



# Surface Characterization and Copper Release of a-C:H:Cu Coatings for Medical Applications



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**Abstract:** This paper focuses on the surface properties of a-C:H:Cu composite coatings for medical devices and how the release of Cu<sup>2+</sup> ions from such coatings can be controlled. The released Cu ions have the potential to act as a bactericidal agent and inhibit bacterial colonization. A PVD–PECVD hybrid process was used to deposit a-C:H:Cu composite coatings onto Ti6Al4V substrates. We examine the layer surface properties using atomic force microscopy and static contact angle measurements. An increasing surface roughness and increasing contact angle of Ringer's solution was measured with increasing copper mole fraction ( $X_{Cu}$ ) in the coatings. The contact angle decreased when a supplementary bias voltage of -50 V was used during the a-C:H:Cu deposition. These findings are in line with earlier published results regarding these types of coatings. The release of Cu<sup>2+</sup> ions from a-C:H:Cu coatings in Ringer's solution was measured by anodic stripping voltammetry. Different layer structures were examined to control the time-resolved Cu release. It was found that the Cu release depends on the overall  $X_{Cu}$  in the a-C:H:Cu coatings and that an additional a-C:H barrier layer on top of the a-C:H:Cu layer effectively delays the release of Cu ions.

Keywords: DLC; copper filled; ion release; medical; coating; amorphous carbon

# 1. Introduction

Regardless of aseptic surgical conditions and perioperative antibiotic prophylaxis, aseptic infection and loosening of implants are still serious potential complications in orthopedic surgery. Early infections can occur up to 90 days after operation and are usually acquired during the surgical procedure or as a result of implant-associated infections (peri-implantitis) [1]. Infection rates range between 0.3% and 1.7% after primary total hip replacement [2–4] and approximately 0.8% to 1.9% after primary total knee replacement [2,5,6]. When focusing on screw implants in orthodontics, the average success rate of dental mini- and microimplants is 84–88% according to various meta-analyses of clinical trials [7–9]. To a significant extent (7% [10]) there is a loss of primary stability from peri-implant infections due to an exposed germination site at the entry to the gingiva. This leads to a reduction of the success rate by 10.9 [11] and 32.2 [12] percentage points. According to a meta-analysis of 15 studies,

the mean prevalence of peri-implantitis is 22% (CI: 14–30%) [13]. Such implants are often made out of titanium alloys like Ti6Al4V.

The treatment of implant-related infections has become more and more difficult, since the excessive treatment with antibiotics has led to the formation of multi-resistant germs through spontaneous mutation or by DNA transfer [13,14]. This is one of the greatest current threats in the clinical practice.

To prevent these implant-related infections, a number of implant surface modifications that inhibit bacterial colonization through the release of antibacterial substances, like coupling of antibiotics [15–20], have been developed. Since ancient times, silver and copper have been known as an effective antimicrobial agent, with a large spectrum against multiple drug-resistant bacteria combined with a reduced cytotoxicity [21–24]. Therefore, titanium surfaces were already modified by adding metal ions of Zn, Ag and Cu [20,23,25,26].

When using such surface modification for implant devices, the biocompatibility and, particularly, the cytotoxicity of the surface must be considered. The release of Ag or Cu ions creates an antibacterial effect of the coating; however, silver ions show higher dose-related cytotoxicity when compared to copper ions [27], which can have several detrimental effects [28]. Moreover, while silver tends to accumulate in the human body and increases the silver serum levels [29] copper is a metabolizable element [30]. Therefore, the two opposing objectives of antibacterial activity and biocompatibility have to be well-balanced when developing coatings for implant devices. To do so, antibacterial effects have to be based on a time-controlled release of the specific antibacterial substances.

Furthermore, the success of an antibacterial coating, like other release-based systems, depends on the ability to control and tune the time frame and kinetics of the release of metal ions from the coating. A critical step for controlling ion release from metal filled coatings depends on the oxidization mechanism in which the metal is transformed into reactive and soluble metal ions. To this effect, part of the research has been focused on various approaches to control and enhance the oxidative dissolution [31]. The surface-dependent factors like distribution, size, concentration, shape or charge of the antibacterial substance, the porosity and roughness of the coating's matrix as well as the overall micro- and nanostructure of the coating, are known to influence the ion release kinetics [32]. In this regard, a fine particle size and the incorporation of oxygen species can improve the Ag ions dissolution rate [33]. Additionally, nano-structuring the surface is also a valuable method to control antibacterial activity due to the high surface to volume ratio, which increases the surface reactivity [34].

In order to better control the ion release, the embedding of metal ions or nanoparticles in different organic or inorganic matrixes like PMMA [35,36], PAA/PAH [28], PVA [37], Si-based sol-gel [38] and gelatin fibers [39] has been investigated. Furthermore, silver was embedded in plasma-based coatings like oxygen-containing [40] or nitrogen-containing [41] plasma polymer films. The use of metal containing amorphous carbon coatings like Cu/DLC or Ag/DLC is also under investigation. Therefore, metal nanoparticles are embedded into the a-C:H matrix. Modifying DLC with different metallic elements such as Cr [42,43], Ti [42,44], W [45,46] and Mo [46] can lead to new functions of the DLC thin films. Adding non-carbide-forming elements like Ag [47,48], Cu [49], Al [50] and Ni [50] into the DLC matrix can reduce residual stress [43,45,47–50] and improve electric conductivity [44,51]. Furthermore, copper- or silver-containing DLC thin films can show a significant antibacterial effect [52–56]. These composite coatings have been deposited by Arc evaporation with metal cathodes [57], magnetron sputtering [51], ion plating/sputtering [58], pulsed laser deposition [21,59–61], laser ablation [62, 63], plasma-immersion ion implantation [64], ion deposition [54], plasma-enhanced chemical vapor deposition (PECVD) [47] and variations thereof. Their use for antibacterial surface modification of biomedical implants and environmental surfaces by the release of antibacterial Ag and Cu ions has been widely studied [20,54,56,65-67].

The release of copper ions from these thin composite films is influenced by the film thickness, substrate roughness and crystallographic structure of the deposited film [68]. Furthermore, the copper release from a surface depends on the exposed area used in the application and on the availability of

proposed to use additional top layers that act as a diffusion barrier [41,72]. This paper reports on the surface properties like roughness and wettability and the time-resolved release kinetics of copper ions (Cu<sup>2+</sup>) from a-C:H:Cu coatings, which were deposited onto Ti6Al4V substrates. A hydrophobic a-C:H barrier layer should be able to reduce the initial burst release and sustain a sufficient release over several days.

# 2. Materials and Methods

# 2.1. Deposition Methods

Coated cylindrical titanium alloy Ti6Al4V specimens (Ti grade 23 ELI,  $\emptyset = 11$  mm) with a thickness of 2 mm were used for the release measurements (Figure 1a). Here only a short summary of the deposition steps is given, as the detailed deposition processes are already described in another paper [73]. The whole process consisted of three deposition steps (Figure 1b). At first, a chemical gradient Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti bonding layer system was deposited by DC magnetron sputtering of a Ti target (99.99% purity;  $\emptyset = 50$  mm) in Ar and Ar/C<sub>2</sub>H<sub>2</sub> atmosphere (99.999%/99.5% purity). The deposition time of 15 min with a deposition rate of 1.2 nm/s led to a Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti layer system thickness of 1 µm. Afterwards, a 400 nm thick a-C:H diffusion barrier layer was deposited in a radio frequency magnetron plasma-enhanced chemical vapor deposition (RF-magPECVD) process at a pressure of 0.7 Pa for a duration of 8 min (0.8 nm/s). The structure of the chemical gradient Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti/a-C:H layer system is shown in Figure 3. The Ti<sub>x</sub>C<sub>y</sub> has a columnar structure which changes to amorphous carbon at the surface.

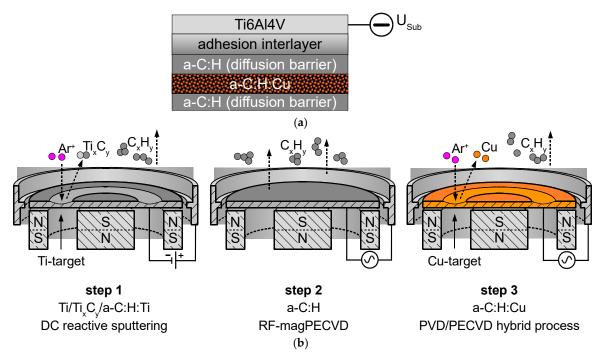


Figure 1. Schematic layer system (a) and illustration of utilized process steps (b).

Next, the top a-C:H:Cu layer was deposited in an  $Ar/C_2H_2$  atmosphere with a Cu target (99.98% purity;  $\emptyset = 50$  mm) by using a RF-PVD/PECVD hybrid process. Additionally, a supplementary bias voltage of -50 V was applied at the substrate holder by a DC power supply, which was deliberately kept low (<-100 V) because a higher electric potential leads to significant resputtering of the coatings

by Ar ions. The process pressure during a-C:H:Cu deposition was kept constant at 1 Pa for all coating experiments.  $X_{Cu}$  was varied by using different  $C_2H_2$  partial pressures (8.3–40 MPa) during deposition. The deposition time was 30 min leading to an a-C:H:Cu layer thickness in the range of 710–1750 nm. It is worth noting that, despite similar deposition times, different coating thicknesses were obtained, mainly as a result of differences in the sputtering rate of the Cu target with the variation of the  $C_2H_2$  partial pressure, in order to influence  $X_{Cu}$ .

#### 2.2. Chemical and Structural Characterisation

The layer's structural properties were examined using scanning electron microscope SEM (Zeiss Auriga Compact, Oberkochen, Germany) with an accelerating voltage of 20 kV. Layer cross sections were examined to determine the layer thickness by SEM.

The overall elemental composition of the deposited a-C:H:Cu layers was evaluated by the energy dispersive X-ray spectroscopy (EDX) method. Since parts of the (titanium carbide) TiC-based interlayer were also probed by EDX, there was a risk that the C content was slightly overestimated for the examined a-C:H:Cu layers. The accelerating voltage was 20 kV for all EDX measurements. The obtained spectra were analyzed using the eZAF routine, provided within the EDAX TEAM software (V 4.2), and results were calculated as molar fraction ( $X_i$ ) of species i (numerically identical with at %).

The elemental depth profiles of the coatings were obtained by glow discharge optical emission spectroscopy (GDOES) [74] using a Horiba Scientific GD Profiler 2 instrument (HORIBA Scientific, Palaiseau, France). A pulsed radio frequency (RF) source (700 Pa, 40 W, 3000 Hz), with anode diameter equal to 4 mm (analyzed surface area equal to 12.56 mm<sup>2</sup>), was used.

#### 2.3. Surface Characterization

Atomic force microscope (AFM) measurements (intermittent contact mode, Air) of surface roughness were conducted with a NanoWizard AFM (JPK instruments AG, 2004, Berlin, Germany). The measurement area was 10  $\mu$ m × 10  $\mu$ m and the tip velocity 17.96  $\mu$ m/s. Measurements were evaluated and visualized using JPK Data Processing software (V 4.3.55). The obtained mean roughness ( $R_a$ ) and mean square roughness ( $R_q$ ) were calculated from a histogram of pixel values.

To examine the surface's wetting behaviour, static contact angle measurements where taken using a DSA 100 (Krüss, Hamburg, Germany). The biomedical application required the use of an artificial serum, Ringer's solution, as the contact medium. Statistical significance between measured samples was calculated using two sample t-tests for unequal sample size and unequal variance ( $\alpha = 0.05$ ).

# 2.4. Release of $Cu^{2+}$ Ions

Quantitative measurements of dissolved copper ions ( $Cu^{2+}$ ) in Ringer's solution (B. Braun, Melsungen, Germany) were performed using an electrochemical detection method called square-wave anodic stripping voltammetry. The ion species was determined by the position of the peak potential in the I/V-curve (-0.3--0.2 V for Cu<sup>2+</sup>) [75]. The standard addition method using standard solutions with different Cu concentration (Ringer's solution with copper(II) sulfate) was used.

Measurements were conducted using screen-printed electrodes IS-HM1 (ItalSens) and EmStat potentiostat (PalmSens BV, Houten, The Netherlands). To measure the Cu<sup>2+</sup> release from the material surface, coated Ti6Al4V specimens were immersed in 10 mL of Ringer's solution with a constant temperature of 37 °C. For this purpose, a peristaltic pump was connected to the measuring cell (Figure 2), which kept the release medium in a constant flow. After various time points, the samples were analyzed using a standard protocol. The release medium was changed every hour or day, depending on measurement conditions.

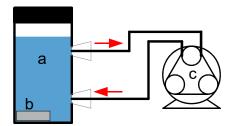
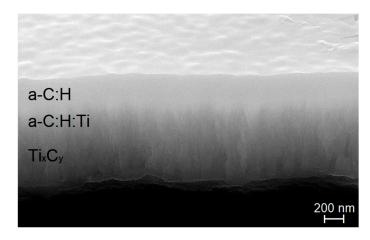


Figure 2. Measuring cell with Ringer's solution (a), a-C:H:Cu specimen (b) and peristaltic pump (c).

#### 3. Results and Discussion

### 3.1. Surface Topography

Besides the coated samples, an uncoated but polished Ti6Al4V sample with  $R_q = 1.94$  nm was examined for reference. When applying the first layer system with its columnar grown Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti bonding layer (see Figure 3) the surface roughness increased to  $R_q = 3.61$  nm.

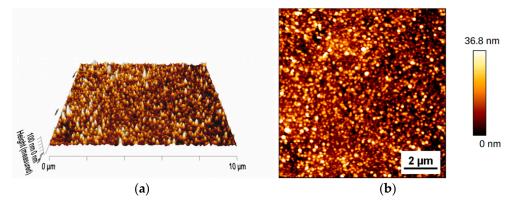


**Figure 3.** SEM image of cross section of the underlying  $\text{Ti}/\text{Ti}_xC_y/a$ -C:H:Ti/a-C:H layer system (perspective view).

The adding of Cu to the a-C:H matrix led to a further increase in roughness, depending on the overall Cu content of the a-C:H:Cu coating (see Table 1). This can been related to the increase in the size of Cu clusters in the a-C:H matrix. It can be noted that the particle diameter on the surface, measured with AFM, ranged between 100 and 400 nm (Figure 4), while scanning TEM (STEM) examinations in previous work [73] showed Cu particle size of 1.5–22 nm. The relative height values of the surface topography, ~30–40 nm, also indicated that the particles on the surface were not spherical. They were rather like flat elevations of the a-C:H matrix, whose surface morphology could have been influenced by small changes in the distribution and size of the Cu nanoparticles in the matrix.

**Table 1.** Roughness of the substrate and the different coating systems (C.I. = 3 for  $\sigma_R$ ).

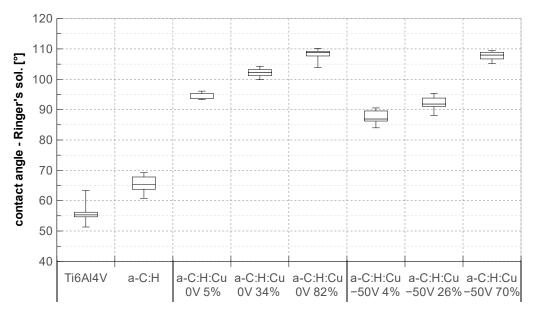
Sample	R <sub>a</sub> (nm)	$\sigma_{ m R}$ (nm)	R <sub>q</sub> (nm)
polished Ti6Al4V substrate	1.36	0.017	1.94
Ti/Ti <sub>x</sub> C <sub>y</sub> /a-C:H:Ti/a-C:H	2.23	0.012	3.61
a-C:H:Cu X <sub>Cu</sub> = 12%	3.01	0.013	5.35
a-C:H:Cu X <sub>Cu</sub> = 31%	3.11	0.018	6.23
a-C:H:Cu X <sub>Cu</sub> = 55%	4.04	0.014	6.79



**Figure 4.** Exemplary AFM image of a Ti/Ti<sub>*x*</sub> $C_y$ /a-C:H:Ti/a-C:H/a-C:H:Cu layer system with  $X_{Cu} = 31\%$  perspective view (**a**) and top view (**b**).

#### 3.2. Contact Angle

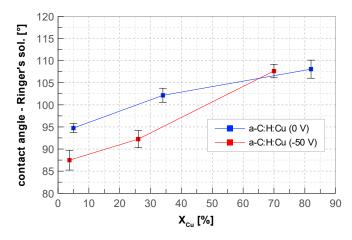
In this section, the static contact angle of Ringer's solution on the surfaces of coated and uncoated samples is evaluated. Figure 5 gives an overview of those measured contact angles. The uncoated Ti6Al4V reference sample showed a contact angle of  $56.2^{\circ} \pm 3.4^{\circ}$  and the Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti/a-C:H interlayer system showed a slightly increased contact angle of  $65.6^{\circ} \pm 2.6^{\circ}$ . Both samples could therefore be described as hydrophilic. When adding the a-C:H:Cu layer, even small, single-digit Cu contents increased the contact angle to over 90° and the surfaces showed increasingly hydrophobic character, which is consistent with the literature [63,76]. By adding more Cu to the a-C:H matrix, the surface became more and more hydrophobic, as shown by the higher contact angles of up to  $107.5^{\circ} \pm 1.5^{\circ}$ . This relationship has already been described in the literature for Ag- and Cu-doped DLC layers and was measured independently of an applied substrate bias voltage in this work. An increase in the amount of Cu mole fraction mainly resulted in a reduction in the polar component of the surface energy [76,77]. This is explained by the increase in the sp<sup>2</sup>-hybridization of the a-C:H matrix by the Cu doping and formation of Cu–O bonds. Furthermore, permanent dipoles that determine the polar part were missing because Cu and C have antibonding orbitals.



**Figure 5.** Contact angle of Ringer's solution on polished Ti6Al4V substrate and on the Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti/a-C:H interlayer and a-C:H:Cu layers with different  $X_{Cu}$  (%). a-C:H:Cu layers were deposited with (-50 V) and without (0 V) substrate bias voltage as indicated at the x-axis. (box plot).

In conjunction with the AFM results and the relationships described by Wenzel and Cassie for textured surfaces [53], it can be concluded that this trend was also based on the increasing surface roughness.

Upon closer examination of the coatings deposited with and without applying a substrate bias voltage in Figure 6, the coatings deposited with a substrate bias voltage had a slightly reduced contact angle. This can be attributed to an increase in the sp<sup>3</sup>-content in the a-C:H matrix. With high amounts of  $X_{Cu} > 70\%$ , no significant difference in contact angle with applied substrate bias was observed [(p = 0.204) > ( $\alpha = 0.05$ )]. It is possible that the influence of the Cu was predominant and thus the different contents of the carbon hybridization no longer had any significant influence on the surface energy.



**Figure 6.** Comparison of contact angles of Ringer's solution on a-C:H:Cu layers deposited with (-50 V) and without (0 V) substrate bias voltage; error bars indicate 5 different measured spots on each sample.

Ma et al. found that an increasing surface roughness and energy leads to increased macrophage vitality on hydrophilic DLC coatings [78]. The hemocompatibility of DLC is related to lower protein adsorption on the hydrophilic surface [78–81]. Further investigations suggest that the biocompatible properties also depend on the hydrogen content and the ratio of  $sp^2/sp^3$  hybridization in the DLC layer. The latter characterizes the material structure with regard to graphitic and diamond-like properties. The  $sp^2/sp^3$  ratio has been shown to affect the vitality of macrophages, as well as bacterial adhesion [82], hemocompatibility [83] and dermal fibroblast adhesion [84]. Wei et al. found that a high surface roughness of the substrate before coating, hydrophobicity, low surface free energy, low hydrogen content, and high residual stresses of the DLC layers negatively impacted cell vitality [85]. Chan et al. found that Cu-containing DLC layers with different  $w_{Cu}$  significantly reduced the adhesion of E. coli on coated glass substrates. The activity of the bacteria on Cu-free DLC layers was about 40%, whereas samples with a mass fraction of  $w_{Cu} > 58.76\%$  showed an antibacterial activity of 99.9%. However, they did not demonstrate the antibacterial influence of surface roughness [55]. Tsai et al. also investigated Cu-containing DLC layers and found an increase in the hydrophobicity of the sample surface with increasing  $w_{Cu}$ , which is consistent with the results obtained in this work [86].

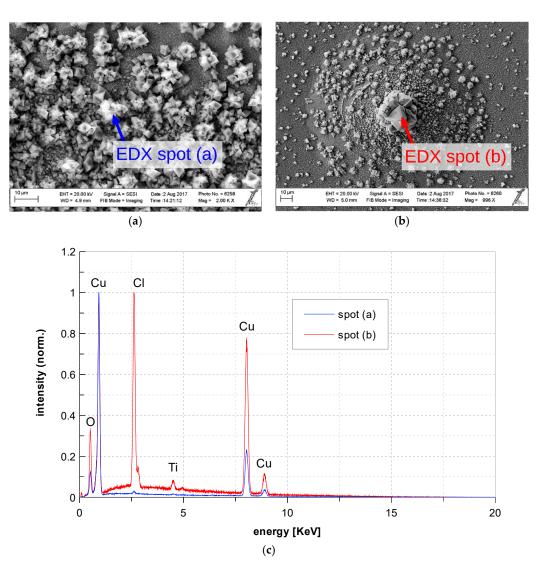
It can be concluded that a surface's biocompatibility is best in a narrow regime of hydrophilicity, at least in vitro. Accordingly, the low wettability of the a-C:H:Cu layers is critical in terms of their biocompatibility but beneficial to their antibacterial properties. A supplementary bias voltage can increase their wettability and therefore biocompatibility.

# 3.3. Release Kinetics of $Cu^{2+}$ Ions

The release of Cu<sup>2+</sup> ions (furthermore referred to as Cu release) was measured by the anodic stripping voltammetry (ASV) method at different time intervals. At first, an a-C:H:Cu coated sample

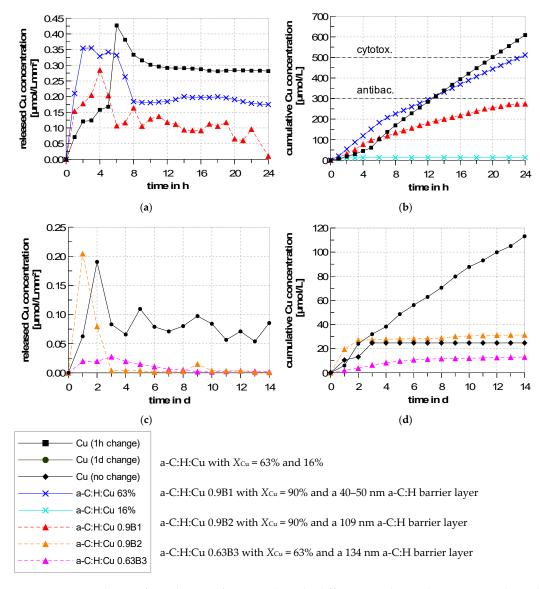
with  $X_{Cu} = 80\%$  was put into a vessel and immersed in Ringer's solution. After a few minutes, the surface color turned blue as a result of the oxidation of the Cu-containing surface.

After 24 h of immersion, EDX measurements on the sample surface were conducted showing high proportions of oxygen and chlorine (Figure 7c). SEM images revealed crystalline structures grown on the sample surface (Figure 7a,b). This suggested that the high concentration of Cu in the interface between the surface and Ringer's solution had formed a mixture of copper oxides, hydroxides and chlorides that had precipitated on the sample surface. The measured Cu molar concentration in Ringer's solution were correspondingly low, since the formation of the passivation layer inhibited the release of Cu<sup>2+</sup> ions. In the human body there is a constant gas, mass and fluid exchange between the blood and surrounding tissue which prevents the formation of such passivation layers. Therefore, a steady exchange of the medium during the release measurements was simulated by a peristaltic pump, by which the release medium was kept in constant flow. The continuous movement of the liquid reduced the formation of the passivation layer. The influence of a regular media change was also examined. For this purpose, pure Cu layers of 1 µm thickness were deposited on Ti6Al4V substrates by sputtering. This results in a calculated total Cu amount of  $n_{Cu} = 10.47$  µmol and surface concentration of 110.16 nmol/mm<sup>2</sup>. A density of  $\varrho = 7$  g/cm<sup>3</sup> for sputtered copper [71] was used for calculation.



**Figure 7.** SEM images of the copper chloride and hydroxide crystals on an a-C:H:Cu layer with  $X_{Cu} = 80\%$  after immersion in Ringer's solution (spot (**a**), red curve) and (spot (**b**), blue curve) and associated EDX spectra (**c**).

Two samples were immersed in Ringer's solution for 14 days. For one sample the release medium was not changed, for the second sample the medium was changed daily. A third sample was immersed for 24 h with an hourly medium change. The Cu molar concentration of Ringer's solution was measured at each medium change. The individually measured Cu releases and the cumulative molar concentrations are shown in Figure 8. When the release medium was not changed, the passivation layer inhibited the Cu release after 5 days. With a daily medium change, there was an increase in Cu release until the second day to  $0.19 \,\mu mol/L \cdot mm^2$ . This dropped to approximately  $0.05-0.1 \,\mu mol/L \cdot mm^2$  per day and remained constant over the entire measurement period of 14 days (see Figure 8c,d). This was due to the formation of the passivation layer within the first two days, which reduced the Cu release in the following days.



**Figure 8.** Cu release of Cu layers ( $d = 1 \mu m$ ) with different medium change intervals and different a-C:H:Cu layer systems ( $X_{Cu} = 63\%$  and 16%) in Ringer's solution over 24 h (**a**) and corresponding cumulative Cu concentration (**b**), or over 14 days (**c**) and corresponding cumulative Cu concentration (**d**), respectively.

If the release medium was changed hourly, an increased release of copper over 24 h was measured. Within the first 5 h the release increased up to 0.168  $\mu$ mol/Lmm<sup>2</sup>. From 6 h the release increased 2.5-fold to 0.426  $\mu$ mol/L·mm<sup>2</sup> and then dropped to an approximately constant 0.28  $\mu$ m/L·mm<sup>2</sup> (see

Figure 8a). The reasons for the strong increase in Cu release that occurred within the first 6 h or 2 days should be further investigated. It is possible that with the dissolution of Cu the roughness of the surface increased. This expanded the contact area between the Cu layer and the release medium, thereby enhancing the Cu release. The subsequent decrease of the Cu release can again be explained by the formation of the passivation layer. A direct comparison of both the 24 h and 14 days samples shows that the hourly medium change led to an approximately 4-fold higher release. If one considers the cumulative molar concentration in Figure 8b it becomes clear that more than the same amount of material was released (0.14 mmol/L) after 7 h with an hourly medium change as was released after 14 days with a daily medium change (0.11 mmol/L). Overall, a cumulative molar concentration of 0.61 mmol/L was achieved after 24 h. This corresponds to a released amount of  $n_{Cu,R} = 6.1 \mu mol$  and thus about 58% of the calculated total Cu amount of the Cu layer ( $n_{Cu} = 10.47 \mu mol$ ). With this Cu layer, the antibacterial concentration for *Staphylococcus aureus* in PBS (phosphate buffered saline) of 300 µmol/L was exceeded after 13 h and the cytotoxic concentration of 500 µmol/L was exceeded after 20 h [67].

To study the Cu release from a-C:H:Cu layers, two samples were prepared with  $X_{Cu} = 63\%$  (a-C:H:Cu<sub>0.63</sub>) and  $X_{Cu} = 16\%$  (a-C:H:Cu<sub>0.16</sub>) and immersed in Ringer's solution for 24 h. The time intervals of the measured Cu releases are shown in Figure 8a. For both a-C:H:Cu layers, the Cu release was the highest in the first 5 h (0.35 and 0.075 µmol/Lmm<sup>2</sup>), in contrast to the abrupt increase after just 6 h for the pure Cu layer. This can be explained by the weakly bounded, near-surface Cu nanoparticles in the a-C:H matrix, which rapidly desorbed. At the same time, the Cu nanoparticles had a higher surface area–volume ratio than a closed Cu layer. After 5 h, the Cu release of the sample a-C:H:Cu<sub>0.63</sub> dropped to an approximately constant 0.18–0.2 µmol/Lmm<sup>2</sup> and was thus below that of a pure Cu layer. For sample a-C:H:Cu<sub>0.16</sub>, the Cu release stopped after 6 h. This difference between the two a-C:H:Cu layers can be explained by the fact that after dissolving the Cu nanoparticles at high  $X_{Cu}$  a porous a-C:H matrix remained. On the one hand, the release medium could penetrate through these pores and thus dissolve Cu in greater depth of the a-C:H:Cu coating; on the other hand, Cu located in deeper layer regions could diffuse through the pores to the surface. The corresponding chemical mechanism was the oxidation of the Cu nanoparticles by hydronium ions and dissolved oxygen in the release medium according to the reaction equation:

$$2Cu(s) + 4H_3O^+ + O_2(aq) + \rightarrow Cu^{2+}(aq) + 6H_2O$$
(aq) aqua = dissolved (s) solid = film (1)

Considering the cumulative molar Cu concentration in Figure 8b, it becomes clear that the a-C:H:Cu<sub>0.63</sub> layer, despite its low Cu amount of  $n_{Cu} = 6.6 \ \mu mol$ , released 0.5 mmol/L within 24 h ( $n_{Cu,R} = 5 \ \mu mol$ ), which is an almost equal amount of copper released as the pure Cu layer. This was mainly due to the higher release within the first 6 h. The a-C:H:Cu<sub>0.63</sub> layer exceeded the antibacterial concentration of 300  $\ \mu mol/L$  for *S. aureus* after 13 h. The a-C:H:Cu<sub>0.16</sub> layer released only 0.0126 mmol/L ( $n_{Cu,R} = 0.126 \ \mu mol$ ) within 24 h and did not reach an antibacterial concentration in the observed period of time.

EDX measurements showed Cu mole fractions of  $X_{Cu,0.63} = 19\%$  and  $X_{Cu,0.16} = 15\%$  for the a-C:H:Cu layers after immersion. Thus, for the a-C:H:Cu<sub>0.63</sub> layer,  $X_{Cu}$  was reduced by 44 percentage points and consequently 70% of the total amount of contained Cu was released. The EDX result is in good agreement with the measured amount of Cu release compared to the original total Cu amount of the layer ( $n_{Cu,R}/n_{Cu} = 6.6 \mu mol/5 \mu mol \times 100\% = 76\%$  of the original total amount of Cu). The small discrepancy can be explained by the fact that the EDX measures an integral signal which is depth-dependent. It is possible that Cu that previously was deeper in the a-C:H:Cu layer diffused to the surface and thereby amplified the Cu signal in the spectrum. Furthermore, the calculated total Cu amount of the layers was only estimated values and subject to errors.

Additionally, the influence of an a-C:H barrier layer on the Cu release was investigated. For this purpose, the  $C_2H_2$  partial pressure was increased towards the end of the deposition phase. In this

way, an a-C:H barrier layer (B1) of 40–50 nm thickness was deposited on an a-C:H:Cu layer with  $X_{Cu} \approx 90\%$  (Figure 9a). Figure 8a shows an unsteady Cu release for the sample a-C:H:Cu<sub>0.9B1</sub> within the first 24 h. Within the first 4 h, the Cu release increased up to 0.285 µmol/Lmm<sup>2</sup> and then dropped to values around 0.1 µmol/Lmm<sup>2</sup>. Over the last 20 h the release showed a constant reduction to 0.0077 µmol/Lmm<sup>2</sup> and after 24 h a cumulative molar Cu concentration of 0.27 mmol/L ( $n_{Cu,R} = 2.7 \mu$ mol) was released, which was just below the antibacterial concentration of 300 µmol/L in PBS. Accordingly, the cumulative Cu release was halved when compared to the a-C:H:Cu<sub>0.63</sub> layer without the barrier layer. Thus, the a-C:H barrier layer effectively reduced the release of the copper.

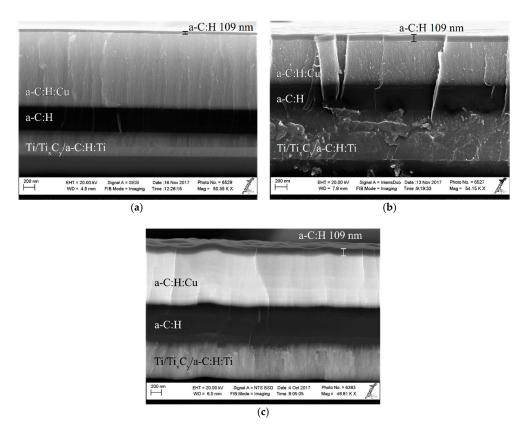


Figure 9. Layer system structure of sample a-C:H:Cu<sub>0.9B1</sub> (a), a-C:H:Cu<sub>0.9B2</sub> (b) and a-C:H:Cu<sub>0.63B3</sub> (c).

Next, the thickness of the a-C:H barrier layer (B2) was increased to 109 nm (Figure 9b) and its Cu release was measured over 14 days with a daily medium change (Figure 8c). The sample a-C:H:Cu<sub>0.9B2</sub> showed the highest Cu release within the first 24 h (0.2  $\mu$ mol/L·mm<sup>2</sup>), which was in the order of the previous sample a-C:H:Cu<sub>0.9B1</sub> with a barrier layer of 40–50 nm thickness. After 3 days, the Cu release was greatly reduced to values around  $3.5 \times 10^{-3} \mu$ mol/Lmm<sup>2</sup>, and after 14 days a cumulative molar Cu concentration of 0.0313 mmol/L ( $n_{Cu,R} = 0.313 \mu$ mol) was released. As a result, the thicker a-C:H barrier layer significantly reduced the Cu release when compared to the pure Cu layer over 14 days (Figure 8c,d).

Furthermore, an a-C:H barrier layer (B3) of 134 nm thickness was deposited onto an a-C:H:Cu<sub>0.63</sub> layer (Figure 9c). This reduced the Cu release by two orders of magnitude when compared to the layer systems without a barrier layer. The highest Cu release was measured after 3 days with 0.027  $\mu$ mol/Lmm<sup>2</sup>, which decreased approximately linearly to  $1.7 \times 10^{-3}$ – $3.4 \times 10^{-3} \mu$ mol/L·mm<sup>2</sup> within the next 5 days and then remained constant. After 14 days, a cumulative molar Cu concentration of 0.0129 mmol/L ( $n_{Cu,R} = 0.129 \mu$ mol) was released from the sample a-C:H:Cu<sub>0.63B3</sub> with the a-C:H barrier layer. The cumulative Cu concentrations of all samples with a barrier layer were well below the antibacterial concentration of 300  $\mu$ mol/L. The a-C:H barrier layer was therefore suitable for delaying Cu release. However, the concentration was potentially too low for antibacterial activity.

Furthermore, it is clear that  $X_{Cu}$  of the a-C:H:Cu layer and the thickness of the a-C:H barrier layer were the key factors for controlling the Cu release. In addition, the thickness of the a-C:H:Cu layer influenced the Cu release by different diffusion, but a deeper discussion goes beyond the scope of this paper. It should be noted that the additional a-C:H barrier layer prevented the formation of a passivation layer. Overall, in the period under consideration (24 h or 14 days), none of the layers released the Cu completely or reached the end of the Cu release. Therefore, further investigations with longer measurement times are necessary in this regard.

Stranak et al. investigated the Cu release of three different Ti/Cu mixed layers in 700  $\mu$ L DMEM (Dulbecco's Modified Eagle Medium (DMEM)) at 37 °C [23]:

- top-layer ( $X_{Cu} = 90\%$ ,  $n_{Cu} = 1.225 \mu mol$ );
- bottom-layer ( $X_{Cu} = 55\%$ ,  $n_{Cu} = 2.31 \mu mol$ );
- dual-layer ( $n_{Cu} = 3.71 \mu mol$ ) consisting of top and bottom layer.

The top layer released most of the Cu within 24 h and reached a Cu concentration of 1.85 mmol/L (6.71 µmol/L·mm<sup>2</sup>). Although the total Cu concentration achieved was one order of magnitude higher than that of the Cu layer and a-C:H:Cu<sub>0.63</sub> layer investigated in this work (500 and 610 µmol/L), when regarding the different sample surface area, their values do not differ greatly (5.26 µmol/L·mm<sup>2</sup> and 6.42 µmol/Lmm<sup>2</sup>). In contrast, only a fraction of the amount of Cu contained in the bottom layer was released within 24 h, resulting in a Cu concentration of 1 mmol/L (3.63 µmol/L·mm<sup>2</sup>). Further, only a small portion of the Cu (<0.901 µmol/L·mm<sup>2</sup>) was released over the measurement period of 10 days. Therefore, in this case, the Cu release was also slowed down by a mixed layer of Ti and Cu. The different Cu release of top-layer and bottom-layer is also discussed by Stranak et al. in relation to their different morphology. However, they did not take into account the different Cu mole fractions of the layers, which can have a far greater influence. The combination of these two layers as a dual-layer reached a Cu concentration of 4.7 mmol/L (17.04 µmol/L·mm<sup>2</sup>) within 24 h, releasing  $n_{Cu,R} = 3.29$  µmol or 89% of the total Cu. This release is thus three times as high as that of the Cu and a-C:H:Cu<sub>0.63</sub> layer presented in this work. After 48 h, a Cu concentration of 400 µmol/L (1.45 µmol/L·mm<sup>2</sup>) was measured, which steadily decreased during the 50 days.

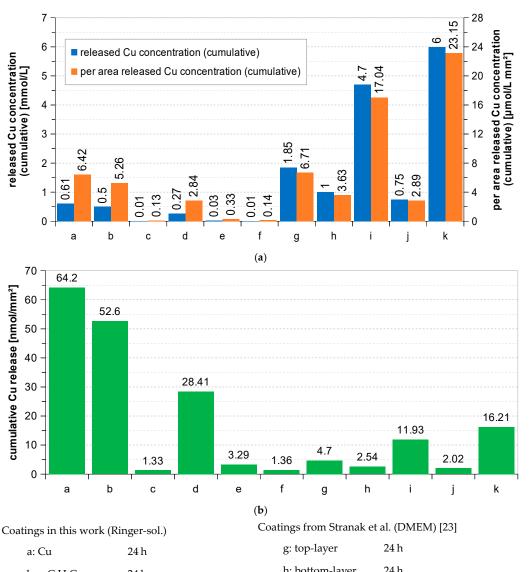
In their previous work, Stranak et al. examined three Ti/Cu layers produced by DC-MS, dual-MS, and dual-HiPIMS [71]. The total amount of Cu in the DC-MS and dual-MS layers was 0.94–1.1  $\mu$ mol, while that of the dual-HiPIMS layer was 3.15  $\mu$ mol. Again, the Cu release was highest within the first 24 h. The dual-MS layer did not release a recognizable amount of Cu, while the DC-MS layer released 750  $\mu$ mol/L (2.89  $\mu$ mol/L·mm<sup>2</sup>) and thus 0.525  $\mu$ mol or 56% of the total amount of Cu. No further Cu release of the sample was measured after 24 h. Due to the low Cu release, there was no antibacterial activity of both layers against *S. epidermidis* or *S. aureus* in DMEM, although the Cu concentration of the DC-MS layer was above the antibacterial Cu concentration of 300  $\mu$ mol/L in PBS [67]. This can be attributed to the fact that DMEM promotes bacterial growth as a nutrient medium. At the same time, in cell culture media Cu ions are bound to proteins and amino acids in the medium, whereby they no longer have an antibacterial effect.

By contrast, the dual-HiPIMS layer released 6 mmol/L (23.15  $\mu$ mol/Lmm<sup>2</sup>) after 24 h and thus  $n_{Cu,R} = 4.2 \mu$ mol. It becomes clear that all Cu was released within the first 24 h. The enhanced Cu release of the dual-HiPIMS layer is also explained by the altered crystallographic properties. It led to an antibacterial effect of the layer against *S. epidermidis* and *S. aureus* in DMEM, which was higher in the planktonic form than in the biofilm-forming. However, their work does not deal with the different Cu mole fraction of the individual layers, which may also be the cause of the different release.

Furthermore, when discussing the results from the literature, it should be noted that different release media volumes were used for Cu release measurements (10 mL, 5 mL, 700  $\mu$ L). Assuming there were equal amounts of dissolved Cu, this resulted in differences in measured concentrations, but when the released Cu concentrations are correlated to the release media volume, the results draw a different picture (Figure 10).

released Cu concentration (cumulative) [mmol/L]

cumulative Cu release [nmol/mm<sup>2</sup>]



a: Cu	24 h	g. top myer		
b: a-C:H:Cu0.63	24 h	h: bottom-layer	24 h	
c: a-C:H:Cu <sub>0.16</sub>	24 h	i: dual-layer	24 h	
d: a-C:H:Cu0.63B	14 days	Coatings from Stranak et al. (DMEM) [71]		
e: a-C:H:Cu0.9B1	14 days	j: DC-MS	24 h	
f: a-C:H:Cu0.9B2	14 days	k: dual-HiPIMS	24 h	

Figure 10. Comparison of the released Cu concentration (cumulative) of different Cu composite layers (a) and comparison of the area-related and volume-adjusted Cu release (cumulative) of different Cu composite layers after 24 h or 14 days (b).

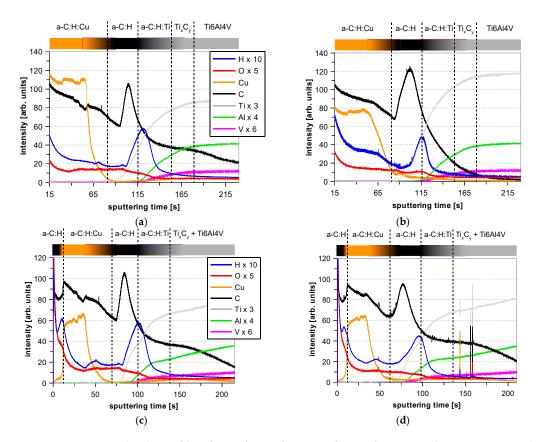
It is clear from the cited work that the control of the release kinetics of the antibacterial substance in medical use is crucial. It must be large enough to stop infection, while maintaining the vitality of the body's own cells. For this purpose, this work can make a significant contribution since it determines how the Cu release can be controlled by a nanoconfigured layer structure. Based on this, coating systems can be modified, expanded and adapted to specific medical requirements. For a final assessment, microbiological studies on the antibacterial efficacy and cytotoxicity of the a-C:H:Cu layers are missing.

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#### 3.4. GDOES Depth Profiles

The GDOES depth profiles in Figure 11 show the distinct layer structures. The layer system consists of C, H (a-C:H) and Cu, as well as significant amounts of O as a contaminant. When reaching the substrate, elements of the Ti6Al4V (Ti, Al, V) were found. Beginning from the Ti6Al4V substrate (right side), the carbon and hydrogen peaks rose as a result of the gradient C deposition from Ti to  $Ti_xC_y$  and a-C:H:Ti. The high amount of H can be attributed to low hydrocarbon ion energies because no substrate voltage was used during this deposition step. When switching to the a-C:H deposition in the rf-magPECVD process, the hydrogen signal decreased. This was a result of the high substrate bias voltage of 1 kV during a-C:H deposition that resulted in high hydrocarbon ion energies, which is known to lower the H deposition in a-C:H coatings [87]. When reaching the a-C:H:Cu layer, the copper content showed a chemical gradient that was a result of the gradient copper deposition process which was used to enhance the adhesion of the a-C:H:Cu to the a-C:H layer. Further information regarding the graded copper deposition is summarized in a further paper [73]. Following that procedure, the copper was homogeneously distributed in the a-C:H:Cu coatings over coating depth. When comparing the GDOES results of the samples with the a-C:H:Cu<sub>0.63</sub> layer before and after release measurement (Figure 11a,b), one can see the reduced amount of copper in the a-C:H:Cu<sub>0.63</sub> layer after release measurement.

In the case of the sample with the barrier layer (Figure 11c,d), the copper content was reduced to 0 at the surface while the carbon content remained high. It is evident that the amount of copper was not significantly reduced, which is consistent with the release measurements in the previous section.



**Figure 11.** GDOES depth profile of a Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti/a-C:H/a-C:H:Cu layer system with a-C:H:Cu<sub>0.63</sub> containing  $X_{Cu}$  = 63% before (**a**) and after (**b**) release measurement, as well as a-C:H:Cu<sub>0.63B3</sub> with an a-C:H barrier layer before (**c**) and after (**d**) release measurement.

#### 4. Conclusions

In this paper we reported on the surface properties and the time-resolved release kinetics of copper ions ( $Cu^{2+}$ ) from a-C:H:Cu coatings that were deposited onto Ti6Al4V substrates, and how the release can be controlled by an a-C:H barrier layer.

Cu containing a-C:H:Cu composite coatings were deposited using a hybrid RF-PVD/PECVD process and grown surface topography, and Cu release in Ringer's solution was investigated. The deposited films consisted of three layers: the Ti/Ti<sub>x</sub>C<sub>y</sub>/a-C:H:Ti/a-C:H interlayer, the Cu-containing a-C:H:Cu layer and an a-C:H barrier layer. The performed AFM measurements showed an increasing roughness of the sample surface with increasing  $X_{Cu}$  in the a-C:H:Cu layer, which is consistent with the literature. This can be attributed to the size of Cu nanoparticles embedded into the a-C:H matrix. The static contact angle of Ringer's solution on the layers also increased with increasing  $X_{Cu}$  and, therefore, Cu nanoparticle size. Thus, the wetting behaviour changed from hydrophilic (polished Ti6Al4V and Cu-free a-C:H layer) to increasingly hydrophobic (a-C:H:Cu). Additionally, an influence of the substrate bias voltage on the wettability was determined. This could be related to a change in content of sp<sup>3</sup> and sp<sup>2</sup> hybridization in the a-C:H matrix [77,87]. The low wettability of the a-C:H:Cu layers is critical in terms of their biocompatibility, but beneficial to their antibacterial properties.

The release of  $Cu^{2+}$  ions from sputtered Cu layers and a-C:H:Cu layers with different  $X_{Cu}$  in Ringer's solution was examined. As a reference, a pure Cu layer was used to investigate the maximum possible Cu release. When using a stationary solution, a passivation layer of sparingly soluble copper oxides, hydroxides and chlorides precipitated on the surface of the Cu and a-C:H:Cu layers, which inhibited further release of Cu. This can be prevented by regularly changing the release medium and generating a continuous flow of the medium.

It has been shown that the release kinetics of the Cu<sup>2+</sup> ions from the a-C:H:Cu layers in the medium over 14 days could be controlled by adjusting  $X_{Cu}$  in the a-C:H:Cu layers and the thickness of an additional a-C:H barrier layer. This enabled a release over a long period of time. The pure Cu layer and a-C:H:Cu<sub>0.63</sub> layer reached a potential antibacterial Cu concentration for *S. aureus* in PBS of at least 300 µmol/L [67] after 13 h. In contrast, this antibacterial concentration was not reached for the a-C:H:Cu<sub>0.16</sub> layer or the samples with a barrier layer in the period considered.

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