



Review

# Review of Antibacterial Activity of Titanium-Based Implants' Surfaces Fabricated by Micro-Arc Oxidation

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**Abstract:** Ti and its alloys are the most commonly-used materials for biomedical applications. However, bacterial infection after implant placement is still one of the significant rising complications. Therefore, the application of the antimicrobial agents into implant surfaces to prevent implant-associated infection has attracted much attention. Scientific papers have shown that inorganic antibacterial metal elements (e.g., Ag, Cu, Zn) can be introduced into implant surfaces with the addition of metal nanoparticles or metallic compounds into an electrolyte via micro-arc oxidation (MAO) technology. In this review, the effects of the composition and concentration of electrolyte and process parameters (e.g., voltage, current density, oxidation time) on the morphological characteristics (e.g., surface morphology, bonding strength), antibacterial ability and biocompatibility of MAO antimicrobial coatings are discussed in detail. Anti-infection and osseointegration can be simultaneously accomplished with the selection of the proper antibacterial elements and operating parameters. Besides, MAO assisted by magnetron sputtering (MS) to endow Ti-based implant materials with superior antibacterial ability and biocompatibility is also discussed. Finally, the development trend of MAO technology in the future is forecasted.

Keywords: micro-arc oxidation; antibacterial ability; Ag; Cu; Zn

#### 1. Introduction

Biomedical titanium (Ti) and its alloys have been widely used in blood vessels, artificial joints, dental implantations and bone screws [1–3] on account of their excellent mechanical properties, good corrosion resistance and favorable biocompatibility. However, implant-associated infection (IAI) remains one of the most devastating postoperative complications [4,5] despite strict sterilization and systemic antibiotic prophylaxis prior to surgery. IAI often commences with bacterial adhesion to the implant and colonization on the implant surface, followed by biofilm formation. The biofilms are extraordinarily resistant to antibiotics and the host immunity defensive system [6–8], leading to further complications. Once the biofilm is formed, it is often not effectively treated except for prosthesis removal and re-implantation [9,10]. This devastating complication may result in chronic suffering and extremely huge medical expenses [11,12]. Therefore, it is highly desirable to introduce antimicrobial agents into implant surfaces to provide antibacterial activities and prevent peri-implant infections [13–17]. In comparison with organic antibiotics, inorganic antibacterial metal elements (e.g., silver (Ag) [18–22], copper (Cu) [23–26] and zinc (Zn) [27–29]) have attracted great attention due to their perfect stabilities, superior broad-spectrum antibacterial properties, relatively low toxicity to human cells and low risk of producing resistant strains [30,31].

Dizaj et al. [32] have reviewed the antimicrobial activity of metal and metal oxide nanoparticles together with their antimicrobial mechanisms and have indicated that the particle size determined the antimicrobial effectiveness of the metal nanoparticles. Rai et al. [33] have extensively reviewed

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the antibacterial, antifungal and antiviral properties of Ag ions, Ag compounds and Ag nanoparticles, but there is no discussion about the antibacterial properties of Cu or Zn. In addition, the toxicities of metal and metal oxide nanoparticles to apply as proper alternatives for antibiotics in biomedical applications were not discussed in the reviews. In order to obtain antibacterial metallic surfaces, numerous surface modifications have been performed. Ferraris et al. [34] have reported the surface modification technologies, such as ion implantation [35,36], ion beam-assisted deposition [37], electrochemical techniques [38], ion exchange [39], sol-gel [40], sputtering [41,42], plasmas pray [43] and chemical vapor deposition [44]. However, micro-arc oxidation (MAO) is not fully described.

MAO, which can produce porous, adhesive and bioactive coatings for implantation, has aroused considerable attention [45-47]. On the one hand, a porous bioactive calcium phosphate-based composite layer can be deposited on Ti-based implant surfaces according to the selected electrolyte, which would enhance the biocompatibility [48,49] and the bonding strength of the coated layer [50]. On the other hand, antibacterial metal elements can be incorporated into implant surfaces to inhibit initial adhesion of bacteria and prevent post-surgery complications, thus enhancing the antibacterial property [34]. Furthermore, the content of bioactive elements and antibacterial metal elements on the MAO coating surface can be tuned by controlling voltage, electrolyte components and MAO time [51]. To our best knowledge, this specific topic has not been reviewed in the recent literature. The aim of the review is to collect and compare the recent scientific papers concerning surface modification of Ti with the incorporation of antibacterial metal elements (e.g., Ag, Cu, Zn) to endow antibacterial properties by MAO. The review focuses mainly on the effects of the composition and concentration of electrolyte and process parameters (voltage, current density, oxidation time) on the morphological characteristics, antibacterial ability and biocompatibility of MAO coating. MAO assisted by magnetron sputtering (MS) to achieve a superior antibacterial property and biocompatibility is also discussed, following the forecast of the development trend of MAO in the future.

# 2. Micro-Arc Oxidation Method

MAO, which is also referred to as plasma electrolyte oxidation (PEO), is a high voltage plasma-assisted anodic oxidation process. MAO is a relatively convenient technique for forming firmly adherent oxide ceramic layers on the surfaces of valve metal, such as Ti, Al, Mg, Zr, Ta and their alloys. As shown in Figure 1, the MAO process was controlled by an MAO power supply. Before MAO, the non-working side of the specimen was connected with a copper conductor and coated with acrylate adhesive. During the MAO process, the specimen was treated as the anode, and the stainless steel electrolytic bath was regarded as the cathode to fill with electrolyte. The stirrer was applied to keep the electrolyte uniform, and the circulating water treatment system was used to sustain the temperature of the electrolyte below 30 °C.

The principle of MAO is explained in Figure 2. At the initial stage of MAO treatment, as soon as the specimen is exposed to the electrolyte, the voltage increases rapidly and linearly with time, and an anodic barrier film, also called a passivating film (Figure 2a), is initially formed on the surface of the specimen. Soon afterwards, with the increase of voltage, some tiny oxygen bubbles can be observed, and a porous insulating oxide layer, which usually grows under conditions of dielectric breakdown, forms on the surface of the specimen [52]. At this stage, the voltage and current flow follow Faraday's law, which corresponds to the conventional anodizing stage (Figure 2b). When the applied voltage on the specimen surpasses the dielectric breakdown of the insulating oxide coating, dielectric breakdown takes place, leading to the formation of spark discharges (Figure 2c). The current flow only concentrates on regions of breakdown, and the Ca and P elements from the electrolyte and the other elements from the specimen enter into regions of breakdown by diffusion and electrophoresis at intense local high temperatures, resulting in localized forming and thickening of the porous structure oxide coating. The discharge sparks gradually grow bigger, and the micro-arcs' discharges are transformed into powerful arc discharges (Figure 2d). When the new generated oxide coating (Figure 2e) is capable of resisting current flow, the other regions are vulnerable to breakdown due to the smaller resistance, and

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finally, the chemical reaction interface would move towards the entire surface [53]. Breakdown of the coating occurs at a vulnerable spot of the growing oxide film. Meanwhile, the new porous structure oxide coating might also be formed and thickened by the emission of gas due to intense local high temperatures at the sites of breakdown [54]. With the prolonging of oxidation time, large discharge channels with intense sparking and gas bubbles would emerge at the surface, leading to the formation of larger protruding pores, even a spongy interconnected microstructure (Figure 2f). The continuous formation and breakdown of the oxide coating (Figure 2g) causes the potential to fluctuate. Both the dissolution of the base material and gasification of the electrolyte enable the formation of the porous ceramic oxide coating (Figure 2h).

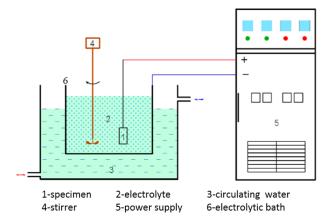
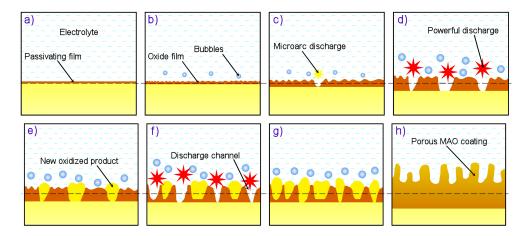


Figure 1. Schematic diagram of the micro-arc oxidation (MAO) experimental device.



**Figure 2.** Schematic of the formation process of MAO porous coating. The passivating film (a), and porous insulating oxide film (b) is initially formed. Then under the action of the spark discharges (c) and, powerful arc discharges (d), the new generated oxide coating (e) is formed and thicken. Finally, the porous ceramic oxide coating is formed (h) with the continuous formation and breakdown of the oxide coating (g) at the large discharge channels (f).

The composition of the MAO electrolytes has a dramatic effect on the morphological characteristics, such as the porosity and thickness of MAO coatings. Hence, the proper selection of the electrolyte composition is imperative to obtain satisfactory performance. Generally, different antibacterial metal elements mixed with calcium acetate (CA) and glycerophosphate disodium (GP) are used as the base electrolyte in the MAO process to enhance the antibacterial property of the implant materials. The characteristics of the fabricated antibacterial coating prepared by MAO techniques are briefly summarized in Table 1, and the details are reviewed according to the difference of the doping method of the antibacterial metal elements.

**Table 1.** Summary of the characteristics of antibacterial coating by MAO.

Ti Alloy	Electrolyte	Electrical Parameter						Release			
		Voltage (V)	Current Density (A·dm <sup>-2</sup> )	Oxidation Time (s)	Surface Topography (Pore Size)	XRD Detected Phase	Surface Content of Ag/Cu/Zn (wt %)	Amount of Ag/Cu/Zn (ppb)	Tested Bacteria	Biocompatibility	References
Ti-6Al-7Nb	$0.02~M~Ca\text{-GP}, 0.15~M~CA,$ and $3.0~g\cdot L^{-1}~Ag~NPs$	<250	20	300	Porous structures (<3 μm) with Ag NP of 37 nm	Ti, rutile and anatase	0.03	=	S. aureus	-	[55]
Ti-6Al-7Nb	$0.02$ M Ca-GP, $0.15$ M CA, and $3.0~g\cdot L^{-1}$ Ag NPs	<250	20	0-300	Porous structures (<5 $\mu$ m) with Ag NP of 7–25 nm	-	-	-	-	-	[56]
Ti-6Al-7Nb	0.02 M Ca-GP, 0.15 M CA, and 0.3 g·L $^{-1}$ Ag NPs	$234\pm3$	20	300	Porous structures (<5 μm)	-	-	12	S. aureus	Human osteoblastic cell	[38]
Ti-6Al-7Nb	$0.02$ M Ca-GP, $0.15$ M CA, and $3.0~g\cdot L^{-1}$ Ag NPs	$237\pm2$	20	300	Porous structures (<5 μm)	-	-	89	S. aureus	Human osteoblastic cell	[38]
Cp-Ti	$2\mathrm{g}\cdot\mathrm{L}^{-1}$ NaOH, $15\mathrm{g}\cdot\mathrm{L}^{-1}$ NaH $_2$ PO $_4$ and $3.0\mathrm{g}\cdot\mathrm{L}^{-1}$ Cu NPs	=	20	300	Porous structures (<5 μm) with Cu NP of 60 nm	=	=	=	Escherichia coli, S. aureus	-	[57]
Cp-Ti	$0.04$ M $\beta\text{-GP}, 0.4$ M CA and $0.004$ M $AgNO_3$	250-350	=	180	Spherical pores (<3 μm)	Ti, rutile, and anatase	-	-	S. aureus	Human osteosarcoma (HOS) cell	[58]
Ср-Ті	0.04 M β-GP, 0.4 M CA and 0.004 M AgNO <sub>3</sub>	420	=	180	Irregular and rough pores with spherical particles and flake	Rutile, $\beta$ -Ca <sub>2</sub> P <sub>2</sub> O <sub>7</sub> , $\alpha$ -TCP and HA	0.21-0.45	-	S. aureus	Human osteosarcoma (HOS) cell	[58]
Cp-Ti	0.04 M β-GP, 0.4 M CA and 0.00006 M AgNO <sub>3</sub>	420	=	180	Irregular and rough pores with spherical particles and flake	Rutile, $\beta$ -Ca <sub>2</sub> P <sub>2</sub> O <sub>7</sub> , $\alpha$ -TCP and HA	0.1	=	S. aureus	Human osteosarcoma (HOS) cell	[58]
Ti6Al4V	β-GP, CA and 0.1 g·L $^{-1}$ AgNO $_3$	400	=	300	Granular morphology with Ag NPs of 20–30 nm	Ti, rutile, anatase, CaTiO <sub>3</sub> and HA	0.6	2500	E. coli	-	[59]
Ti6Al4V	β-GP, CA and $0.4 \text{ g} \cdot \text{L}^{-1} \text{ AgNO}_3$	400	=	300	Needle-like morphology with Ag NPs of 20–30 nm	Ti, rutile, anatase, CaTiO <sub>3</sub> and HA	2.1	8000	E. coli	-	[59]
Cp-Ti	Na <sub>2</sub> HPO <sub>4</sub> , CA, and 0.0025 M CH <sub>3</sub> COOAg	380	_	300	Flake-like morphology with regional Ag particles less than 200 nm	Ti, rutile, anatase, CaTiO <sub>3</sub> and HA	4.6	-	E. coli, S. aureus	-	[60]
Ср-Ті	0.5–1.0 g·L <sup>-1</sup> AgNO <sub>3</sub>	-	65	5–240	Highly ordered nanopores with Ag NPs of 10–30 nm within micropits	-	-	200–450	S. aureus	Newborn mouse pre-osteoblast cells	[61]
Cp-Ti	7.6 g·L $^{-1}$ Na $_3$ PO $_4$ , 9.4 g·L $^{-1}$ Ca(NO $_3$ ) $_2$ and 1.0 g·L $^{-1}$ AgNO $_3$	-	65	240	Highly ordered nanopores with Ag NPs of 10–30 nm within micropits	-	-	-	S. aureus	Newborn mouse pre-osteoblast cells	[61]

 Table 1. Cont.

Electrolyte	Electrical Parameter						Release			
	Voltage (V)	Current Density (A·dm <sup>-2</sup> )	Oxidation Time (s)	Surface Topography (Pore Size)	XRD Detected Phase	of Ag/Cu/Zn (wt %)	Amount of Ag/Cu/Zn (ppb)	Tested Bacteria	Biocompatibility	References
1.0~8.0 g·L <sup>−1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub>	=	65	240	Mesopores (20–40 nm) within micropits	Ti	-	_	-	Osteoblast cells	[62]
$3.8 \sim 7.6 \text{ g} \cdot \text{L}^{-1} \text{ Na}_3 \text{PO}_4 \text{ and} \\ 1.0 \text{ g} \cdot \text{L}^{-1} \text{ Cu(NO}_3)_2$	-	65	240	Mesopores (20–40 nm) within micropits	Ti	-	-	-	Osteoblast cells	[62]
0.05 M β-GP, 0.1 M CA and 0.05 M (CH <sub>3</sub> COO) <sub>2</sub> Cu	-	16.5	240	Micropores or crater structures (3–5 μm) with nano-grains of 30–50 nm	Ti and anatase	1.43	-	S. aureus	Human osteosarcoma cell	[63]
0.02 M β-GP, 0.2 M CA and 0.00125~0.005 M Cu (CH <sub>3</sub> COO) <sub>2</sub>	450	_	90	Micropore structures (1–4 μm)	Rutile and anatase	0.67–1.93	2.8-60.2	S. aureus	Mouse fibroblast cell	[64]
0.15 M Ca-GP, 0.02 M CA and 0.06 M ZA	-	30	300	Porous structures (<5 μm)	Ti, rutile and anatase	8.7	-	E. coli, S. aureus	Osteoblast cells	[65]
0.05 M β-GP, 0.1 M CA and 0.02~0.06 ZA	-	16.5	240	Microporous structures (<5 μm)	-	4.6–9.3	1000–3620	E. coli, S. aureus	Rat bone mesenchymal stem cells	[10]
$0.02 \text{ M}$ β-GP, $0.1 \text{ M}$ CA, $0.1 \text{ M}$ ZA, and $6 \text{ g} \cdot \text{L}^{-1}$ Ag NPs	390	=	30–90	Microporous structures (1–4 µm)	Anatase and rutile	1.06-1.42 (Ag), 22.19-26.93 (Zn)	-	S. aureus	-	[66]
0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs	390	_	120	Microporous structures (1–4 µm) with Ag NPs of 5–10 nm	Anatase, rutile and ZnO	1.56 (Ag), 29.38 (Zn)	684 (Ag), 6880 (Zn)	S. aureus	-	[66]
$0.02~M~\beta$ -GP, $0.1~M~CA$ , $0.1~M~ZA$ , and $6~g\cdot L^{-1}~Ag~NPs$	390	-	240	Microporous structures (1–4 µm) with some deposits	Anatase, rutile, ZnO and Zn <sub>2</sub> TiO <sub>4</sub>	1.58 (Ag), 31.27 (Zn)	-	S. aureus	-	[66]
	1.0~8.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> 3.8~7.6 g·L <sup>-1</sup> Na <sub>3</sub> PO <sub>4</sub> and 1.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> 0.05 M β-GP, 0.1 M CA and 0.05 M (CH <sub>3</sub> COO) <sub>2</sub> Cu  0.02 M β-GP, 0.2 M CA and 0.00125~0.005 M Cu (CH <sub>3</sub> COO) <sub>2</sub> 0.15 M Ca-GP, 0.02 M CA and 0.06 M ZA  0.05 M β-GP, 0.1 M CA and 0.02~0.06 ZA  0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs  0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs	1.0~8.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> - 3.8~7.6 g·L <sup>-1</sup> Na <sub>3</sub> PO <sub>4</sub> and 1.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> - 0.05 M β-GP, 0.1 M CA and 0.05 M (CH <sub>3</sub> COO) <sub>2</sub> Cu - 0.02 M β-GP, 0.2 M CA and 0.00125-0.005 M Cu (CH <sub>3</sub> COO) <sub>2</sub> - 0.15 M Ca-GP, 0.02 M CA and 0.06 M ZA - 0.05 M β-GP, 0.1 M CA and 0.02~0.06 ZA - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs - 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65     3.8~7.6 g·L <sup>-1</sup> Na <sub>3</sub> PO <sub>4</sub> and 1.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub>   - 65     0.05 M β-GP, 0.1 M CA and 0.05 M (CH <sub>3</sub> COO) <sub>2</sub> Cu   - 16.5     0.02 M β-GP, 0.2 M CA and 0.00125~0.005 M Cu (CH <sub>3</sub> COO) <sub>2</sub>   450   - (CH <sub>3</sub> COO) <sub>2</sub>     0.15 M Ca-GP, 0.02 M CA and 0.06 M ZA   - 30     0.05 M β-GP, 0.1 M CA and 0.02~0.06 ZA   - 16.5     0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs   390   - 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        16.5         240           0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs         390         -         30–90           0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs         390         -         120           0.02 M β-GP, 0.1 M CA, 0.1 M ZA, and 6 g·L <sup>-1</sup> Ag NPs         390         -         120	1.0~8.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub>	Density (N)   Density (A-dm^2)   Time (s)   Density (A-dm^2)   Time (s)	1.0~8.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub>	Electrolyte         Voltage (V)         Current Density (N-dmm^-2)         Oxidation Time (s)         Surface Topography (Pore Size)         XRD Detected Phase         Surface Content of Ag/Cu/Zn (wt %)         Amount of Ag/Cu/Zn (wt %)           1.0~8.0 g.L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> -         65         240         Mesopores (20–40 nm) within micropits         Ti         -         -           3.8~7.6 g.L <sup>-1</sup> Na <sub>3</sub> PO <sub>4</sub> and 1.0 g.L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> -         65         240         Mesopores (20–40 nm) within micropits         Ti         -         -           0.05 M β-GP, 0.1 M CA and 0.05 M (CH <sub>3</sub> COO) <sub>2</sub> Cu         -         16.5         240         Micropores or crater structures (3–5 μm) with nano-grains of 30–50 nm         Ti and anatase         1.43         -           0.02 M β-GP, 0.1 M CA and 0.00125-0.005 M Cu (CH <sub>3</sub> COO) <sub>2</sub> Cu         450         -         90         Micropore structures (1–4 μm)         Rutile and anatase         0.67–1.93         2.8–60.2           0.15 M Ca-GP, 0.1 M CA and 0.06 M ZA         -         30         300         Porous structures (<5 μm)	Electrolyte         Voltage (V)         Current Density (Pore Size)         Surface Topography (Pore Size)         XRD Detected Phase         Surface Content of Ag/Cu/Zn (ppb)         Ag/Cu/Zn (ppb)         Tested Bacteria           1.0~8.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> -         65         240         Mesopores (20–40 nm) within micropits         Ti         -         -         -         -           3.8~7.6 g·L <sup>-1</sup> Na <sub>3</sub> PO <sub>4</sub> and 1.0 g·L <sup>-1</sup> Cu(NO <sub>3</sub> ) <sub>2</sub> -         65         240         Mesopores (20–40 nm) within micropits         Ti         -         -         -           0.05 M β·GP, 0.1 M CA and 0.05 M (CH <sub>3</sub> COO) <sub>2</sub> cu         -         16.5         240         Micropores or crater structures (3–5 µm) with nano-grains of 30–50 m) within nano-grains of 30–50 m) anatase         1.43         -         S. aureus           0.02 M β·GP, 0.2 M CA and 0.00125~0.005 M Cu (CH <sub>3</sub> COO) <sub>2</sub> 450         -         90         Micropores tructures (-5 µm) with nano-grains of 30–50 m anatase         Rutile and anatase         0.67–1.93         2.8–60.2         S. aureus           0.15 M Ca-GP, 0.2 M CA and 0.06 M ZA         -         30         300         Porous structures (-5 µm)         Ti, rutile and anatase         8.7         -         E. coli, S. aureus           0.05 M β·GP, 0.1 M CA and 0.02~0.06 ZA         -         16.5         240         Microporous structures (-5 µm)	Flectrolyte   Voltage   Current (Phase)   Voltage (Phase)   Vol

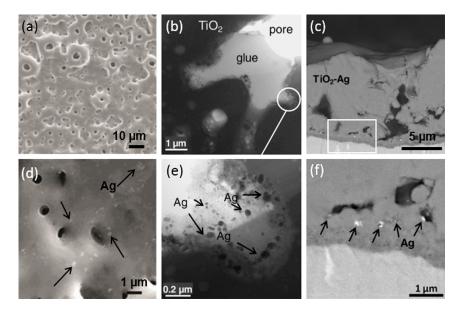
Notes: GP: Glycerophosphate; CA: Calcium acetate; α-TCP: Ca<sub>3</sub>(PO4)<sub>2</sub>; HA: Ca<sub>10</sub>(PO4)<sub>6</sub>(OH)<sub>2</sub>.

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### 2.1. Introduction of Metal Nanoparticles into MAO Electrolyte

NPs present a greater chemical activity than the bulk material for the advantage of their larger ratio of surface area to volume [67]. Ag NPs are the most popular inorganic antimicrobial NPs [68] and can be incorporated into implants by direct dispersion of solid Ag NPs in the electrolyte during MAO. The antibacterial mechanisms as reported are that Ag NPs can directly interact with the microbial cells, leading to the increase of membrane permeability [69], the degradation of lipopolysaccharide molecules with the formation of pits and gaps in the bacterial membrane and the damage of the bacterial outer membrane [70,71]. Ag NPs can also penetrate the bacterial cell envelope and cause DNA damage and protein oxidation by producing secondary products, such as reactive oxygen species [72]. It has also been demonstrated that the antimicrobial activity of Ag NPs is higher than that of Ag ion [73]. The use of Ag NPs will lead to anti-bacterial effects maximized due to the increased numbers of particles per unit area [67].

Necula et al. [55] have successfully fabricated Ag-bearing TiO<sub>2</sub> coatings with different contents of Ag NPs by MAO in the electrolyte, including CA, calcium glycerophosphate (Ca-GP) with Ag NPs. The morphology of metallic Ag NPs is similar to that of the particles in the electrolyte [56]. The average size of Ag NPs was measured to be  $37 \pm 6$  nm. Under the assistance of electrophoresis during MAO, Ag NPs were homogeneously absorbed on the porous surface (Figure 3a,d) [38], transmitted to the inner pore walls of the titanium oxide along the short-circuit paths (Figure 3b,e) [55,56] or embedded in the dense oxide layer (Figure 3c,f) [56]. The incorporation of Ag NPs in the titanium dioxide (TiO<sub>2</sub>) matrix has no effect on the surface morphology of the coatings. All of the layers were porous with well-defined pores ranging in size from a few nanometers up to 5 µm [38], which is the typical morphology of MAO. Moreover, the contents of Ag NPs on the surface went up with the increased amount of Ag NPs in the electrolyte. They found that Ag NPs displayed good antibacterial activity against methicillin-resistant Staphylococcus aureus (S. aureus), one of the most problematic pathogens for orthopedic implants, while showing little cytotoxicity to a human osteoblastic cell line with the adjustment of the content of Ag NPs on TiO<sub>2</sub>/Ag coating by decreasing the concentration of the Ag NPs in the electrolyte [74]. However, the coatings prepared by this method are expensive due to the high price of Ag NPs.

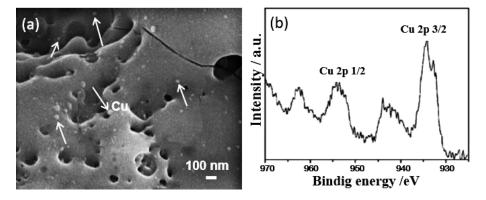


**Figure 3.** Low and high magnification surface SEM images (**a**,**d**); bright field TEM images (**b**,**e**), and cross-sectional SEM images (**c**,**f**) of the bactericidal coatings showing that the Ag NPs are distributed on the porous surface, in the micropores or embedded in the dense oxide layer. (Reprinted with permission from [38,56]; Copyright 2012 Elsevier, Copyright 2011 Elsevier.)

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Cu can inactivate the central catabolic and biosynthetic pathways, namely catalytic clusters of dehydratases [75], which endows Cu with strong antibacterial properties. Yao et al. [57] have prepared Cu-doped TiO<sub>2</sub> coatings by MAO in the working electrolyte containing NaOH, NaH<sub>2</sub>PO<sub>4</sub> and 3 g·L<sup>-1</sup> Cu NPs. The results of top-surface morphologies revealed that Cu NPs distributed both on the Cu-doped coating surface and inside the pores (Figure 4a). The size of most of the NPs was less than 60 nm. High-resolution XPS spectra indicated that Cu mainly existed in the Cu<sup>2+</sup> (from CuO) state (Figure 4b). Cu-doped TiO<sub>2</sub> coatings showed excellent antibacterial properties attributed to the incorporation of Cu NPs with a high surface area to volume ratio, which would directly kill bacteria and release copper ions. A similar mechanism was reported by Raffi et al. [76], in which the reason for the antibacterial property of Cu NPs was mainly ascribed to the adhesion of Cu NPs onto bacteria surfaces, namely direct contact-killing, since Cu NPs are oppositely charged with respect to bacteria. When they encounter each other, the reduction reaction will occur at the bacterial cell wall, resulting in the formation of cavities/pits. Though Usman et al. [77] have also testified to the antimicrobial and antifungal activities of Cu-chitosan NPs against methicillin-resistant S. aureus, B. subtilis, C. albicans, P. aeruginosa and Salmonella choleraesuis, Pape et al. [78] have confirmed that the antibacterial activity of Cu NPs was significantly weaker than that of Ag NPs. Moreover, Cu NPs were rapidly oxidized when exposed to air [79], limiting their application.

With the consideration of the bad dispersion of NPs in solution and the weak adhesion of NPs onto the coating surface, only several valuable studies have been documented in the field of antibacterial NPs by MAO so far.



**Figure 4.** SEM micrographs of Cu-doped coatings revealing the presence of Cu nanoparticles on the surface and inside the pores (**a**); high-resolution XPS spectra of Cu 2p in TiO<sub>2</sub> coating indicated that Cu mainly exists in the CuO state (**b**). (Reprinted with permission from [57]; Copyright 2014 Elsevier.)

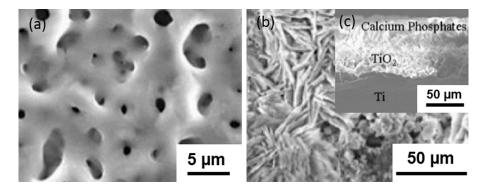
# 2.2. Introduction of Metallic Compounds into MAO Electrolyte

For the biomedical industry, it seems to be an attractive strategy to endow implants' surfaces with metallic coatings. Recent studies [45,80,81] have shown that biological or antibacterial elements can be incorporated into the coating during the MAO process by the introduction of metallic compound into the MAO electrolyte. Ag-containing coatings have occupied the largest share of the global antibacterial market.

Song et al. [58] have incorporated Ag into Ti implants by MAO in  $0.04 \,\mathrm{M}$   $\beta$ -glycerophosphate disodium salt pentahydrate ( $\beta$ -GP),  $0.4 \,\mathrm{M}$  CA and silver nitrate (AgNO<sub>3</sub>) or silver acetate (CH<sub>3</sub>COOAg) electrolytic solution at a fixed applied voltage range from 250 to 450 V. The addition of AgNO<sub>3</sub> reduced the required voltage for the formation of calcium phosphate compounds from 450 down to 400 V [82,83]. When the concentration of AgNO<sub>3</sub> was fixed at  $0.004 \,\mathrm{M}$ , the oxidized layer was a porous microstructure with spherical pores at voltages below 350 V (Figure 5a), while the coating surface became rough and covered with spherical particles and flakes above 380 V (Figure 5b) due to the existence of calcium phosphate compounds (Figure 5c). MAO coatings containing 0.21– $0.45 \,\mathrm{mt}$  % of Ag were cytotoxic

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to human osteosarcoma MG63 cells, while Chen et al. [84] reported that 2.05 wt % Ag displayed no osteoblast cytotoxicity. The comparison results indicated that the cell proliferation not only has a relation to the content of Ag, but also depends on other factors, such as the surface roughness of the coatings [85]. The relatively smooth coating oxidized at 400 V with a lower content of Ag (0.00006 M AgNO<sub>3</sub>) exhibited no cytotoxicity with an unreduced antibacterial property.



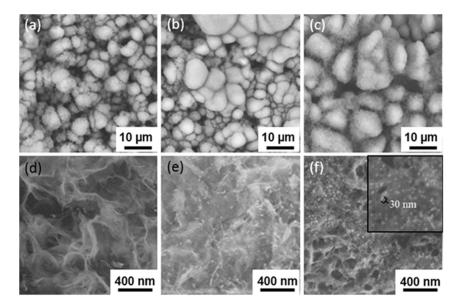
**Figure 5.** Surface morphologies of MAO samples obtained in electrolyte solution containing 0.004 M AgNO3 for 3 min at 350 V (**a**), and 400 V (**b**); (**c**) the cross-section view of the specimen in (**b**) [58]. (Reprinted with permission from [58], Copyright 2009 John Wiley & Sons.)

Muhaffel et al. [59] have also prepared novel multi-layer coatings composed mainly of inner TiO<sub>2</sub> layers and outer HA (HA ( $Ca_{10}(PO_4)_6(OH)_2$ )) layers via MAO in  $\beta$ -GP and CA electrolyte with different addition levels of AgNO<sub>3</sub> (0.1 g·L<sup>-1</sup> and 0.4 g·L<sup>-1</sup>). Unlike the general surface characteristics of MAO coatings, micro-pores were hardly identified owing to the existence of outer granular HA on MAO-0 (Figure 6a), MAO-0.1 (Figure 6b) or needle-like HA layers on MAO-0.4 (Figure 6c). The addition of AgNO<sub>3</sub> can speed up the MAO process at higher current density levels, enhance the crystallinity of the HA layer and precipitation of Ag NPs on the HA layer. Compared to amorphous HA, the crystalline HA exhibits better bioactivity in SBF (simulated body fluid) and biointegration [86]. Ag NPs with a size of 20–30 nm were deposited on the HA layer of MAO-0.1 (Figure 6e) and MAO-0.4 (Figure 6f). The rapid release of Ag<sup>+</sup> ions at the initial periods and the subsequent slower release contributed to long-term antibacterial activity occurred. Moreover, cumulative release of Ag<sup>+</sup> ions (2.5 ppm for MAO-0.1 and 8 ppm for MAO-0.4) was lower than the toxic concentration (10 ppm) of Ag for human cells [87]. Overall consideration,  $0.1 \text{ g} \cdot \text{L}^{-1}$  AgNO<sub>3</sub> appeared to exhibit sufficient antibacterial activity and no risk of cytotoxicity, while preserving the structural characteristics. Muhaffel et al. [60] have further reported a multi-layer coating consisting of subnatant TiO<sub>2</sub> layers and upper bio-mimetic precipitation in disodium hydrogen phosphate (Na<sub>2</sub>HPO<sub>4</sub>), CA and 0.0025 M CH<sub>3</sub>COOAg electrolyte at 380 V. Incorporation of 4.6 wt % Ag into the multi-layer led to superior antibacterial efficiency against Escherichia. coli (E. coli) and S. aureus while conserving the biomimetic apatite precipitation. The multilayer consisted of three layers: A thin compact TiO<sub>2</sub> layer above the substrate showing better substrate bonding, a middle porous and thick TiO<sub>2</sub> layer on the compact layer and a flake-like top layer of biocompatible compound incorporated with regional Ag particles (Figure 7).

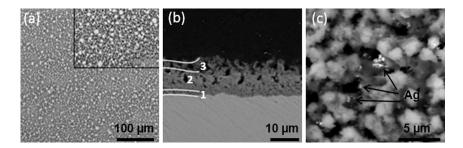
Unlike the common surface micrometer-sized pores of MAO coatings, highly ordered nanopores (Figure 8b,c) within micropits (Figure 8a) were successfully prepared by Chang et al. [61] via a novel one-step high-current anodization (HCA) at the current density of 65 A·dm<sup>-2</sup> in the AgNO<sub>3</sub> electrolyte range from 0.5–16 g·L<sup>-1</sup>. The nanopore diameter decreased and the wall thickness increased with increasing of AgNO<sub>3</sub> concentrations (Figure 8d). Due to the controllable dimensions, the nanoporous structure modulated osteoblast functions, accommodating various clinical needs [88–90]. Scratch tests showed significantly enhanced bonding strength on the HCA treated sample (Figure 8e) in comparison with flaking on the anodized sample (Figure 8f), even at a small normal load. The TEM and XRD showed that Ag was embedded in crystallized TiO<sub>2</sub> in the form of crystallized AgO NPs with

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diameters between 10 nm and 30 nm. The crystallized  $TiO_2$  also contributed to good bioactivity [49]. Gao et al. [91] reported that several ppb of Ag released quantity were capable of killing all the bacteria. Therefore, the high release level of Ag (>200 ppb) revealed strong antimicrobial properties, but resulted in severe cytotoxicity. To improve the cytocompatibility and reduce toxicity,  $Na_3PO_4$  and  $Ca(NO_3)_2$  were added into the electrolyte to inhibit the formation of nanopores to a certain extent and to lower the release of  $Ag^+$  while preserving their antibacterial ability.



**Figure 6.** Low and high magnification SEM micrographs of MAO-0 (**a**,**d**); MAO-0.1 (**b**,**e**) and MAO-0.4 (**c**,**f**) [59]. (Reprinted with permission from [59], Copyright 2016 Elsevier.)



**Figure 7.** Low magnification surface (a) and cross-sectional SEM (b) micrographs for the MAO-Ag samples obtained in electrolyte solution containing 0.0025 M CH<sub>3</sub>COOAg (1: compact TiO<sub>2</sub> layer; 2: porous TiO<sub>2</sub> layer; 3: biocompatible compound containing layer); (c) high magnification surface SEM micrographs indicated that regional Ag particles covered the surface [60]. (Reprinted with permission from [60], Copyright 2015 Elsevier.)

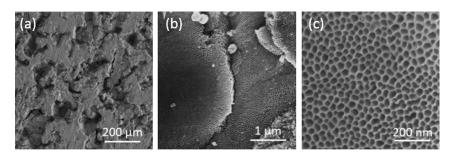
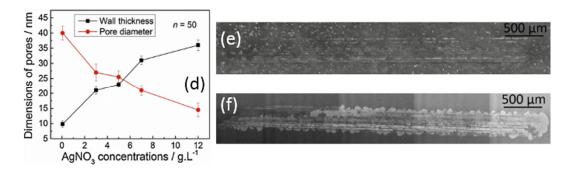
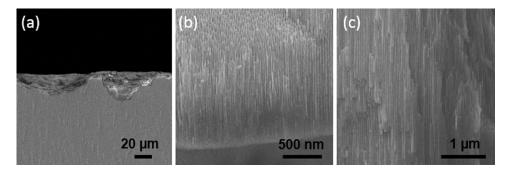


Figure 8. Cont.



**Figure 8.** Low-magnification (**a**) and high-magnification (**b**,**c**) SEM images of the high-current anodization (HCA)-treated sample in AgNO<sub>3</sub> electrolyte; (**d**) variation of the pore diameters and thickness of the HCA-treated samples as a function of AgNO<sub>3</sub>. Scratch SEM images of the sample anodized in the electrolyte with the AgNO<sub>3</sub> concentration of 2 g·L<sup>-1</sup> (**e**) and the sample anodized in ethylene glycol supplemented with 0.5 wt % NH<sub>4</sub>F, 5.0 vol % H<sub>2</sub>O and 5.0 vol % CH<sub>3</sub>OH at an applied voltage of 30 V for 1.5 h at room temperature (**f**) showed the significantly enhanced bonding strength on the HCA-treated sample [61]. (Reprinted with permission from [61], Copyright 2015 Elsevier.)

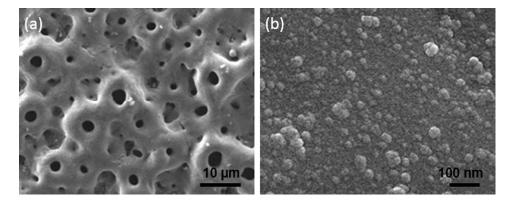
Likewise, a unique structure with a micro-/nano-morphology can also be obtained in  $\text{Cu}(\text{NO}_3)_2$  electrolyte due to field-assisted chemical etching of  $\text{NO}_3^-$ . Huang et al. [62] believed that the use of  $\text{Cu}(\text{NO}_3)_2$  instead of  $\text{Ag}\text{NO}_3$  as the electrolyte may avoid the potential cytotoxicity of heavy metal ions.  $\text{Ag}^+$  was easily oxidized to higher valence  $(\text{Ag}^{2+})$ , which will lead to severe cytotoxicity [61]. However, bivalent  $\text{Cu}^{2+}$  in  $\text{Cu}(\text{NO}_3)_2$  cannot be further oxidized. The results of EDX and XPS confirmed that toxic elements Cu were not incorporated into the coating surface, so copper will not directly contact the cells to produce cell toxicity. Cross-sectional images (Figure 9) also indicated that highly-ordered nano-channels were perpendicular to the substrate, and a compact layer can be observed between the nanopore arrays and the substrate. Besides, the addition of  $\text{PO}_4^{3-}$  in the electrolyte further improved the cellular performance [92,93]. Moreover, A well-defined micro-/nano-morphology can also be acquired by this HCA technique in electrolytes, such as  $\text{Cu}(\text{NO}_3)_2$ ,  $\text{Zn}(\text{NO}_3)_2$ ,  $\text{(Na}_3\text{PO}_4 + \text{Ca}(\text{NO}_3)_2 + \text{Na}_2\text{SiO}_3$ ) and  $\text{(Na}_3\text{PO}_4 + \text{Ag}\text{NO}_3 + \text{Na}_2\text{SiO}_3$ ), which will extend its application.



**Figure 9.** (a) Low and (b,c) high magnification cross-sectional SEM images of the high-current anodization (HCA)-treated sample in Cu(NO<sub>3</sub>)<sub>2</sub> electrolyte indicated the presence of highly-ordered nano-channels [62]. (Reprinted with permission from [62], Copyright 2016 Elsevier.)

As an essential trace element for living organisms, Cu participates in a variety of metabolic activities, so it might be more suitable for antibacterial application compared to Ag. Zhu et al. [63] prepared porous and nano-structured Cu-incorporated TiO<sub>2</sub> coatings by MAO in a novel Cu-containing electrolyte containing 0.05 M  $\beta$ -GP, 0.1 M CA and 0.05 M copper acetate (Cu(CH<sub>3</sub>COO)<sub>2</sub>) at 16.5 A·dm<sup>-2</sup> for 4 min. The result showed that Cu-TiO<sub>2</sub> coatings displayed rough micropores or crater structures with diameters of 3–5  $\mu$ m (Figure 10a), and the coatings were fully covered with nano-grains with a size of 30–50 nm (Figure 10b) at a higher magnification. The antimicrobial activity

of Cu-TiO<sub>2</sub> coatings was improved due to the inhibitory effect of Cu. Meanwhile, the osteoblastic adhesion, spreading, early proliferation and late differentiation on Cu-TiO<sub>2</sub> coatings were significantly enhanced due to the existence of the porous nano-structured surface produced during MAO, just like those found by other researchers [81,94]. Besides, the Cu element also plays a vital role in various cell behaviors, including osteoblasts [95], endothelial cells [96], bone marrow stem cells [63] and fibroblasts [64].

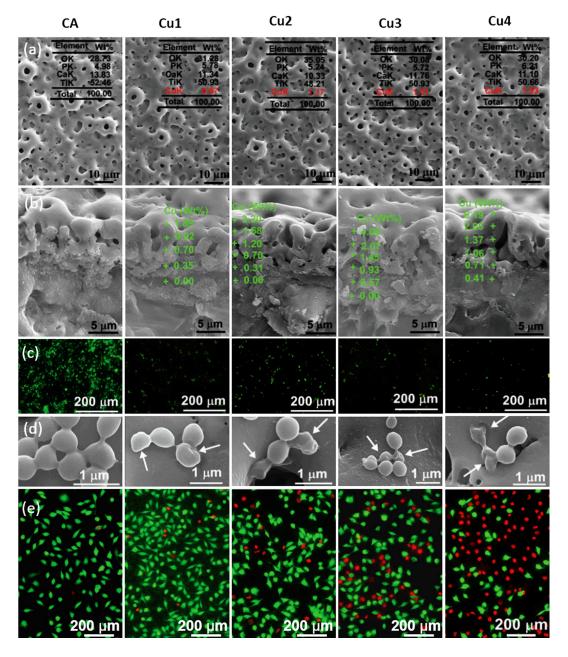


**Figure 10.** Surface morphologies of Cu-TiO<sub>2</sub> coatings at different magnifications showing micropores with a diameter of 3–5  $\mu$ m (a) and nano-grains with a size of 30–50 nm (b) on the surface of Cu-TiO<sub>2</sub> [63]. (Reprinted with permission from [63], Copyright 2013 Elsevier.)

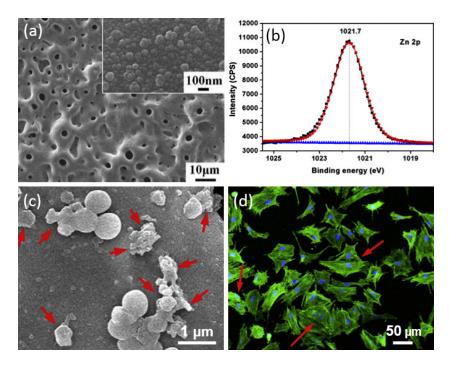
Zhang et al. [64] have investigated the tolerable and safe upper intake level of Cu<sup>2+</sup> in order to improve the response of fibroblasts while maintaining the antibacterial activity. TiO<sub>2</sub> coatings doped with different amounts of Cu<sup>2+</sup> were directly fabricated via MAO in copper acetate 0.02 M β-GP, 0.2 M CA and various amounts of Cu(CH<sub>3</sub>COO)<sub>2</sub> (0.00125 M, 0.0025 M, 0.00375 M and 0.005 M). As shown in Figure 11a, the coatings still maintained the typical porous structure of MAO, and Cu content on the surface of the sample increased with the increase of the concentration of Cu(CH<sub>3</sub>COO)<sub>2</sub> in electrolyte. Cu content also increased gradually from the substrate to the coating surface, and the apparent continuity between the coatings and Ti substrates indicated the strong bonding forces (Figure 11b). As the dose of Cu<sup>2+</sup> was increased, the number of adhering bacteria (Figure 11c) was obviously reduced due to the contact-killing and release-killing mechanism, and the bacterial morphologies and membrane integrities were subject to serious disruption (Figure 11d). Results of biological compatibility experiments showed that the appropriate dose of Cu (0.67 wt %, Cu1) can improve the adhesion and proliferation of fibroblasts and significantly support denser collagen deposition, while excess Cu<sup>2+</sup> was poisonous to the cells (Figure 11e), will disrupt fundamental cellular processes and trigger apoptosis, just as mesenchymal stem cells [97]. Cu1 prepared in 0.00125 M Cu(CH<sub>3</sub>COO)<sub>2</sub> electrolyte with a Cu mass fraction of 0.67 delivered the best compromise between antibacterial effectiveness and cytotoxicity.

As one of the most important trace elements in the human body, Zn also has been recognized recently [98], although the antibacterial activity of Zn is relatively weaker than that of Ag and Cu. For example, both Zhang et al. [65] and Hu et al. [10] used zinc acetate (ZA) solution to modify a titanium surface by MAO. Zhang et al. [65] prepared Zn-incorporated TiO<sub>2</sub> coatings in 0.06 M ZA, 0.02 M CA and 0.15 M Ca-GP solution at a current density of 30 A·dm<sup>-2</sup> for 5 min. Hu et al. [10] obtained Zn-incorporated TiO<sub>2</sub> coatings with different Zn contents (Z0, Z2, Z4, Z6) in mixture solutions of different concentrations (0 M, 0.02 M, 0.04 M, 0.06 M) of ZA, 0.1 M CA and 0.05 M  $\beta$ -GP at 16.5 A·dm<sup>-2</sup> for 4 min. It has been found that the incorporation of Zn in TiO<sub>2</sub> did not apparently change the surface topography; all of the TiO<sub>2</sub> coatings were still of a typical porous surface morphology, and the micropores with a pore size less than 5  $\mu$ m were uniformly distributed on the coating surface (Figure 12a). The coatings were primarily composed of the anatase and rutile phase. Increasing the

content of ZA in the electrolyte led to a slight increase in the Zn content of the oxide layer. However, the feature peaks of Zn-containing compounds were not detected due to the low concentration of Zn (4.6–9.3 wt %). The XPS showed that Zn existed in the form of ZnO (Figure 12b). The antibacterial mechanism of ZnO was attributed to the generation of reactive oxygen species (ROS) [99] on the coating surface with higher Zn content (Figure 12c), rather than the low zinc ions release (<3.62 ppm) [100]. The accumulated released Zn ions not only facilitated the initial adhesion (Figure 12d), spreading and proliferation of bone marrow stem cells (bMSC) [98,101], but also upregulated the gene and protein expression of bMSC while producing no cytotoxicity.



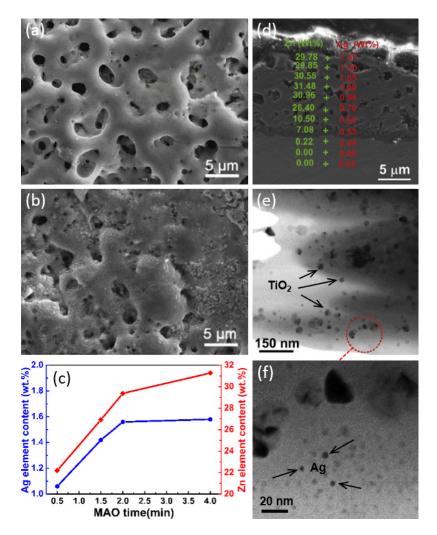
**Figure 11.** Surface (**a**) and cross-sectional (**b**) SEM morphologies images of the CA, Cu1, Cu2, Cu3 and Cu4 coatings showed that Cu content increases with the increasing concentration of Cu(CH<sub>3</sub>COO)<sub>2</sub> in electrolyte. Live-dead assay (**c**), the corresponding bacterial morphologies (**d**) of *S. aureus* after 24 h of incubation and cell viabilities (**e**) of the cells cultured for three days are highly dependent on the amount of Cu content incorporated [64]. (Reprinted with permission from [64], Copyright 2016 Royal Society of Chemistry.)



**Figure 12.** Surface SEM morphologies of Z6 coatings samples obtained in electrolyte solution containing 0.06 M ZA (zinc acetate) (a); high-resolution XPS spectra of Zn 2p in Z6 coating indicated that Zn mainly exists in the ZnO state (b); the obvious bacterial cell debris and completely lysed cells (indicated by red arrows) showed the improved antimicrobial property of Z6 coatings surfaces (c). The initial adhesion and spreading activity of bMSC cells after 24 h incubation on Z6 coatings were verified by the extraordinarily stretched cell morphology structure (d) and the presence of mitosis phase cells (indicated by red arrows) [10]. (Reprinted with permission from [10], Copyright 2012 Elsevier.)

# 2.3. Introduction of Both Metal Nanoparticles and Metallic Compound into MAO Electrolyte

Recently, it has found that Zn<sup>2+</sup> co-doped with small amount of Ag<sup>+</sup> may lead to an obvious increase of the antibacterial ability [102] in comparison with single Zn<sup>2+</sup>. Zhang et al. [66] fabricated micro-porous TiO<sub>2</sub> coatings co-doped with Zn<sup>2+</sup> and Ag NPs by MAO in the electrolyte composed of 0.1 M CA, 0.1 M ZA,  $0.02 \text{ M } \beta$ -GP,  $6 \text{ g} \cdot \text{L}^{-1}$  Ag NPs and  $0.25 \text{ g} \cdot \text{L}^{-1}$  sodium dodecyl benzene sulfonate for 0.5, 1.5, 2 and 4 min. All of the coatings were micro-porous with pore diameters of 1–4  $\mu m$ (Figure 13a), but some deposits were displayed on micro-pores at 4 min (Figure 13b). The contents of Ag and Zn on the coating surfaces increased with prolonged MAO time (Figure 13c). Continuity between coating and substrate indicated strong bonding force by MAO, and the Ag and Zn contents increased gradually from the substrate to the coating surface (Figure 13d). Ag was composed of 73% Ag<sub>2</sub>O and 27% metallic Ag [103,104], while Zn existed in the form of ZnO [105]. TEM images indicated that many large rutile or anatase grains of 20-40 nm in size (Figure 13e,f) and many relatively small Ag nano-grains with sizes of 5–10 nm (Figure 13f) were homogenously embedded in amorphous TiO<sub>2</sub> matrix. However, the diffraction spots of ZnO with trace amounts were not detected. The cumulative ion releases of Zn<sup>2+</sup> (6.88 ppm) [10] and Ag<sup>+</sup> (0.684 ppm) played an important role in inhibiting the growth of bacteria. Besides, the secondary products such as reactive oxygen species produced by Zn<sup>2+</sup> and Ag<sup>+</sup> were also harmful to bacteria. Carvalho et al. [106] and Jin et al. [28] have also proven the enhanced antibacterial activities of simultaneous incorporation of Zn and Ag into Ti. Jin et al. [28] believed that the interactions between Zn ions and bacteria were long-range based on the release of Zn ions [27], while the interactions between embedded Ag NPs and bacteria were short-range via the micro-galvanic effects without the release of Ag ions [37,107]. Moreover, an appropriate amount of released Zn<sup>2+</sup> also accelerated the osteogenic activity of rat bone mesenchymal stem cells (rBMSCs) via the long-range interactions.



**Figure 13.** MAO treated coatings at 2 min (**a**), and 4 min (**b**) showed different surface morphologies in electrolyte solution containing 0.1 M ZA (zinc acetate), 6 g·L<sup>-1</sup> Ag NPs. Cross-sectional morphology of the 2 min coating showed that the contents of Ag and Zn on the coating surfaces increased with prolonged MAO time (**c**), and increased gradually from the substrate to the coating surface (**d**); low (**e**), and high (**f**) magnification TEM images showed the TiO<sub>2</sub> grains of 20–40 nm in size and nano-grains with a size of 5–10 nm [66]. (Reprinted with permission from [66], Copyright 2016 Elsevier.)

# 2.4. MAO Assisted by MS

MS magnetron sputtering (MS) is a deposition technique belonging to the sputtering processes and is one of the physical vapor deposition (PVD) technologies. The MS deposition process is conducted in an inert gas (usually high-purity argon (Ar) gas) atmosphere regulated by the flow meter in a vacuum chamber. The specimens were placed in the sputtering chamber, and high-purity Ar (99.999%) is introduced when the chamber sputtering is evacuated to a base pressure below  $6 \times 10^{-3}$  Pa. Films are deposited at various sputtering parameters, such as gas flow, working pressure, sputtering power and sputtering time, in order to acquire the desired properties of the material. For the sputtering process, there exists an electric field between the target material and the substrates. Under the action of the electric field, Ar is ionized and bombards the target material located in the cathode of the discharge, which often causes the ejection of ions or atoms from the surface of the target material toward that of the substrates. However, the lower ionization and bombardment rates of Ar often lead to the slower deposition rates and lower film densities. Given this, MS technology is introduced by installing a set of magnets beneath the target. The schematic of the MS deposition system has been described

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in Figure 14. The magnets are used to provide a magnetic field to capture the secondary electrons emitted by the target. Under the action of the magnetic field, secondary electrons are bound in the high density plasma region near the target surface, perform a circular motion around the target surface and constantly impact ionized large amounts of argon ion in the process of movement, thus improving the intensity of Ar ion bombardment and the deposition rate of the films.

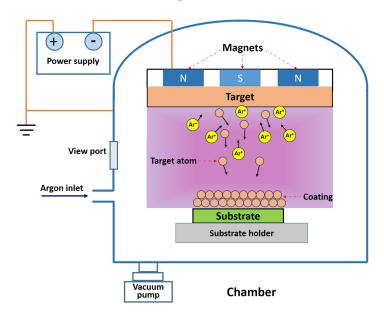
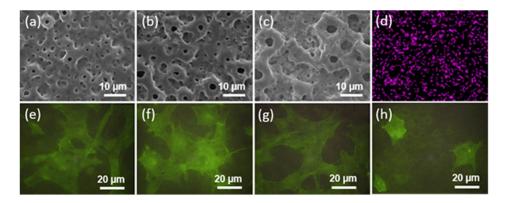


Figure 14. The schematic of the magnetron sputtering (MS) system.

MS is also an effective technique to incorporate antibacterial agents into the implant materials. The desired antibacterial ability can be obtained only by the choice of the optimal processing parameters. Bai et al. [108] have fabricated Ti-Ag composite coatings with different Ag contents (1.2–21.6 at %) using co-sputtering Ti-Ag targets by MS. The composite coating showed long-term antibacterial ability. However, only the Ag on the surface of Ti-Ag composite coatings contributed to the antibacterial activity [109]. It is hence necessary to carry out a post-treatment to fully use the Ag on Ti-Ag composite coatings while the antibacterial activity is preserved.

By MS, the deposited AgTi layers cover the entire surface and are firmly combined with the substrate. The concentration of Ag in the layers can be accurately controlled only by adjusting the technological parameters. Though Ag will be dissolved in the electrolyte during the MAO process, a great quantity of Ag is still reserved in the MAO coating. MAO coating shows a dense inner layer and a rough porous outer structure. The dense inner layer can improve the mechanical interlocking ability and prevent the corrosion from body fluid, while the porous outer structure may increase the surface roughness, thus promoting cell reaction. The desirable antibacterial ability and compatibility can achieve a balance by this duplex technique of combining MS and MAO.

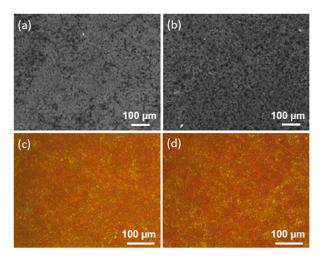
Zhang et al. [110] have developed the novel duplex-treatment technique combining MS with MAO to prepare the Ag-containing bioactive antibacterial coatings. AgTi layers were first deposited onto Ti by MS, then subjected to MAO at  $20~\text{A}\cdot\text{dm}^{-2}$  for 2, 5 and 8 min in the electrolyte containing 26.4 g·L $^{-1}$  CA and 4.3 g·L $^{-1}$   $\beta$ -GP. The presence of Ag did not exert a significant influence on the voltage and surface morphology. Micron-sized pores were still distributed over the surfaces, and pore sizes tend to increase (Figure 15a–c) due to the generated larger breakdown channels and more intense sparking with the increase of oxidation time. The coatings were comprised of rutile, anatase and a little Ag. However, Ag was uniformly distributed throughout the coating surface (Figure 15d), but in the form of Ag NPs [56]. During MAO, a large amount of Ag was dissolved in the electrolyte, which caused a sharp decrease in the Ag content of the MAO coatings. Ag, existing as the metallic Ag $^0$  state, also showed excellent antibacterial ability.



**Figure 15.** SEM surface micrographs of M-AgTi coating oxidation for 2 min (a), 5 min (b) and 8 min (c) indicated that the pore size increase with the increasing oxidation time. EDX maps showed that Ag were uniformly distributed throughout the coating surface (d). Fluorescence microscopy images of F-actin cytoskeleton of pre-osteoblast cells cultured for 48 h on M0 (e), M1 (f), M2 (g) and M3 (h) coatings indicated that a high concentration of Ag exhibited cytotoxic effect [110,111]. (Reprinted with permission from [110,111]; Copyright 2013 Elsevier, Copyright 2014 Elsevier.)

Furthermore, Zhang et al. [111] produced TiO<sub>2</sub> coatings with different Ag concentrations (M0, M1, M2, M3) by the above duplex-treatment technique. The concentration of Ag in the TiO<sub>2</sub> coatings increased with the increase of Ag incorporated in the AgTi layers. The initially high level and subsequently low level of Ag release provided relatively long-term antibacterial activities and met the clinical requirements. Nonetheless, M2 and M3 with 1.36 and 1.93 wt % (Figure 15g,h) Ag exhibited cytotoxic effect, which restricted cell proliferation and caused cell death. M0 with 0.95 wt % Ag (Figure 15f) achieved a balance between antibacterial properties and good pre-osteoblast cell viability.

On this basis, He et al. [112] fabricated porous Ag/Sr-containing  $TiO_2$  coatings via a similar method in the MAO electrolyte containing 0.167 M CA, 0.02 M  $\beta$ -GP and 0.033 M strontium acetate hemihydrate (SA). The coating displayed long-lasting antibacterial ability even up to 28 days due to the incorporation of Ag. An addition of the bioactive element Sr alleviated the potential cytotoxic effect of the excessive Ag and further facilitated the proliferation and differentiation of MC3T3-E1 osteoblasts (Figure 16).



**Figure 16.** SEM images of MC3T3-E1 cells after culturing for 3 days and quantitative results of collagen secreted by MC3T3-E1 cells after culturing for 7 days on M-Ag0.40 (**a**,**c**) and M-Sr/Ag0.40 (**b**,**d**) indicated that the bioactive element Sr is able to facilitate cell proliferation and differentiation of MC3T3-E1 osteoblasts and alleviate the potential cytotoxic effect of excessive Ag [112]. (Reprinted with permission from [112], Copyright 2016 IOP.)

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#### 3. Conclusions and Future Outlook

Implant-associated infections are still one of the critical issues for dental and orthopedic implantology. The surface modification of Ti and Ti alloy seems to be a challenging strategy to prevent the development of peri-implant infection. In the present paper, the MAO technique effectively and versatilely introduces inorganic antibacterial metal elements (e.g., Ag, Cu, Zn) into biomedical implants, which successfully solves the growing problem of peri-implant infection. In this mini-review, the research advance in the fabrication of anti-microbial surfaces by MAO is summarized. Antibacterial metal elements were introduced into the electrolyte in the form of either metal NPs or metallic compounds.

However, the main open problems related to this strategy are that the antibacterial effect increases in a dose-dependent manner, but coupled to cytotoxic behavior at a higher concentration. In fact, many Ti-based implants with strong antibacterial properties are still not available in the field of medicine, probably due to the problem mentioned above. In this review, we can also see that many studies have only focused on the development of Ti-based surfaces with an effective antibacterial effect, but ignored the study of bioactivity and toxicity. Therefore, it is necessary to precisely control the content of the antibacterial metal elements and MAO parameters towards an optimal trade-off between antibacterial properties and cytotoxicity. It is a potential way to achieve an enhancement of both the antibacterial property and bioactivity directly by the simultaneous addition of antibacterial metal elements and bioactive elements, such as strontium (Sr) and silicon (Si) in the MAO electrolyte.

In addition, the MAO method combined with MS was introduced to obtain the desired antibacterial property and fabricate bioactive coatings in this review. Ca-P ions can be formed in the porous bio-ceramic layer on the Ti surface by MAO and further transformed into crystal HA through other treatment, such as heat treatment or electrophoretic deposition. The good crystallinity of HA is conducive to cell reaction and bioactivity. Therefore, it is still a trend to develop new methods that can promote the formation of high crystallinity HA.

In conclusion, in order to overcome the toxicity concerns and guarantee the prolonging antibacterial properties, it seems to be a future research trend in the biomaterial field to add antibacterial metal elements and bioactive elements by MAO directly or assisted by other surface modification techniques, such as heat treatment, hydrothermal treatment and plasma spraying, in the near future.

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