

Supplementary Table S1: Quality data of the contigs based on the QUAST (v5.2.) software analysis.

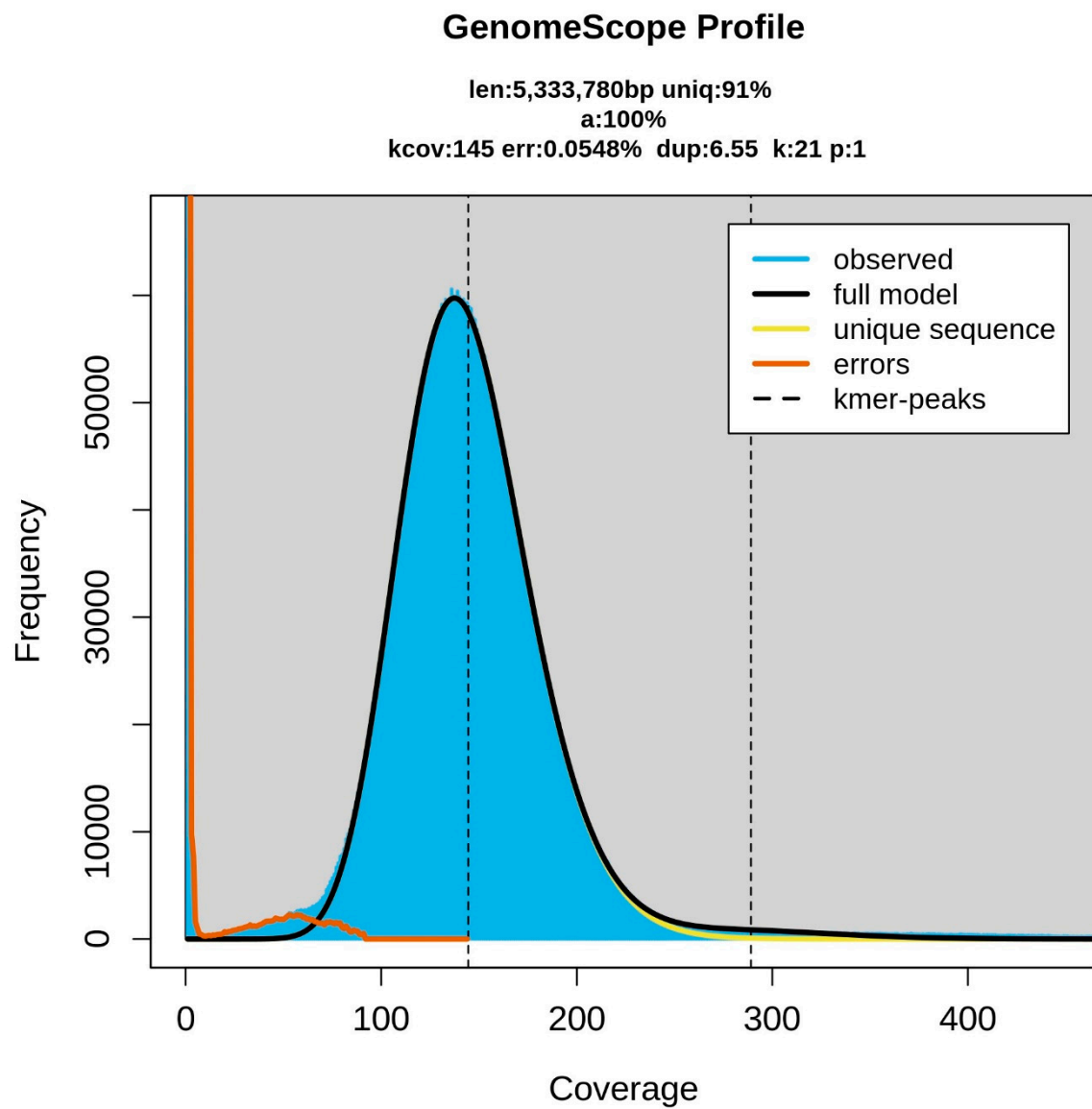
Strain	No of trimmed reads	No of contigs	Coverage	N50	N75	L50	L75
0× AMX	4956318	109	180.18	231989	121408	6	14
1× AMX	6628030	111	259.102	205034	121530	7	14
10× AMX	4791998	126	168.046	194885	107365	9	18
100× AMX	5440885	193	134.166	160547	84576	12	24
1000× AMX	4221108	102	161.305	202436	130873	8	16
0× CTX	2277416	249	55.8348	120 956	65 648	14	27
1× CTX	2421900	253	59.5407	132 252	57 752	14	27
10× CTX	2444649	237	59.7927	155 433	84 590	12	24
100× CTX	2315672	260	56.5416	120 953	50 443	15	29
1000× CTX	1413575	327	34.4063	73 430	29 355	21	48

AMX-amoxicillin, CTX-cefotaxime

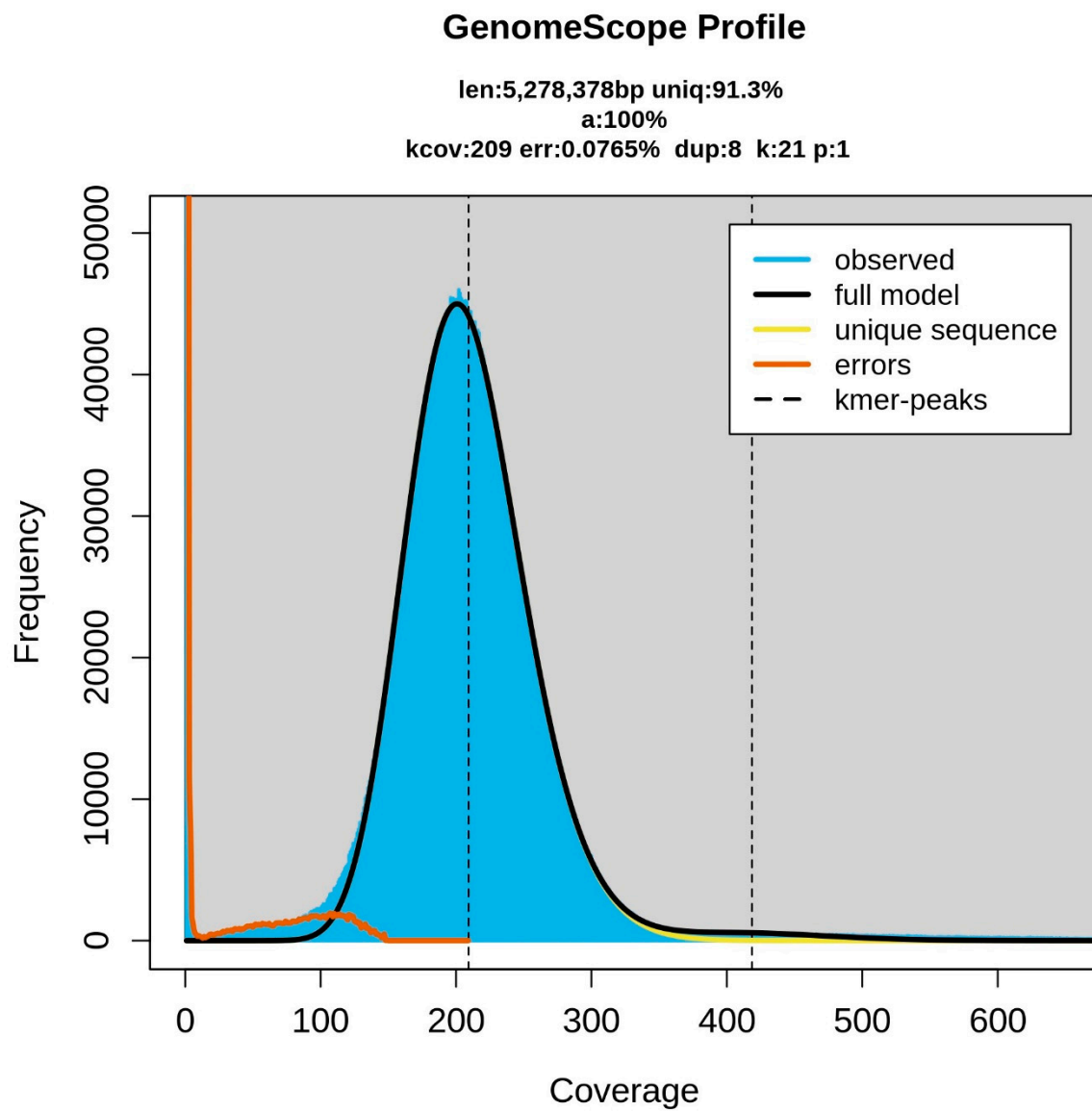
Supplementary Table S2: The set of 44 ARGs identified in the next-generation sequencing was the same for all samples.

Gene	Coverage %	Identity %	Mechanism	Resistance
<i>acrA</i>	100.00	99.16	antibiotic efflux	cephalosporin, fluoroquinolone, glycylicycline, penam, phenicol, rifamycin, tetracycline and triclosan
<i>acrB</i>	100.00	98.64	antibiotic efflux	
<i>acrD</i>	100.00	98.04	antibiotic efflux	
<i>acrE</i>	100.00	98.79	antibiotic efflux	
<i>acrF</i>	100.00	96.49	antibiotic efflux	
<i>acrS</i>	100.00	98.34	antibiotic efflux	cephalosporin, cephamycin, fluoroquinolone, glycylicycline, penam, phenicol, rifamycin, tetracycline and triclosan
<i>ampC</i>	100.00	98.15	antibiotic inactivation	cephalosporin and penam
<i>ampH</i>	100.00	97.50	antibiotic inactivation	
<i>bacA</i>	99.76	98.17	target alteration	peptide
<i>baeR</i>	99.86	96.81	antibiotic efflux	aminocoumarin andaminoglycoside
<i>baeS</i>	100.00	90.53	antibiotic efflux	
<i>cpxA</i>	100.00	98.47	antibiotic efflux	
<i>CRP</i>	100.00	99.21	antibiotic efflux	fluoroquinolone, macrolide and penam
<i>emrA</i>	100.00	98.21	antibiotic efflux	fluoroquinolone
<i>emrB</i>	100.00	96.95	antibiotic efflux	
<i>emrE</i>	100.00	92.19	antibiotic efflux	macrolide
<i>emrK</i>	100.00	97.73	antibiotic efflux	tetracycline
<i>emrR</i>	100.00	98.68	antibiotic efflux	fluoroquinolone
<i>emrY</i>	100.00	97.73	antibiotic efflux	tetracycline
<i>eptA</i>	100.00	91.85	target alteration	peptide
<i>evgA</i>	100.00	99.02	antibiotic efflux	fluoroquinolone, macrolide, penam and tetracycline
<i>evgS</i>	100.00	96.19	antibiotic efflux	
<i>gadW</i>	100.00	99.86	antibiotic efflux	fluoroquinolone, macrolide and penam
<i>gadX</i>	100.00	93.82	antibiotic efflux	

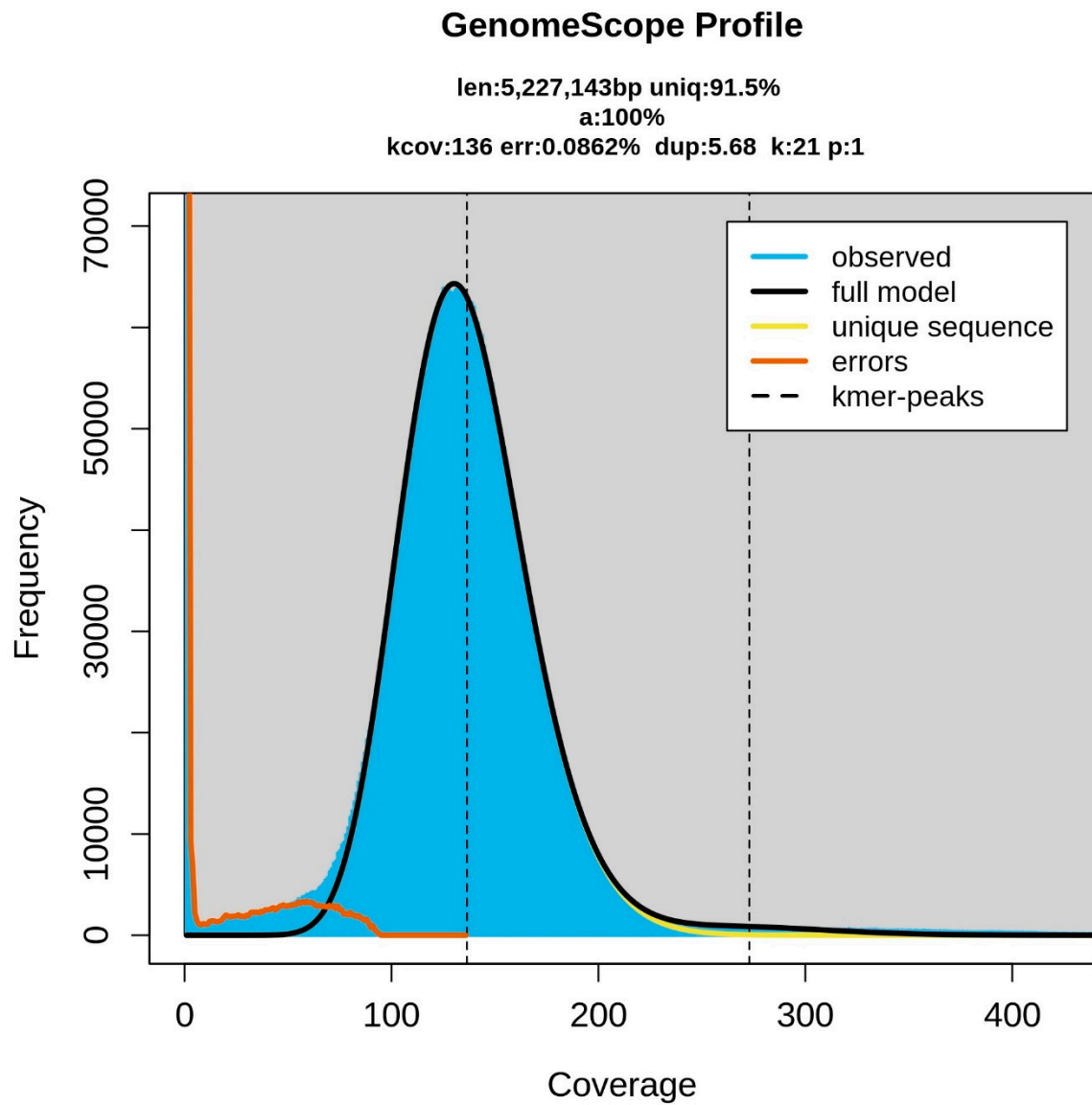
<i>H-NS</i>	100.00	99.28	antibiotic efflux	cephalosporin, cephamycin, fluoroquinolone, macrolide, penam and tetracycline
<i>kdpE</i>	99.26	95.84	antibiotic efflux	aminoglycoside
<i>marA</i>	100.00	98.70	reduced permeability	carbapenem, cephalosporin, cephamycin, fluoroquinolone, glycylcycline, monobactam, penam, penem, phenicol, rifamycin, tetracycline and triclosan
<i>mdfA</i>	100.00	96.59	antibiotic efflux	benzalkonium chloride, rhodamine and tetracycline
<i>mdtA</i>	100.00	95.11	antibiotic efflux	aminocoumarin
<i>mdtB</i>	100.00	96.29	antibiotic efflux	
<i>mdtC</i>	100.00	94.15	antibiotic efflux	
<i>mdtE</i>	100.00	98.62	antibiotic efflux	fluoroquinolone, macrolide and penam
<i>mdtF</i>	100.00	97.33	antibiotic efflux	
<i>mdtG</i>	100.00	98.21	antibiotic efflux	
<i>mdtH</i>	100.00	98.26	antibiotic efflux	fosfomycin
<i>mdtM</i>	100.00	95.05	antibiotic efflux	fluoroquinolone
<i>mdtN</i>	100.00	95.64	antibiotic efflux	acridine dye, fluoroquinolone, lincosamide, nucleoside and phenicol
<i>mdtO</i>	100.00	97.08	antibiotic efflux	acridine dye and nucleoside
<i>mdtP</i>	100.00	97.61	antibiotic efflux	
<i>msbA</i>	100.00	98.06	antibiotic efflux	
<i>pmrF</i>	100.00	97.63	target alteration	nitroimidazole
<i>tolC</i>	100.00	97.98	antibiotic efflux	peptide
<i>ugd</i>	100.00	96.92	target alteration	aminocoumarin, aminoglycoside, carbapenem, cephalosporin, cephamycin, fluoroquinolone, glycylcycline, macrolide, penam, penem, peptide, phenicol, rifamycin, tetracycline and triclosan
<i>yoiI</i>	100.00	98.05	antibiotic efflux	peptide



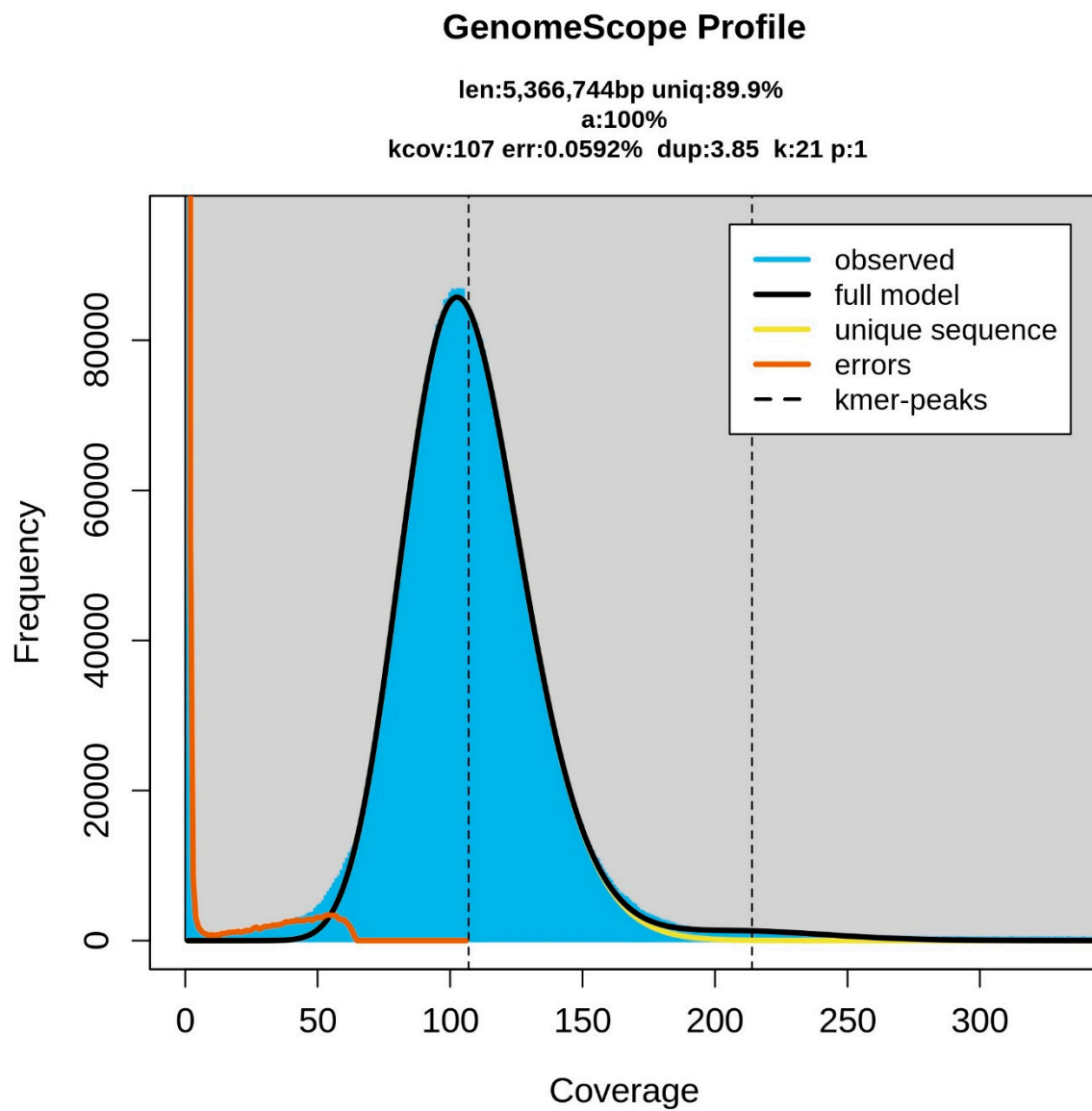
Supplementary Figure S1: GenoScope profile of 0× AMX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



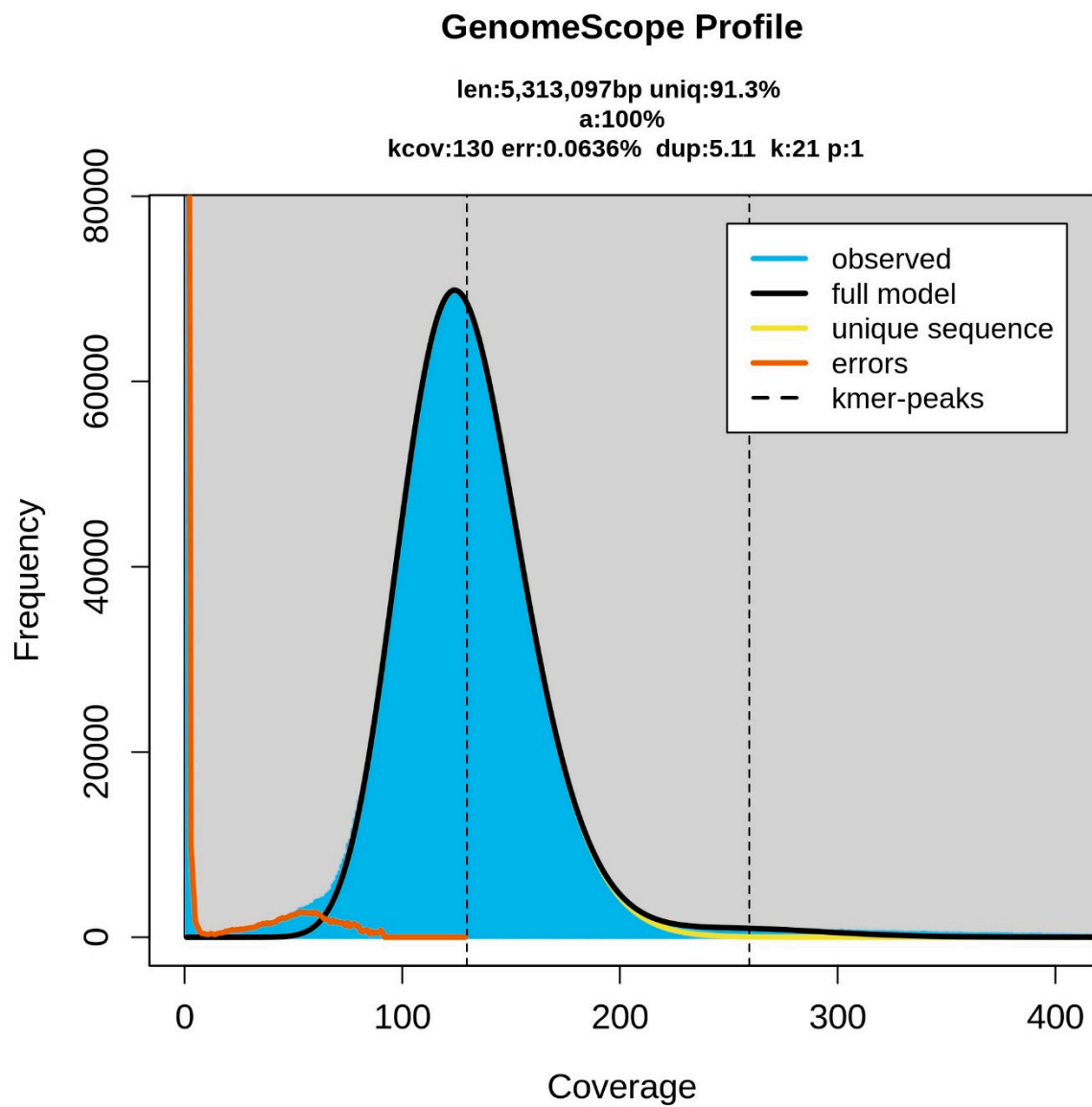
Supplementary Figure S2: GenoScope profile of 1× AMX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



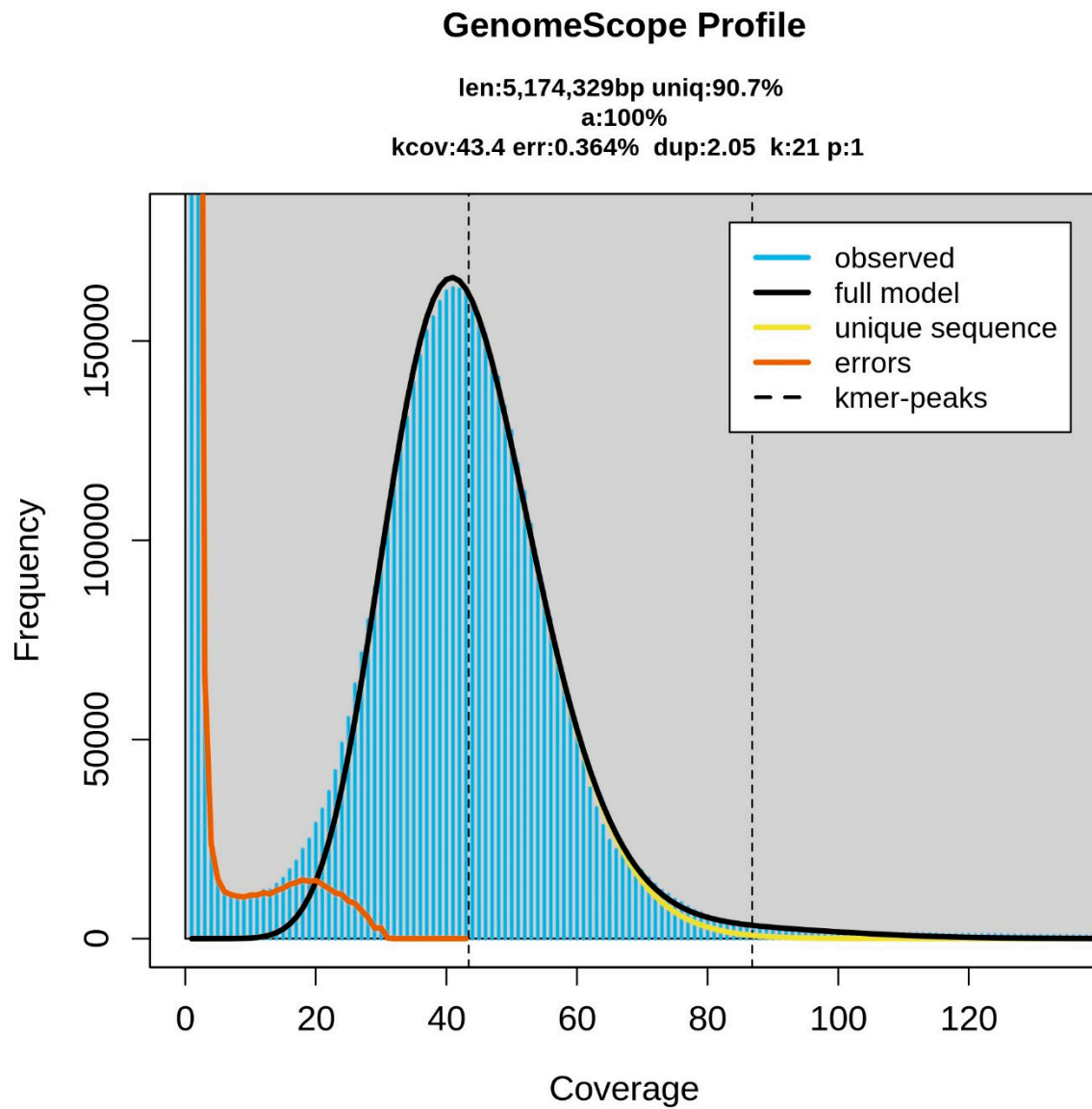
Supplementary Figure S3: GenoScope profile of 10× AMX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



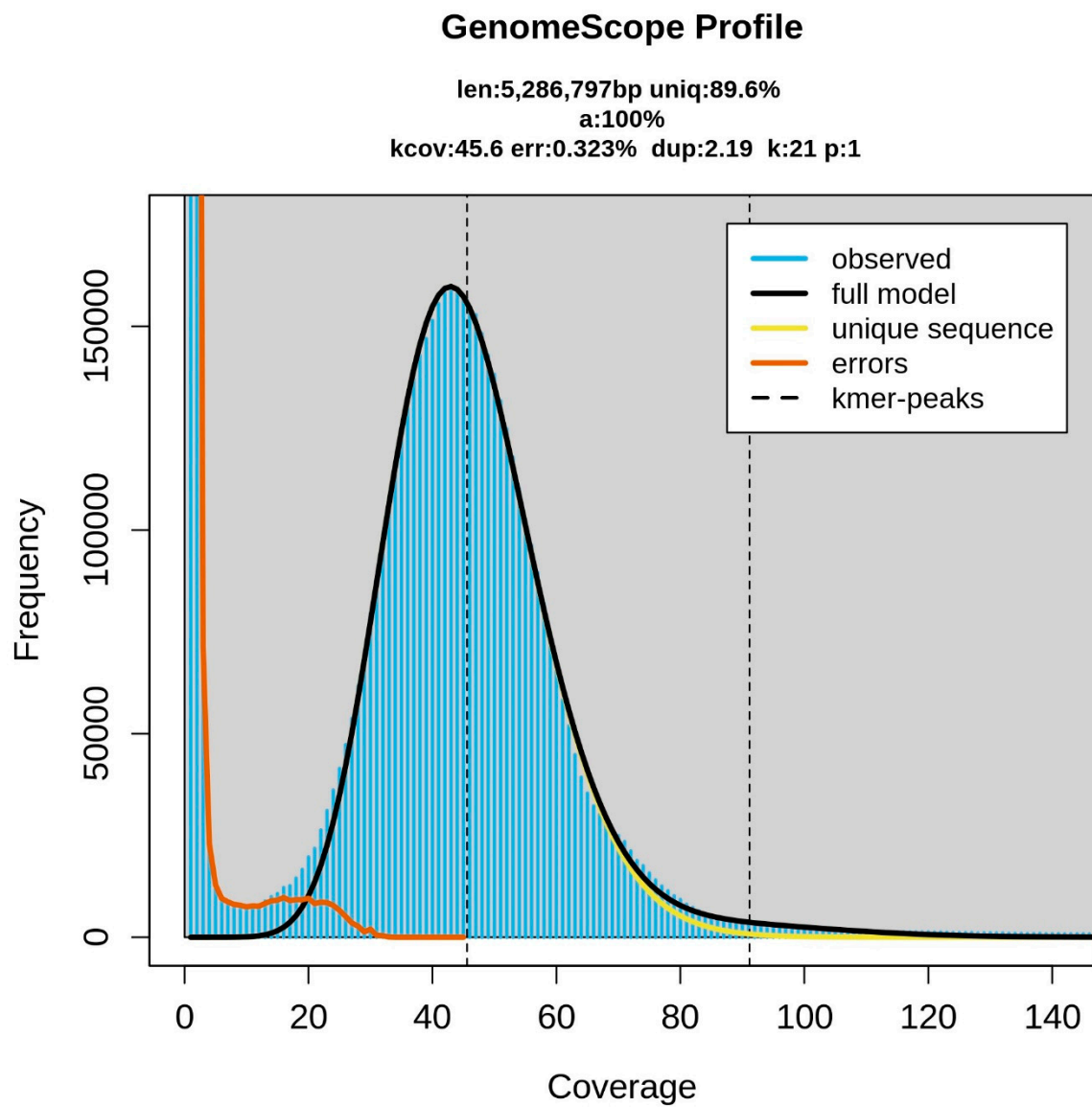
Supplementary Figure S4: GenoScope profile of 100× AMX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



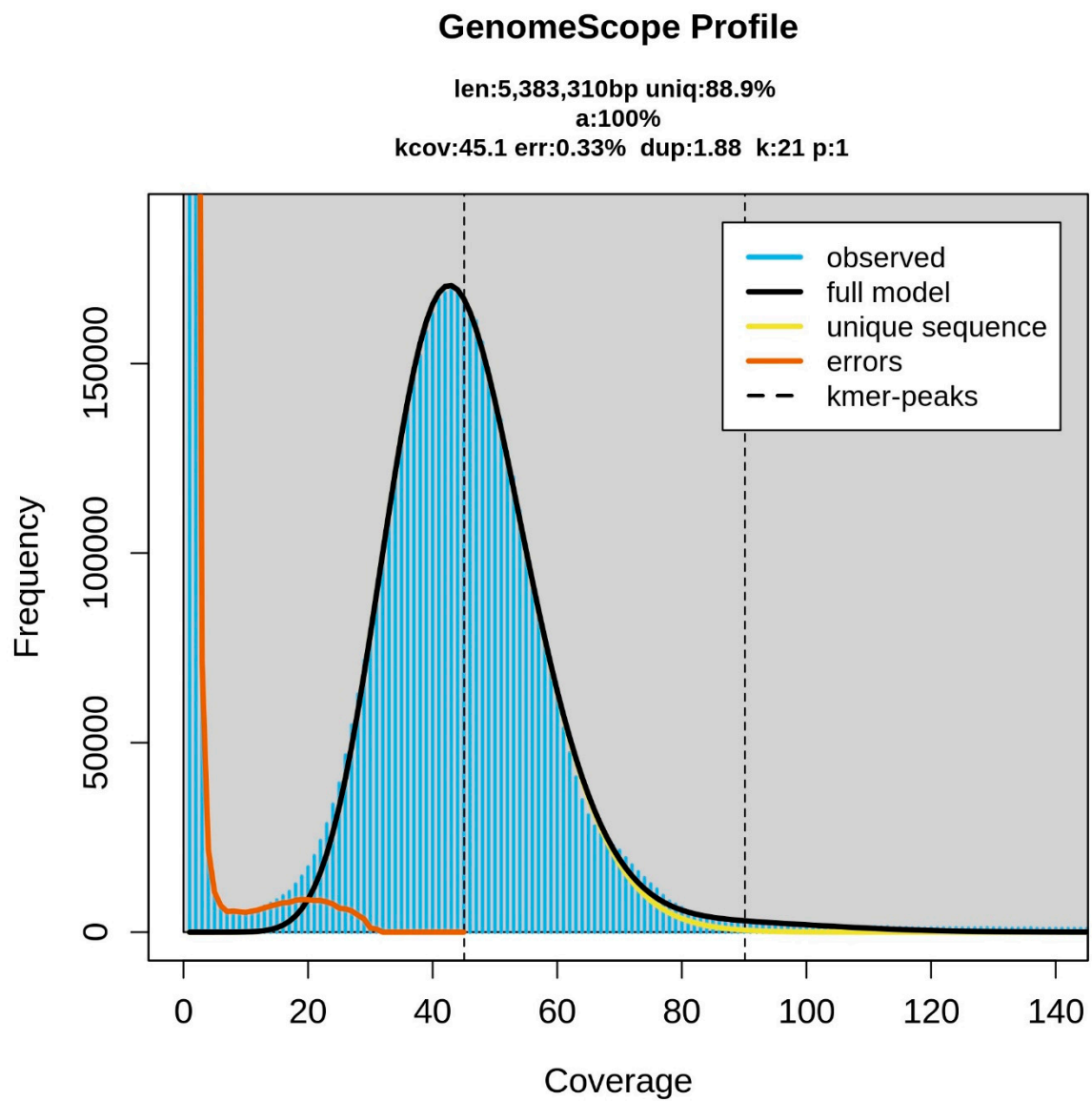
Supplementary Figure S5: GenoScope profile of 1000× AMX sample. The fit of the model (black line) to the observed kmer fre-quencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



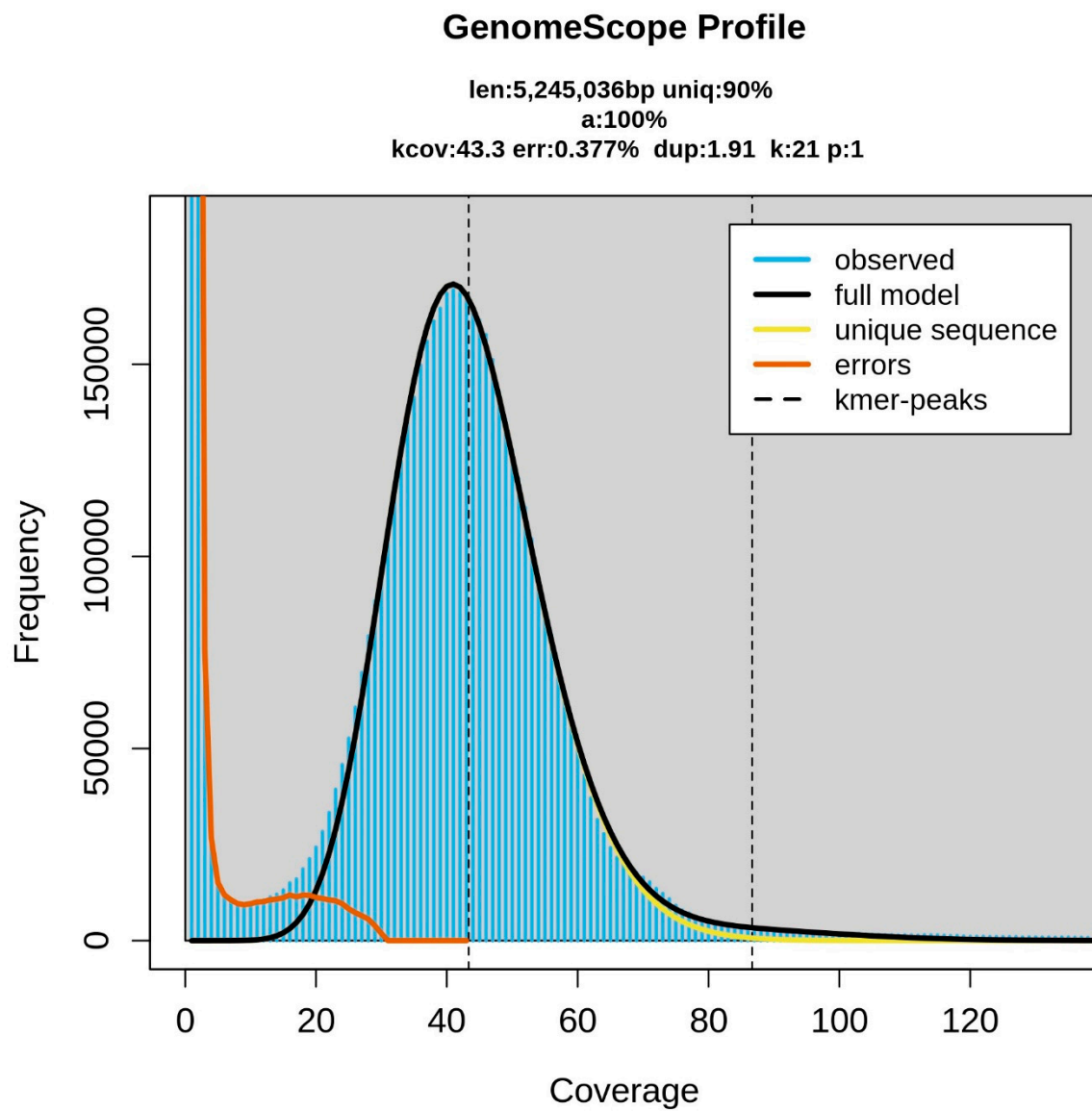
Supplementary Figure S6: GenoScope profile of 0× CTX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



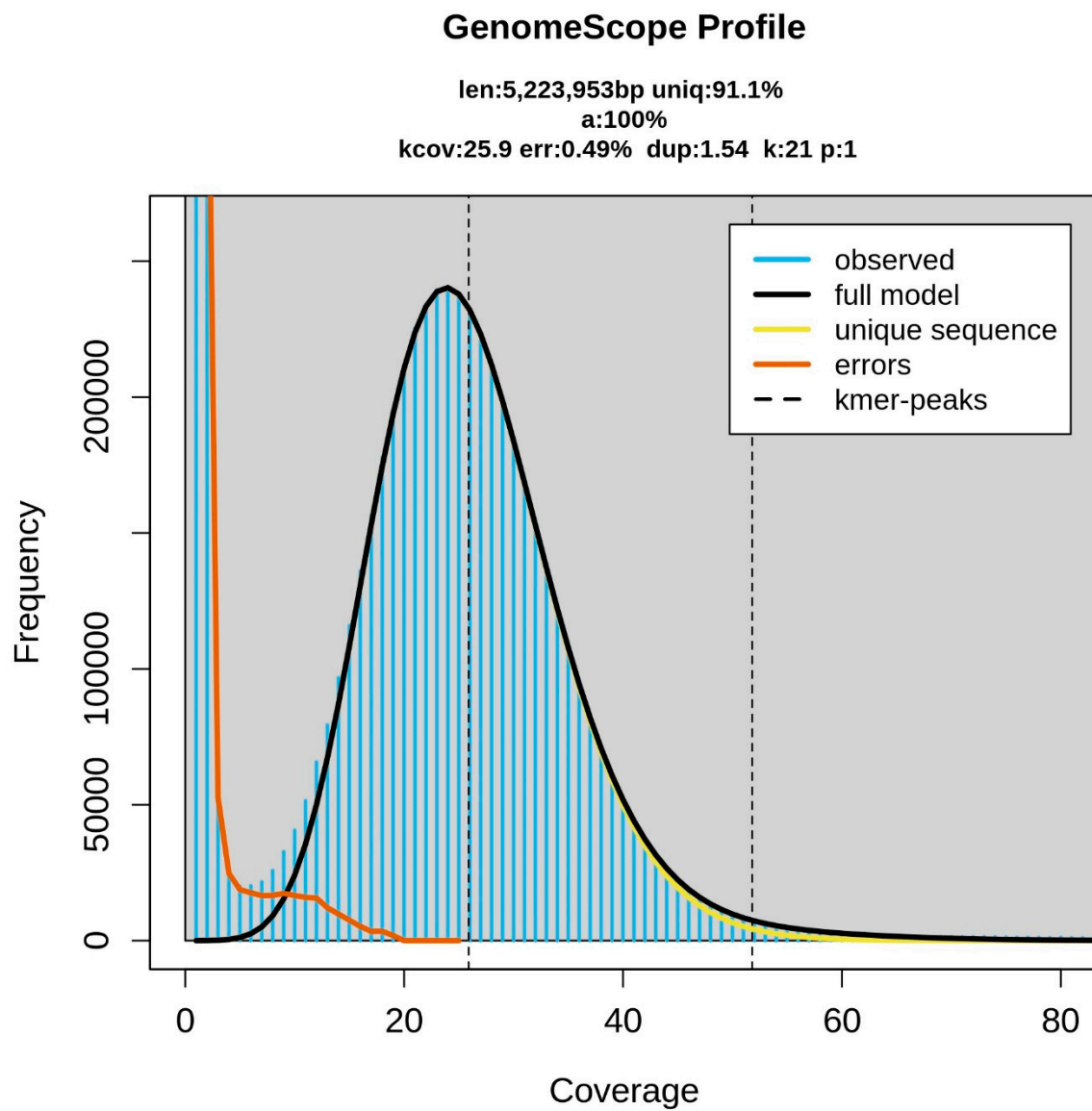
Supplementary Figure S7: GenoScope profile of 1× AMX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



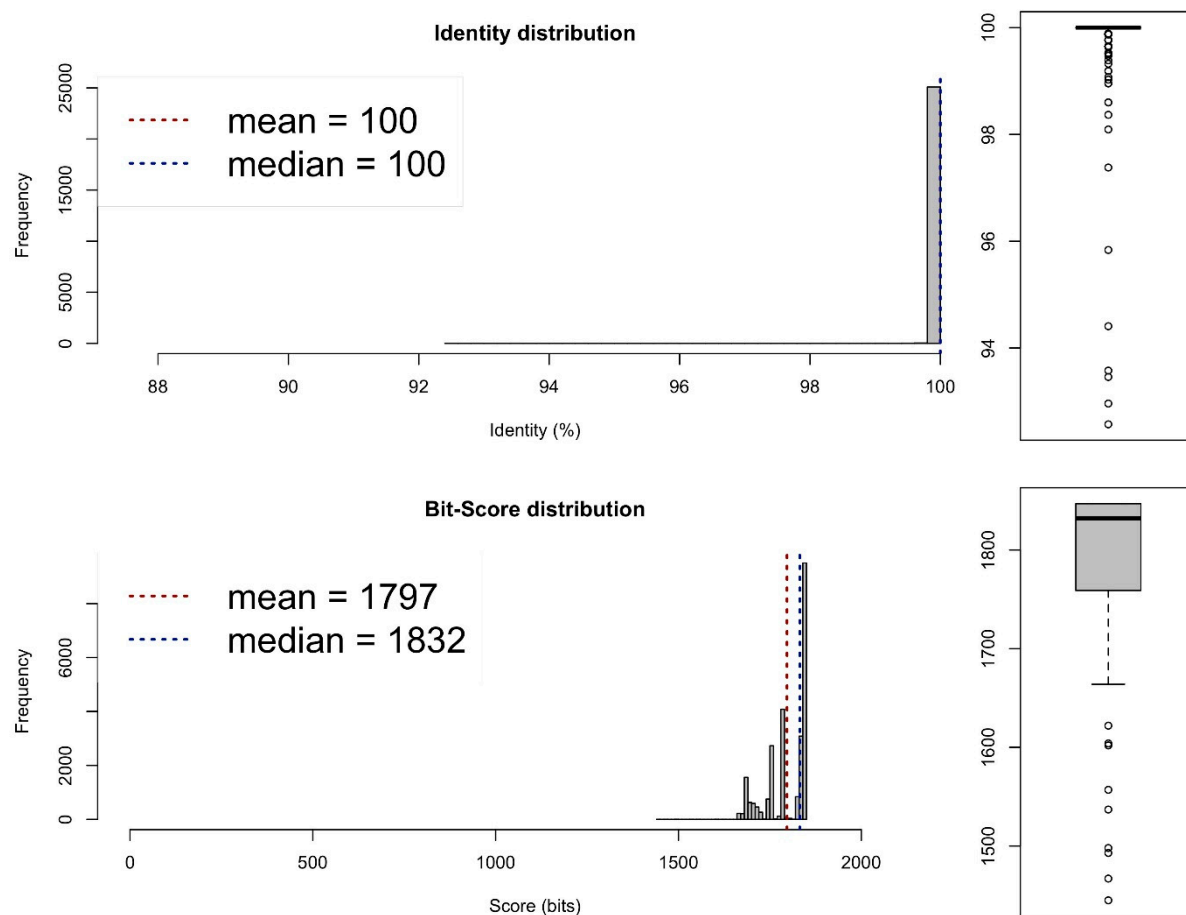
Supplementary Figure S8: GenoScope profile of 10× AMX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



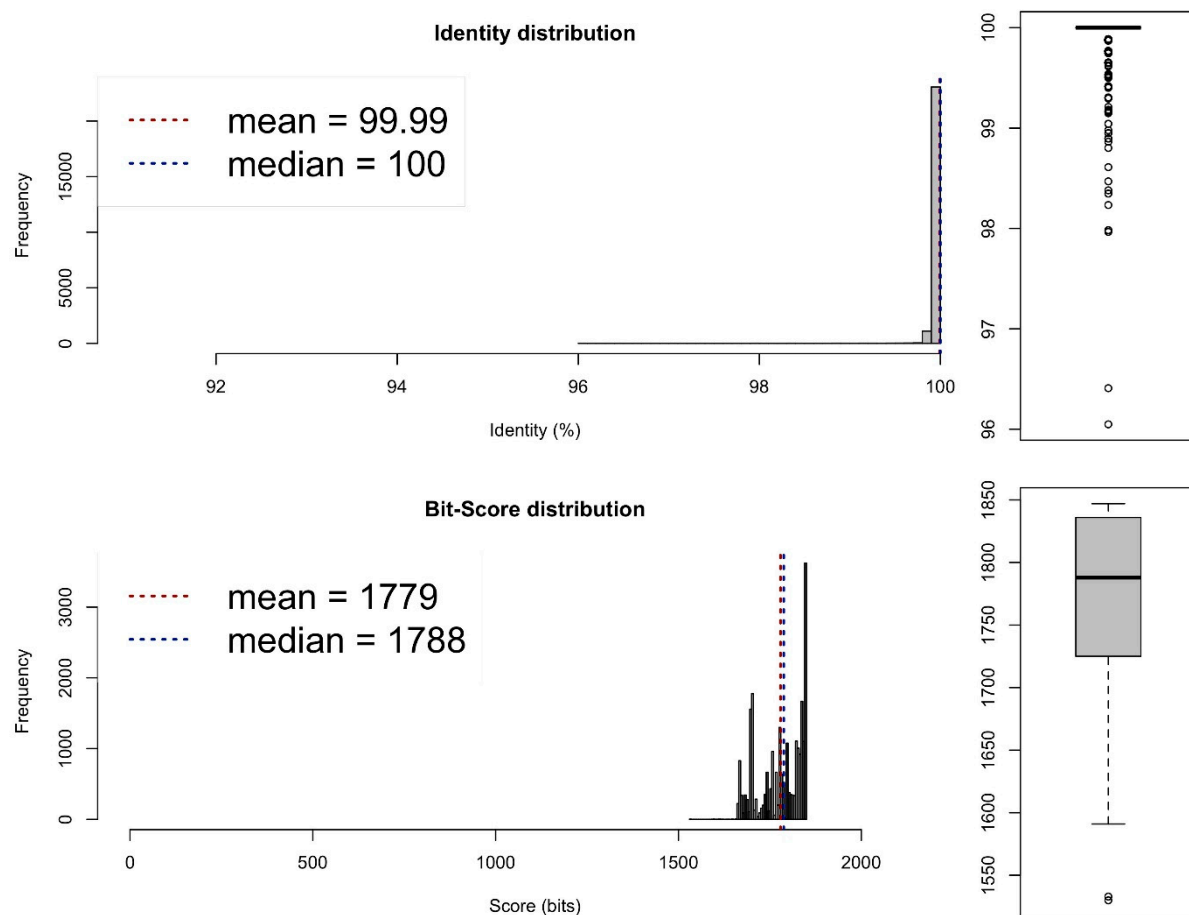
Supplementary Figure S9: GenoScope profile of 100× AMX sample. The fit of the model (black line) to the observed kmer frequencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



Supplementary Figure S10: GenoScope profile of 1000× AMX sample. The fit of the model (black line) to the observed kmer fre-quencies (blue area) is shown for *E. coli* sample. The genome size, heterozygote ratio and repetitive content of unprocessed short reads can be read.



Supplementary Figure S11: The Average Nucleotide Identity (ANI) values between 0× and 1000× amoxicillin genomes of the same species are above 95%. One-way ANI 1: 99.99% (SD: 0.23%), from 25210 fragments. One-way ANI 2: 99.99% (SD: 0.15%), from 25203 fragments. Two-way ANI: 100.00% (SD: 0.10%), from 25145 fragments.



Supplementary Figure S12: The Average Nucleotide Identity (ANI) values between 0× and 1000× cefotaxime genomes of the same species are above 95%. One-way ANI 1: 99.99% (SD: 0.17%), from 24483 fragments. One-way ANI 2: 99.99% (SD: 0.07%), from 24366 fragments. Two-way ANI: 99.99% (SD: 0.07%), from 24307 fragments.