI3K/AKT pathway and directly inhibiting GSK-3ß [7,225,241,255,256,260]. Consequently, proper BDNF levels can be restored to replenish neuronal plasticity in the AD brain. Similar chemicals, such as epigallocatechin-3-gallate (EGCG), attenuated tau hyperphosphorylation in a similar mechanism [23,268–270]. Thus, quercetin clearly has dual neuroprotective and pro-neuroplastic mechanisms in cells [33,65,252], and the clinical outcomes of quercetin's pro-neuroplastic mechanisms were supported by its memory and cognition-boosting effects in rodent models of AD and Parkinson's disease [23,38,44,271–276]. Select molecular targets of quercetin are described in Table 1.

Kaempferol may have similar pro-neuroplastic mechanisms to quercetin, and some of its molecular targets are outlined in Table 1. First, kaempferol improved hippocampal plasticity following traumatic brain injury in young rodents [277] and improved memory in rodents [29,57] and Drosophila [173]. Moreover, kaempferol dose-dependently maintained cell viability following A β treatment in multiple studies [29,149,195,248]. This could be due to kaempferol's inhibition of BACE1-mediated A β synthesis [253,254] or the activation of the

PI3K/AKT signaling pathway, enhancing CREB-mediated BDNF transcription [175,211,258]. Although one molecular docking study suggested that kaempferol may have minimal affinity for GSK-3β [250], kaempferol likely inhibits GSK-3β indirectly by first binding and activating PI3K [256] or AKT [175,185,257]. Via this mechanism, kaempferol prevents tau hyperphosphorylation, protecting neuronal morphology and function [47,278–281]. Then, AKT can activate CREB-mediated BDNF transcription [217]. Supporting this pro-neuroplastic mechanism, kaempferol and its metabolite, kaempferide, produced similar effects that resulted in Trkβ signaling [171,210] and enhanced BDNF expression in Aβ-treated mice [243]. Taken together, these data suggest that kaempferol enhances neuroplasticity to reverse Aβ damage by activating the PI3K/AKT cascade, which potentiates CREB-mediated BDNF transcription. However, kaempferol produces the opposite effect on this signaling pathway in microglial cells [167] and cancer cells [282]. Thus, kaempferol's effects on the PI3K/AKT signaling cascade are dynamic and depend on cell lineage.

Despite the lack of literature demonstrating a direct modulation of tau by kaempferol, there is plenty of evidence to support the possibility that kaempferol inhibits tau hyperphosphorylation via the PI3K/AKT pathway and by antagonizing A β -mediated GSK-3 β signaling [29,149,195]. This mechanism prevents neuronal degeneration and a loss of synaptic plasticity. Thus, the pro-neuroplastic effect of kaempferol requires the inhibition of GSK-3 β and CREB phosphorylation. Remarkably, a recent molecular docking study suggested that kaempferol could bind to NMDAR [259]. However, in vivo studies are still required to confirm this effect.

These data suggest a clear anti-AD mechanism of quercetin and kaempferol, as outlined in Figure 4B. First, quercetin and kaempferol could enter the cell cytoplasm due to their lipophilic polyphenolic structure. Quercetin and kaempferol scavenge ROS and activate PI3K/AKT signaling to inhibit GSK-3β. Specifically, they can bind directly to PI3K or AKT to activate protective signaling, inhibiting GSK-3 β and preventing tau hyperphosphorylation. This signaling cascade reduces the formation of NFTs in the AD brain. GSK-3β inhibition can also antagonize Aβ-NMDAR interactions. Thus, downstream pro-apoptotic signaling mediators are also inhibited by quercetin and kaempferol treatment. Due to reduced NFT and amyloid plaque formation, microglial hyperactivity decreases in the absence of the burden of clearance. Thus, progressive neuroinflammatory signaling is slowed, allowing surrounding neuronal synapses to survive. After chronic quercetin treatment, progressive elevations in BDNF release rebuild damaged synapses by favoring neurotrophic signaling over cytotoxic A β signaling, improving memory and cognition. Of note, molecular docking studies have not supported the possibility that kaempferol and quercetin can directly bind to tau protein, supporting their indirect inhibitory mechanism via GSK-3 β inhibition. Taken together, kaempferol and quercetin share multiple mechanisms that slow AD progression by first limiting ROS activity, NFT aggregation, and Aβ-mediated toxic signaling, slowing neurodegeneration.



Figure 4. (**A**) Neuroplasticity deficits accelerate AD progression and must be treated. Impaired PI3K-AKT signaling facilitates GSK3 β -mediated phosphorylation of tau. A β may potentiate tau hyperphosphorylation via GSK3 β . (**B**) Kaempferol and quercetin (K/Q) invoke the PI3K/AKT pathway to antagonize A β and reduce tau hyperphosphorylation in neurons. As a result, neuroplasticity is increased in the AD brain [283].

6. Quercetin and Kaempferol in Common Herbs

Although data on the co-treatment of quercetin and kaempferol are still somewhat limited, the abundance of both compounds in several common herbs requires the investigation of the synergistic effects of both flavonoids, in addition to their interactions with other herbal phytochemicals. Flavonoid-rich herbs are commonly employed in traditional Chinese medicine (TCM), in which an emphasis is placed on the utility of natural treatments. Moreover, these herbs are generally safe for consumption [224]. Kaempferol is the second most common flavonoid in traditional medicinal herbs, following quercetin [225,284]. Other reviews have assessed the efficacy and safety of natural medicine in the treatment of neurodegenerative diseases [7,224], highlighting the potential medicinal properties of herbs in treating AD. Flavonoids are commonly found in herbs such as *Schima wallichii* Korth, *Maesa membranacea, Ginkgo biloba,* and many more [175,225,278]. These phytochemicals could work synergistically with each other and with other herbal components to invoke anti-AD effects. Thus, we explore common herbal sources of kaempferol and quercetin, describe the anti-AD mechanisms of herbs, and propose a design for a future AD treatment based on the current evidence of these effects.

Ginkgo biloba is a quercetin- and kaempferol-rich herb proposed to treat AD [285]. G. biloba improves memory and cognition by inhibiting ROS, facilitating hippocampal neuron proliferation, halting Aß plaque accumulation, and reducing tau hyperphosphorylation [47,286–288]. Moreover, this effect is associated with reduced GSK-3ß activity and the increased expression of PSD-95 and synapsin-1 [47]. As seen with kaempferol and quercetin alone, G. biloba potentiates PI3K/AKT relative to CREB signaling to promote neuroplasticity [287,289–293]. Hippophae rhamnoides extracts are also rich in quercetin and kaempferol, and they enhanced neuronal differentiation and neurite outgrowth via PI3K/AKT and ERK signaling [294,295]. However, clinical trials have revealed the inconsistent efficacy of *G. biloba* on cognition and other AD-related parameters [296]. Camellia sinensis is another kaempferol- and quercetin-rich herb commonly grown to produce black and green tea [297,298]. C. sinensis extracts improved spatial memory and reduced hippocampal A^β fibrillization in AD rodents and had greater antioxidant effects compared to other herbs [298,299]. Kaempferol and its derivatives are found in the leaves of Maesa membranacea, Schima wallichii Korth, Carthamus tinctorius, Panax ginseng, and several other herbs [175,188,225,278]. S. wallichii was neuroprotective due to the promotion of hippocampal and cortical AKT signaling [175], and M. membranacea could protect H202treated SH-SY5Y cells [225] and hippocampal tissue [300] via the same pathway due to their kaempferol abundance. C. tinctorus is rich in kaempferol, produces a similar effect, and invokes protective AMPK signaling [188]. Finally, recent studies also suggested that other herbs such as Morenga oleifera, Cuscuta chinensis, Allium cepa, Litchi chinensis, Prakia roxburghii, Radix astragali, Acoritatan Fagopyrum tataricum, Carthami flos, Punica granatum, and Cyperi *rhizoma* [251,257,264,301–305] may also be great sources of kaempferol and/or quercetin and produce anti-AD effects. Their medicinal properties and expression of kaempferol and quercetin are outlined in Table 2.

Polyherbal cocktails, such as Chaihu shugan san (CSS) and Huangqi Sijunzi (HQSJDZ), could treat AD and its risk factors. CSS is abundant in kaempferol and quercetin and contains herbs such as *Glycyrrhiza uralensis*, *Cyperus rotundus*, and *Buplerum falcatum* [256]. Specifically, the antidepressant effect of CSS is mediated by increased PI3K/AKT/BDNF signaling and decreased GSK-3 β and IL-2 activity [256], suggesting that polyherbal cocktails may be protected from AD development. HQSJDZ, rich in kaempferol and quercetin, had cholinergic, anti-inflammatory, and anti-GSK-3 β effects [278,306]. Moreover, a cocktail of *C. sinensis*, Hypericum perforatum, and Bacopa monnieri produced robust antioxidant effects compared to single-herb treatment [298]. These data suggest that polyherbal treatment may be superior to single-herb therapy.

Due to the well-documented effects of quercetin and kaempferol on A β , GSK-3 β , PI3K/AKT, and multiple pro-inflammatory molecules, it is possible that both phytochemicals, given their abundance, contribute vastly to the anti-AD effects of several herbs. Such

herbs include *Ginkgo biloba*, *Camellia sinensis*, *Glycyrrhiza uralensis*, *Cyperus rotundus*, and *Buplerum falcatum*. The herbal sources outlined in Table 2 may also be great additions to the treatment protocol that can enhance the dietary intake of kaempferol and quercetin. According to the practice of TCM, it is possible that a multi-herb cocktail containing varying amounts of these herbs could alleviate AD symptoms, as seen with current medications, but it may also halt progression relative to a unique multi-modal mechanism. Multiple studies have suggested that the synergistic effects of polyherbal treatments produce greater anti-AD efficacy compared to single-herb treatment [256,278,298]. Thus, the research and development of future AD drugs should consider the applications of these common herbs in future drug cocktails. On the other hand, since clinical trials featuring *Ginkgo biloba* extracts have demonstrated controversial results on the progression of AD [296], single-herb treatments may be insufficient to treat AD.

Table 2. Plant sources of kaempferol and quercetin and/or their metabolites and a description of reported herbal health effects.

Species Name	Kaempferol	Quercetin	Example Health Effects	Reference
Ginkgo biloba	+	+	Memory and cognition improvement	[285,296,307,308]
Camellia sinensis	+	+	Improved memory and antioxidant effects	[297–299]
Maesa membranacea	+	+	Neuroprotective	[175,188,225,278]
Schima wallichii Korth	+	_	Neuroprotective	[175,187,225,278]
Carthamus tinctorius	+	+	Neuroprotective	[175,187,225,278,309]
Panax ginseng	+	+	Neuroprotective	[175,187,225,278,310]
Morenga oleifera	+	+	Memory improvement	[300-302]
Cuscuta chinensis	+	+	Memory improving, Neuroprotective, Hepatoprotective, Immunomodulatory	[311]
Allium cepa	+	+	Anti-inflammatory	[312,313]
Hippophae rhamnoides L.	+	+	Anti-inflammatory	[294,295]
Litchi chinensis	+	+	Neuroprotective	[303,314]
Prakia roxburghii	-	+	Neuroprotective	[304]
Radix astragali	+	+	Neuroprotective	[213]
Fagopyrum tataricum (L.)	+	+	Decrease neurotoxicity	[251]
Carthami flos	+	+	Anti-ischemic	[213]
Punica granatum	+	+	Anti-inflammatory	[264,315]
Cyperi rhizoma	+	+	Antidepressant	[257]

7. Limitations of Kaempferol and Quercetin Treatment

7.1. Bioavailability

Despite the promising effects of these herbs and flavonoids in AD treatment, low bioavailability and blood-brain barrier (BBB) permeability are common obstacles interfering with drug delivery to the brain [316–318]. Thus, structural manipulations are commonly required to improve the bioavailability of flavonoids. Moreover, the varying dietary intake of macromolecules like fats and carbohydrates also impacts BBB permeability relative to polyphenols [44]. Other factors, such as aging or diagnosis with AD, may increase BBB permeability to peripheral chemicals [39,319–321]. However, tau hyperphosphorylation

and astrocytic hyperactivity invoke neuroinflammatory signaling that damages BBB integrity and increases its permeability [112,322–324]. The limited BBB permeability may also explain the lack of clinical trials in humans [68].

Despite its lipophilicity and easy oral administration in common foods, quercetin treatment for AD may be challenged by its limited bioavailability relative to the brain [3,258]. Since quercetin absorption is predominantly mediated by the small intestine, it is vulnerable to extensive first-pass metabolism [133,258,325]. While its distribution was evidenced in the plasma, liver, heart, spleen, kidneys, and lungs, quercetin levels were non-detectable in the rat brain [326,327]. Hence, it has around 65% BBB permeability [321,328] and is absorbed in the stomach with 24–53% bioavailability [329]. The P-glycoprotein transporter, which is a BBB efflux transporter, has a high affinity for free, unaltered quercetin and greatly reduces its bioavailability by pumping quercetin away from the brain [51,330]. While in vitro studies showed the promising antioxidant effects of quercetin, most studies in animal models have demonstrated limited efficacy [3,331]. These data show that quercetin's limited bioavailability could debilitate anti-AD effects [258].

Chemical modifications are necessary to ensure quercetin distribution to the brain, as some metabolites may also have higher efficacy than quercetin alone. For instance, quercetin–glucoside conjugation enhanced its bioavailability [129]. Quercetin glycosides are commonly available in fruits and vegetables, improving its delivery to the CNS [51,332]. Glucuronidation in the liver also increased the distribution of quercetin to the brain in oxidative stress models [23]. Moreover, in vivo studies showed that lipid nanoparticle-loaded quercetin enhances its entry into the brain [39,44,146,158,163,333,334]. Moreover, quercetin loading into selenium nanoparticles improved brain distribution and anti-A β mechanisms [335]. However, excess selenium levels in the body can produce oxidative stress [336,337], potentially limiting the clinical efficacy of this approach.

Like quercetin, free kaempferol generally has low oral bioavailability due to metabolic degradation [324,338,339]. Kaempferol is generally slowly absorbed in the GI tract and can be distributed to several tissues [326,340], suggesting that the primary limitation of kaempferol treatment is limited bioavailability. However, several modifications to improve its BBB permeability have been proposed. First, nanoparticle loading also improves kaempferol bioavailability [194,334,341–344], and kaempferol-sugar conjugates also demonstrate superior protective efficacy [36]. For instance, nanoparticle-loaded kaempferol has more robust anti-inflammatory effects than kaempferol alone [68]. Clinical trials revealed that quercetin had superior memory-modulating activity in AD patients compared to healthy elderly controls [345–347], suggesting that the increased BBB permeability in AD may, in turn, improve flavonoid bioavailability and efficacy in neurodegenerative brains. Several other forms of delivery have been proposed for both flavonoids, including gold-infused nanoparticles [348,349], multi-targeted drugs [350], extracellular vesicles [351], and intranasal administration [352]. Finally, other proposed nanoformulation delivery systems include nanomatrixes, nanoemulsions, nanostructured lipid carriers, and nanocomplexes [343,344,353].

7.2. Adverse Health Effects and Other Limitations

Most studies show promising medical benefits for kaempferol and quercetin and suggest that they are safe in a variety of doses. For example, quercetin is included in the Food and Drug Administration's Generally Recognized as Safe (GRAS) list for supplemental use of up to 500 mg per serving in foods and beverages [129,354]. However, flavonoids' clinical efficacy may also be limited by adverse effects [329]. While the Ames test suggested that quercetin could have carcinogenic properties, most studies have opposed this finding and suggested that quercetin is safe [355]. One study suggested that high-dose quercetin treatment reduced neuronal survival, induced oxidative stress, and inhibited AKT [356]. Thus, physicians should carefully manage the abundance of quercetin in the AD patient's diet to maintain its proper anti-degenerative effects. Moreover, the efficacy of quercetin may be limited in AD patients who are also diagnosed with leukemia, as quercetin inhibits the PI3K/AKT signaling pathway in HG3 cells [282]. It is possible that, since most dietary quercetin is distributed to peripheral sites, lower concentrations in the brain may decrease its efficacy in AD.

Although kaempferol is most likely safe to consume [339] and most studies showed low toxicity in mice [357–359], some studies have reported concerns about potential mutagenic effects in people with iron and folic acid deficiencies [338,339,360]. Since the excess inhibition of GSK-3 β may produce toxic effects in cells [233], kaempferol's low-affinity GSK-3 β interactions may underlie its generally low toxicity. In a 4-week randomized, double-blind clinical trial, participants were divided into a group that received 50 mg of kaempferol daily and a placebo group; kaempferol was reported as mostly safe, but the small sample size of 24 in each group limits this study [359]. Overall, the majority of work on the herb suggests it to be safe, even in high doses, but more clinical trials are highly recommended.

8. Discussion

Since AD still lacks a true cure, and currently available medications are insufficient to halt disease progression, the field has sought out multimodal treatments for AD. However, little progress in drug development has been made in recent decades, necessitating new alternative treatments. Thus, the objective of this review was to deduce the anti-AD mechanisms of kaempferol and quercetin. These phytochemicals were selected for multiple reasons, including their abundance [38,116] and their multimodal mechanisms (Figure 5) that include antioxidant, anti-inflammatory, pro-neuroplastic, and neuroprotective effects. Thus, quercetin and kaempferol may treat Alzheimer's disease, and we aimed to explore their anti-amyloidogenic, antioxidant, anti-inflammatory, anti-tau, and pro-neuroplastic mechanisms [6,29,38,39,51,127,128,149,159,167,361]. In turn, phytochemicals may not only reduce AD symptoms [29,33,132] but also delay the progression of the disorder. Of note, the efficacy of these flavonoids to produce the effects outlined in this review depends on any chemical modifications that may occur throughout the absorption and distribution of phytochemicals to the brain.

Perhaps the most significant contribution of this review is the complex anti-degenerative mechanism of kaempferol. We utilized the available literature to show that kaempferol's dual anti-tau and anti-A β mechanisms are due to its modulation of the PI3K/AKT/GSK-3 β signaling pathway. Both phytochemicals resolve oxidative stress by increasing antioxidant levels and inhibiting ROS signaling [119]. Meanwhile, they halt inflammatory signaling [29,38] to commence a neuroprotective effect. Then, resolved microglial and astrocytic activity facilitates proper A β clearance from the brain [6] and reduces continued neuronal damage due to the neuroinflammatory environment [122,188,195]. The modulation of PI3K/AKT/GSK-3 β and Trk β /BDNF signaling potentiates neuroplasticity and protects neurons from insults like A β [10,240], decreasing tau hyperphosphorylation and preserving the neuronal cytoskeletal structure. These phytochemicals, in turn, protect neuronal networks [33,40], improving memory and cognitive function in AD patients. Other flavonoids with heterocyclic structures [362], including morin [363–366], rutin [367,368], and luteolin [369–371], share many similar anti-AD properties relative to kaempferol and quercetin. However, rutin [368] failed to increase BDNF levels, like kaempferol and quercetin.

Due to the superior efficacy of polyherbal treatments, such as HQSJDZ and CSS [256,278,298], we proposed that polyherbal treatment, containing quercetin- and kaempferol-rich herbs like *Ginkgo biloba*, *Camellia sinensis*, *Glycyrrhiza uralensis*, *Cyperus rotundus*, and *Buplerum falcatum* may produce superior anti-AD efficacy compared to single-herb supplements. Recent studies also suggested that herbs such as Morenga oleifera, Cuscuta chinensis, Allium cepa, Hippophae rhamnoides, Litchi chinensis, Prakia roxburghii, Radix astragali, Fagopyrum tataricum, and Carthami flos [251,294,301–304] may also be candidates for polyherbal treatment. However, a recent review noted that kaempferol and quercetin are widely available in hundreds of herbs, and it is possible that they may not

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be as abundant as other phytochemicals in some species [372], supporting the necessity of polyherbal treatment to obtain biologically effective concentrations.

Figure 5. A graphical summary of the underlying mechanisms behind AD progression (pathogenesis), the proposed mechanisms of kaempferol and quercetin (K/Q), where K/Q represents kaempferol and quercetin, and the impact of these molecular changes on behavior and disease progression (outcomes). Each category is presented in a top-to-bottom chronological order.

As previously mentioned, clinical trials suggest that kaempferol and quercetin could treat AD in humans [135,346,347,373,374], but single-herb treatment was unsuccessful in clinical trials [296]. Future trials should assess bioavailability-enhancing delivery methods for quercetin and kaempferol. However, recent studies also suggested that both quercetin and kaempferol have the ability to maintain and protect BBB integrity [375–379]. This could possibly be due to their anti-inflammatory properties that could be invoked if they reach the brain. Of course, clinical trials should continue to assess the efficacy of herbal sources in AD-related symptoms. However, the misuse of herbal treatments may produce side effects, including gastrointestinal discomfort, insomnia, and tachycardia [298]. Thus, studies assessing these side effects are limited and require further investigation [36,37]. Nonetheless, these natural herbs are generally considered safe, and toxic effects are uncommon [51,116,142]. Finally, an investigation of interactions between these polyphenols and other drugs commonly prescribed to AD patients is required.

Although the data presented in this review showcase the great potential of these herbs in AD treatment, a few limitations have impacted this review. Specifically, studies investigating the tau hyperphosphorylation-inhibiting mechanisms of these herbs may be limited due to the rapid dephosphorylation of the protein in postmortem AD tissues [15,279]. Moreover, the abundantly described bioavailability limitations of both herbs critically limit the efficiency of human studies. This could be one reason underlying the lack of kaempferol and quercetin's clinical efficacy to date. Clinical trials investigating compounds that increase the bioavailability of these phytochemicals are still needed. Since quercetin and kaempferol are naturally abundant in the average diet, future clinical trials can be easily conducted. Finally, while molecular docking studies show the potential pharmacodynamic interactions between kaempferol/quercetin and the outlined pro-neuroplastic targets, these approaches are merely estimates of binding affinity based on the crystal structures of the target protein and the molecular structures of the ligand, and they could be vulnerable to mispredictions [380]. Thus, future studies must either employ competition assays or ligand inhibitor/antagonist studies to confidently elucidate the true affinity of kaempferol and quercetin for the targets of interest. Nonetheless, recent data support the exciting potential of kaempferol and quercetin to slow the progression of AD and alleviate the symptoms.

9. Conclusions

Kaempferol and quercetin clearly exhibit multimodal mechanisms that halt AD progression and alleviate symptoms. Given the multifaceted nature of AD pathogenesis, future treatments need to adopt a multimodal approach that targets the A β -tau signaling pathway via the modulation of the PI3K/AKT/GSK3 β signaling cascade, leading to a pro-neuroplastic effect via enhanced BDNF signaling. To our knowledge, our review demonstrates how kaempferol and quercetin address various aspects of AD, including neuroinflammation, oxidative stress, reduced plasticity, and A β and tau signaling. Notably, our review is the first to propose that kaempferol can mitigate both tau hyperphosphorylation and A β toxicity by directly targeting the PI3K/AKT/GSK3 β pathway. Additionally, we suggest that polyherbal cocktails rich in kaempferol and quercetin may yield robust anti-AD effects, and we identified potential herbal sources of kaempferol and quercetin. Finally, we discuss the limitations that currently impede the efficacy of kaempferol/quercetin treatment, and suggest potential adjustments to circumvent these challenges. Together, these changes can improve the anti-AD efficacy of natural flavonoids and could be ideal adjunctive or alternative treatments to currently available drugs.

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Herbs Manuscript Abbreviations

Αβ	amyloid beta
AChEIs	acetylcholinesterase inhibitors
AChE	acetylcholinesterase
ACh	acetylcholine
AD	Alzheimer's disease
AKT	protein Kinase B

AMPK	AMP-activated protein kinase
APP	amyloid precursor protein
BACE1	beta-site APP cleaving enzyme 1
Bax	bcl-2-like protein 4
BBB	blood-brain barrier
BDNF	brain-derived neurotrophic factor
Cdk5	cyclin-dependent kinase 5
p-Cdk5	phosphorvlated forms of Cdk5
CNS	central nervous system
COX-1	cvclooxygenase-1
COX-2	cvclooxygenase-2
CREB	cAMP response element-binding protein
CSS	Chaihu shugan san
EGCG	epigallocatechin-3-gallate
EPM	elevated plus-maze test
ERK1/2	extracellular receptor signal-regulated kinase 1&2
GLUT4	glucose transporter type 4
GSH	glutathione
GSK3ß	glycogen synthase kinase-3 beta
GPx	glutathione peroxidase
HT22	immortalized mouse hippocampal cell line
HO-1	heme oxygenase-1
HOSIDZ	Huangai Sijunzi
H2O2	hydrogen peroxide
IDF	insulin-degrading enzyme
IEE IFN-y	interferon gamma
П1В	interleukin-1ß
IL-1p II_2	interleukin-1
iNOS	inducible nitric oxide synthese
ICR	strain of Swiss mice produced at the Institute of Cancer Research
IP	insulin registance
I/P	corobral ischemia /reportucion
IPS1	insulin response substrate-1
INK	c-Jun N-terminal kinase
KAC	k_{2} appropriate k_{1} (k_{2}) k_{2} (k_{1}) k_{2} (
KAG	lysing acetylase
KDAC	lysine descatylase
KDAC K 3 Rh	kaompforol 3 O rhamposido
K-5-Kii	kaempferol and quaractin so treatment
K/Q I DC	linonolyzacharida
DCE2	npopolysacchande prostaglandin E2
DI2V	phosphoinositide 2 kinasos
LOK	phosphoinositide 2 kinase / protoin kinase B / alwagen synthese kinase 2
PI3K/AKT/GSK-3β	phospholiosinde 5-kinase/protein kinase B/giycogen synnase kinase-5
DVC	protoin kinasa C
	protein kinase C
DCD 05	protein prospiratase 2
MAD	migratubula associated protein
MAR	mitogen activated metain kinage
MLK2	
IVILNZ	nuxeu mieage kinase 2 nudear factor kappa B
INT-KD NET-	nuclear factor kappa D
	N methyl D comertete recorders
NIVIDAKS	ny-memyi-D-aspartate receptors
NU Nuf2	nitric oxide
INTIZ NIDOD	N methyl D concrete to recenter subtract 2
INIXZD	iv-memyr D-aspartate receptor subtype 2B

PET	positron emission tomography
PON2	paroxonase 2
ROS	reactive oxygen species
SIRT1	Sirtuin 1
Ser	serine
Ser9	serine 9
SOD	superoxide dismutase
STZ	streptozotocin
TCM	traditional Chinese medicine
Thr	threonine
TLRs	toll-like receptors
TLR2	toll-like receptor 2
TLR4	toll-like receptor 4
TLR9	toll-like receptor 9
TNF-α	tumor necrosis factor- α
Trkβ	tropomycin-related kinase β
$3 \times Tg AD$ mice	triple transgenic Alzheimer's disease mice

References

- Breijyeh, Z.; Karaman, R. Comprehensive Review on Alzheimer's Disease: Causes and Treatment. *Molecules* 2020, 25, 5789. [CrossRef] [PubMed]
- 2. Tamagno, E.; Guglielmotto, M.; Vasciaveo, V.; Tabaton, M. Oxidative Stress and Beta Amyloid in Alzheimer's Disease. Which Comes First: The Chicken or the Egg? *Antioxidants* **2021**, *10*, 1479. [CrossRef] [PubMed]
- 3. Riche, K.; Lenard, N.R. Quercetin's Effects on Glutamate Cytotoxicity. *Molecules* 2022, 27, 7620. [CrossRef]
- 4. Yu, H.; Wu, J. Amyloid-β: A double agent in Alzheimer's disease? *Biomed. Pharmacother.* 2021, 139, 111575. [CrossRef] [PubMed]
- Ozben, T.; Ozben, S. Neuro-inflammation and anti-inflammatory treatment options for Alzheimer's disease. *Clin. Biochem.* 2019, 72, 87–89. [CrossRef] [PubMed]
- 6. Minter, M.R.; Taylor, J.M.; Crack, P.J. The contribution of neuroinflammation to amyloid toxicity in Alzheimer's disease. *J. Neurochem.* **2016**, 136, 457–474. [CrossRef]
- 7. Kitagishi, Y.; Nakanishi, A.; Ogura, Y.; Matsuda, S. Dietary regulation of PI3K/AKT/GSK-3β pathway in Alzheimer's disease. *Alzheimer's Res. Ther.* **2014**, *6*, 35. [CrossRef]
- Gabbouj, S.; Ryhänen, S.; Marttinen, M.; Wittrahm, R.; Takalo, M.; Kemppainen, S.; Martiskainen, H.; Tanila, H.; Haapasalo, A.; Hiltunen, M.; et al. Altered Insulin Signaling in Alzheimer's Disease Brain—Special Emphasis on PI3K-Akt Pathway. *Front. Neurosci.* 2019, *13*, 629. [CrossRef]
- 9. Bhaskar, K.; Miller, M.; Chludzinski, A.; Herrup, K.; Zagorski, M.; Lamb, B.T. The PI3K-Akt-mTOR pathway regulates Abeta oligomer induced neuronal cell cycle events. *Mol. Neurodegener.* 2009, *4*, 14. [CrossRef]
- Hardy, J.; Selkoe, D.J. The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* 2002, 297, 353–356. [CrossRef]
- Moldogazieva, N.T.; Mokhosoev, I.M.; Mel'nikova, T.I.; Porozov, Y.B.; Terentiev, A.A. Oxidative Stress and Advanced Lipoxidation and Glycation End Products (ALEs and AGEs) in Aging and Age-Related Diseases. *Oxid. Med. Cell. Longev.* 2019, 2019, 3085756. [CrossRef]
- 12. Uddin, M.S.; Kabir, M.T. Oxidative Stress in Alzheimer's Disease: Molecular Hallmarks of Underlying Vulnerability. In *Biological*, *Diagnostic and Therapeutic Advances in Alzheimer's Disease*; Ashraf, G., Alexiou, A., Eds.; Springer: Singapore, 2019. [CrossRef]
- 13. Youssef, P.; Chami, B.; Lim, J.; Middleton, T.; Sutherland, G.T.; Witting, P.K. Evidence supporting oxidative stress in a moderately affected area of the brain in Alzheimer's disease. *Sci. Rep.* **2018**, *8*, 11553. [CrossRef]
- Gao, W.; Wang, W.; Peng, Y.; Deng, Z. Antidepressive effects of kaempferol mediated by reduction of oxidative stress, proinflammatory cytokines and up-regulation of AKT/β-catenin cascade. *Metab. Brain Dis.* 2019, 34, 485–494. [CrossRef] [PubMed]
- 15. Alquezar, C.; Arya, S.; Kao, A.W. Tau Post-translational Modifications: Dynamic Transformers of Tau Function, Degradation, and Aggregation. *Front. Neurol.* **2021**, *11*, 595532. [CrossRef] [PubMed]
- Merino-Serrais, P.; Benavides-Piccione, R.; Blazquez-Llorca, L.; Kastanauskaite, A.; Rábano, A.; Avila, J.; DeFelipe, J. The influence of phospho-tau on dendritic spines of cortical pyramidal neurons in patients with Alzheimer's disease. *Brain* 2013, *136*, 1913–1928. [CrossRef] [PubMed]
- 17. Spittaels, K.; Haute, C.V.D.; Van Dorpe, J.; Geerts, H.; Mercken, M.; Bruynseels, K.; Lasrado, R.; Vandezande, K.; Laenen, I.; Boon, T.; et al. Glycogen synthase kinase-3β phosphorylates protein tau and rescues the axonopathy in the central nervous system of human four-repeat tau transgenic mice. *J. Biol. Chem.* 2000, 275, 41340–41349. [CrossRef]
- Tatebayashi, Y.; Haque, N.; Tung, Y.-C.; Iqbal, K.; Grundke-Iqbal, I. Role of tau phosphorylation by glycogen synthase kinase-3β in the regulation of organelle transport. *J. Cell Sci.* 2004, 117, 1653–1663. [CrossRef]
- 19. Jaworski, T.; Kügler, S.; van Leuven, F. Modeling of tau-mediated synaptic and neuronal degeneration in Alzheimer's disease. *Int. J. Alzheimer's Dis.* **2010**, 2010, 573138. [CrossRef]

- Hoover, B.R.; Reed, M.N.; Su, J.; Penrod, R.D.; Kotilinek, L.A.; Grant, M.K.; Pitstick, R.; Carlson, G.A.; Lanier, L.M.; Yuan, L.-L.; et al. Tau mislocalization to dendritic spines mediates synaptic dysfunction independently of neurodegeneration. *Neuron* 2010, 68, 1067–1081. [CrossRef]
- 21. Thies, E.; Mandelkow, E.-M. Missorting of tau in neurons causes degeneration of synapses that can be rescued by the kinase MARK2/Par-1. *J. Neurosci.* 2007, 27, 2896–2907. [CrossRef]
- Lyman, M.; Lloyd, D.G.; Ji, X.; Vizcaychipi, M.P.; Ma, D. Neuroinflammation: The role and consequences. *Neurosci. Res.* 2013, 79, 1–12. [CrossRef] [PubMed]
- Sabogal-Guáqueta, A.M.; Muñoz-Manco, J.I.; Ramírez-Pineda, J.R.; Lamprea-Rodriguez, M.; Osorio, E.; Cardona-Gómez, G.P. The flavonoid quercetin ameliorates Alzheimer's disease pathology and protects cognitive and emotional function in aged triple transgenic Alzheimer's disease model mice. *Neuropharmacology* 2015, *93*, 134–145. [CrossRef]
- 24. Reitz, C.; Brayne, C.; Mayeux, R. Epidemiology of Alzheimer disease. Nat. Rev. Neurol. 2011, 7, 137–152. [CrossRef] [PubMed]
- 25. Joe, E.; Ringman, J.M. Cognitive symptoms of Alzheimer's disease: Clinical management and prevention. *BMJ* **2019**, *367*, l6217. [CrossRef] [PubMed]
- 26. Casey, D.A.; Antimisiaris, D.; O'Brien, J. Drugs for Alzheimer's disease: Are they effective? *Pharm. Ther.* 2010, 35, 208–211.
- 27. Francis, P.T. The interplay of neurotransmitters in Alzheimer's disease. CNS Spectrums 2005, 10 (Suppl. S18), 6–9. [CrossRef]
- Wang, R.; Reddy, P.H. Role of Glutamate and NMDA Receptors in Alzheimer's Disease. J. Alzheimer's Dis. 2017, 57, 1041–1048. [CrossRef]
- 29. Kouhestani, S.; Jafari, A.; Babaei, P. Kaempferol attenuates cognitive deficit via regulating oxidative stress and neuroinflammation in an ovariectomized rat model of sporadic dementia. *Neural Regen. Res.* **2018**, *13*, 1827–1832. [CrossRef]
- Farlow, M.R.; Miller, M.L.; Pejovic, V. Treatment options in Alzheimer's disease: Maximizing benefit, managing expectations. Dement. Geriatr. Cogn. Disord. 2008, 25, 408–422. [CrossRef]
- 31. Tian, J.; Shi, J.; Zhang, X.; Wang, Y. Herbal therapy: A new pathway for the treatment of Alzheimer's disease. *Alzheimer's Res. Ther.* **2010**, *2*, 30. [CrossRef]
- Lee, J.; Jin, C.; Cho, S.Y.; Park, S.U.; Jung, W.S.; Moon, S.K.; Park, J.M.; Ko, C.N.; Cho, K.H.; Kwon, S. Herbal medicine treatment for Alzheimer disease: A protocol for a systematic review and meta-analysis. *Medicine* 2020, 99, e21745. [CrossRef]
- 33. Chen, M.M.; Yin, Z.Q.; Zhang, L.Y.; Liao, H. Quercetin promotes neurite growth through enhancing intracellular cAMP level and GAP-43 expression. *Chin. J. Nat. Med.* 2015, *13*, 667–672. [CrossRef]
- Zhang, X.W.; Chen, J.Y.; Ouyang, D.; Lu, J.H. Quercetin in Animal Models of Alzheimer's Disease: A Systematic Review of Preclinical Studies. Int. J. Mol. Sci. 2020, 21, 493. [CrossRef]
- 35. Scarmeas, N.; Stern, Y.; Tang, M.X.; Mayeux, R.; Luchsinger, J.A. Mediterranean diet and risk for Alzheimer's disease. *Ann. Neurol.* **2006**, *59*, 912–921. [CrossRef]
- 36. Ren, J.; Lu, Y.; Qian, Y.; Chen, B.; Wu, T.; Ji, G. Recent progress regarding kaempferol for the treatment of various diseases. *Exp. Ther. Med.* **2019**, *18*, 2759–2776. [CrossRef]
- 37. Sreenivasmurthy, S.G.; Liu, J.Y.; Song, J.X.; Yang, C.B.; Malampati, S.; Wang, Z.Y.; Huang, Y.Y.; Li, M. Neurogenic traditional Chinese medicine as a promising strategy for the treatment of Alzheimer's disease. *Int. J. Mol. Sci.* **2017**, *18*, 272. [CrossRef]
- Khan, A.; Ali, T.; Rehman, S.U.; Khan, M.S.; Alam, S.I.; Ikram, M.; Muhammad, T.; Saeed, K.; Badshah, H.; Kim, M.O. Neuroprotective Effect of Quercetin Against the Detrimental Effects of LPS in the Adult Mouse Brain. *Front. Pharmacol.* 2018, 9, 1383. [CrossRef]
- Testa, G.; Gamba, P.; Badilli, U.; Gargiulo, S.; Maina, M.; Guina, T.; Calfapietra, S.; Biasi, F.; Cavalli, R.; Poli, G.; et al. Loading into nanoparticles improves quercetin's efficacy in preventing neuroinflammation induced by oxysterols. *PLoS ONE* 2014, 9, e96795. [CrossRef]
- Ma, Z.X.; Zhang, R.Y.; Rui, W.J.; Wang, Z.Q.; Feng, X. Quercetin alleviates chronic unpredictable mild stress-induced depressivelike behaviors by promoting adult hippocampal neurogenesis via FoxG1/CREB/ BDNF signaling pathway. *Behav. Brain Res.* 2021, 406, 113245. [CrossRef]
- Das, D.; Biswal, S.; Barhwal, K.K.; Chaurasia, O.P.; Hota, S.K. Kaempferol Inhibits Extra-synaptic NMDAR-Mediated Downregulation of TRkβ in Rat Hippocampus During Hypoxia. *Neuroscience* 2018, 392, 77–91. [CrossRef]
- 42. Hussein, R.M.; Mohamed, W.R.; Omar, H.A. A neuroprotective role of kaempferol against chlorpyrifos-induced oxidative stress and memory deficits in rats via GSK3β-Nrf2 signaling pathway. *Pestic. Biochem. Physiol.* **2018**, 152, 29–37. [CrossRef] [PubMed]
- Yu, L.; Chen, C.; Wang, L.F.; Kuang, X.; Liu, K.; Zhang, H.; Du, J.R. Neuroprotective effect of kaempferol glycosides against brain injury and neuroinflammation by inhibiting the activation of NF-κB and STAT3 in transient focal stroke. *PLoS ONE* 2013, *8*, e55839. [CrossRef] [PubMed]
- Azam, S.; Jakaria, M.; Kim, I.S.; Kim, J.; Haque, M.E.; Choi, D.K. Regulation of Toll-Like Receptor (TLR) Signaling Pathway by Polyphenols in the Treatment of Age-Linked Neurodegenerative Diseases: Focus on TLR4 Signaling. *Front. Immunol.* 2019, 10, 1000. [CrossRef] [PubMed]
- Hou, Y.; Aboukhatwa, M.A.; Lei, D.L.; Manaye, K.; Khan, I.; Luo, Y. Anti-depressant natural flavonols modulate BDNF and beta amyloid in neurons and hippocampus of double TgAD mice. *Neuropharmacology* 2010, *58*, 911–920. [CrossRef]
- Kim, J.H.; Kim, H.Y.; Cho, E.J. Protective effects of kaempferol, quercetin, and its glycosides on amyloid beta-induced neurotoxicity in C6 glial cell. J. Appl. Biol. Chem. 2019, 62, 327–332. [CrossRef]

- Zeng, K.; Li, M.; Hu, J.; Mahaman, Y.A.R.; Bao, J.; Huang, F.; Xia, Y.; Liu, X.; Wang, Q.; Wang, J.Z.; et al. *Ginkgo biloba* Extract EGb761 Attenuates Hyperhomocysteinemia-induced AD Like Tau Hyperphosphorylation and Cognitive Impairment in Rats. *Curr. Alzheimer Res.* 2018, 15, 89–99. [CrossRef]
- Ly, P.T.; Wu, Y.; Zou, H.; Wang, R.; Zhou, W.; Kinoshita, A.; Zhang, M.; Yang, Y.; Cai, F.; Woodgett, J.; et al. Inhibition of GSK3β-mediated BACE1 expression reduces Alzheimer-associated phenotypes. J. Clin. Investig. 2013, 123, 224–235. [CrossRef]
- 49. Latta, C.H.; Brothers, H.M.; Wilcock, D.M. Neuroinflammation in Alzheimer's disease; A source of heterogeneity and target for personalized therapy. *Neuroscience* 2015, 302, 103–111. [CrossRef]
- 50. Karuppagounder, S.S.; Madathil, S.K.; Pandey, M.; Haobam, R.; Rajamma, U.; Mohanakumar, K.P. Quercetin up-regulates mitochondrial complex-I activity to protect against programmed cell death in rotenone model of Parkinson's disease in rats. *Neuroscience* **2013**, 236, 136–148. [CrossRef]
- 51. Zaplatic, E.; Bule, M.; Shah, S.Z.A.; Uddin, M.S.; Niaz, K. Molecular mechanisms underlying protective role of quercetin in attenuating Alzheimer's disease. *Life Sci.* 2019, 224, 109–119. [CrossRef]
- 52. Uttara, B.; Singh, A.V.; Zamboni, P.; Mahajan, R. Oxidative stress and neurodegenerative diseases: A review of upstream and downstream antioxidant therapeutic options. *Curr. Neuropharmacol.* **2009**, *7*, 65–74. [CrossRef] [PubMed]
- Wang, X.; Wang, W.; Li, L.; Perry, G.; Lee, H.G.; Zhu, X. Oxidative stress and mitochondrial dysfunction in Alzheimer's disease. Biochim. Biophys. Acta 2014, 1842, 1240–1247. [CrossRef] [PubMed]
- 54. Beach, T.G.; Walker, R.; McGeer, E.G. Patterns of gliosis in Alzheimer's disease and aging cerebrum. *Glia* **1989**, *2*, 420–436. [CrossRef]
- 55. Delacourte, A. General and dramatic glial reaction in Alzheimer brains. *Neurology* **1990**, 40, 33. [CrossRef] [PubMed]
- 56. Arends, Y.M.; Duyckaerts, C.; Rozemuller, J.M.; Eikelenboom, P.; Hauw, J.J. Microglia, amyloid and dementia in alzheimer disease. A correlative study. *Neurobiol. Aging* **2000**, *21*, 39–47. [CrossRef]
- Kim, J.K.; Choi, S.J.; Cho, H.Y.; Hwang, H.J.; Kim, Y.J.; Lim, S.T.; Kim, C.J.; Kim, H.K.; Peterson, S.; Shin, D.H. Protective effects of kaempferol (3,4',5,7-tetrahydroxyflavone) against amyloid beta peptide (Abeta)-induced neurotoxicity in ICR mice. *Biosci. Biotechnol. Biochem.* 2010, 74, 397–401. [CrossRef]
- 58. Shen, X.Y.; Luo, T.; Li, S.; Ting, O.Y.; He, F.; Xu, J.; Wang, H.Q. Quercetin inhibits okadaic acid-induced tau protein hyperphosphorylation through the Ca2+-calpain-p25-CDK5 pathway in HT22 cells. *Int. J. Mol. Med.* **2018**, *41*, 1138–1146. [CrossRef]
- 59. Busche, M.A.; Hyman, B.T. Synergy between amyloid-β and tau in Alzheimer's disease. *Nat. Neurosci.* **2020**, *23*, 1183–1193. [CrossRef]
- Sperling, R.A.; Mormino, E.C.; Schultz, A.P.; Betensky, R.A.; Papp, K.V.; Amariglio, R.E.; Hanseeuw, B.J.; Buckley, R.; Chhatwal, J.; Hedden, T.; et al. The impact of amyloid-beta and tau on prospective cognitive decline in older individuals. *Ann. Neurol.* 2019, 85, 181–193. [CrossRef]
- 61. Pievani, M.; de Haan, W.; Wu, T.; Seeley, W.W.; Frisoni, G.B. Functional network disruption in the degenerative dementias. *Lancet Neurol.* **2011**, *10*, 829–843. [CrossRef]
- 62. Sakakibara, R.; Kawai, T. Cerebrospinal fluid oxidative stress markers in Alzheimer's disease. *Neurol. Clin. Neurosci.* 2020, *8*, 232–240. [CrossRef]
- 63. Willette, A.A.; Li, T.; Willette, S.A.; Larsen, B.A.; Pollpeter, A.; Klinedinst, B.S.; Moody, S.; Barnett, N.; Parvin, M.; Pappas, C.; et al. Oxidative stress biomarkers and longitudinal changes in human brain imaging across the Alzheimer's disease continuum. *Alzheimer's Dement.* **2022**, *18*, e068364. [CrossRef]
- 64. Kim, H.G.; Ju, M.S.; Shim, J.S.; Kim, M.C.; Lee, S.H.; Huh, Y.; Kim, S.Y.; Oh, M.S. Mulberry fruit protects dopaminergic neurons in toxin-induced Parkinson's disease models. *Br. J. Nutr.* **2010**, *104*, 8–16. [CrossRef] [PubMed]
- 65. Song, K.S.; Yang, E.J.; Kim, G.S.; Kim, J.A. Protective effects of onion-derived quercetin on glutamate-mediated hippocampal neuronal cell death. *Pharmacogn. Mag.* 2013, *9*, 302–308. [CrossRef] [PubMed]
- Greenwood, S.M.; Connolly, C.N. Dendritic and mitochondrial changes during glutamate excitotoxicity. *Neuropharmacology* 2007, 53, 891–898. [CrossRef]
- 67. Mattson, M.P. Apoptosis in neurodegenerative disorders. Nat. Rev. Mol. Cell Biol. 2000, 1, 120–130. [CrossRef]
- 68. Simpson, D.S.; Oliver, P.L. ROS generation in microglia: Understanding oxidative stress and inflammation in neurodegenerative disease. *Antioxidants* **2020**, *9*, 743. [CrossRef]
- 69. Di Filippo, M.; Sarchielli, P.; Picconi, B.; Calabresi, P. Neuroinflammation and synaptic plasticity: Theoretical basis for a novel, immune-centred, therapeutic approach to neurological disorders. *Trends Pharmacol. Sci.* **2008**, *29*, 402–412. [CrossRef]
- Chen, W.W.; Zhang, X.; Huang, W.J. Role of neuroinflammation in neurodegenerative diseases. *Mol. Med. Rep.* 2016, 13, 3391–3396. [CrossRef]
- 71. Kempuraj, D.; Thangavel, R.; Selvakumar, G.P.; Zaheer, S.; Ahmed, M.E.; Raikwar, S.P.; Zahoor, H.; Saeed, D.; Natteru, P.A.; Iyer, S.; et al. Brain and peripheral atypical inflammatory mediators potentiate neuroinflammation and neurodegeneration. *Front. Cell. Neurosci.* 2017, *11*, 216. [CrossRef]
- Kempuraj, D.; Thangavel, R.; Natteru, P.A.; Selvakumar, G.P.; Saeed, D.; Zahoor, H.; Zaheer, S.; Iyer, S.S.; Zaheer, A. Neuroinflammation induces neurodegeneration. J. Neurol. Neurosurg. Spine 2016, 1, 1003. [PubMed]
- Cheignon, C.; Tomas, M.; Bonnefont-Rousselot, D.; Faller, P.; Hureau, C.; Collin, F. Oxidative stress and the amyloid beta peptide in Alzheimer's disease. *Redox Biol.* 2018, 14, 450–464. [CrossRef] [PubMed]

- 74. Paris, D.; Mathura, V.; Ait-Ghezala, G.; Beaulieu-Abdelahad, D.; Patel, N.; Bachmeier, C.; Mullan, M. Flavonoids lower Alzheimer's Aβ production via an NFκB dependent mechanism. *Bioinformation* **2011**, *6*, 229–236. [CrossRef] [PubMed]
- 75. Fang, E.F.; Hou, Y.; Palikaras, K.; Adriaanse, B.A.; Kerr, J.S.; Yang, B.; Lautrup, S.; Hasan-Olive, M.M.; Caponio, D.; Dan, X.; et al. Mitophagy inhibits amyloid-β and tau pathology and reverses cognitive deficits in models of Alzheimer's disease. *Nat. Neurosci.* 2019, 22, 401–412. [CrossRef]
- 76. Huang, W.J.; Zhang, X.; Chen, W.W. Role of oxidative stress in Alzheimer's disease. Biomed. Rep. 2016, 4, 519–522. [CrossRef]
- Cai, Z.; Zhao, B.; Ratka, A. Oxidative stress and β-amyloid protein in Alzheimer's disease. *NeuroMolecular Med.* 2011, 13, 223–250. [CrossRef]
- 78. Kim, H.R.; Lee, P.; Seo, S.W.; Roh, J.H.; Oh, M.; Oh, J.S.; Oh, S.J.; Kim, J.S.; Jeong, Y. Comparison of Amyloid β and Tau Spread Models in Alzheimer's Disease. *Cereb. Cortex* **2019**, *29*, 4291–4302. [CrossRef]
- 79. Ismail, R.; Parbo, P.; Madsen, L.S.; Hansen, A.K.; Hansen, K.V.; Schaldemose, J.L.; Kjeldsen, P.L.; Stokholm, M.G.; Gottrup, H.; Eskildsen, S.F.; et al. The relationships between neuroinflammation, beta-amyloid and tau deposition in Alzheimer's disease: A longitudinal PET study. J. Neuroinflammation 2020, 17, 151. [CrossRef]
- 80. Wang, D.M.; Li, S.Q.; Wu, W.L.; Zhu, X.Y.; Wang, Y.; Yuan, H.Y. Effects of long-term treatment with quercetin on cognition and mitochondrial function in a mouse model of Alzheimer's disease. *Neurochem. Res.* 2014, *39*, 1533–1543. [CrossRef]
- 81. Cha, M.Y.; Han, S.H.; Son, S.M.; Hong, H.S.; Choi, Y.J.; Byun, J.; Mook Jung, I. Mitochondria-specific accumulation of amyloid beta induces mitochondrial dysfunction leading to apoptotic cell death. *PLoS ONE* **2012**, *7*, e34929. [CrossRef]
- 82. Moreira, P.I.; Santos, M.S.; Moreno, A.; Rego, A.C.; Oliveira, C. Effect of amyloid beta-peptide on permeability transition pore: A comparative study. *J. Neurosci. Res.* 2002, *69*, 257–267. [CrossRef] [PubMed]
- 83. Beal, M.F. Mitochondria take centre stage in aging and neurodegeneration. Ann. Neurol. 2005, 58, 495–505. [CrossRef] [PubMed]
- 84. Li, Y.; Rusinek, H.; Butler, T.; Glodzik, L.; Pirraglia, E.; Babich, J.; Mozley, P.D.; Nehmeh, S.; Pahlajani, S.; Wang, X.; et al. Decreased CSF clearance and increased brain amyloid in Alzheimer's disease. *Fluids Barriers CNS* **2022**, *19*, 21. [CrossRef]
- 85. Heneka, M.T.; Kummer, M.P.; Latz, E. Innate immune activation in neurodegenerative disease. *Nat. Rev. Immunol.* 2014, 14, 463–477. [CrossRef]
- Heneka, M.T.; Carson, M.J.; Khoury, J.E.; Landreth, G.E.; Brosseron, F.; Feinstein, D.L.; Jacobs, A.H.; Wyss-Coray, T.; Vitorica, J.; Ransohoff, R.M.; et al. Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 2015, 14, 388–405. [CrossRef] [PubMed]
- 87. Arai, H.; Suzuki, H.; Yoshiyama, T.; Lobello, K.; Peng, Y.; Liu, E.; Ketter, N.; Margolin, R.; Jackson, N.; Fujimoto, Y. Safety, tolerability and immunogenicity of an immunotherapeutic vaccine (vanutide cridificar [ACC-001]) and the QS-21 adjuvant in Japanese individuals with mild-to-moderate Alzheimer's disease: A phase IIa, multicenter, randomized, adjuvant and placebo clinical trial. *Alzheimer's Dement.* 2013, *9*, 282.
- 88. Doody, R.S.; Raman, R.; Farlow, M.; Iwatsubo, T.; Vellas, B.; Joffe, S.; Kieburtz, K.; He, F.; Sun, X.; Thomas, R.G.; et al. A phase 3 trial of semagacestat for treatment of Alzheimer's disease. *N. Engl. J. Med.* **2013**, *369*, 341–350. [CrossRef]
- 89. Doody, R.S.; Farlow, M.; Aisen, P.S. Alzheimer's Disease Cooperative Study Data Analysis and Publication Committee. Phase 3 trials of solanezumab and bapineuzumab for Alzheimer's disease. *N. Engl. J. Med.* **2014**, *370*, 1460.
- Li, S.; Selkoe, D.J. A mechanistic hypothesis for the impairment of synaptic plasticity by soluble Aβ oligomers from Alzheimer's brain. J. Neurochem. 2020, 154, 583–597. [CrossRef]
- 91. Barbier, P.; Zejneli, O.; Martinho, M.; Lasorsa, A.; Belle, V.; Smet-Nocca, C.; Tsvetkov, P.O.; Devred, F.; Landrieu, I. Role of Tau as a Microtubule-Associated Protein: Structural and Functional Aspects. *Front. Aging Neurosci.* **2019**, *11*, 204. [CrossRef]
- 92. Stancu, I.C.; Vasconcelos, B.; Terwel, D.; Dewachter, I. Models of β-amyloid induced Tau-pathology: The long and "folded" road to understand the mechanism. *Mol. Neurodegener.* **2014**, *9*, 51. [CrossRef] [PubMed]
- Takashima, A.; Honda, T.; Yasutake, K.; Michel, G.; Murayama, O.; Murayama, M.; Ishiguro, K.; Yamaguchi, H. Activation of tau protein kinase I/glycogen synthase kinase-3beta by amyloid beta peptide (25–35) enhances phosphorylation of tau in hippocampal neurons. *Neurosci. Res.* 1998, 31, 317–323. [CrossRef]
- 94. Ferreira, A.; Lu, Q.; Orecchio, L.; Kosik, K.S. Selective phosphorylation of adult tau isoforms in mature hippocampal neurons exposed to fibrillar A beta. *Mol. Cell. Neurosci.* **1997**, *9*, 220–234. [CrossRef]
- 95. Zheng, W.H.; Bastianetto, S.; Mennicken, F.; Ma, W.; Kar, S. Amyloid beta peptide induces tau phosphorylation and loss of cholinergic neurons in rat primary septal cultures. *Neuroscience* **2002**, *115*, 201–211. [CrossRef]
- 96. Ma, Q.L.; Lim, G.P.; Harris-White, M.E.; Yang, F.; Ambegaokar, S.S.; Ubeda, O.J.; Glabe, C.G.; Teter, B.; Frautschy, S.A.; Cole, G.M. Antibodies against beta-amyloid reduce Abeta oligomers, glycogen synthase kinase-3beta activation and tau phosphorylation in vivo and in vitro. J. Neurosci. Res. 2006, 83, 374–384. [CrossRef]
- Tackenberg, C.; Grinschgl, S.; Trutzel, A.; Santuccione, A.C.; Frey, M.C.; Konietzko, U.; Grimm, J.; Brandt, R.; Nitsch, R.M. NMDA receptor subunit composition determines beta-amyloid-induced neurodegeneration and synaptic loss. *Cell Death Dis.* 2013, 4, e608. [CrossRef]
- Wang, W.Y.; Tan, M.S.; Yu, J.T.; Tan, L. Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease. *Ann. Transl. Med.* 2015, *3*, 136. [CrossRef] [PubMed]
- 99. Vogel, J.W.; Iturria-Medina, Y.; Strandberg, O.T.; Smith, R.; Levitis, E.; Evans, A.C.; Hansson, O.; Alzheimer's Disease Neuroimaging Initiative; Swedish BioFinder Study. Spread of pathological tau proteins through communicating neurons in human Alzheimer's disease. *Nat. Commun.* **2020**, *11*, 2612, Erratum in *Nat. Commun.* **2021**, *12*, 4862. [CrossRef] [PubMed]
- 100. Giacobini, E.; Gold, G. Alzheimer disease therapy-Moving from amyloid-β to tau. Nat. Rev. Neurol. 2013, 9, 677-686. [CrossRef]

- 101. Amaral, A.C.; Perez-Nievas, B.G.; Chong, M.S.T.; Gonzalez-Martinez, A.; Argente-Escrig, H.; Rubio-Guerra, S.; Commins, C.; Muftu, S.; Eftekharzadeh, B.; Hudry, E.; et al. Isoform-selective decrease of glycogen synthase kinase-3-beta (GSK-3β) reduces synaptic tau phosphorylation, transcellular spreading, and aggregation. *Iscience* 2021, 24, 102058. [CrossRef]
- 102. Mandelkow, E.-M.; Mandelkow, E. Biochemistry and cell biology of tau protein in neurofibrillary degeneration. *Cold Spring Harb. Perspect. Med.* **2012**, *2*, a006247. [CrossRef] [PubMed]
- 103. Roberson, E.D.; Scearce-Levie, K.; Palop, J.J.; Yan, F.; Cheng, I.H.; Wu, T.; Gerstein, H.; Yu, G.Q.; Mucke, L. Reducing endogenous tau ameliorates amyloid β-induced deficits in an Alzheimer's disease mouse model. *Science* 2007, 316, 750–754. [CrossRef] [PubMed]
- 104. Duchen, M.R. Mitochondria and calcium: From cell signaling to cell death. J. Physiol. 2000, 529, 57–68. [CrossRef] [PubMed]
- 105. Squier, M.K.; Miller, A.C.; Malkinson, A.M.; Cohen, J.J. Calpain activation in apoptosis. J. Cell. Physiol. 1994, 159, 229–237. [CrossRef]
- 106. Maher, P.; Schubert, D. Signaling by reactive oxygen species in the nervous system. *Cell. Mol. Life Sci.* 2000, 57, 1287–1305. [CrossRef] [PubMed]
- 107. Darling, A.L.; Uversky, V.N. Intrinsic disorder and posttranslational modifications: The darker side of the biological dark matter. *Front. Genet.* **2018**, *9*, 158. [CrossRef]
- 108. Barber, K.W.; Rinehart, J. The ABCs of PTMs. Nat. Chem. Biol. 2018, 14, 188–192. [CrossRef]
- Buee, L.; Bussiere, T.; Buee-Scherrer, V.; Delacourte, A.; Hof, P.R. Tau protein isoforms, phosphorylation and role in neurodegenerative disorders. *Brain Res. Rev.* 2000, 33, 95–130. [CrossRef]
- 110. Xia, C.; Makaretz, S.J.; Caso, C.; McGinnis, S.; Gomperts, S.N.; Sepulcre, J.; Gomez-Isla, T.; Hyman, B.T.; Schultz, A.; Vasdev, N.; et al. Association of in vivo [¹⁸F]AV-1451 tau PET imaging results with cortical atrophy and symptoms in typical and atypical Alzheimer disease. *JAMA Neurol.* 2017, 74, 427–436. [CrossRef]
- 111. Bejanin, A.; Schonhaut, D.R.; La Joie, R.; Kramer, J.H.; Baker, S.L.; Sosa, N.; Ayakta, N.; Cantwell, A.; Janabi, M.; Lauriola, M.; et al. Tau pathology and neurodegeneration contribute to cognitive impairment in Alzheimeras disease. *Brain* 2017, 140, 3286–3300. [CrossRef]
- 112. Fleeman, R.M.; Proctor, E.A. Astrocytic Propagation of Tau in the Context of Alzheimer's Disease. *FFront. Cell. Neurosci.* 2021, 15, 645233. [CrossRef] [PubMed]
- 113. Wegmann, S.; Bennett, R.E.; Delorme, L.; Robbins, A.B.; Hu, M.; MacKenzie, D.; Kirk, M.J.; Schiantarelli, J.; Tunio, N.; Amaral, A.C.; et al. Experimental evidence for the age dependence of tau protein spread in the brain. *Sci. Adv.* 2019, *5*, eaaw6404. [CrossRef] [PubMed]
- 114. Wei, Y.; Liu, M.; Wang, D. The propagation mechanisms of extracellular tau in Alzheimer's disease. *J. Neurol.* **2022**, 269, 1164–1181. [CrossRef]
- 115. Kazim, S.F.; Sharma, A.; Saroja, S.R.; Seo, J.H.; Larson, C.S.; Ramakrishnan, A.; Wang, M.; Blitzer, R.D.; Shen, L.; Peña, C.J.; et al. Chronic Intermittent Hypoxia Enhances Pathological Tau Seeding, Propagation, and Accumulation and Exacerbates Alzheimer-like Memory and Synaptic Plasticity Deficits and Molecular Signatures. *Biol. Psychiatry* 2022, *91*, 346–358. [CrossRef] [PubMed]
- 116. Liu, R.H. Health-promoting components of fruits and vegetables in the diet. Adv. Nutr. Int. Rev. J. 2013, 4, 384S–392S. [CrossRef]
- Hertog, M.G.L.; Feskens, E.J.M.; Hollman, P.C.H.; Katan, M.B.; Kromhout, D. Dietary antioxidant flavonoids and risk of coronary heart disease: The Zutphen Elderly Study. *Lancet* 1993, 342, 1007–1011. [CrossRef]
- 118. Anhê, G.F.; Okamoto, M.M.; Kinote, A.; Sollon, C.; Lellis-Santos, C.; Anhê, F.F.; Lima, G.A.; Hirabara, S.M.; Velloso, L.A.; Bordin, S.; et al. Quercetin decreases inflammatory response and increases insulin action in skeletal muscle of ob/ob mice and in L6 myotubes. *Eur. J. Pharmacol.* 2012, 689, 285–293. [CrossRef]
- 119. Simunkova, M.; Alwasel, S.H.; Alhazza, I.M.; Jomova, K.; Kollar, V.; Rusko, M.; Valko, M. Management of oxidative stress and other pathologies in Alzheimer's disease. *Arch. Toxicol.* **2019**, *93*, 2491–2513. [CrossRef]
- 120. Hanaki, M.; Murakami, K.; Akagi, K.; Irie, K. Structural insights into mechanisms for inhibiting amyloid β42 aggregation by non-catechol-type flavonoids. *Bioorganic Med. Chem.* **2016**, *24*, 304–313. [CrossRef]
- 121. Porat, Y.; Abramowitz, A.; Gazit, E. Inhibition of amyloid fibril formation by polyphenols: Structural similarity and aromatic interactions as a common inhibition mechanism. *Chem. Biol. Drug Des.* **2006**, *67*, 27–37. [CrossRef]
- Ono, K.; Yoshiike, Y.; Takashima, A.; Hasegawa, K.; Naiki, H.; Yamada, M. Potent anti-amyloidogenic and fibril-destabilizing effects of polyphenols in vitro: Implications for the prevention and therapeutics of Alzheimer's disease. *J. Neurochem.* 2003, 87, 172–181. [CrossRef]
- Jiménez-Aliaga, K.; Bermejo-Bescós, P.; Benedí, J.; Martín-Aragón, S. Quercetin and rutin exhibit antiamyloidogenic and fibrildisaggregating effects in vitro and potent antioxidant activity in APPswe cells. *Life Sci.* 2011, 89, 939–945. [CrossRef]
- 124. Sato, M.; Murakami, K.; Uno, M.; Nakagawa, Y.; Katayama, S.; Akagi, K.; Masuda, Y.; Takegoshi, K.; Irie, K. Site-specific inhibitory mechanism for amyloid β42 aggregation by catechol-type flavonoids targeting the Lys residues. *J. Biol. Chem.* 2013, 288, 23212–23224. [CrossRef]
- 125. Yu, X.; Li, Y.; Mu, X. Effect of Quercetin on PC12 Alzheimer's Disease Cell Model Induced by Aβ25-35 and Its Mechanism Based on Sirtuin1/Nrf2/HO-1 Pathway. *BioMed Res. Int.* 2020, 2020, 8210578. [CrossRef] [PubMed]
- Kumar, S.; Krishnakumar, V.G.; Morya, V.; Gupta, S.; Datta, B. Nanobiocatalyst facilitated aglycosidic quercetin as a potent inhibitor of tau protein aggregation. *Int. J. Biol. Macromol.* 2019, 138, 168–180. [CrossRef] [PubMed]

- 127. Luo, C.; Yang, H.; Tang, C.; Yao, G.; Kong, L.; He, H.; Zhou, Y. Kaempferol alleviates insulin resistance via hepatic IKK/NF-κB signal in type 2 diabetic rats. *Int. Immunopharmacol.* **2015**, *28*, 744–750. [CrossRef]
- Peng, J.; Li, Q.; Li, K.; Zhu, L.; Lin, X.; Lin, X.; Shen, Q.; Li, G.; Xie, X. Quercetin Improves Glucose and Lipid Metabolism of Diabetic Rats: Involvement of Akt Signaling and SIRT1. J. Diabetes Res. 2017, 2017, 3417306. [CrossRef]
- 129. Dabeek, W.M.; Marra, M.V. Dietary Quercetin and Kaempferol: Bioavailability and Potential Cardiovascular-Related Bioactivity in Humans. *Nutrients* **2019**, *11*, 2288. [CrossRef] [PubMed]
- 130. Panche, A.N.; Diwan, A.D.; Chandra, S.R. Flavonoids: An overview. J. Nutr. Sci. 2016, 5, e47. [CrossRef]
- Crozier, A.; Lean, M.E.J.; McDonald, M.S.; Black, C. Quantitative analysis of the flavonoid content of commercial tomatoes, onions, lettuce, and celery. J. Agric. Food Chem. 1997, 45, 590–595. [CrossRef]
- Xu, M.; Huang, H.; Mo, X.; Zhu, Y.; Chen, X.; Li, X.; Peng, X.; Xu, Z.; Chen, L.; Rong, S.; et al. Quercetin-3-O-Glucuronide Alleviates Cognitive Deficit and Toxicity in Aβ1-42 -Induced AD-Like Mice and SH-SY5Y Cells. *Mol. Nutr. Food Res.* 2021, 65, e2000660. [CrossRef]
- Shen, P.; Lin, W.; Deng, X.; Ba, X.; Han, L.; Chen, Z.; Qin, K.; Huang, Y.; Tu, S. Potential Implications of Quercetin in Autoimmune Diseases. Front. Immunol. 2021, 12, 689044. [CrossRef]
- Babaei, F.; Mirzababaei, M.; Nassiri-Asl, M. Quercetin in Food: Possible Mechanisms of Its Effect on Memory. J. Food Sci. 2018, 83, 2280–2287. [CrossRef] [PubMed]
- Yang, D.; Wang, T.; Long, M.; Li, P. Quercetin: Its Main Pharmacological Activity and Potential Application in Clinical Medicine. Oxidative Med. Cell. Longev. 2020, 2020, 8825387. [CrossRef] [PubMed]
- 136. Nishihira, J.; Nishimura, M.; Kurimoto, M.; Kagami-Katsuyama, H.; Hattori, H.; Nakagawa, T.; Muro, T.; Kobori, M. The effect of 24-week continuous intake of quercetin-rich onion on age-related cognitive decline in healthy elderly people: A randomized, double-blind, placebo-controlled, parallel-group comparative clinical trial. J. Clin. Biochem. Nutr. 2021, 69, 203–215. [CrossRef]
- 137. Bayazid, A.B.; Lim, B.O. Quercetin Is An Active Agent in Berries against Neurodegenerative Diseases Progression through Modulation of Nrf2/HO1. *Nutrients* 2022, 14, 5132. [CrossRef]
- Islam, M.S.; Quispe, C.; Hossain, R.; Islam, M.T.; Al-Harrasi, A.; Al-Rawahi, A.; Martorell, M.; Mamurova, A.; Seilkhan, A.; Altybaeva, N.; et al. Neuropharmacological Effects of Quercetin: A Literature-Based Review. *Neuropharmacology* 2022, 12, 665031. [CrossRef] [PubMed]
- Wu, Q.; Naeem, A.; Zou, J.; Yu, C.; Wang, Y.; Chen, J.; Ping, Y. Isolation of Phenolic Compounds from Raspberry Based on Molecular Imprinting Techniques and Investigation of Their Anti-Alzheimer's Disease Properties. *Molecules* 2022, 27, 6893. [CrossRef]
- 140. Ulusoy, H.G.; Sanlier, N. A minireview of quercetin: From its metabolism to possible mechanisms of its biological activities. *Crit. Rev. Food Sci. Nutr.* **2020**, *60*, 3290–3303. [CrossRef]
- Xiao, L.; Luo, G.; Tang, Y.; Yao, P. Quercetin and iron metabolism: What we know and what we need to know. *Food Chem. Toxicol.* 2018, 114, 190–203. [CrossRef]
- 142. Lesjak, M.; Beara, I.; Simin, N.; Pintać, D.; Majkić, T.; Bekvalac, K.; Orčić, D.; Mimica-Dukić, N. Antioxidant and anti-inflammatory activities of quercetin and its derivatives. J. Funct. Foods 2018, 40, 68–75. [CrossRef]
- 143. Chahar, M.K.; Sharma, N.; Dobhal, M.P.; Joshi, Y.C. Flavonoids: A versatile source of anticancer drugs. *Pharmacogn. Rev.* 2011, 5, 1.
- 144. Nakagawa, T.; Ohta, K. Quercetin Regulates the Integrated Stress Response to Improve Memory. *Int. J. Mol. Sci.* 2019, 20, 2761. [CrossRef] [PubMed]
- 145. Liu, Y.W.; Liu, X.L.; Kong, L.; Zhang, M.Y.; Chen, Y.J.; Zhu, X.; Hao, Y.C. Neuroprotection of quercetin on central neurons against chronic high glucose through enhancement of Nrf2/ARE/glyoxalase-1 pathway mediated by phosphorylation regulation. *Biomed. Pharmacother.* 2019, 109, 2145–2154. [CrossRef]
- 146. Costa, L.G.; Garrick, J.M.; Roquè, P.J.; Pellacani, C. Mechanisms of Neuroprotection by Quercetin: Counteracting Oxidative Stress and More. *Oxidative Med. Cell. Longev.* **2016**, 2016, 2986796. [CrossRef]
- 147. Wei, C.; Li, S.; Zhu, Y.; Chen, W.; Li, C.; Xu, R. Network pharmacology identify intersection genes of quercetin and Alzheimer's disease as potential therapeutic targets. *Front. Aging Neurosci.* **2022**, *14*, 902092. [CrossRef] [PubMed]
- 148. García-Mediavilla, M.V.; Crespo, I.; Collado, P.S.; Esteller, A.; Sánchez-Campos, S.; Tuñón, M.J.; González-Gallego, J. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. *Eur. J. Pharmacol.* 2007, 557, 221–229. [CrossRef] [PubMed]
- 149. Song, K.S.; Jeong, W.S.; Jun, M. Inhibition of β-amyloid peptide-induced neurotoxicity by kaempferol 3-O-(6"-acetyl)-βglucopyranoside from butterbur (Petasites japonicus) leaves in B103 cells. *Food Sci. Biotechnol.* **2012**, 21, 845–851. [CrossRef]
- 150. Kaypee, S.; Singh, S.; Swarnkar, S.; Kundu, T.K. Emerging epigenetic therapies—Lysine acetyltransferase inhibitors. In *Epigenetic Cancer Therapy*; Academic Press: Cambridge, MA, USA, 2023; pp. 459–505.
- 151. Xiao, X.; Shi, D.; Liu, L.; Wang, J.; Xie, X.; Kang, T.; Deng, W. Quercetin suppresses cyclooxygenase-2 expression and angiogenesis through inactivation of P300 signaling. *PLoS ONE* **2011**, *6*, e22934. [CrossRef]
- 152. Pei, Y.; Parks, J.S.; Kang, H.W. Quercetin alleviates high-fat diet-induced inflammation in brown adipose tissue. *J. Funct. Foods* **2021**, *85*, 104614. [CrossRef]

- 153. Son, S.M.; Park, S.J.; Fernandez-Estevez, M.; Rubinsztein, D.C. Autophagy regulation by acetylation-implications for neurodegenerative diseases. *Exp. Mol. Med.* **2021**, *53*, 30–41. [CrossRef] [PubMed]
- 154. Fiorentino, F.; Mai, A.; Rotili, D. Lysine Acetyltransferase Inhibitors From Natural Sources. *Front. Pharmacol.* **2020**, *11*, 1243. [CrossRef] [PubMed]
- Liao, Y.; Mai, X.; Wu, X.; Hu, X.; Luo, X.; Zhang, G. Exploring the Inhibition of Quercetin on Acetylcholinesterase by Multispectroscopic and In Silico Approaches and Evaluation of Its Neuroprotective Effects on PC12 Cells. *Molecules* 2022, 27, 7971. [CrossRef] [PubMed]
- 156. Alghamdi, A.; Birch, D.J.; Vyshemirsky, V.; Rolinski, O.J. Impact of the Flavonoid Quercetin on β-Amyloid Aggregation Revealed by Intrinsic Fluorescence. *J. Phys. Chem. B* **2022**, *126*, 7229–7237. [CrossRef]
- Ho, C.L.; Kao, N.J.; Lin, C.I.; Cross, T.L.; Lin, S.H. Quercetin Increases Mitochondrial Biogenesis and Reduces Free Radicals in Neuronal SH-SY5Y Cells. *Nutrients* 2022, 14, 3310. [CrossRef] [PubMed]
- Bao, D.; Wang, J.; Pang, X.; Liu, H. Protective Effect of Quercetin against Oxidative Stress-Induced Cytotoxicity in Rat Pheochromocytoma (PC-12) Cells. *Molecules* 2017, 22, 1122. [CrossRef]
- 159. Jiang, W.; Luo, T.; Li, S.; Zhou, Y.; Shen, X.-Y.; He, F.; Xu, J.; Wang, H.Q. Quercetin Protects against Okadaic Acid-Induced Injury via MAPK and PI3K/Akt/GSK3β Signaling Pathways in HT22 Hippocampal Neurons. *PLoS ONE* **2016**, *11*, e0152371. [CrossRef]
- Paula, P.C.; Maria, S.G.; Luis, C.H.; Patricia, C.G. Preventive Effect of Quercetin in a Triple Transgenic Alzheimer's Disease Mice Model. *Molecules* 2019, 24, 2287. [CrossRef]
- Molaei, A.; Hatami, H.; Dehghan, G.; Sadeghian, R.; Khajehnasiri, N. Synergistic effects of quercetin and regular exercise on the recovery of spatial memory and reduction of parameters of oxidative stress in animal model of Alzheimer's disease. *EXCLI J.* 2020, *19*, 596–612. [CrossRef]
- 162. Dhawan, S.; Kapil, R.; Singh, B. Formulation development and systematic optimization of solid lipid nanoparticles of quercetin for improved brain delivery. *J. Pharm. Pharmacol.* **2011**, *63*, 342–351. [CrossRef]
- 163. Chen, J.; Deng, X.; Liu, N.; Li, M.; Liu, B.; Fu, Q.; Qu, R.; Ma, S. Quercetin attenuates tau hyperphosphorylation and improves cognitive disorder via suppression of ER stress in a manner dependent on AMPK pathway. J. Funct. Foods 2016, 22, 463–476. [CrossRef]
- National Center for Biotechnology Information. PubChem Compound Summary for CID 5280343, Quercetin. Retrieved 9 November 2023. 2023. Available online: https://pubchem.ncbi.nlm.nih.gov/compound/Quercetin (accessed on 6 October 2023).
- 165. Dimitrić Marković, J.M.; Milenković, D.; Amić, D.; Popović-Bijelić, A.; Mojović, M.; Pašti, I.A.; Marković, Z.S. Energy requirements of the reactions of kaempferol and selected radical species in different media: Towards the prediction of the possible radical scavenging mechanisms. *Struct. Chem.* 2014, 25, 1795–1804. [CrossRef]
- Wang, L.; Tu, Y.C.; Lian, T.W.; Hung, J.T.; Yen, J.H.; Wu, M.J. Distinctive antioxidant and antiinflammatory effects of flavonols. J. Agric. Food Chem. 2006, 54, 9798–9804. [CrossRef] [PubMed]
- Park, S.E.; Sapkota, K.; Kim, H.; Kim, S.J. Kaempferol acts through mitogen-activated protein kinases and protein kinase B/AKT to elicit protection in a model of neuroinflammation in BV2 microglial cells. Br. J. Pharmacol. 2011, 164, 1008–1025. [CrossRef]
- Olszewska, M. Separation of quercetin, sexangularetin, kaempferol and isorhamnetin for simultaneous HPLC determination of flavonoid aglycones in inflorescences, leaves and fruits of three Sorbus species. J. Pharm. Biomed. Anal. 2008, 48, 629–635. [CrossRef]
- Kiziltaş, H. Comprehensive evaluation of Reseda lutea L. (Wild Mignonette) and 7 isolated flavonol glycosides: Determination of antioxidant activity, anti-Alzheimer, antidiabetic and cytotoxic effects with in vitro and in silico methods. *Turk. J. Chem.* 2022, 46, 1185–1198. [CrossRef]
- 170. Sulfahri; Wardhani, R.; Makatita, F.A.; Iskandar, I.W. Utilization of Nypa fruit in Alzheimer's Disease: An In Silico Approach. J. Phys. Conf. Ser. 2019, 1341, 022003. [CrossRef]
- 171. Yuan, Y.; Zhai, Y.; Chen, J.; Xu, X.; Wang, H. Kaempferol Ameliorates Oxygen-Glucose Deprivation/Reoxygenation-Induced Neuronal Ferroptosis by Activating Nrf2/SLC7A11/GPX4 Axis. *Biomolecules* **2021**, *11*, 923. [CrossRef]
- Uysal, M.; Celikten, M.; Beker, M.; Polat, N.; Huseyinbas, O.; Terzioglu-Usak, S.; Elibol, B. Kaempferol treatment ameliorates memory impairments in STZ-induced neurodegeneration by acting on reelin signaling. *Acta Neurobiol. Exp. (Wars)* 2023, 83, 236–245. [CrossRef]
- 173. Beg, T.; Jyoti, S.; Naz, F.; Rahul, X.; Ali, F.; Ali, S.K.; Reyad, A.M.; Siddique, Y.H. Protective Effect of Kaempferol on the Transgenic Drosophila Model of Alzheimer's Disease. CNS Neurol. Disord. Drug Targets 2018, 17, 421–429. [CrossRef]
- 174. Zhang, N.; Xu, H.; Wang, Y.; Yao, Y.; Liu, G.; Lei, X.; Sun, H.; Wu, X.; Li, J. Protective mechanism of kaempferol against Aβ25-35-mediated apoptosis of pheochromocytoma (PC-12) cells through the ER/ERK/MAPK signalling pathway. *Arch. Med Sci.* 2020, 17, 406–416. [CrossRef]
- 175. Sun, J.; Wang, J.; Hu, L.; Yan, J. K-3-Rh Protects Against Cerebral Ischemia/Reperfusion Injury by Anti-Apoptotic Effect Through PI3K-Akt Signaling Pathway in Rat. *Neuropsychiatr. Dis. Treat.* **2020**, *16*, 1217–1227. [CrossRef]
- 176. Al-Brakati, A.; Albarakati, A.J.A.; Lokman, M.S.; Theyab, A.; Algahtani, M.; Menshawi, S.; AlAmri, O.D.; Al Omairi, N.E.; Essawy, E.A.; Kassab, R.B.; et al. Possible Role of Kaempferol in Reversing Oxidative Damage, Inflammation, and Apoptosis-Mediated Cortical Injury Following Cadmium Exposure. *Neurotox. Res.* 2021, 39, 198–209. [CrossRef]
- 177. Ai, R.; Zhuang, X.X.; Anisimov, A.; Lu, J.H.; Fang, E.F. A synergized machine learning plus cross-species wet-lab validation approach identifies neuronal mitophagy inducers inhibiting Alzheimer disease. *Autophagy* **2022**, *18*, 939–941. [CrossRef]

- Zarei, M.; Mohammadi, S.; Komaki, A.; Golipour Choshali, Z. Antidepressant-like Effects of Intra-cerebroventricular Microinjection of Kaempferol in Male Rats: Involvement of 5-HT2 Receptors. *Avicenna J. Neuro Psycho Physiol.* 2022, 9, 23–30.
- 179. Rita, L.; Neumann, N.R.; Laponogov, I.; Gonzalez, G.; Veselkov, D.; Pratico, D.; Aalizadeh, R.; Thomaidis, N.S.; Thompson, D.C.; Vasiliou, V.; et al. Alzheimer's disease: Using gene/protein network machine learning for molecule discovery in olive oil. *Hum. Genom.* 2023, 17, 57. [CrossRef]
- 180. Karunakaran, K.B.; Thiyagaraj, A.; Santhakumar, K. Novel insights on acetylcholinesterase inhibition by Convolvulus pluricaulis, scopolamine and their combination in zebrafish. *Nat. Prod. Bioprospecting* **2022**, *12*, 6. [CrossRef] [PubMed]
- 181. Simunkova, M.; Barbierikova, Z.; Jomova, K.; Hudecova, L.; Lauro, P.; Alwasel, S.H.; Alhazza, I.; Rhodes, C.J.; Valko, M. Antioxidant vs. Prooxidant Properties of the Flavonoid, Kaempferol, in the Presence of Cu(II) Ions: A ROS-Scavenging Activity, Fenton Reaction and DNA Damage Study. Int. J. Mol. Sci. 2021, 22, 1619. [CrossRef] [PubMed]
- 182. Ajiboye, B.O.; Ojo, O.A.; Okesola, M.A.; Akinyemi, A.J.; Talabi, J.Y.; Idowu, O.T.; Fadaka, A.O.; Boligon, A.A.; de Campos, M.M.A. In vitro antioxidant activities and inhibitory effects of phenolic extract of *Senecio biafrae* (Oliv and Hiern) against key enzymes linked with type II diabetes mellitus and Alzheimer's disease. *Food Sci. Nutr.* 2018, *6*, 1803–1810. [CrossRef] [PubMed]
- 183. Shabir, I.; Pandey, V.K.; Shams, R.; Dar, A.H.; Dash, K.K.; Khan, S.A.; Bashir, I.; Jeevarathinam, G.; Rusu, A.V.; Esatbeyoglu, T.; et al. Promising bioactive properties of quercetin for potential food applications and health benefits: A review. *Front. Nutr.* 2022, 9, 999752. [CrossRef]
- 184. Álvarez-Berbel, I.; Espargaró, A.; Viayna, A.; Caballero, A.B.; Busquets, M.A.; Gámez, P.; Luque, F.J.; Sabaté, R. Three to Tango: Inhibitory Effect of Quercetin and Apigenin on Acetylcholinesterase, Amyloid-β Aggregation and Acetylcholinesterase-Amyloid Interaction. *Pharmaceutics* 2022, 14, 2342. [CrossRef] [PubMed]
- 185. Wang, J.; Mao, J.; Wang, R.; Li, S.; Wu, B.; Yuan, Y. Kaempferol Protects Against Cerebral Ischemia Reperfusion Injury Through Intervening Oxidative and Inflammatory Stress Induced Apoptosis. *Front. Pharmacol.* **2020**, *11*, 424. [CrossRef] [PubMed]
- 186. Dong, X.; Zhou, S.; Nao, J. Kaempferol as a therapeutic agent in Alzheimer's disease: Evidence from preclinical studies. *Ageing Res. Rev.* **2023**, *87*, 101910. [CrossRef] [PubMed]
- Li, W.H.; Cheng, X.; Yang, Y.L.; Liu, M.; Zhang, S.S.; Wang, Y.H.; Du, G.H. Kaempferol attenuates neuroinflammation and blood brain barrier dysfunction to improve neurological deficits in cerebral ischemia/reperfusion rats. *Brain Res.* 2019, 1722, 146361. [CrossRef]
- 188. El-Kott, A.F.; Abd-Lateif, A.-E.M.; Khalifa, H.S.; Morsy, K.; Ibrahim, E.H.; Bin-Jumah, M.; Abdel-Daim, M.M.; Aleya, L. Kaempferol protects against cadmium chloride-induced hippocampal damage and memory deficits by activation of silent information regulator 1 and inhibition of poly (ADP-Ribose) polymerase-1. *Sci. Total. Environ.* 2020, 728, 138832. [CrossRef]
- Lin, H.; Wang, X.; Zhao, J.; Lin, Z. Protective effect of kaempferol against cognitive and neurological disturbances induced by d-galactose and aluminum chloride in mice. J. Funct. Foods 2023, 100, 105385. [CrossRef]
- 190. Selvi, R.B.; Swaminathan, A.; Chatterjee, S.; Shanmugam, M.K.; Li, F.; Ramakrishnan, G.B.; Siveen, K.S.; Chinnathambi, A.; Zayed, M.E.; Alharbi, S.A.; et al. Inhibition of p300 lysine acetyltransferase activity by luteolin reduces tumor growth in head and neck squamous cell carcinoma (HNSCC) xenograft mouse model. *Oncotarget* 2015, *6*, 43806–43818. [CrossRef]
- 191. Zhou, Y.P.; Li, G.C. Kaempferol protects cell damage in in vitro ischemia reperfusion model in rat neuronal PC12 cells. *BioMed Res. Int.* 2020, 2020, 2461079. [CrossRef]
- 192. Kadioglu, O.; Nass, J.; Saeed, M.E.; Schuler, B.; Efferth, T. Kaempferol Is an Anti-Inflammatory Compound with Activity towards NF-κB Pathway Proteins. *Anticancer Res.* **2015**, *35*, 2645–2650.
- Devi, K.P.; Malar, D.S.; Nabavi, S.F.; Sureda, A.; Xiao, J.; Nabavi, S.M.; Daglia, M. Kaempferol and inflammation: From chemistry to medicine. *Pharmacol. Res.* 2015, 99, 1–10. [CrossRef]
- Alam, W.; Khan, H.; Shah, M.A.; Cauli, O.; Saso, L. Kaempferol as a Dietary Anti-Inflammatory Agent: Current Therapeutic Standing. *Molecules* 2020, 25, 4073. [CrossRef] [PubMed]
- 195. Sharoar, G.; Thapa, A.; Shahnawaz, M.; Ramasamy, V.S.; Woo, E.-R.; Shin, S.Y.; Park, I.-S. Keampferol-3-O-rhamnoside abrogates amyloid beta toxicity by modulating monomers and remodeling oligomers and fibrils to non-toxic aggregates. *J. Biomed. Sci.* 2012, 19, 104. [CrossRef] [PubMed]
- 196. Chowdhury, M.A.; Ko, H.J.; Lee, H.; Aminul Haque, M.; Park, I.S.; Lee, D.S.; Woo, E.R. Oleanane triterpenoids from Akebiae Caulis exhibit inhibitory effects on Aβ42 induced fibrillogenesis. *Arch. Pharm. Res.* **2017**, *40*, 318–327. [CrossRef] [PubMed]
- 197. Guo, Q.; Sebastian, L.; Sopher, B.L. Increased vulnerability of hippocampal neurons from presenilin-1 mutant knock-in mice to amyloid-β peptide toxicity. *J. Neurochem.* **1999**, *72*, 1019–1029. [CrossRef] [PubMed]
- 198. Ishige, K.; Schubert, D.; Sagara, Y. Flavonoids protect neuronal cells from oxidative stress by three distinct mechanisms. *Free. Radic. Biol. Med.* **2001**, *30*, 433–446. [CrossRef]
- 199. Miranda, S.; Opazo, C.; Larrondo, L.F.; Munoz, F.J. The role of oxidative stress in the toxicity induced by amyloid β-peptide in Alzheimer's disease. *Prog. Neurobiol.* **2000**, *62*, 633–648. [CrossRef]
- Jafari, A.; Babaei, P.; Rohampour, K.; Rashtiani, S. The Effect of Kaempferol on Autophagy and Nrf-2 Signaling in a Rat Model of Aβ1-42-induced Alzheimer's Disease. *Casp. J. Neurol. Sci.* 2022, 8, 7–16. [CrossRef]
- 201. Xie, C.; Zhuang, X.X.; Niu, Z.; Ai, R.; Lautrup, S.; Zheng, S.; Jiang, Y.; Han, R.; Gupta, T.S.; Cao, S.; et al. Amelioration of Alzheimer's disease pathology by mitophagy inducers identified via machine learning and a cross-species workflow. *Nat. Biomed. Eng.* 2022, *6*, 76–93. [CrossRef]

- Kaempferol: National Center for Biotechnology Information. PubChem Compound Summary for CID 5280863, Kaempferol. Retrieved 9 November 2023. 2023. Available online: https://pubchem.ncbi.nlm.nih.gov/compound/Kaempferol (accessed on 6 October 2023).
- Kouhestani, S.; Zare, S.; Babaei, P. Effects of pure flavonoid of medlar leaves on passive avoidance learning and memory in Alzheimer model of ovariectomized rats. J. Guilan Univ. Med. Sci. 2017, 26, 62–71.
- 204. Krishnaveni, M. Flavonoid in enhancing memory function. J. Pharm. Res. 2012, 5, 3870–3874.
- 205. Spencer, J.P. The impact of fruit flavonoids on memory and cognition. Br. J. Nutr. 2010, 104, 40–47. [CrossRef] [PubMed]
- 206. Liu, L.; Liu, Y.; Zhen, Y.; Guo, T.; Wang, C.; Shen, L.; Li, W. Quercetin inhibits cytotoxicity of PC12 cells induced by amyloid-beta 25–35 via stimulating estrogen receptor α, activating ERK1/2, and inhibiting apoptosis. Open Life Sci. 2022, 17, 230–242. [CrossRef]
- 207. Jin, S.; Zhang, L.; Wang, L. Kaempferol, a potential neuroprotective agent in neurodegenerative diseases: From chemistry to medicine. *Biomed. Pharmacother.* 2023, *165*, 115215. [CrossRef]
- Damirchi, A.; Hosseini, F.; Babaei, P. Mental Training Enhances Cognitive Function and BDNF More Than Either Physical or Combined Training in Elderly Women With MCI: A Small-Scale Study. *Am. J. Alzheimers Dis. Other Demen.* 2018, 33, 20–29. [CrossRef] [PubMed]
- Lee, J.; Fukumoto, H.; Orne, J.; Klucken, J.; Raju, S.; Vanderburg, C.R.; Irizarry, M.C.; Hyman, B.T.; Ingelsson, M. Decreased levels of BDNF protein in Alzheimer temporal cortex are independent of BDNF polymorphisms. *Exp. Neurol.* 2005, 194, 91–96. [CrossRef] [PubMed]
- Yan, T.; He, B.; Xu, M.; Wu, B.; Xiao, F.; Bi, K.; Jia, Y. Kaempferide prevents cognitive decline via attenuation of oxidative stress and enhancement of brain-derived neurotrophic factor/tropomyosin receptor kinase B/cAMP response element-binding signaling pathway. *Phytotherapy Res.* 2019, 33, 1065–1073. [CrossRef]
- 211. Amidfar, M.; de Oliveira, J.; Kucharska, E.; Budni, J.; Kim, Y.K. The role of CREB and BDNF in neurobiology and treatment of Alzheimer's disease. *Life Sci.* 2020, 257, 118020. [CrossRef]
- 212. Walton, M.R.; Dragunow, M. Is CREB a key to neuronal survival? Trends Neurosci. 2000, 23, 48–53. [CrossRef]
- Gao, Q.; Tian, D.; Han, Z.; Lin, J.; Chang, Z.; Zhang, D.; Ma, D. Network pharmacology and molecular docking analysis on molecular targets and mechanisms of buyang huanwu decoction in the treatment of ischemic stroke. *Evid. -Based Complement. Altern. Med.* 2021, 2021, 1–15. [CrossRef]
- 214. Wang, Z.-H.; Xiang, J.; Liu, X.; Yu, S.P.; Manfredsson, F.P.; Sandoval, I.M.; Wu, S.; Wang, J.Z.; Ye, K. Deficiency in BDNF/TrkB neurotrophic activity stimulates δ-secretase by upregulating C/EBPβ in Alzheimer's disease. *Cell Rep.* 2019, 28, 655–669. [CrossRef]
- Connor, B.; Young, D.; Yan, Q.; Faull, R.L.M.; Synek, B.; Dragunow, M. Brain-derived neurotrophic factor is reduced in Alzheimer's disease. *Mol. Brain Res.* 1997, 49, 71–81. [CrossRef] [PubMed]
- Levenga, J.; Wong, H.; Milstead, R.; LaPlante, L.; Hoeffer, C.A. Immunohistological Examination of AKT Isoforms in the Brain: Cell-Type Specificity That May Underlie AKT's Role in Complex Brain Disorders and Neurological Disease. *Cereb. Cortex Commun.* 2021, 2, tgab036. [CrossRef] [PubMed]
- Zarneshan, S.N.; Fakhri, S.; Khan, H. Targeting Akt/CREB/BDNF signaling pathway by ginsenosides in neurodegenerative diseases: A mechanistic approach. *Pharmacol. Res.* 2022, 177, 106099. [CrossRef] [PubMed]
- Pak, M.E.; Yang, H.J.; Li, W.; Kim, J.K.; Go, Y. Yuk-Gunja-Tang attenuates neuronal death and memory impairment via ERK/CREB/BDNF signaling in the hippocampi of experimental Alzheimer's disease model. *Front. Pharmacol.* 2022, 13, 1014840. [CrossRef] [PubMed]
- 219. Jain, V.; Baitharu, I.; Prasad, D.; Ilavazhagan, G. Enriched environment prevents hypobaric hypoxia induced memory impairment and neurodegeneration: Role of BDNF/PI3K/GSK3β pathway coupled with CREB activation. *PLoS ONE* 2013, *8*, e62235. [CrossRef]
- 220. Gao, L.; Zhang, Y.; Sterling, K.; Song, W. Brain-derived neurotrophic factor in Alzheimer's disease and its pharmaceutical potential. *Transl. Neurodegener.* 2022, 11, 4. [CrossRef]
- Rada, P.; Rojo, A.I.; Chowdhry, S.; McMahon, M.; Hayes, J.D.; Cuadrado, A. SCF/{beta}-TrCP promotes glycogen synthase kinase 3-dependent degradation of the Nrf2 transcription factor in a Keap1-independent manner. *Mol. Cell. Biol.* 2011, 31, 1121–1133. [CrossRef]
- 222. Kume, T.; Kouchiyama, H.; Kaneko, S.; Maeda, T.; Kaneko, S.; Akaike, A.; Shimohama, S.; Kihara, T.; Kimura, J.; Wada, K.; et al. BDNF prevents NO mediated glutamate cytotoxicity in cultured cortical neurons. *Brain Res.* **1997**, *756*, 200–204. [CrossRef]
- Mercado-Gómez, O.; Hernández-Fonseca, K.; Villavicencio-Queijeiro, A.; Massieu, L.; Chimal-Monroy, J.; Arias, C. Inhibition of Wnt and PI3K signaling modulates GSK-3beta activity and induces morphological changes in cortical neurons: Role of tau phosphorylation. *Neurochem. Res.* 2008, 33, 1599–1609. [CrossRef]
- Long, H.Z.; Cheng, Y.; Zhou, Z.W.; Luo, H.Y.; Wen, D.D.; Gao, L.C. PI3K/AKT Signal Pathway: A Target of Natural Products in the Prevention and Treatment of Alzheimer's Disease and Parkinson's Disease. *Front. Pharmacol.* 2021, 12, 648636. [CrossRef]
- 225. Jantas, D.; Malarz, J.; Le, T.N.; Stojakowska, A. Neuroprotective Properties of Kempferol Derivatives from Maesa membranacea against Oxidative Stress-Induced Cell Damage: An Association with Cathepsin D Inhibition and PI3K/Akt Activation. *Int. J. Mol. Sci.* 2021, 22, 10363. [CrossRef] [PubMed]
- 226. Wu, J.; Liu, H.; Chu, T.; Jiang, P.; Li, S.T. Neuregulin-1β attenuates sepsis-induced diaphragm atrophy by activating the PI3K/Akt signaling pathway. J. Muscle Res. Cell Motil. 2019, 40, 43–51. [CrossRef] [PubMed]

- 227. Kandezi, N.; Mohammadi, M.; Ghaffari, M.; Gholami, M.; Motaghinejad, M.; Safari, S. Novel Insight to Neuroprotective Potential of Curcumin: A Mechanistic Review of Possible Involvement of Mitochondrial Biogenesis and PI3/Akt/GSK3 or PI3/Akt/CREB/BDNF Signaling Pathways. Int. J. Mol. Cell. Med. 2020, 9, 1–32. [CrossRef] [PubMed]
- 228. Tanqueiro, S.R.; Ramalho, R.M.; Rodrigues, T.M.; Lopes, L.V.; Sebastião, A.M.; Diógenes, M.J. Inhibition of NMDA Receptors Prevents the Loss of BDNF Function Induced by Amyloid β. *Front. Pharmacol.* **2018**, *9*, 237. [CrossRef]
- Garzon, D.J.; Fahnestock, M. Oligomeric amyloid decreases basal levels of brain-derived neurotrophic factor (BDNF) mRNA via specific downregulation of BDNF transcripts IV and V in differentiated human neuroblastoma cells. *J. Neurosci.* 2007, 27, 2628–2635. [CrossRef]
- Tong, L.; Thornton, P.L.; Balazs, R.; Cotman, C.W. β-amyloid-(1–42) impairs activity-dependent cAMP-response element-binding protein signaling in neurons at concentrations in which cell survival is not compromised. *J. Biol. Chem.* 2001, 276, 17301–17306. [CrossRef]
- Cowansage, K.K.; LeDoux, J.E.; Monfils, M.H. Brain-derived neurotrophic factor: A dynamic gatekeeper of neural plasticity. *Curr. Mol. Pharmacol.* 2010, 3, 12–29. [CrossRef]
- Rosa, E.; Fahnestock, M. CREB expression mediates amyloid β-induced basal BDNF downregulation. *Neurobiol. Aging* 2015, 36, 2406–2413. [CrossRef]
- 233. DaRocha-Souto, B.; Coma, M.; Perez-Nievas, B.; Scotton, T.; Siao, M.; Sánchez-Ferrer, P.; Hashimoto, T.; Fan, Z.; Hudry, E.; Barroeta, I. Activation of glycogen synthase kinase-3 beta mediates β-amyloid induced neuritic damage in Alzheimer's disease. *Neurobiol. Dis.* 2012, 45, 425–437. [CrossRef]
- 234. Barco, A.; Pittenger, C.; Kandel, E.R. CREB, memory enhancement and the treatment of memory disorders: Promises, pitfalls and prospects. *Expert Opin. Ther. Targets* 2003, 7, 101–114. [CrossRef]
- 235. Christensen, R.; Marcussen, A.B.; Wörtwein, G.; Knudsen, G.; Aznar, S. Aβ (1–42) injection causes memory impairment, lowered cortical and serum BDNF levels, and decreased hippocampal 5-HT2A levels. *Exp. Neurol.* 2008, 210, 164–171. [CrossRef] [PubMed]
- 236. Ciaramella, A.; Salani, F.; Bizzoni, F.; Orfei, M.D.; Langella, R.; Angelucci, F.; Spalletta, G.; Taddei, A.R.; Caltagirone, C.; Bossù, P. The stimulation of dendritic cells by amyloid beta 1–42 reduces BDNF production in Alzheimer's disease patients. *Brain Behav. Immun.* 2013, 32, 29–32. [CrossRef] [PubMed]
- 237. Zussy, C.; Brureau, A.; Keller, E.; Marchal, S.; Blayo, C.; Delair, B.; Ixart, G.; Maurice, T.; Givalois, L. Alzheimer's disease related markers, cellular toxicity and behavioral deficits induced six weeks after oligomeric amyloid-β peptide injection in rats. *PLoS* ONE 2013, 8, e53117. [CrossRef] [PubMed]
- Wu, H.; Lu, D.; Jiang, H.; Xiong, Y.; Qu, C.; Li, B.; Mahmood, A.; Zhou, D.; Chopp, M. Simvastatin-mediated upregulation of VEGF and BDNF, activation of the PI3K/AKT pathway, and increase of neurogenesis are associated with therapeutic improvement after traumatic brain injury. J. Neurotrauma 2008, 25, 130–139. [CrossRef]
- Hu, Y.S.; Long, N.; Pigino, G.; Brady, S.T.; Lazarov, O. Molecular mechanisms of environmental enrichment: Impairments in AKT/GSK3β, neurotrophin-3 and CREB signaling. *PLoS ONE*. 2013, *8*, e64460. [CrossRef] [PubMed]
- 240. Nordberg, A. PET imaging of amyloid in Alzheimer's disease. *Lancet Neurol.* 2004, *3*, 519–527. [CrossRef]
- Caricasole, A.; Copani, A.; Caruso, A.; Caraci, F.; Iacovelli, L.; Sortino, M.A.; Terstappen, G.C.; Nicoletti, F. The Wnt pathway, cell-cycle activation and beta-amyloid: Novel therapeutic strategies in Alzheimer's disease? *Trends Pharmacol. Sci.* 2003, 24, 233–238. [CrossRef]
- Lee, C.W.; Lau, K.F.; Miller, C.C.; Shaw, P.C. Glycogen synthase kinase-3 beta-mediated tau phosphorylation in cultured cell lines. *Neuroreport* 2003, 14, 257–260. [CrossRef]
- Yao, R.Q.; Qi, D.S.; Yu, H.L.; Liu, J.; Yang, L.H.; Wu, X.X. Quercetin attenuates cell apoptosis in focal cerebral ischemia rat brain via activation of BDNF-TrkB-PI3K/Akt signaling pathway. *Neurochem. Res.* 2012, 37, 2777–2786. [CrossRef]
- 244. Datta, S.R.; Dudek, H.; Tao, X.; Masters, S.; Fu, H.; Gotoh, Y.; Greenberg, M.E. Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery. *Cell* **1997**, *91*, 231–241. [CrossRef]
- Cardone, M.H.; Roy, N.; Stennicke, H.R.; Salvesen, G.S.; Franke, T.F.; Stanbridge, E.; Frisch, S.; Reed, J.C. Regulation of cell death protease caspase-9 by phosphorylation. *Science* 1998, 282, 1318–1321. [CrossRef] [PubMed]
- 246. Cheng, B.; Martinez, A.A.; Morado, J.; Scofield, V.; Roberts, J.L.; Maffi, S.K. Retinoic acid protects against proteasome inhibition associated cell death in SH-SY5Y cells via the AKT pathway. *Neurochem. Int.* **2013**, *62*, 31–42. [CrossRef] [PubMed]
- Lee, K.Y.; Koh, S.H.; Noh, M.Y.; Park, K.W.; Lee, Y.J.; Kim, S.H. Glycogen synthase kinase-3beta activity plays very important roles in determining the fate of oxidative stress-inflicted neuronal cells. *Brain Res.* 2007, 1129, 89–99. [CrossRef] [PubMed]
- 248. Gu, Y.; Zhang, X.; Xu, A.; Chen, W.; Liu, K.; Wu, L.; Mo, S.; Hu, Y.; Liu, M.; Luo, Q. Protein-ligand binding affinity prediction with edge awareness and supervised attention. *iScience* 2022, *26*, 105892. [CrossRef]
- Jain, A.N. Surflex: Fully automatic flexible molecular docking using a molecular similarity-based search engine. *J. Med. Chem.* 2003, 46, 499–511. [CrossRef]
- 250. Simayi, J.; Bayinsang; Nuermaimaiti, M.; Hailati, S.; Han, M.; Reheman, Z.; Wumaier, A.; Zhou, W. A Network Pharmacology-Based Study on the Mechanism of Dibutyl Phthalate of Ocimum basilicum L. against Alzheimer's Disease through the AKT/GSK-3β Pathway. *BioMed Res. Int.* 2022, 2022, 9494548. [CrossRef]
- 251. Rahmatkar, S.N.; Rana, A.K.; Kumar, R.; Singh, D. Fagopyrum tataricum (L.) Gaertn interacts with Gsk-3β/Nrf-2 signalling to protect neurotoxicity in a zebrafish model. J. Ethnopharmacol. 2023, 319, 117187. [CrossRef]

- 252. Chiu, Y.J.; Teng, Y.S.; Chen, C.M.; Sun, Y.C.; Hsieh-Li, H.M.; Chang, K.H.; Lee-Chen, G.J. A Neuroprotective Action of Quercetin and Apigenin through Inhibiting Aggregation of Aβ and Activation of TRKB Signaling in a Cellular Experiment. *Biomol. Ther.* 2023, *31*, 285–297. [CrossRef]
- 253. Das, S.; Sengupta, S.; Chakraborty, S. Scope of β-secretase (bace1)-targeted therapy in alzheimer's disease: Emphasizing the flavonoid based natural scaffold for bace1 inhibition. *CS Chem. Neurosci.* **2020**, *11*, 3510–3522. [CrossRef]
- 254. Shimmyo, Y.; Kihara, T.; Akaike, A.; Niidome, T.; Sugimoto, H. Flavonols and flavones as BACE-1 inhibitors: Structure–activity relationship in cell-free, cell-based and in silico studies reveal novel pharmacophore features. *Biochim. Et Biophys. Acta (BBA)-Gen. Subj.* 2008, 1780, 819–825. [CrossRef]
- 255. Li, E.; Yan, K.; Zhang, R.; Zou, P.; Li, S.; Ma, Q.; Liao, B. Kaempferol Protects Against Apoptosis in PC12 Cells Exposed to Hydrogen Peroxide by Activating Akt1. Nat. Prod. Commun. 2023, 18, 1934578X231170448. [CrossRef]
- 256. Zhang, S.; Lu, Y.; Chen, W.; Shi, W.; Zhao, Q.; Zhao, J.; Li, L. Network Pharmacology and Experimental Evidence: PI3K/AKT Signaling Pathway is Involved in the Antidepressive Roles of Chaihu Shugan San. Drug Des. Dev. Ther. 2021, 15, 3425–3441. [CrossRef]
- 257. Shi, Y.; Chen, M.; Zhao, Z.; Pan, J.; Huang, S. Network pharmacology and molecular docking analyses of mechanisms underlying effects of the cyperi rhizoma-chuanxiong rhizoma herb pair on depression. *Evid. -Based Complement. Altern. Med.* 2021, 2021, 5704578. [CrossRef] [PubMed]
- 258. Zhang, S.S.; Liu, M.; Liu, D.N.; Shang, Y.F.; Du, G.H.; Wang, Y.H. Network pharmacology analysis and experimental validation of kaempferol in the treatment of ischemic stroke by inhibiting apoptosis and regulating neuroinflammation involving neutrophils. *Int. J. Mol. Sci.* 2022, 23, 12694. [CrossRef] [PubMed]
- Touati, I.; Abdalla, M.; Ali, N.H.; AlRuwaili, R.; Alruwaili, M.; Britel, M.R.; Maurady, A. Constituents of Stachys plants as potential dual inhibitors of AChE and NMDAR for the treatment of Alzheimer's disease: A molecular docking and dynamic simulation study. J. Biomol. Struct. Dyn. 2023, 1–17. [CrossRef]
- Rasouli, H.; Hosseini Ghazvini, S.M.B.; Yarani, R.; Altıntaş, A.; Jooneghani, S.G.N.; Ramalho, T.C. Deciphering inhibitory activity of flavonoids against tau protein kinases: A coupled molecular docking and quantum chemical study. *J. Biomol. Struct. Dyn.* 2022, 40, 411–424. [CrossRef]
- 261. Omar, S.H.; Scott, C.J.; Hamlin, A.S.; Obied, H.K. Biophenols: Enzymes (β-secretase, Cholinesterases, histone deacetylase and tyrosinase) inhibitors from olive (*Olea europaea* L.). *Fitoterapia* 2018, 128, 118–129. [CrossRef]
- Chukwuma, I.F.; Ezeorba, T.P.C.; Nworah, F.N.; Apeh, V.O.; Khalid, M.; Sweilam, S.H. Bioassay-guided identification of potential Alzheimer's disease therapeutic agents from Kaempferol-Enriched fraction of Aframomum melegueta seeds using in vitro and chemoinformatics approaches. *Arab. J. Chem.* 2023, *16*, 105089. [CrossRef]
- Pandey, D.; Pal, T.; Sharma, A. Phytochemicals as Potential Anti-Alzheimer's Agents- An In-Silico Evidence. J. Dis. Markers 2022, 7, 1047.
- 264. Grewal, A.K.; Singh, T.G.; Sharma, D.; Sharma, V.; Singh, M.; Rahman, M.H.; Najda, A.; Walasek-Janusz, M.; Kamel, M.; Albadrani, G.M.; et al. Mechanistic insights and perspectives involved in neuroprotective action of quercetin. *Biomed. Pharmacother.* 2021, 140, 111729. [CrossRef]
- 265. Gong, P.; Wang, D.; Cui, D.; Yang, Q.; Wang, P.; Yang, W.; Chen, F. Anti-aging function and molecular mechanism of Radix Astragali and Radix Astragali preparata via network pharmacology and PI3K/Akt signaling pathway. *Phytomedicine* 2021, 84, 153509. [CrossRef]
- 266. Sadighparvar, S.; Darband, S.G.; Yousefi, B.; Kaviani, M.; Ghaderi-Pakdel, F.; Mihanfar, A.; Mobaraki, K.; Majidinia, M. Combination of quercetin and exercise training attenuates depression in rats with 1,2-dimethylhydrazine-induced colorectal cancer: Possible involvement of inflammation and BDNF signalling. *Exp. Physiol.* 2020, 105, 1598–1609. [CrossRef]
- 267. Khan, H.; Singh, A.; Thapa, K.; Garg, N.; Grewal, A.K.; Singh, T.G. Therapeutic modulation of the phosphatidylinositol 3-kinases (PI3K) pathway in cerebral ischemic injury. *Brain Res.* 2021, 1761, 147399. [CrossRef] [PubMed]
- Rezai-Zadeh, K.; Arendash, G.W.; Hou, H.; Fernandez, F.; Jensen, M.; Runfeldt, M.; Shytle, R.D.; Tan, J. Green tea epigallocatechin-3-gallate (EGCG) reduces beta-amyloid mediated cognitive impairment and modulates tau pathology in Alzheimer transgenic mice. *Brain Res.* 2008, 1214, 177–187. [CrossRef] [PubMed]
- Gong, E.J.; Park, H.R.; Kim, M.E.; Piao, S.; Lee, E.; Jo, D.G.; Chung, H.Y.; Ha, N.C.; Mattson, M.P.; Lee, J. Morin attenuates tau hyperphosphorylation by inhibiting GSK3beta. *Neurobiol. Dis.* 2011, 44, 223–230. [CrossRef] [PubMed]
- Miyai, S.; Yamaguchi, A.; Iwasaki, T.; Shamsa, F.; Ohtsuki, K. Biochemical characterization of epigallocatechin-3-gallate as an effective stimulator for the phosphorylation of its binding proteins by glycogen synthase kinase-3beta in vitro. *Biol. Pharm. Bull.* 2010, 33, 1932–1937. [CrossRef]
- Ashrafpour, M.; Parsaei, S.; Sepehri, H. Quercetin improved spatial memory dysfunctions in rat model of intracerebroventricular streptozotocin-induced Alzheimer's disease. *Natl. J. Physiol. Pharm. Pharmacol.* 2015, 5, 411. [CrossRef]
- 272. Tong-un, T.; Muchimapura, S.; Phachonpai, W.; Wattanathorn, J. Effects of quercetin encapsulated liposomes via nasal administration: A novel cognitive enhancer. *Am. J. Appl. Sci.* 2010, *7*, 906–913. [CrossRef]
- 273. Sriraksa, N.; Wattanathorn, J.; Muchimapura, S.; Tiamkao, S.; Brown, K.; Chaisiwamongkol, K. Cognitive-enhancing effect of quercetin in a rat model of Parkinson's disease induced by 6-hyrodoxydopamine. *J. Evid. Based Complement. Alternat. Med.* 2011, 2012, 823206. [CrossRef]

- 274. Elreedy, H.A.; Elfiky, A.M.; Ahmed Mahmoud, A.; Ibrahim, K.S.; Ghazy, M.A. Effect Of Quercetin As Therapeutic And Protective Agent In Aluminum Chloride-Induced Alzheimer's Disease Rats. *Egypt. J. Chem.* 2022, 65, 633–641. [CrossRef]
- 275. Elfiky, A.M.; Mahmoud, A.A.; Elreedy, H.A.; Ibrahim, K.S.; Ghazy, M.A. Quercetin stimulates the non-amyloidogenic pathway via activation of ADAM10 and ADAM17 gene expression in aluminum chloride-induced Alzheimer's disease rat model. *Life Sci.* 2021, 285, 119964. [CrossRef] [PubMed]
- 276. Singh, N.K.; Garabadu, D. Quercetin Exhibits α7nAChR/Nrf2/HO-1-Mediated Neuroprotection Against STZ-Induced Mitochondrial Toxicity and Cognitive Impairments in Experimental Rodents. *Neurotox. Res.* 2021, 39, 1859–1879. [CrossRef]
- 277. Parent, M.; Chitturi, J.; Santhakumar, V.; Hyder, F.; Sanganahalli, B.G.; Kannurpatti, S.S. Kaempferol Treatment after Traumatic Brain Injury during Early Development Mitigates Brain Parenchymal Microstructure and Neural Functional Connectivity Deterioration at Adolescence. J. Neurotrauma 2020, 37, 966–974. [CrossRef] [PubMed]
- 278. Zhang, W.; Lv, M.; Shi, Y.; Mu, Y.; Yao, Z.; Yang, Z. Network Pharmacology-Based Study of the Underlying Mechanisms of Huangqi Sijunzi Decoction for Alzheimer's Disease. *Evid. Based Complement. Altern. Med.* 2021, 2021, 6480381. [CrossRef] [PubMed]
- Matsuo, E.S.; Shin, R.W.; Billingsley, M.L.; van de Voorde, A.; O'Connor, M.; Trojanowski, J.Q.; Lee, V.M. Biopsy-derived adult human brain tau is phosphorylated at many of the same sites as Alzheimer's disease paired helical filament tau. *Neuron* 1994, 13, 989–1002. [CrossRef]
- 280. Iqbal, K.; Liu, F.; Gong, C.X. Tau and neurodegenerative disease: The story so far. Nat. Rev. Neurol. 2016, 12, 15–27. [CrossRef]
- Xia, Y.; Liu, R.; Chen, R.; Tian, Q.; Zeng, K.; Hu, J.; Liu, X.; Wang, Q.; Wang, P.; Wang, X.C.; et al. Novel multipotent AChEI-CCB attenuates hyperhomocysteinemia-induced memory deficits and Neuropathologies in rats. J. Alzheimer's Dis. 2014, 42, 1029–1039. [CrossRef]
- Russo, M.; Milito, A.; Spagnuolo, C.; Carbone, V.; Rosén, A.; Minasi, P.; Lauria, F.; Russo, G.L. CK2 and PI3K are direct molecular targets of quercetin in chronic lymphocytic leukaemia. *Oncotarget* 2017, *8*, 42571–42587. [CrossRef]
- 283. Images Created. Available online: BioRender.com (accessed on 6 October 2023).
- 284. Fang, J.; Wang, L.; Wu, T.; Yang, C.; Gao, L.; Cai, H.; Liu, J.; Fang, S.; Chen, Y.; Tan, W.; et al. Network pharmacology-based study on the mechanism of action for herbal medicines in Alzheimer treatment. J. Ethnopharmacol. 2017, 196, 281–292. [CrossRef]
- 285. Luo, Y.; Smith, J.V.; Paramasivam, V.; Burdick, A.; Curry, K.J.; Buford, J.P.; Khan, I.; Netzer, W.J.; Xu, H.; Butko, P. Inhibition of amyloid-β aggregation and caspase-3 activation by the *Ginkgo biloba* extract EGb761. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 12197–12202. [CrossRef]
- Longpré, F.; Garneau, P.; Christen, Y.; Ramassamy, C. Protection by EGb 761 against beta-amyloid-induced neurotoxicity: Involvement of NF-kappaB, SIRT1, and MAPKs pathways and inhibition of amyloid fibril formation. *Free. Radic. Biol. Med.* 2006, 41, 1781–1794. [CrossRef]
- Tchantchou, F.; Xu, Y.; Wu, Y.; Christen, Y.; Luo, Y. EGb 761 enhances adult hippocampal neurogenesis and phosphorylation of CREB in transgenic mouse model of Alzheimer's disease. *FASEB J.* 2007, 21, 2400–2408. [CrossRef] [PubMed]
- 288. Kwon, K.J.; Lee, E.J.; Cho, K.S.; Cho, D.H.; Shin, C.Y.; Han, S.H. *Ginkgo biloba* extract (Egb761) attenuates zinc-induced tau phosphorylation at Ser262 by regulating GSK3β activity in rat primary cortical neurons. *Food Funct.* 2015, *6*, 2058–2067. [CrossRef] [PubMed]
- Lejri, I.; Grimm, A.; Eckert, A. *Ginkgo biloba* extract increases neurite outgrowth and activates the Akt/mTOR pathway. *PLoS ONE*. 2019, 14, e0225761. [CrossRef] [PubMed]
- 290. Rhein, V.; Song, X.; Wiesner, A.; Ittner, L.M.; Baysang, G.; Meier, F.; Ozmen, L.; Bluethmann, H.; Dröse, S.; Brandt, U.; et al. Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proc. Natl. Acad. Sci. USA* 2009, 106, 20057–20062. [CrossRef]
- 291. Muller, W.E.; Heiser, J.; Leuner, K. Effects of the standardized *Ginkgo biloba* extract EGb 761(R) on neuroplasticity. *Int. Psychogeriatrics* 2012, 24 (Suppl. S1), S21–S24. [CrossRef]
- Xu, Y.; Cui, C.; Pang, C.; Christen, Y.; Luo, Y. Restoration of impaired phosphorylation of cyclic AMP response element-binding protein (CREB) by EGb 761 and its constituents in Abeta-expressing neuroblastoma cells. *Eur. J. Neurosci.* 2007, 26, 2931–2939. [CrossRef]
- 293. Tchantchou, F.; Lacor, P.N.; Cao, Z.; Lao, L.; Hou, Y.; Cui, C.; Klein, W.L.; Luo, Y. Stimulation of neurogenesis and synaptogenesis by bilobalide and quercetin via common final pathway in hippocampal neurons. J. Alzheimer's Dis. 2009, 18, 787–798. [CrossRef]
- 294. Tian, R.; Wang, H.; Xiao, Y.; Hu, P.; Du, R.; Shi, X.; Wang, Z.; Xie, Y. Fabrication of nanosuspensions to improve the oral bioavailability of total flavones from Hippophae rhamnoides L. and their comparison with an inclusion complex. *AAPS Pharmscitech* **2020**, *21*, 249. [CrossRef]
- 295. Xia, C.X.; Gao, A.X.; Zhu, Y.; Dong, T.T.; Tsim, K.W. Flavonoids from Seabuckthorn (*Hippophae rhamnoides* L.) restore CUMSinduced depressive disorder and regulate the gut microbiota in mice. *Food Funct.* 2023, 14, 7426–7438. [CrossRef]
- Liu, H.; Ye, M.; Guo, H. An Updated Review of Randomized Clinical Trials Testing the Improvement of Cognitive Function of Ginkgo biloba Extract in Healthy People and Alzheimer's Patients. Front. Pharmacol. 2020, 10, 1688. [CrossRef] [PubMed]
- 297. Jeganathan, B.; Punyasiri, P.A.; Kottawa-Arachchi, J.D.; Ranatunga, M.A.; Abeysinghe, I.S.; Gunasekare, M.T.; Bandara, B.M. Genetic Variation of Flavonols Quercetin, Myricetin, and Kaempferol in the Sri Lankan Tea (*Camellia sinensis* L.) and Their Health-Promoting Aspects. *Int. J. Food Sci.* 2016, 2016, 6057434. [CrossRef] [PubMed]

- 298. Ishan, T.; Lalit, S.; Girdhari, G.; Gupta, G.L. Synergistic antioxidant activity of three medicinal plants Hypericum perforatum, Bacopa monnieri, and *Camellia sinensis*. *Indo Am. J. Pharm. Res.* **2014**, *4*, 2563–2568.
- 299. Mahmoodzadeh, T.; Kashani, M.H.K.; Ramshini, H.; Moslem, A.; Mohammad-Zadeh, M. Effect of *Camellia sinensis* on Spatial Memory in a Rat Model of Alzheimer's Disease. J. Biomed. 2016, 1, e5340. [CrossRef]
- Onasanwo, S.A.; Adamaigbo, V.O.; Adebayo, O.G.; Eleazer, S.E. Moringa oleifera-supplemented diet protect against corticohippocampal neuronal degeneration in scopolamine-induced spatial memory deficit in mice: Role of oxido-inflammatory and cholinergic neurotransmission pathway. *Metab. Brain Dis.* 2021, *36*, 2445–2460. [CrossRef]
- Sermkaew, N.; Plyduang, T. Self-microemulsifying drug delivery systems of Moringa oleifera extract for enhanced dissolution of kaempferol and quercetin. Acta Pharm. 2020, 70, 77–88. [CrossRef]
- Liu, W.L.; Wu, B.F.; Shang, J.H.; Wang, X.F.; Zhao, Y.L.; Huang, A.X. Moringa oleifera seed ethanol extract and its active component kaempferol potentiate pentobarbital-induced sleeping behaviours in mice via a GABAergic mechanism. *Pharm. Biol.* 2022, 60, 810–824. [CrossRef]
- 303. Sun, Y.; Wu, A.; Li, X.; Qin, D.; Jin, B.; Liu, J.; Tang, Y.; Wu, J.; Yu, C. The seed of Litchi chinensis fraction ameliorates hippocampal neuronal injury in an Aβ25-35-induced Alzheimer's disease rat model via the AKT/GSK-3β pathway. *Pharm. Biol.* 2020, 58, 35–43. [CrossRef]
- 304. El Gizawy, H.A.; Abo-Salem, H.M.; Ali, A.A.; Hussein, M.A. Phenolic Profiling and Therapeutic Potential of Certain Isolated Compounds from Parkia roxburghii against AChE Activity as well as GABAA α5, GSK-3β, and p38α MAP-Kinase Genes. ACS Omega 2021, 6, 20492–20511. [CrossRef]
- Singh, S.; Singh, T.G.; Mahajan, K.; Dhiman, S. Medicinal plants used against various inflammatory biomarkers for the management of rheumatoid arthritis. J. Pharm. Pharmacol. 2020, 72, 1306–1327. [CrossRef]
- Liu, Y.; Xue, Q.; Li, A.; Li, K.; Qin, X. Mechanisms exploration of herbal pair of HuangQi-DanShen on cerebral ischemia based on metabonomics and network pharmacology. J. Ethnopharmacol. 2020, 253, 112688. [CrossRef] [PubMed]
- 307. Moriwaki, M.; Tominaga, E.; Kito, K.; Nakagawa, R.; Kapoor, M.P.; Matsumiya, Y.; Fukuhara, T.; Kobayashi, J.; Satomoto, K.; Yamagata, H.; et al. Bioavailability of Flavonoids in *Ginkgo biloba* Extract-γ-Cyclodextrin Complex. *Nat. Prod. Commun.* 2023, 18, 1934578X231170221. [CrossRef]
- 308. Zubčić, K.; Radovanović, V.; Vlainić, J.; Hof, P.R.; Oršolić, N.; Šimić, G.; Jazvinšćak Jembrek, M. PI3K/Akt and ERK1/2 Signalling Are Involved in Quercetin-Mediated Neuroprotection against Copper-Induced Injury. Oxidative Med. Cell. Longev. 2020, 2020, 9834742. [CrossRef] [PubMed]
- Permana, A.D.; Maddeppungeng, N.M.; Asma, N.; Rahim, A.; Nainu, F.; Bahar, M.A.; Yulianty, R. Validation of HPLC-UV method for simultaneous determination of quercetin and luteolin from chartamus tinctorius L in solid lipid nanoparticles incorporated in floating gel in situ formulation. *Microchem. J.* 2023, 194, 109373. [CrossRef]
- 310. Cho, K.M.; Lee, H.Y.; Cho, D.Y.; Jung, J.G.; Kim, M.J.; Bin Jeong, J.; Jang, S.N.; Lee, G.O.; Sim, H.-S.; Kang, M.J.; et al. Comprehensive Comparison of Chemical Composition and Antioxidant Activity of Panax ginseng Sprouts by Different Cultivation Systems in a Plant Factory. *Plants* 2022, *11*, 1818. [CrossRef]
- Lin, M.K.; Lee, M.S.; Huang, H.C.; Cheng, T.J.; Cheng, Y.D.; Wu, C.R. Cuscuta chinensis and C. campestris attenuate scopolamineinduced memory deficit and oxidative damage in mice. *Molecules* 2018, 23, 3060. [CrossRef]
- Marefati, N.; Ghorani, V.; Shakeri, F.; Boskabady, M.; Kianian, F.; Rezaee, R.; Boskabady, M.H. A review of anti-inflammatory, antioxidant, and immunomodulatory effects of Allium cepa and its main constituents. *Pharm. Biol.* 2021, 59, 285–300. [CrossRef]
- 313. Sagar, N.A.; Pareek, S.; Benkeblia, N.; Xiao, J. Onion (*Allium cepa* L.) bioactives: Chemistry, pharmacotherapeutic functions, and industrial applications. *Food Front.* **2022**, *3*, 380–412. [CrossRef]
- 314. Tan, S.; Tang, J.; Shi, W.; Wang, Z.; Xiang, Y.; Deng, T.; Gao, X.; Li, W.; Shi, S. Effects of three drying methods on polyphenol composition and antioxidant activities of Litchi chinensis Sonn. *Food Sci. Biotechnol.* **2019**, *29*, 351–358. [CrossRef]
- Singh, B.; Singh, J.P.; Kaur, A.; Singh, N. Phenolic compounds as beneficial phytochemicals in pomegranate (*Punica granatum* L.) peel: A review. *Food Chem.* 2018, 261, 75–86. [CrossRef]
- 316. Chen, Z.; Liu, L.; Gao, C.; Chen, W.; Vong, C.T.; Yao, P.; Yang, Y.; Li, X.; Tang, X.; Wang, S.; et al. Astragali Radix (Huangqi): A promising edible immunomodulatory herbal medicine. J. Ethnopharmacol. 2020, 258, 112895. [CrossRef]
- Pool, H.; Mendoza, S.; Xiao, H.; McClements, D.J. Encapsulation and release of hydrophobic bioactive components in nanoemulsion-based delivery systems: Impact of physical form on quercetin bioaccessibility. *Food Funct.* 2013, 4, 162–174. [CrossRef]
- 318. Bangar, S.P.; Chaudhary, V.; Sharma, N.; Bansal, V.; Ozogul, F.; Lorenzo, J.M. Kaempferol: A flavonoid with wider biological activities and its applications. *Crit. Rev. Food Sci. Nutr.* **2022**, 1–25. [CrossRef]
- Elahy, M.; Jackaman, C.; Mamo, J.C.; Lam, V.; Dhaliwal, S.S.; Giles, C.; Nelson, D.; Takechi, R. Blood-brain barrier dysfunction developed during normal aging is associated with inflammation and loss of tight junctions but not with leukocyte recruitment. *Immun. Ageing* 2015, 12, 2. [CrossRef] [PubMed]
- Popescu, B.O.; Toescu, E.C.; Popescu, L.M.; Bajenaru, O.; Muresanu, D.F.; Schultzberg, M.; Bogdanovic, N. Blood-brain barrier alterations in ageing and dementia. J. Neurol. Sci. 2009, 283, 99–106. [CrossRef] [PubMed]
- 321. Leoni, V.; Masterman, T.; Patel, P.; Meaney, S.; Diczfalusy, U.; Björkhem, I. Side chain oxidized oxysterols in cerebrospinal fluid and the integrity of blood-brain and blood-cerebrospinal fluid barriers. J. Lipid Res. 2003, 44, 793–799. [CrossRef] [PubMed]

- 322. Blair, L.J.; Frauen, H.D.; Zhang, B.; Nordhues, B.A.; Bijan, S.; Lin, Y.-C.; Zamudio, F.; Hernandez, L.D.; Sabbagh, J.J.; Selenica, M.-L.B.; et al. Tau depletion prevents progressive blood-brain barrier damage in a mouse model of tauopathy. *Acta Neuropathol. Commun.* **2015**, *3*, 8. [CrossRef]
- 323. Majerova, P.; Michalicova, A.; Cente, M.; Hanes, J.; Vegh, J.; Kittel, A.; Kosikova, N.; Cigankova, V.; Mihaljevic, S.; Jadhav, S.; et al. Trafficking of immune cells across the blood-brain barrier is modulated by neurofibrillary pathology in tauopathies. *PLoS ONE* 2019, 14, e0217216. [CrossRef]
- 324. Moradi-Afrapoli, F.; Oufir, M.; Walter, F.R.; Deli, M.A.; Smiesko, M.; Zabela, V.; Butterweck, V.; Hamburger, M. Validation of UHPLC-MS/MS methods for the determination of kaempferol and its metabolite 4-hydroxyphenyl acetic acid, and application to in vitro blood-brain barrier and intestinal drug permeability studies. J. Pharm. Biomed. Anal. 2016, 128, 264–274. [CrossRef]
- 325. Gupta, A.; Kaur, C.D.; Saraf, S.; Saraf, S. Formulation, characterization, and evaluation of ligand-conjugated biodegradable quercetin nanoparticles for active targeting. *Artif. Cells Nanomed. Biotechnol.* **2016**, *44*, 960–970. [CrossRef]
- 326. Dong, Y.; Tao, B.; Xue, X.; Feng, C.; Ren, Y.; Ma, H.; Zhang, J.; Si, Y.; Zhang, S.; Liu, S.; et al. Molecular mechanism of Epicedium treatment for depression based on network pharmacology and molecular docking technology. *BMC Complement. Med. Ther.* 2021, 21, 222. [CrossRef] [PubMed]
- 327. Wu, D.; Zhang, L.; Zhang, S. Distribution of quercetin in plasma and tissues in rats. Chin. J. Hosp. Pharm. 2008, 28, 1822–1824.
- 328. Orhan, I.E. Cholinesterase Inhibitory Potential of Quercetin towards Alzheimer's Disease—A Promising Natural Molecule or Fashion of the Day?—A Narrowed Review. *Curr. Neuropharmacol.* **2021**, *19*, 2205–2213. [CrossRef] [PubMed]
- 329. Imran, M.; Thabet, H.K.; Alaqel, S.I.; Alzahrani, A.R.; Abida, A.; Alshammari, M.K.; Kamal, M.; Diwan, A.; Asdaq, S.M.B.; Alshehri, S. The Therapeutic and Prophylactic Potential of Quercetin against COVID-19: An Outlook on the Clinical Studies, Inventive Compositions, and Patent Literature. *Antioxidants* 2022, 11, 876. [CrossRef] [PubMed]
- 330. Huebbe, P.; Wagner, A.E.; Boesch-Saadatmandi, C.; Sellmer, F.; Wolffram, S.; Rimbach, G. Effect of dietary quercetin on brain quercetin levels and the expression of antioxidant and Alzheimer's disease relevant genes in mice. *Pharmacol. Res.* **2010**, *61*, 242–246. [CrossRef]
- Crozier, A.; Del Rio, D.; Clifford, M.N. Bioavailability of Dietary Flavonoids and Phenolic Compounds. *Mol. Asp. Med.* 2010, 31, 446–467. [CrossRef]
- 332. Guo, Y.; Bruno, R.S. Endogenous and exogenous mediators of quercetin bioavailability. J. Nutr. Biochem. 2015, 26, 201–210. [CrossRef]
- 333. Ebrahimpour, S.; Zakeri, M.; Esmaeili, A. Crosstalk between obesity, diabetes, and alzheimer's disease: Introducing quercetin as an effective triple herbal medicine. *Ageing Res. Rev.* **2020**, *62*, 101095. [CrossRef]
- 334. Pinheiro, R.; Granja, A.; Loureiro, J.; Pereira, M.; Pinheiro, M.; Neves, A.; Reis, S. RVG29-Functionalized Lipid Nanoparticles for Quercetin Brain Delivery and Alzheimer's Disease. *Pharm. Res.* 2020, *37*, 139. [CrossRef]
- 335. Qi, Y.; Yi, P.; He, T.; Song, X.; Liu, Y.; Li, Q.; Zheng, J.; Song, R.; Liu, C.; Zhang, Z.; et al. Quercetin-loaded selenium nanoparticles inhibit amyloid-β aggregation and exhibit antioxidant activity. *Colloids Surf. A Physicochem. Eng. Asp.* 2020, 602, 125058. [CrossRef]
- 336. Chen, G.; Yang, F.; Fan, S.; Jin, H.; Liao, K.; Li, X.; Liu, G.-B.; Liang, J.; Zhang, J.; Xu, J.F.; et al. Immunomodulatory roles of selenium nanoparticles: Novel arts for potential immunotherapy strategy development. *Front. Immunol.* 2022, 13, 956181. [CrossRef] [PubMed]
- Kieliszek, M.; Błażejak, S.; Bzducha-Wróbel, A.; Kot, A.M. Effect of selenium on growth and antioxidative system of yeast cells. *Mol. Biol. Rep.* 2019, 46, 1797–1808. [CrossRef] [PubMed]
- Colombo, M.; de Lima Melchiades, G.; Michels, L.R.; Figueiró, F.; Bassani, V.L.; Teixeira, H.F.; Koester, L.S. Solid Dispersion of Kaempferol: Formulation Development, Characterization, and Oral Bioavailability Assessment. AAPS PharmSciTech 2019, 20, 106. [CrossRef] [PubMed]
- 339. Alkandahri, M.Y.; Pamungkas, B.T.; Oktoba, Z.; Shafirany, M.Z.; Sulastri, L.; Arfania, M.; Anggraeny, E.N.; Pratiwi, A.; Astuti, F.D.; Indriyani, D.S.Y.; et al. Hepatoprotective Effect of Kaempferol: A Review of the Dietary Sources, Bioavailability, Mechanisms of Action, and Safety. *Adv. Pharmacol. Pharm. Sci.* 2023, 2023, 1387665. [CrossRef] [PubMed]
- 340. Zhang, M.; Kong, L.; Luo, C.; Li, X.; Zhou, Y. Pharmacokinetic Study of Keampferol in Rabbits. China Pharm. 2014, 25, 4040–4042.
- 341. Salehi, B.; Machin, L.; Monzote, L.; Sharifi-Rad, J.; Ezzat, S.M.; Salem, M.A.; Merghany, R.M.; El Mahdy, N.M.; Kılıç, C.S.; Sytar, O.; et al. Therapeutic Potential of Quercetin: New Insights and Perspectives for Human Health. ACS Omega 2020, 5, 11849–11872. [CrossRef]
- 342. Liu, Y.; Gong, Y.; Xie, W.; Huang, A.; Yuan, X.; Zhou, H.; Zhu, X.; Chen, X.; Liu, J.; Liu, J.; et al. Microbubbles in combination with focused ultrasound for the delivery of quercetin-modified sulfur nanoparticles through the blood brain barrier into the brain parenchyma and relief of endoplasmic reticulum stress to treat Alzheimer's disease. *Nanoscale* 2020, 12, 6498–6511.
- Chandekar, L.; Katgeri, R.; Takke, A. The Potential Clinical Uses and Nanoformulation Strategies of Kaempferol, a Dietary Flavonoid. *Rev. Bras. de Farm.* 2022, 32, 693–707.
- Benameur, T.; Soleti, R.; Porro, C. The Potential Neuroprotective Role of Free and Encapsulated Quercetin Mediated by miRNA against Neurological Diseases. *Nutrients* 2021, 13, 1318. [CrossRef]
- 345. Nishimura, M.; Ohkawara, T.; Nakagawa, T.; Muro, T.; Sato, Y.; Satoh, H.; Kobori, M.; Nishihira, J. A randomized, double-blind, placebo-controlled study evaluating the effects of quercetin-rich onion on cognitive function in elderly subjects. *Funct. Foods Heal. Dis.* 2017, 7, 353. [CrossRef]

- 346. Broman-Fulks, J.J.; Canu, W.H.; Trout, K.L.; Nieman, D.C. The effects of quercetin supplementation on cognitive functioning in a community sample: A randomized, placebo-controlled trial. *Ther. Adv. Psychopharmacol.* **2012**, *2*, 131–138. [CrossRef]
- 347. Nakagawa, T.; Itoh, M.; Ohta, K.; Hayashi, Y.; Hayakawa, M.; Yamada, Y.; Akanabe, H.; Chikaishi, T.; Nakagawa, K.; Itoh, Y.; et al. Improvement of memory recall by quercetin in rodent contextual fear conditioning and human early-stage Alzheimer's disease patients. *Neuroreport* 2016, 27, 671–676. [CrossRef] [PubMed]
- Liu, Y.; Zhou, H.; Yin, T.; Gong, Y.; Yuan, G.; Chen, L.; Liu, J. Quercetin-modified gold-palladium nanoparticles as a potential autophagy inducer for the treatment of Alzheimer's disease. J. Colloid Interface Sci. 2019, 552, 388–400. [CrossRef] [PubMed]
- Rananaware, P.; Pandit, P.; Naik, S.; Mishra, M.; Keria, R.S.; Brahmkhatri, V.P. Anti-amyloidogenic property of gold nanoparticle decorated querc. RSC Adv. 2022, 12, 23661–23674. [CrossRef] [PubMed]
- 350. Zhu, C.; Yang, Y.; Li, X.; Chen, X.; Lin, X.; Wu, X. Develop potential multi-target drugs by self-assembly of quercetin with amino acids and metal ion to achieve significant efficacy in anti-Alzheimer's disease. *Nano Res.* **2022**, *15*, 5173–5182.
- Cui, Z.; Zhao, X.; Amevor, F.K.; Du, X.; Wang, Y.; Li, D.; Shu, G.; Tian, Y.; Zhao, X. Therapeutic application of quercetin in aging-related diseases: SIRT1 as a potential mechanism. *Front. Immunol.* 2022, *13*, 943321. [CrossRef] [PubMed]
- 352. Karthika, C.; Appu, A.P.; Akter, R.; Rahman, H.; Tagde, P.; Ashraf, G.M.; Abdel-Daim, M.M.; Ul Hassan, S.S.; Abid, A.; Bungau, S. Potential innovation against Alzheimer's disorder: A tricomponent combination of natural antioxidants (vitamin E, quercetin, and basil oil) and the development of its intranasal delivery. *Environ. Sci. Pollut. Res.* 2022, 29, 10950–10965.
- Alaqeel, N.K.; AlSheikh, M.H.; Al-Hariri, M.T. Quercetin Nanoemulsion Ameliorates Neuronal Dysfunction in Experimental Alzheimer's Disease Model. *Antioxidants* 2022, 11, 19866. [CrossRef]
- 354. Center for Food Safety and Applied Nutrition. (n.d.). Gras Notice Inventory—Agency Response Letter Gras Notice no. GRN 000341. Archive. Available online: https://www.cfsanappsexternal.fda.gov/scripts/fdcc/index.cfm?set=GRASNotices&id=341 (accessed on 6 October 2023).
- 355. Batiha, G.E.; Beshbishy, A.M.; Ikram, M.; Mulla, Z.S.; El-Hack, M.E.A.; Taha, A.E.; Algammal, A.M.; Elewa, Y.H.A. The Pharmacological Activity, Biochemical Properties, and Pharmacokinetics of the Major Natural Polyphenolic Flavonoid: Quercetin. *Foods* 2020, 9, 374. [CrossRef]
- Spencer, J.P.; Rice-Evans, C.; Williams, R.J. Modulation of pro-survival Akt/protein kinase B and ERK1/2 signaling cascades by quercetin and its in vivo metabolites underlie their action on neuronal viability. J. Biol. Chem. 2003, 278, 34783–34793. [CrossRef]
- 357. Kimoto, H.; Fujiwara, S.; Koyama, N.; Uesugi, T. Genotoxicity and subchronic toxicity of a kaempferol aglycone-rich product produced from horseradish leaves. *Fundam. Toxicol. Sci.* **2022**, *9*, 71–83, 2189-115X. [CrossRef]
- 358. Yangzom, P.; Amruthanand, S.; Sharma, M.; Mahajan, S.; Lingaraju, M.C.; Parida, S.; Sahoo, M.; Kumar, D.; Singh, T.U. Subacute 28 days oral toxicity study of kaempferol and biochanin-A in the mouse model. *J. Biochem. Mol. Toxicol.* 2022, 36, e23090. [CrossRef] [PubMed]
- Akiyama, M.; Mizokami, T.; Ito, H.; Ikeda, Y. A randomized, placebo-controlled trial evaluating the safety of excessive administration of kaempferol aglycone. *Food Sci. Nutr.* 2023, 11, 5427–5437. [CrossRef] [PubMed]
- Hart, J.J.; Tako, E.; Wiesinger, J.; Glahn, R.P. Polyphenolic Profiles of Yellow Bean Seed Coats and Their Relationship with Iron Bioavailability. J. Agric. Food Chem. 2020, 68, 769–778. [CrossRef]
- Vaez, S.; Parivr, K.; Amidi, F.; Rudbari, N.H.; Moini, A.; Amini, N. Quercetin and polycystic ovary syndrome; inflammation, hormonal parameters and pregnancy outcome: A randomized clinical trial. Am. J. Reprod. Immunol. 2023, 89, e13644. [CrossRef]
- 362. Thakur, K.; Zhu, Y.Y.; Feng, J.Y.; Zhang, J.G.; Hu, F.; Prasad, C.; Wei, Z.J. Morin as an imminent functional food ingredient: An update on its enhanced efficacy in the treatment and prevention of metabolic syndromes. *Food Funct.* 2020, 11, 8424–8443. [CrossRef]
- Carmona, V.; Martín-Aragón, S.; Goldberg, J.; Schubert, D.; Bermejo-Bescós, P. Several targets involved in Alzheimer's disease amyloidogenesis are affected by morin and isoquercitrin. *Nutr. Neurosci.* 2020, 23, 575–590. [CrossRef]
- 364. Gur, C.; Kandemir, F.M.; Darendelioglu, E.; Caglayan, C.; Kucukler, S.; Kandemir, O.; Ileriturk, M. Morin protects against acrylamide-induced neurotoxicity in rats: An investigation into different signal pathways. *Environ. Sci. Pollut. Res.* 2021, 28, 49808–49819. [CrossRef]
- 365. Mohammadi, N.; Asle-Rousta, M.; Rahnema, M.; Amini, R. Morin attenuates memory deficits in a rat model of Alzheimer's disease by ameliorating oxidative stress and neuroinflammation. *Eur. J. Pharmacol.* 2021, 910, 174506. [CrossRef]
- 366. Soubh, A.A.; El-Gazar, A.A.; Mohamed, E.A.; Awad, A.S.; El-Abhar, H.S. Further insights for the role of Morin in mRTBI: Implication of non-canonical Wnt/PKC-α and JAK-2/STAT-3 signaling pathways. *Int. Immunopharmacol.* 2021, 100, 108123. [CrossRef]
- 367. Thabet, N.M.; Moustafa, E.M. Protective effect of rutin against brain injury induced by acrylamide or gamma radiation: Role of PI3K/AKT/GSK-3β/NRF-2 signalling pathway. Arch. Physiol. Biochem. 2018, 124, 185–193. [CrossRef]
- 368. Çelik, H.; Kandemir, F.M.; Caglayan, C.; Özdemir, S.; Çomaklı, S.; Kucukler, S.; Yardım, A. Neuroprotective effect of rutin against colistin-induced oxidative stress, inflammation and apoptosis in rat brain associated with the CREB/BDNF expressions. *Mol. Biol. Rep.* 2020, 47, 2023–2034. [CrossRef]
- 369. Ahmad, S.; Jo, M.H.; Ikram, M.; Khan, A.; Kim, M.O. Deciphering the potential neuroprotective effects of luteolin against Aβ1–42-Induced alzheimer's disease. *Int. J. Mol. Sci.* 2021, 22, 9583. [CrossRef] [PubMed]
- He, H.; Chen, X. Luteolin Attenuates Cognitive Dysfunction Induced By Chronic Cerebral Hypoperfusion Through the Modulation of The PI3K/Akt Pathway in Rats. J. Vet. Res. 2021, 65, 341–349. [CrossRef] [PubMed]

- 371. Sawmiller, D.; Li, S.; Shahaduzzaman; Smith, A.J.; Obregon, D.; Giunta, B.; Borlongan, C.V.; Sanberg, P.R.; Tan, J. Luteolin reduces Alzheimer's disease pathologies induced by traumatic brain injury. *Int. J. Mol. Sci.* **2014**, *15*, 895–904. [CrossRef]
- 372. Zeng, P.; Su, H.F.; Ye, C.Y.; Qiu, S.W.; Shi, A.; Wang, J.Z.; Zhou, X.W.; Tian, Q. A Tau Pathogenesis-Based Network Pharmacology Approach for Exploring the Protections of Chuanxiong Rhizoma in Alzheimer's Disease. *Front. Pharmacol.* 2022, 13, 877806. [CrossRef]
- 373. Yi, H.; Peng, H.; Wu, X.; Xu, X.; Kuang, T.; Zhang, J.; Du, L.; Fan, G. The Therapeutic Effects and Mechanisms of Quercetin on Metabolic Diseases: Pharmacological Data and Clinical Evidence. Oxidative Med. Cell. Longev. 2021, 2021, 6678662. [CrossRef] [PubMed]
- 374. Michala, A.S.; Pritsa, A. Quercetin: A Molecule of Great Biochemical and Clinical Value and Its Beneficial Effect on Diabetes and Cancer. *Diseases* **2022**, *10*, 37. [CrossRef]
- 375. Cheng, X.; Yang, Y.L.; Yang, H.; Wang, Y.H.; Du, G.H. Kaempferol alleviates LPS-induced neuroinflammation and BBB dysfunction in mice via inhibiting HMGB1 release and down-regulating TLR4/MyD88 pathway. *Int. Immunopharmacol.* 2018, 56, 29–35. [CrossRef]
- 376. Yang, Y.L.; Cheng, X.; Li, W.H.; Liu, M.; Wang, Y.H.; Du, G.H. Kaempferol Attenuates LPS-Induced Striatum Injury in Mice Involving Anti-Neuroinflammation, Maintaining BBB Integrity, and Down-Regulating the HMGB1/TLR4 Pathway. Int. J. Mol. Sci. 2019, 20, 491. [CrossRef]
- 377. Silva Dos Santos, J.; Gonçalves Cirino, J.P.; de Oliveira Carvalho, P.; Ortega, M.M. The Pharmacological Action of Kaempferol in Central Nervous System Diseases: A Review. *Front. Pharmacol.* 2021, 11, 565700. [CrossRef] [PubMed]
- 378. Rifaai, R.A.; Mokhemer, S.A.; Saber, E.A.; El-Aleem, S.A.; El-Tahawy, N.F. Neuroprotective effect of quercetin nanoparticles: A possible prophylactic and therapeutic role in Alzheimer's disease. J. Chem. Neuroanat. 2020, 107, 101795. [CrossRef]
- Xu, D.; Hu, M.J.; Wang, Y.Q.; Cui, Y.L. Antioxidant Activities of Quercetin and Its Complexes for Medicinal Application. *Molecules* 2019, 24, 1123. [CrossRef] [PubMed]
- 380. Pantsar, T.; Poso, A. Binding Affinity via Docking: Fact and Fiction. Molecules 2018, 23, 1899. [CrossRef]

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