

**Table S1.** Plasmids used in this study.

Mutants	Original sequences	Mutant sequences
pR1-T <sub>m1</sub>	CTAGC <sup>3294</sup> <u>A</u> CAACGTGCTAG	CTAGC <sup>3294</sup> <u>A</u> <b>GGATCC</b> ACTAGTCCCGGGTGCTAG
pR1-T <sub>m2</sub>	CTAGC <sup>3294</sup> <u>A</u> CAACGTGCTAG	CTAGC <sup>3294</sup> <u>A</u> <b>GGATCC</b> ACTAGTCCCGGGCAACGTGCTAG
pR2-T <sub>m1</sub>	CTAGC <sup>2988</sup> <u>A</u> CAACGTGCTAG	CTAGC <sup>2988</sup> <u>A</u> <b>GGATCC</b> ACTAGTCCCGGGTGCTAG
pR2-T <sub>m2</sub>	CTAGC <sup>2988</sup> <u>A</u> CAACGTGCTAG	CTAGC <sup>2988</sup> <u>A</u> <b>GGATCC</b> ACTAGTCCCGGGCAACGTGCTAG
pR3-T <sub>m1</sub>	CTAGC <sup>2152</sup> <u>A</u> CAACGTGCTAG	CTAGC <sup>2152</sup> <u>A</u> <b>GGATCC</b> ACTAGTCCCGGGTGCTAG
pR3-T <sub>m2</sub>	CTAGC <sup>2152</sup> <u>A</u> CAACGTGCTAG	CTAGC <sup>2152</sup> <u>A</u> <b>GGATCC</b> ACTAGTCCCGGGCAACGTGCTAG
pR2-2bPT	TCTGA <sup>2661</sup> <u>G</u> GCCTCT	TCTGA <sup>2661</sup> <u>G</u> <b>TAATAGGGATCC</b> ACTAGTCCCGGGGCCTCT
pR3-M5	CCGAC <sup>548</sup> <u>G</u> TATGATTGTCCTAT	CCGAC <sup>548</sup> <u>G</u> <b>gcTGcaTGc</b> CCTAT
pR2-2bPT-1	TCTGA <sup>2661</sup> <u>G</u> GCCTCT	TCTGA <sup>2661</sup> <u>G</u> <b>TAATAGGGATCC-100 bp fragment from</b> <b>TVBMV-CCCGGGGCCTCT</b>
pR2-2bPT-2	TCTGA <sup>2661</sup> <u>G</u> GCCTCT	TCTGA <sup>2661</sup> <u>G</u> <b>TAATAGGGATCC-200 bp fragment from</b> <b>TVBMV-CCCGGGGCCTCT</b>

Remarks: The inserted bases were bold, and mutated sites were indicated by low case.

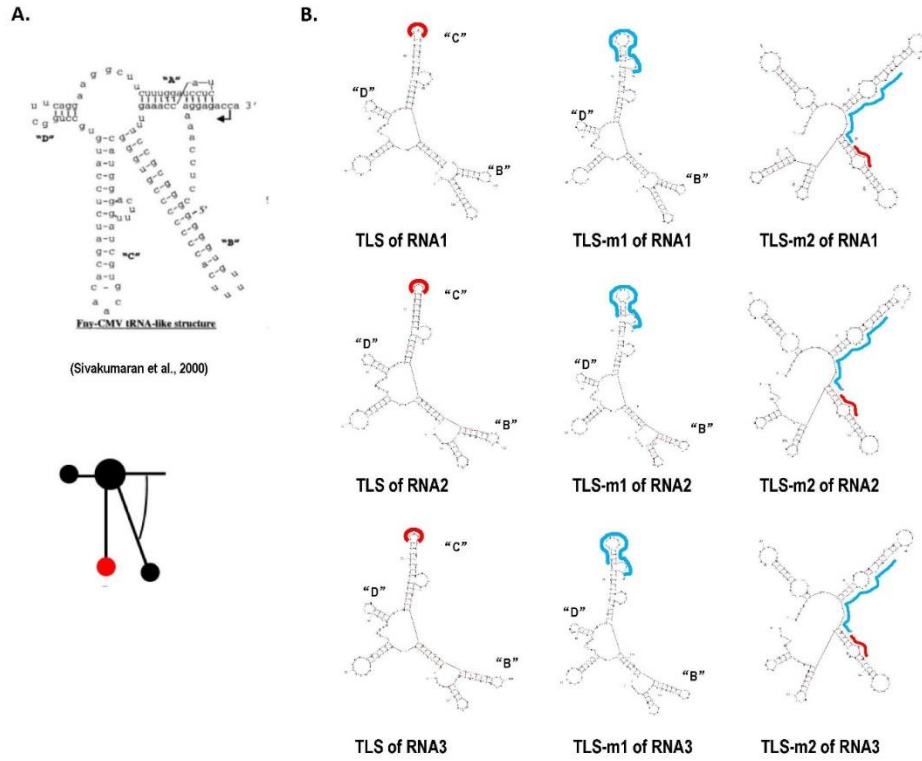
**Table S2.** Primers used in this study.

DNA Oligos	Sequence (5'-3')	Purpose
R1-T <sub>m1</sub> -F	<u>TCGGATCC</u> ACTAGTCCCGGTGCTAGTTTCAGGGTACGGGTGC	Construction of R1-T <sub>m1</sub>
R1-T <sub>m1</sub> -R	ATGGATCCTGCTAGAAGTACACGGACCGAAG	
R1-T <sub>m2</sub> -F	<u>TCGGATCC</u> ACTAGTCCCGGGCAACGTGCTAGTTTCAGGGTACGGGTG	Construction of R1-T <sub>m2</sub>
R1-T <sub>m2</sub> -R	ATGGATCCTGCTAGAAGTACACGGACCGAAG	
R2-T <sub>m1</sub> -F	<u>TCGGATCC</u> ACTAGTCCCGGTGCTAGTTTCAGGGTACGGGTGC	Construction of R2-T <sub>m1</sub>
R2-T <sub>m1</sub> -R	ATGGATCCTGCTAGAAGTACACGGACCGAAG	
R2-T <sub>m2</sub> -F	<u>TCGGATCC</u> ACTAGTCCCGGGCAACGTGCTAGTTTCAGGGTACGGGTG	Construction of R2-T <sub>m2</sub>
R2-T <sub>m2</sub> -R	ATGGATCCTGCTAGAAGTACACGGACCGAAG	
R3-T <sub>m1</sub> -F	<u>TCGGATCC</u> ACTAGTCCCGGTGCTAGTTTCAGGGTACGGGTGC	Construction of R3-T <sub>m1</sub>
R3-T <sub>m1</sub> -R	ATGGATCCTGCTAGAGGTACACGGACCGAAG	
R3-T <sub>m2</sub> -F	<u>TCGGATCC</u> ACTAGTCCCGGGCAACGTGCTAGTTTCAGGGTACGGGTG	Construction of R3-T <sub>m2</sub>
R3-T <sub>m2</sub> -R	ATGGATCCTGCTAGAGGTACACGGACCGAAG	
R2-2bPT-F	<u>TCGGATCC</u> ACTAGTCCCGGGCCTCTCGTTAGAGTTATCGGCG	Construction of R2-2bPT
R2-2bPT-R	ATGGATCCCATTACTCAGACTCGGGTAACTCCGCC	
TVBMV-7600-F	ATGGATCCCAGCACCTCTGGATACTCTACTAG	Construction of R2-2bPT-1 and R2-2bPT-2
TVBMV-7699-R	ATCCCGGGATCCCGACACTCCATGGTC	
TVBMV-7799-R	ATCCCGGGTGTC AATGAGCTGTCAAAACGACT	Construction of R3-M5
R3-M5-F	CAACCGACGGCTGCATGCCCTATGGAA	
R3-M5-R	TTCCATAGGGCATGCAGCCGTCGGTTG	
Fny3-pCB301-F	AGTTCATTTCAATTGGAGAGGGTAATCTTACCACTGTGTGTGTGC	
Fny3-pCB301-R	GTGGAGATGCCATGCCGACCCTGGTCTCCTTTTGGAGGCC	
pCB301-800-F	GGGTCGGCATGGCATCTCCA	
pCB301-767-R	CCTCTCCAAATGAAATGAACTTCCTTATATAGAGGA	
R1-2658-F	ATGGCTACGAAGGCCCTTC	Detection of TLS mutants in RNA1
R2-2510-F	AGAATCGACGGAACGAGGT	Detection of TLS and 2b mutants in RNA2
R2-2760-R	GGGGAGGTTTCAGAAAGCACC	Detection of 2b mutants in RNA2
R3-1385-F	CCAACTATTAACCAACCAACCTTTGT	Detection of TLS mutants in RNA3
R1-R	TGGTCTCCTTTTAGAGACCCACG	Detection of TLS mutants in RNA1
R2/3-R	TGGTCTCCTTTTGGAGGCCCC	Detection of TLS mutants in RNA2/3
R3-1883-F	GTTATTGTCTACTGACTATATAGAGAGTGTT	Clone of the 3' half of larger RNA3
R3-198-F	AAATGGCTACTGAGTGTGACC	Detection of 3a mutation in RNA3
R3-1040-R	GCATCGCGTCACAGATGTCTAC	
R1-NT-F	TGTGACCTGTTAAACAGTTTCTTATTG	RNA1 probe for northern blot
R1-NT-R	ACACAATGTGTTTAGTGACTTCAGAC	

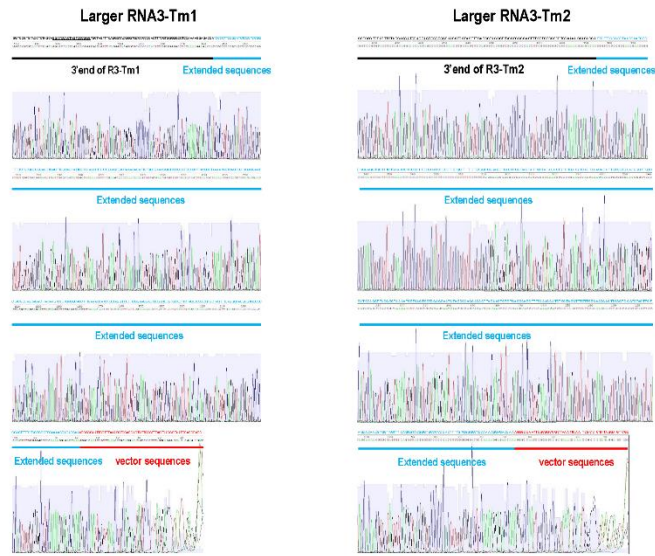
R2-NT-F	AGAGCGCGTTCAAATCTCAGA	RNA2 probe for northern blot
R2-NT-R	TTCGTTACCGGCGAACCAAT	
R3-NT-F	GTACTGGTTTATCAGTATGCCGC	RNA3 and RNA4 probe for northern blot
R3-NT-R	GACTGGGAGCACTCCAGAT	
CM-NT-F	AAACTGTCTGAAGTCACTAAAC	CMV genome and subgenome probe for northern blot
CM-NT-R	TGGTCTCCTTTTGAGGC	

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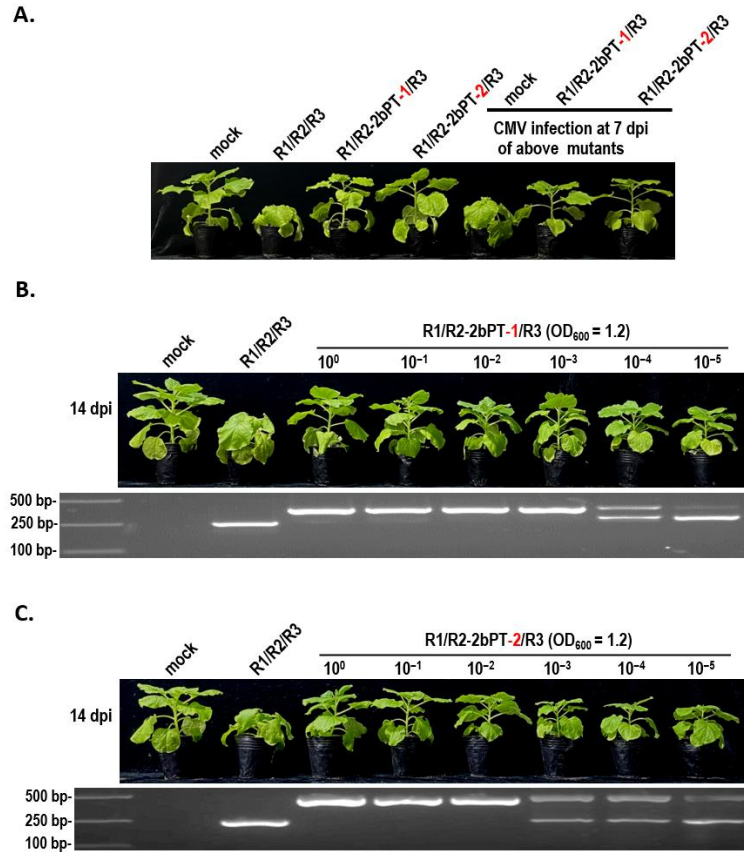
Remarks: Protective bases were displayed in a bold font. Endonuclease sites was displayed by underline. Stop codons was displayed in italics.



**Figure S1.** Alignment of tRNA-like structure (TLS) region among wt RNAs,  $T_{m1}$  and  $T_{m2}$  mutants of cucumber mosaic virus. Secondary structure of TLS in CMV and corresponding schematic representations. caacg loop was indicated by red color. B. Alignment of TLS among wt RNAs,  $T_{m1}$  and  $T_{m2}$  mutants of CMV. caacg loop sequences were indicated by opened red circles in wt RNAs and red lines in  $T_{m2}$  mutants. 18 nt new sequences were indicated by blue lines.



**Figure S2.** Sequencing results of RT-PCR clones: Red nucleotides indicate 18nt insertion sequences in R3-T<sub>m1</sub> and R3-T<sub>m2</sub>. Blue nucleotides indicate the extended sequences. Purple nucleotides indicate vector sequences.



**Figure S3.** Dilution inoculation of attenuated vaccines of CMV and mutant repair assay. (A) Mutants cross protection assay of CMV<sub>Fny</sub>. R1, RNA1 of CMV<sub>Fny</sub>; R2, RNA2 of CMV<sub>Fny</sub>; R3, RNA3 of CMV<sub>Fny</sub>. (B) Mutation repair assay under dilution inoculation of R1/R2-2bPT-1/R3. The PCR products showed a band of 350 bp indicating all mutants were existent; the PCR products showed two bands of 250 bp and 350 bp indicating part of the mutants were repaired to wild-type RNA2. (C) Mutation repair assay under dilution inoculation of R1/R2-2bPT-2/R3. The PCR products showed a band of 450 bp indicating all mutants were existent; the PCR products showed two bands of 250 bp and 450 bp indicating part of the mutants were repaired to wild-type RNA2.