

Supplementary Materials

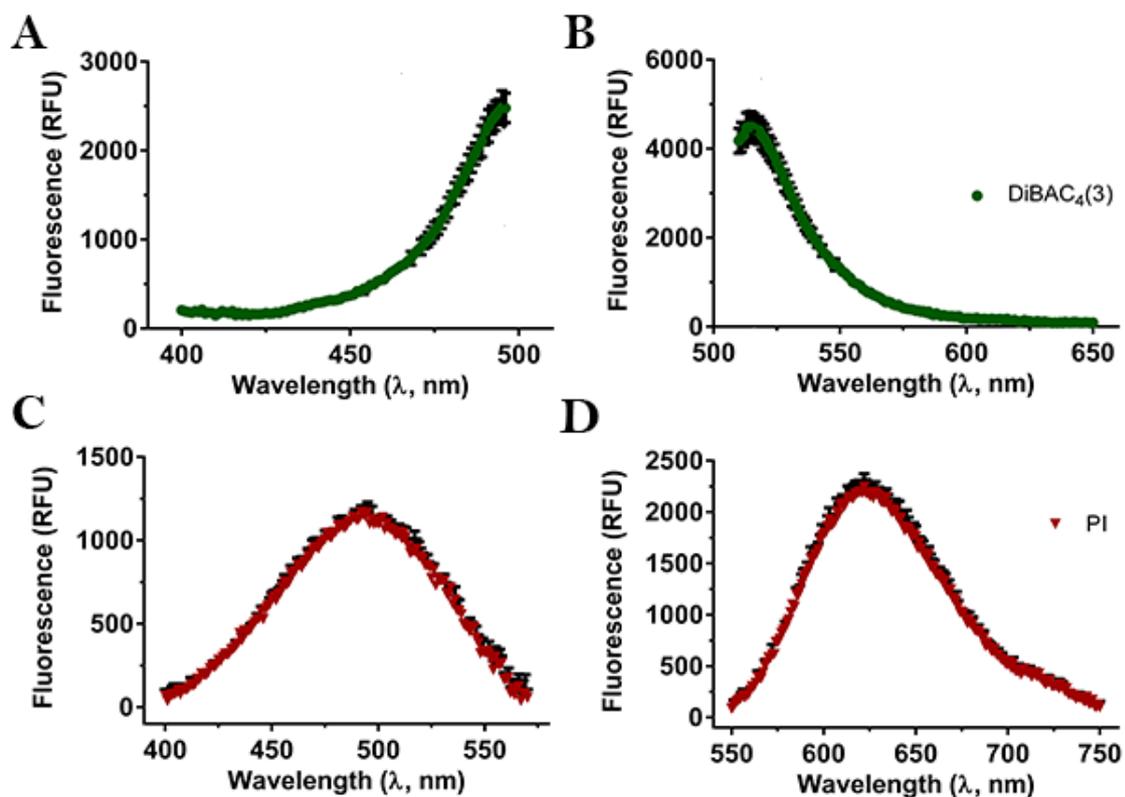


Figure S1. Separate fluorescence Excitation (A, C) and Emission (B, D) spectra of bis-(1,3-dibutylbarbituric acid)trimethine oxonol (DiBAC₄(3)) (A–B) and propidium iodide (PI) (C–D) in phosphate-buffered saline supplemented with 25 mM glucose (PBS-glc). (A, C) Excitation spectra of 500 nM DiBAC₄(3) with 516 nm (A) and 20 μ g/mL PI with 617 nm (C) detection wavelengths. (B, D) Emission spectra of 500 nM DiBAC₄(3) with 496 nm (B) and 20 μ g/mL PI with 490 nm (D) excitation wavelengths. For technical reasons due to instrument settings, spectra could not be measured at wavelengths > 496 nm (A) and < 510 nm (B).

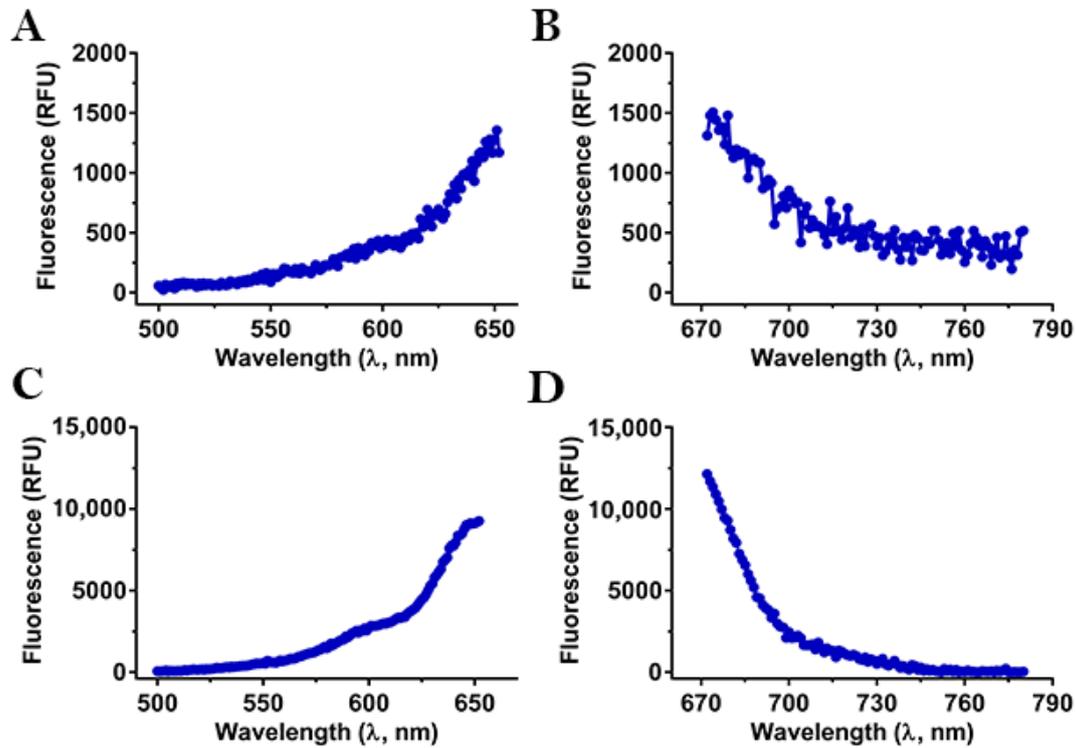


Figure S2. Fluorescence Excitation and Emission spectra of 400 nM 3,3'-dipropylthiadicarbocyanine Iodide (diSC₅) in black wellplates (A–B) and low-binding OptiPlate (C–D). (A, C) Excitation spectra with 672 nm detection wavelength, and (B, D) emission spectra with 652 nm excitation wavelength (error bars were omitted for clarity). For technical reasons due to instrument settings, spectra could not be measured at wavelengths > 650 nm (A, C) and < 670 nm (B, D).

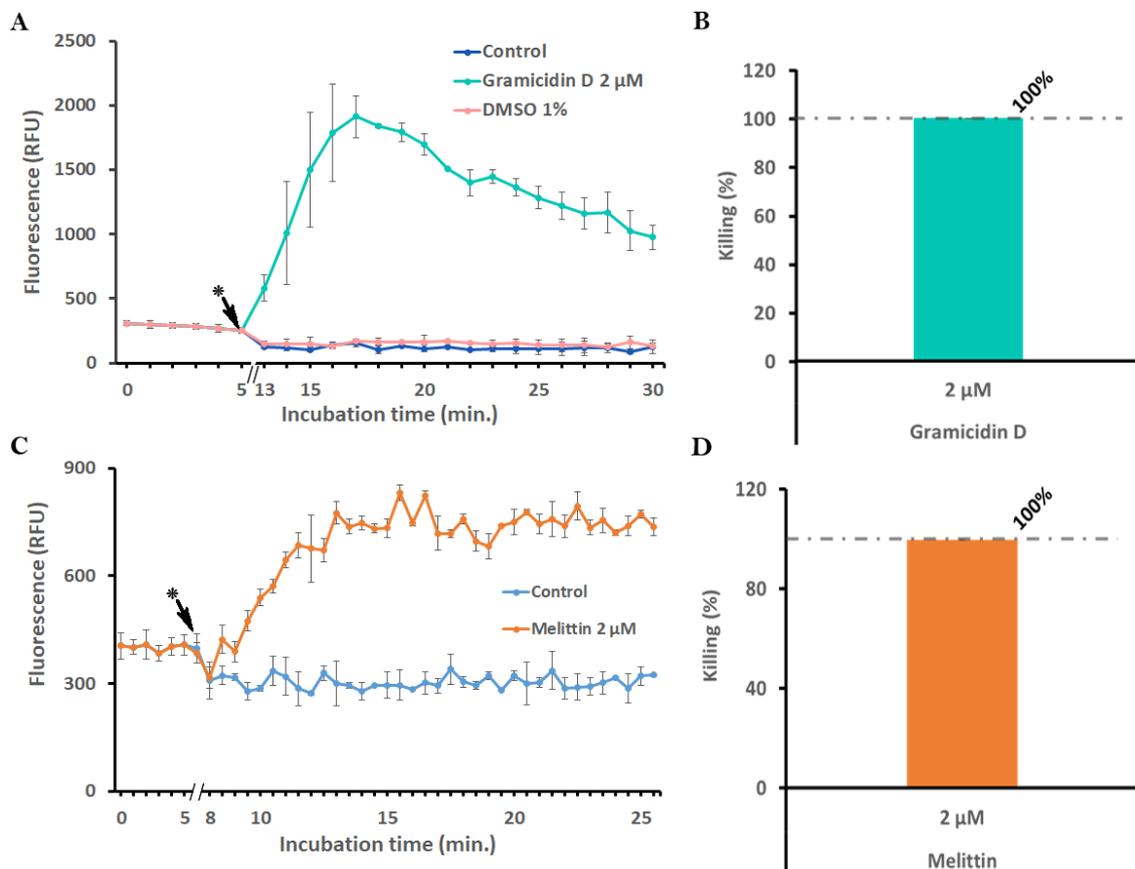


Figure S3. Time-course of fluorescence and killing percentage for Gramicidin D and Melittin. (A, C) Time-course of fluorescence (RFU) vs. incubation time (min.) for Gramicidin D (A) and Melittin (C). (B, D) Killing percentage (%) vs. concentration (μM) for Gramicidin D (B) and Melittin (D). Error bars represent standard deviation.

Figure S3. Membrane depolarization (diSC₃(5)) (A) and Killing (B) of *S. epidermidis* ATCC 35984 caused by gramicidin D and membrane permeabilization (PI) (C) and Killing (D) of *E.coli* ATCC 25922 caused by melittin. Experiments were performed with 10⁷ CFU/mL of the indicated strains in PBS-glc with 400 nM diSC₃(5) ($\lambda_{\text{ex}} = 652 \text{ nm}$, $\lambda_{\text{em}} = 672 \text{ nm}$) and 5 $\mu\text{g/mL}$ PI ($\lambda_{\text{ex}} = 535 \text{ nm}$, $\lambda_{\text{em}} = 617 \text{ nm}$) at 37°C. CFU counts were determined at 30 min incubation. (*) peptide addition.