



Supplementary Materials: Figures S1–S6

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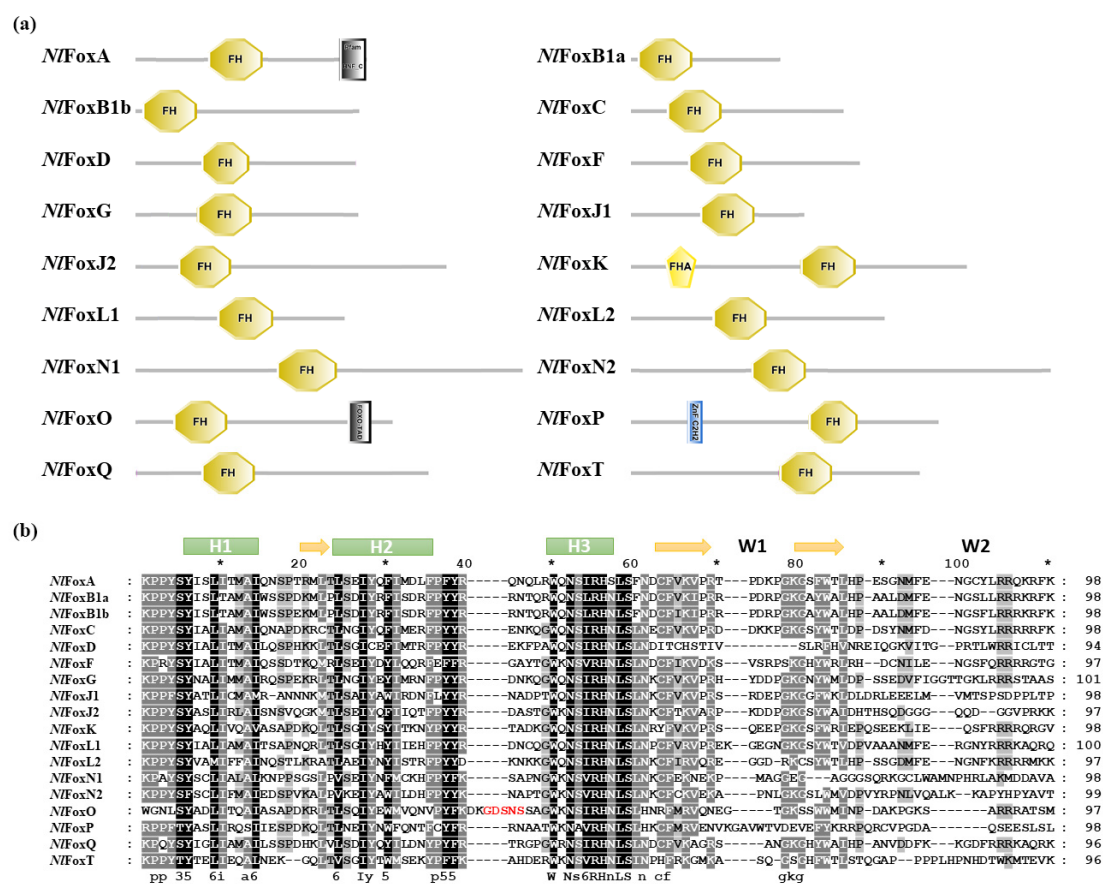
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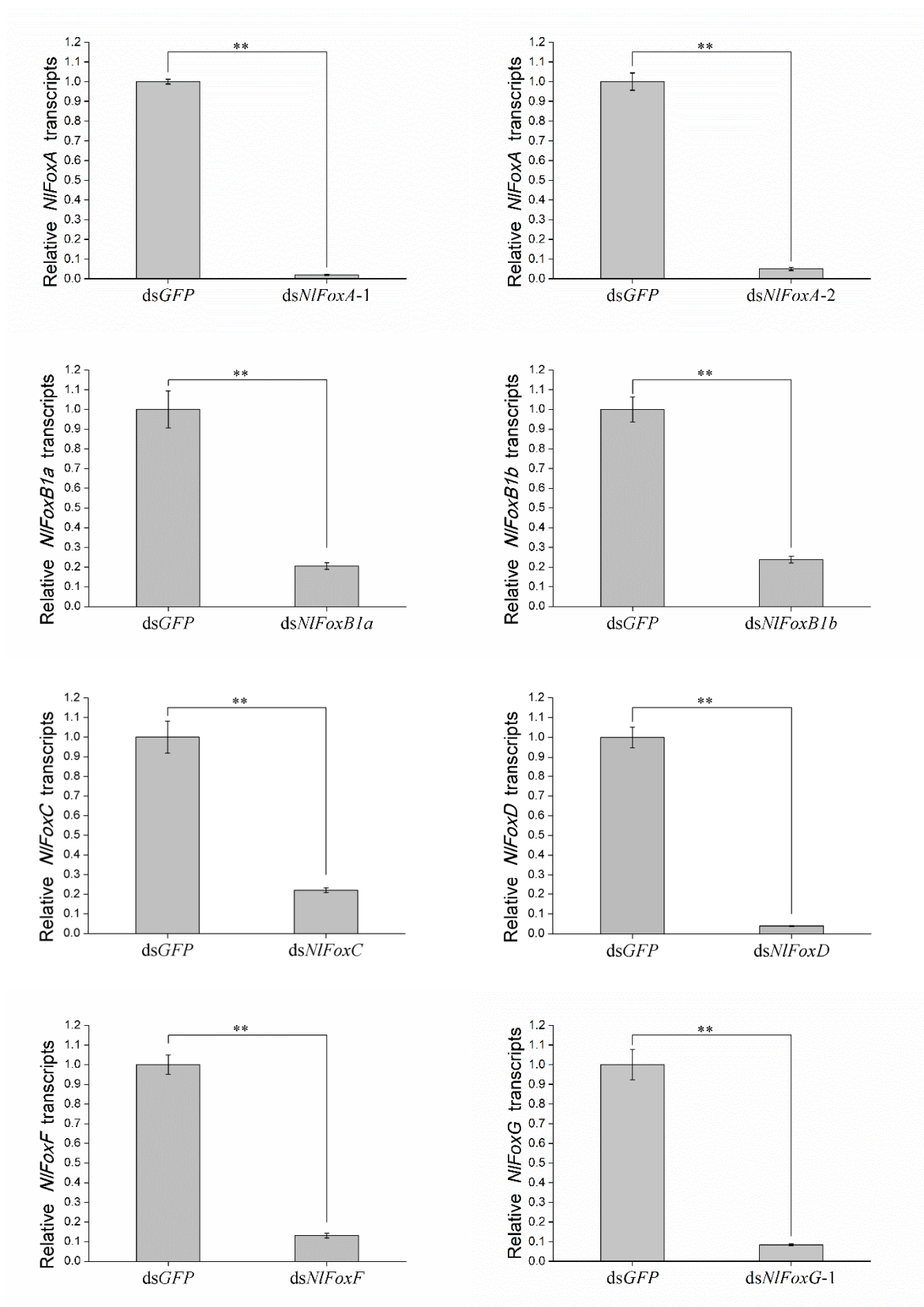
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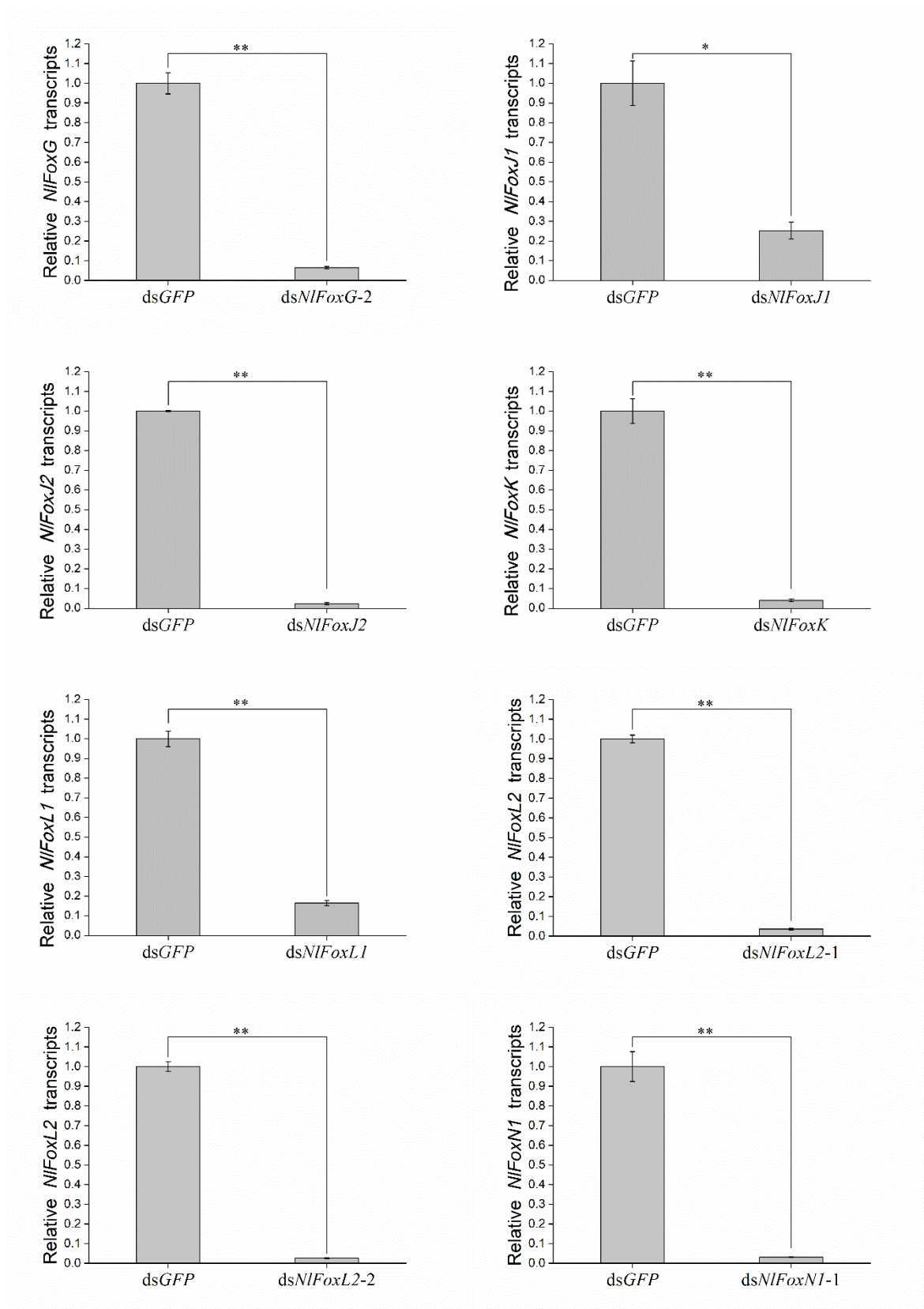
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Figure S1. The cDNA sequences of 18 BPH Fox genes were listed below. The initiators and terminators were boxed. The primers used for cloning, qRT-PCR, RNAi were highlighted in blue, red and yellow, respectively. The primers used for RNAi targeting the second non-overlapping region were highlighted in green. The sequences of FHD were underlined.







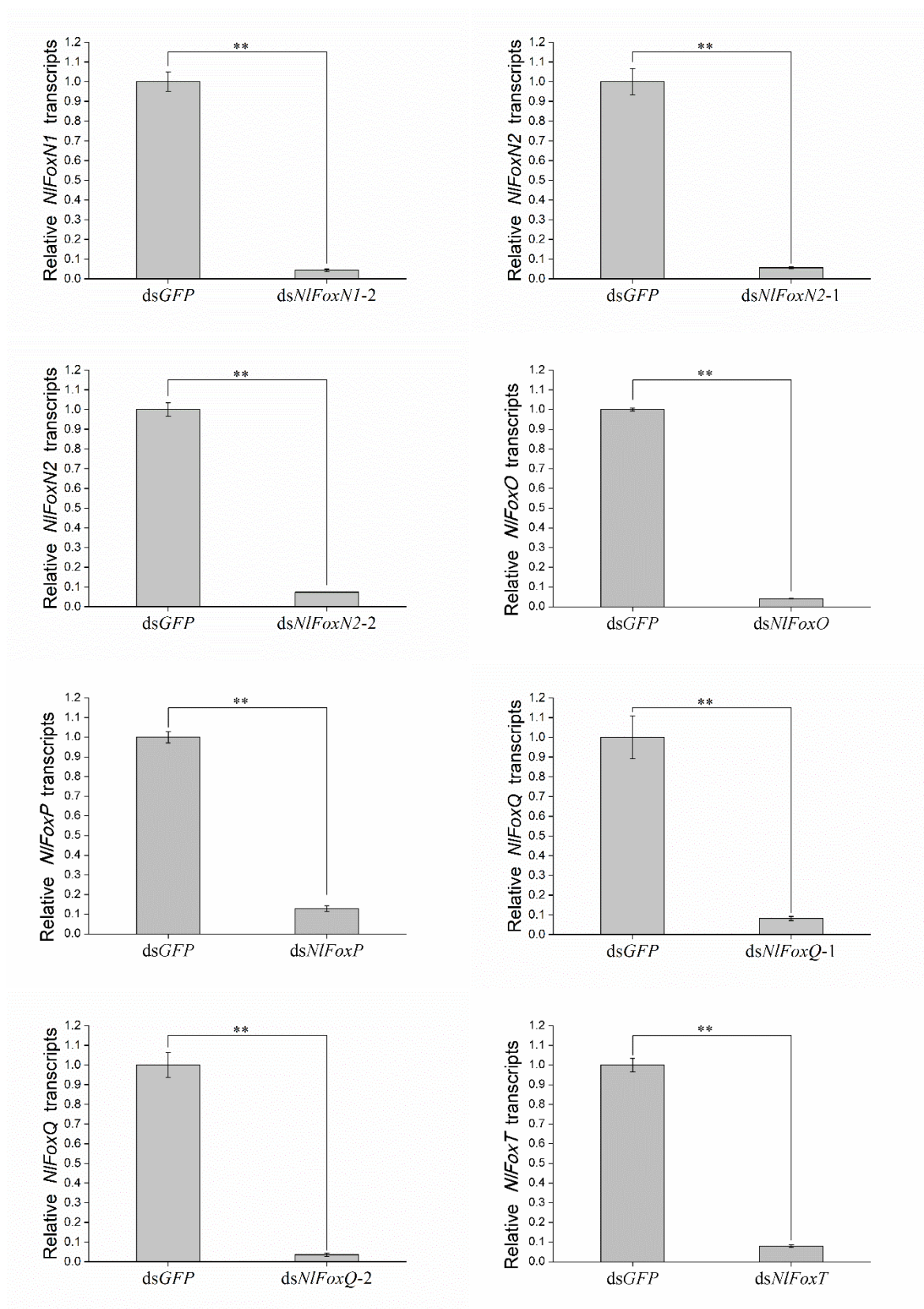


Figure S3. RNAi efficiency by was determined by qRT-PCR. dsRNA (50 ng per insect; $n = 100$) was injected into BPHs at the very beginning of newly emerged adults. Total RNA samples for *NIfoxA*, *NIfoxJ2*, *NIfoxK*, *NIfoxN1*, *NIfoxN2* and *NIfoxO* were extracted from eggs within 2h after laid. Total RNA samples for *NIfoxD*, *NIfoxG* and *NIfoxQ* were extracted from eggs 48h after laid. Total RNA samples for *NIfoxB1a*, *NIfoxC*, *NIfoxF*, *NIfoxJ1* and *NIfoxL1* were extracted from eggs 72h after laid. Total RNA samples for *NIfoxB1b* and *NIfoxP* were extracted from eggs 96h after laid. Total RNA sample for *NIfoxL2* and *NIfoxT* were extracted from BPHs 3 days after injection. dsGFP was injected as negative control for the nonspecific effects of dsRNA. *NI18S* was used as an internal control gene. Mean \pm s.e.m. from three experiments. * $P < 0.05$, ** $P < 0.01$ (Student's *t*-test), difference from dsGFP.

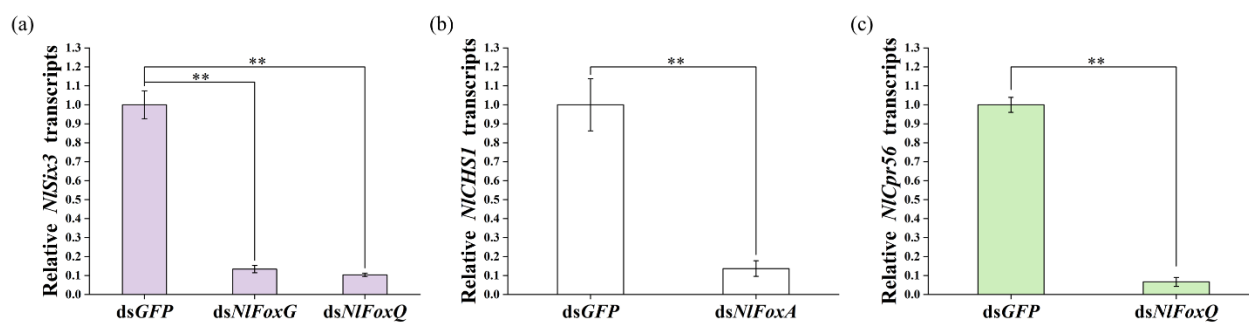


Figure S4. (a) Knockdown of *NIFoxG* or *NIFoxQ* both decreased expression levels of *NISix3* in the eggs; (b) Knockdown of *NIFoxA* decreased expression levels of *NICH51* in the nymphs; (c) Knockdown of *NIFoxQ* decreased expression levels of cuticular protein genes in the nymphs. dsRNA for a (50 ng per insect; $n = 100$) was injected into newly emerged female adults (within two hours). Total RNA samples were extracted from eggs 48 h after laid. dsRNA for b-c (50 ng per insect; $n = 100$) was injected into BPHs at the very beginning of the 4th instar. Total RNA samples were extracted from BPHs 3 days after injection. dsGFP was injected as negative control for the nonspecific effects of dsRNA. Mean \pm s.e.m. from three experiments. $^{***}P < 0.01$ (Student's *t*-test), difference from dsGFP.

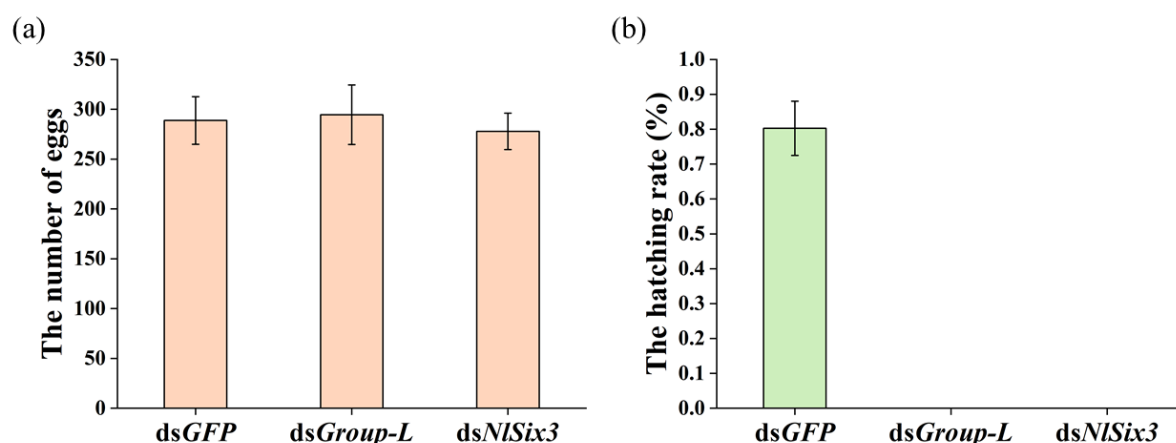


Figure S5. Knockdown of *Group-Late* or *NISix3* prevented the hatchability. (a) The number of eggs; (b) The hatching rate. dsRNA (50 ng per insect; $n = 100$) was injected into newly emerged female adults (within two hours). dsGFP was injected as negative control for the nonspecific effects of dsRNA. Mean \pm standard error of the mean (s.e.m.) from three experiments.

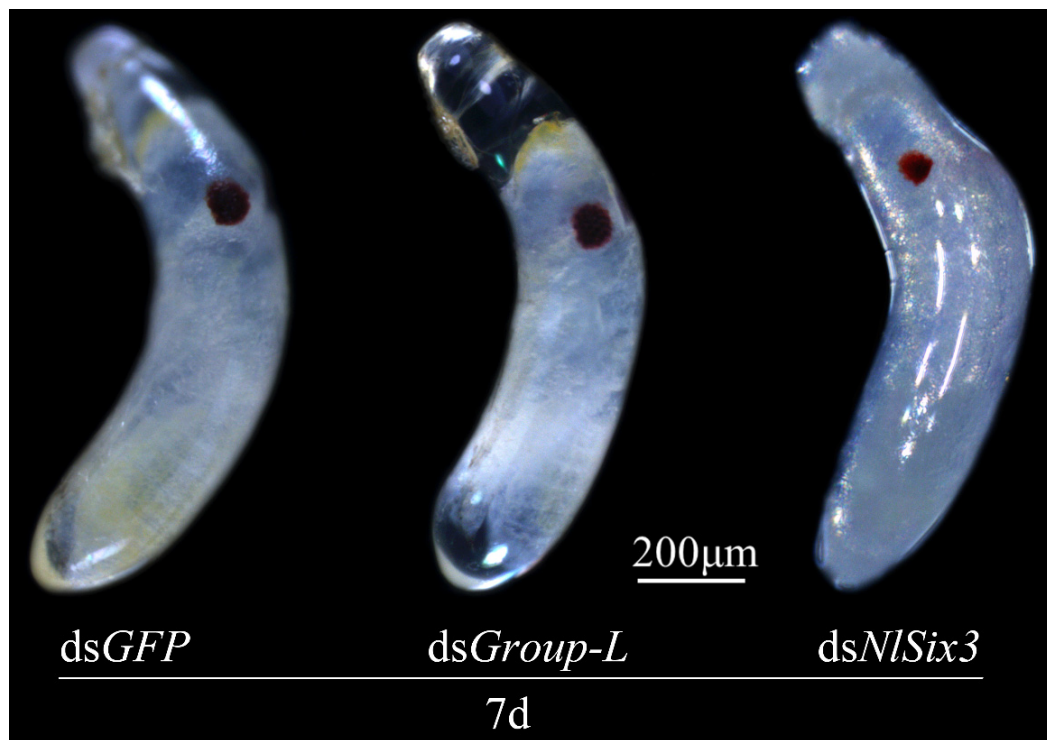


Figure S6. Lethal phenotypes of eggs injected with dsRNA for *Group-Late* or *NISix3*. dsRNA (50 ng per insect; $n = 100$) was injected into newly emerged female adults (within two hours). *dsGFP* was injected as negative control for the non-specific effects of dsRNA.