



Article Impact of Beta-Lactam Target Attainment on Resistance Development in Patients with Gram-Negative Infections

Nicole F. Maranchick ^{1,2,*}, Jessica Webber ³, Mohammad H. Alshaer ^{1,2}, Timothy W. Felton ^{4,5} and Charles A. Peloquin ^{1,2}

- ¹ Infectious Disease Pharmacokinetics Lab, Department of Pharmacotherapy and Translational Research, College of Pharmacy, University of Florida, Gainesville, FL 32610, USA
- ² Emerging Pathogens Institute, University of Florida, Gainesville, FL 32610, USA
- ³ College of Pharmacy, University of Florida, Gainesville, FL 32610, USA
- ⁴ North West Ventilation Unit, Manchester University NHS Foundation Trust, Manchester M23 9LT, UK
- ⁵ Division of Infection, Immunity and Respiratory Medicine, School of Biological Sciences, Faculty of Biology,
 - Medicine and Health, The University of Manchester, Manchester M13 9NT, UK
- * Correspondence: n.maranchick@cop.ufl.edu

Abstract: Background: The objective was to identify associations between beta-lactam pharmacokinetic/pharmacodynamic (PK/PD) targets and Gram-negative bacteria resistance emergence in patients. Methods: Retrospective data were collected between 2016 to 2019 at the University of Florida Health-Shands Hospital in Gainesville, FL. Adult patients with two Gram-negative isolates receiving cefepime, meropenem, or piperacillin-tazobactam and who had plasma beta-lactam concentrations were included. Beta-lactam exposures and time free drug concentrations that exceeded minimum inhibitory concentrations (fT > MIC), four multiples of MIC ($fT > 4 \times MIC$), and free area under the time concentration curve to MIC (fAUC/MIC) were generated. Resistance emergence was defined as any increase in MIC or two-fold increase in MIC. Multiple regression analysis assessed the PK/PD parameter impact on resistance emergence. Results: Two hundred fifty-six patients with 628 isolates were included. The median age was 58 years, and 59% were males. Cefepime was the most common beta-lactam (65%) and Pseudomonas aeruginosa the most common isolate (43%). The mean daily $fAUC/MIC \ge 494$ was associated with any increase in MIC (p = 0.002) and two-fold increase in MIC (p = 0.004). The daily fAUC/MIC \geq 494 was associated with decreased time on antibiotics (p = 0.008). *P. aeruginosa* was associated with any increase in MIC (OR: 6.41, 95% CI [3.34–12.28]) or $2 \times$ increase in MIC (7.08, 95% CI [3.56–14.07]). Conclusions: $fAUC/MIC \ge 494$ may be associated with decreased Gram-negative resistance emergence.

Keywords: drug resistance; gram-negative bacteria; beta-lactams; pharmacokinetic/pharmacodynamic

1. Introduction

Antimicrobial resistance puts millions of lives at risk, and in the United States, antimicrobial-resistant bacteria and fungi contribute to more than 35,000 deaths each year [1,2]. Antimicrobial resistance occurs when changes in bacteria cause the drugs used to treat them to become less effective [3]. Gram-negative resistance is especially a concern regarding Enterobacterales and *Pseudomonas aeruginosa*, which have demonstrated resistance to all available antibiotics through varying mechanisms [1,4]. Gram-negative bacteria are highly adaptable to antibiotics, warranting a judicious use to minimize resistance emergence [5]. In addition, given the slow development of new antibiotics, alternative strategies to minimize resistance emergence are necessary, such as optimizing pharmacokinetic/pharmacodynamic (PK/PD) drug targets [6–9].

Beta-lactams are the most commonly used class of antibiotics, and their bacterial killing depends upon the percent of time that free drug concentrations remain above the bacteria minimum inhibitory concentration (fT > MIC) [10]. Clinical data have suggested



Citation: Maranchick, N.F.; Webber, J.; Alshaer, M.H.; Felton, T.W.; Peloquin, C.A. Impact of Beta-Lactam Target Attainment on Resistance Development in Patients with Gram-Negative Infections. *Antibiotics* 2023, *12*, 1696. https://doi.org/ 10.3390/antibiotics12121696

Academic Editor: Ilias Karaiskos

Received: 26 October 2023 Revised: 28 November 2023 Accepted: 29 November 2023 Published: 3 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). that PK/PD targets such as 100% $fT > 4-5 \times$ MIC may be necessary to improve clinical and microbiological outcomes [11–13]. Given that beta-lactams are widely used due to their broad efficacy and excellent safety profile, resistance to them is particularly concerning [6]. Consensus is currently lacking as to what PK/PD target maximizes beta-lactams' clinical efficacy while also minimizing resistance emergence [14]. It may be expected that the pharmacodynamic driver for resistance suppression would be the same as for bacterial cell killing, but this is not always the case [15]. Given the lack of evidence, the purpose of this study was to identify associations between beta-lactam PK/PD targets and resistance emergence in patients with Gram-negative bacterial infections.

2. Results

In total, 256 patients with 628 Gram-negative isolates were included. The median (interquartile range) age and body mass index (BMI) were 58 (42–69) years and 26 (21.4–33.4) kg/m², respectively (Table 1). Seventy-nine percent of patients were critically ill at the start of beta-lactam antibiotics. The most common Gram-negative isolates were *P. aeruginosa* (43%), *Escherichia coli* (14%), and *Klebsiella pneumoniae* (8%). The most common culture types were from the lung (38%) and blood (38%). Multi-drug resistance was identified in 9.8% of isolates. Most patients received cefepime (65%). Fifteen (6%) patients received more than one study antibiotic, but not concurrently.

Table 1. Patient characteristics, *n* = 256 patients.

Clinical Characteristics	Median (IQR) or <i>n</i> (%)
Age, years	58 (42–69)
Sex, Male	151 (59)
Weight, kg	73.6 (60.9–94.3)
Serum creatinine, mg/dL	0.81 (0.57–1.27)
BMI, kg/m ²	26 (21.4–33.4)
Renal replacement therapy	44 (17)
Liver Disease	78 (30)
COPD	55 (21)
Diabetes	113 (44)
Heart failure	105 (41)
Mechanical Ventilation	66 (26)
Patient in ICU at beta-lactam initiation	203 (79)
Length of stay, days	
ICU	14 (2–29)
Hospital	25 (16–47)
Mortality	122 (48)
Gram-negative isolates, <i>n</i>	628
Common isolated bacteria, <i>n</i> (% developing resistance)	
Pseudomonas aeruginosa	134 (35.6)
Escherichia coli	30 (6.7)
Klebsiella pneumoniae	26 (11.5)
Enterobacter cloacae	20 (0)
Serratia marcescens	19 (5.2)
Proteus mirabilis	14 (0)
Acinetobacter baumannii	14 (14.2)
Klebsiella aerogenes	10 (10)

Table 1. Cont.

Clinical Characteristics	Median (IQR) or <i>n</i> (%)
Multi-drug resistant isolates #	62 (9.8)
All Culture Sources	
Lung	233
Blood	230
Wound	80
Urinary Tract	36
Abscess/body fluid	32
Other	17
Culture Sources (Same Final and Initial Source)	
Lung	204
Blood	214
Wound	46
Urinary Tract	32
Abscess/body fluid	16
Other	6
Beta-lactam Received	
Cefepime	179 (65)
Meropenem	54 (20)
Piperacillin/tazobactam	41 (15)
Number of samples	
Cefepime	316
Meropenem	91
Piperacillin/tazobactam	73
Beta-lactam therapy duration, days	8 (5–14)
Time between cultures, days	7 (4–15)
Time between start of beta-lactam therapy and TDM, days	3 (2–8)
Concomitant Antibiotics	
-Aminoglycoside	124 (48)
-Fluoroquinolone	57 (22)
-Polymyxin	30 (12)

IQR—interquartile range; BMI—body mass index; COPD—chronic obstructive pulmonary disease; ICU—intensive care unit; TDM—therapeutic drug monitoring; ^ Total = 274. Fifteen patients received more than one beta-lactam during the study period; [#] Multidrug resistant (MDR) isolates include extended spectrum beta-lactamase (ESBL) *Enterobacterales*, carbapenem-resistant (CR) *Enterobacterales*, MDR-*Pseudomonas aeruginosa*, MDR-*Acinetobacter baumannii*, or any bacteria non-susceptible to ≥ 1 agent in ≥ 3 antimicrobial categories.

Figure 1 shows beta-lactam target attainment stratified by beta-lactam and time window (0–24 h, 0–7 days, and duration of therapy). In the first 24 h, 81% and 45% (cefepime), 70% and 48% (meropenem), and 37% and 16% (piperacillin) achieved 100% *f*T > MIC and 100% *f*T > 4× MIC, respectively. In the first 7 days, 61% and 34% (cefepime), 55% and 38% (meropenem), and 18% and 2% (piperacillin) achieved 100% *f*T > MIC and 100% *f*T > 4× MIC. For the total duration of antibiotics, 49% and 24% (cefepime), 50% and 34% (meropenem), and 14% and 2% (piperacillin) achieved 100% *f*T > MIC and 100% *f*T > 4× MIC. For the total duration of antibiotics, 49% and 24% (cefepime), 50% and 34% (meropenem), and 14% and 2% (piperacillin) achieved 100% *f*T > MIC and 100% *f*T > 4× MIC, respectively. Figure 2 shows the mean (SD) daily *f*AUC/MIC stratified by beta-lactam. Meropenem had the highest mean free area under the time concentration curve to MIC (*f*AUC/MIC) of 708 (999), followed by cefepime with a mean daily *f*AUC/MIC of 500 (499), and then piperacillin with a mean AUC/MIC of 145 (175).

Nineteen percent of isolates had an increase in MIC between cultures, of which 77% was due to *P. aeruginosa*. After testing for associations between baseline characteristics and resistance, renal replacement therapy (RRT) during admission, intensive care unit (ICU) length of stay (LOS), hospital LOS, mechanical ventilation, and days between first and second culture were significantly associated with resistance (Table 2). The Sequential Organ Failure Assessment (SOFA) score was associated with a two-fold increase in MIC.

These covariates were controlled for in the multiple regression analysis (Table 3). Classification and regression tree (CART) analysis determined a significant *f*AUC/MIC target of 494 associated with resistance emergence that was included in the models. Subjects with a mean daily *f*AUC/MIC \geq 494 had significantly less resistance emergence when defined as any increase in MIC (aOR 0.25, 95% CI [0.11–0.61]) or at least a two-fold increase in MIC (aOR 0.27, 95% CI [0.11–0.67]). These relationships held true even in a subgroup analysis including only initial and final culture types of the same source. In the subgroup analysis, a daily *f*AUC/MIC \geq 494 was significantly associated with an increased risk of Gram-negative resistance emergence when defined as any increase in MIC (aOR 0.33, 95% CI [0.13–0.87]) or at least a two-fold increase in MIC (aOR 0.38, 95% CI [0.14–0.99]). No associations were found between *f*T > MIC and Gram-negative resistance emergence.



Figure 1. Violin plots showing PK/PD target attainment stratified by beta-lactam. PK/PD—pharmacokinetic/pharmacodynamic; MIC—minimum inhibitory concentration; fT > MIC—time free drug concentrations exceed minimum inhibitory concentration; $fT > 4 \times$ MIC—time free drug concentrations exceed four multiples of MIC.

Table 2. Univariate analysis.

	Resistance (Any Increase in MIC)		Resistance (≥2× MIC Increase)	
Covariates	OR	<i>p</i> -Value	OR	<i>p</i> -Value
Age (per 1 year)	0.99	0.08	0.99	0.35
BMI (per 1 kg/m ²)	1.00	0.73	1.00	0.82
RRT during admission	2.24	0.03	2.50	0.01
Days on antibiotic therapy (per 1 day)	1.01	0.06	1.01	0.06
Days between cultures (per 1 day)	1.06	< 0.0001	1.06	< 0.0001
Mechanical Ventilation	2.27	0.007	2.61	0.002

	Resistance (Any Increase in MIC)		Resistance (≥2× MIC Increase)	
Covariates	OR	<i>p</i> -Value	OR	<i>p</i> -Value
Hospital LOS (per 1 day)	1.01	<0.0001	1.01	<0.0001
ICU LOS (per 1 day)	1.02	< 0.0001	1.02	< 0.0001
Diabetes	1.14	0.67	0.94	0.88
Liver Disease	1.38	0.28	1.30	0.43
COPD	1.38	0.39	1.38	0.38
Heart Failure	1.18	0.56	1.36	0.30
SOFA Score (per 1 point)	1.05	0.12	1.08	0.02

Note: MIC—minimum inhibitory concentration; BMI—body mass index; RRT—renal replacement therapy; LOS—length of stay; ICU—intensive care unit; COPD—chronic obstructive pulmonary disease; SOFA—sequential organ failure assessment.



Figure 2. Bar graphs showing mean (standard deviation) daily *f*AUC/MIC stratified by beta-lactam. *f*AUC/MIC—free area under the time concentration curve to minimum inhibitory concentration.

P. aeruginosa was found to be significantly associated with bacterial resistance emergence for both any increase in MIC (OR: 6.41, 95% CI [3.34–12.28]) and at least a two-fold increase in MIC (7.08, 95% CI [3.56–14.07]).

Figure 3 shows the Kaplan–Meier curve for the time on beta-lactams based upon the mean daily *f*AUC/MIC target attainment. Thirty-one percent achieved a mean daily *f*AUC/MIC \geq 494. Patients achieving *f*AUC/MIC \geq 494 had a significantly shorter time on antibiotics (*p* = 0.008).

Table 2. Cont.

	Resistance (Any Increase in MIC)		Resistance (≥2× MIC Increase)	
PK/PD Parameter	aOR (95% CI)	<i>p</i> -Value	aOR (95% CI)	<i>p</i> -Value
% <i>f</i> T > MIC 0–24 h (per 10%)	0.96 (0.88–1.07)	0.50	1.06 (0.95–1.20)	0.34
% $fT > 4 \times MIC 0-24 h$ (per 10%)	0.98 (0.91–1.06)	0.64	1.02 (0.94–1.10)	0.71
% <i>f</i> T > MIC 0–7 d (per 10%)	1.06 (0.95–1.21)	0.31	1.08 (0.94–1.24)	0.27
% <i>f</i> T > 4× MIC 0–7 d (per 10%)	1.03 (0.94–1.12)	0.51	1.08 (0.99–1.19)	0.08
% $fT > MIC$ duration of therapy (per 10%)	0.98 (0.89–1.08)	0.67	1.03 (0.92–1.14)	0.63
% $fT > 4 \times$ MIC duration of therapy (per 10%)	0.99 (0.92–1.09)	0.95	1.03 (0.94–1.13)	0.53
Mean daily <i>f</i> AUC/MIC (per increments of 10)	0.99 (0.98–1.00)	0.08	1.00 (0.99–1.001)	0.17
Mean daily <i>f</i> AUC/MIC of 494 achieved (yes)	0.25 (0.11-0.61)	0.002	0.27 (0.11–0.67)	0.004

Table 3. Final statistical models with PK/PD predictors.

Note: MIC—minimum inhibitory concentration; PK/PD—pharmacokinetic/pharmacodynamic; aOR—adjusted odds ratio; CI—confidence interval.



Figure 3. Kaplan–Meier curve for beta-lactam duration of therapy comparing patients based on *f*AUC/MIC target attainment of 494.

3. Discussion

This study's objective was to identify associations between beta-lactam target attainment and the prevention of resistance emergence in patients with Gram-negative infections. In the covariate analysis, RRT, mechanical ventilation, ICU LOS, hospital LOS, and days between first and last culture were associated with resistance. The SOFA score was associated with a two-fold increase in MIC. The mean daily $fAUC/MIC \ge 494$ was associated with a decreased risk of resistance development. Time-to-event analysis showed that patients achieving a mean daily $fAUC/MIC \ge 494$ had less time on beta-lactam therapy. These

results indicate that optimizing daily beta-lactam *f*AUC/MIC exposure may minimize Gram-negative resistance and the duration of therapy.

In a previous study of 76 patients receiving either cefepime or ceftazidime, researchers aimed to characterize the relationship between PD parameters and clinical and microbiological outcomes. Patients were included if they had sepsis and suspected or proven infection due to a pathogen susceptible to cefepime/ceftazidime. Patient PK parameters were estimated from doses administered and patient-specific data. Patients achieving *f*AUC/MIC ratios ≥ 250 had significantly higher rates of clinical cure (p = 0.002) and bacteriological eradication (p < 0.001) [16]. In a study by Schentag et al., researchers performed simulations to evaluate *f*AUC/MIC targets for cefmenoxime, tobramycin, and ciprofloxacin. They found that for cefmenoxime, a cephalosporin, an *f*AUC/MIC ratio of 540 per 24 h was required for bacterial eradication. They also proposed that each antibiotic has a unique 24-h *f*AUC/MIC value associated with bacterial eradication at 4 days [17]. Our PK/PD target of 494 falls within the ranges from the previously published literature and may be beneficial for clinical outcomes, bacterial eradication, and/or resistance suppression.

In general, the efficacy of beta-lactams depends upon fT > MIC, with 40–70% fT > MICproposed as the minimum threshold for bactericidal activity [18]. In a study by Gatti et al., 116 ICU patients receiving beta-lactam continuous infusions for Gram-negative infections with at least one therapeutic drug monitoring in the first 72 h of treatment were assessed for PK/PD target thresholds. Steady state concentration/MIC ratios \leq 5 were associated with microbiological failure. In addition, *P. aeruginosa* infection was associated with microbiological failure [14]. Felton et al. compared piperacillin/tazobactam PK/PD indices to suppress bacterial resistance in both high and low burdens of *P. aeruginosa*. A Cmin/MIC of 3.4 was required, unless the bacterial burden was high, in which case a Cmin/MIC of 4.6 was needed [19]. In our study, we did not find an association between fT > MIC and $fT > 4 \times MIC$ with the emergence of Gram-negative resistance. However, based upon the findings of the previous study and increased target concentrations with a high bacterial burden, there is the potential that our targets of fT > MIC and $fT > 4 \times MIC$ were not sufficient to suppress bacterial resistance. In addition, P. aeruginosa was responsible for approximately 43% of the isolates in our study and was an independent risk factor for resistance emergence, which may have impacted the ability of PK/PD target attainment to suppress resistance emergence.

Beta-lactam target attainment (fT > MIC, $fT > 4 \times MIC$, and $fAUC/MIC \ge 494$) for patients in the present study was in general poor, especially for piperacillin. The low target attainment could be due to the high percentage of ICU patients (approximately 79%) who have an increased risk of pharmacokinetic variability. Of note, while patients achieving daily $fAUC/MIC \ge 494$ had significantly less time on antibiotics, only 31% of patients met this target. In a previous study of 80 ICU patients receiving cefepime, meropenem, and piperacillin-tazobactam, researchers found that serum concentrations remained $4 \times$ above target concentrations for the *P. aeruginosa* breakpoint for 34% (cefepime), 57% (meropenem), and 33% (piperacillin-tazobactam) of the dosing interval. They concluded that only meropenem had acceptable serum concentrations and that more aggressive dosing may be needed to empirically cover pathogens, especially in critically ill patients [20]. The EXPAT study was a prospective observational study in two ICUs. Researchers enrolled patients receiving beta-lactam antibiotics and collected drug samples on day 2 of therapy. Of 147 patients, researchers concluded that 63.3% and 36.7% of patients achieved 100% fT > MIC and 100% fT > 4× MIC, respectively. They identified male gender, high BMI, and elevated eGFR as risk factors for target non-attainment [21]. While our study did not find an association between fT > MIC or $fT > 4 \times MIC$, we have demonstrated that fAUC/MIC target attainment decreased time on antibiotics, which provides further support for early target attainment in clinical practice. Therefore, due to the risks of subtherapeutic beta-lactam concentrations and the benefits of early target attainment, especially in critically ill patients, therapeutic drug monitoring would ideally be initiated on day 1 of beta-lactam therapy to improve target attainment.

There are limitations to this study. This was a retrospective study utilizing data from a single center with a high percentage of critically ill patients, which may limit external generalizability to other sites. In addition, protein binding values were estimated and used to calculate the unbound fraction, as total drug concentrations were measured. Unbound fractions may be highly variable due to critical illness, hypoalbuminemia, and chronic kidney disease. Sixteen percent of patients received renal replacement therapy during beta-lactam therapy. However, the models used to generate beta-lactam exposures do not account for patients receiving renal replacement therapy, so beta-lactam exposures may have been impacted. In addition, it is generally accepted that MICs have variability, with repeat assessments potentially producing MIC values that differ by 200% [22]. Additionally, due to the retrospective nature of the study, bacterial cultures were ordered by the treating team and were not taken at scheduled intervals. Although controlled for in analysis, future studies should utilize scheduled intervals to obtain cultures to assess resistance emergence.

4. Materials and Methods

4.1. Data Collection

This retrospective study utilized pooled data from the UF Health-Shands hospital in Gainesville, Florida, USA between 2016 to 2019. Patients were included if \geq 18 years old, had two separate cultures positive for Gram-negative bacteria during the same hospitalization, were receiving cefepime, meropenem, or piperacillin (administered with tazobactam), and had beta-lactam concentrations measured during therapy as part of the usual therapeutic drug monitoring service [23]. Cultures were included if collected from the same site at least one day apart. The first culture was included if it was before or at the start of antibiotic therapy, and subsequent cultures were included age, BMI, ICU LOS, hospital LOS, SOFA score, days on antibiotic therapy, days between cultures, and clinical factors, such as renal replacement therapy, mechanical ventilation, diabetes, liver disease, chronic obstructive pulmonary disease (COPD), and heart failure. These covariates were chosen due to their potential to be associated with resistance emergence [24–27].

Resistance was defined as any increase in minimum inhibitory concentration (MIC) or at least a two-fold increase in MIC. If the second culture had no growth, it was considered susceptible. If multiple MICs were available after the first culture, the highest available MIC was used. MICs were determined by the UF-Health Shands microbiology. Methods to identify bacteria and MICs included VITEK[®] Mass Spectrometry and Vitek[®] II (bioMérieux, Inc., Durham, NC, USA). Etest was utilized for MIC quantification for the following bacteria (beta-lactam) combinations: *Acinetobacter* spp. (cefepime, meropenem, and piperacillin), *Burkholderia cepacia complex* (meropenem), and Gram-negative non-fermenters (cefepime, meropenem, and piperacillin). Multidrug-resistant (MDR) isolates were defined as extended spectrum beta-lactamase (ESBL) *Enterobacterales*, carbapenem-resistant (CR) *Enterobacterales*, MDR-*Pseudomonas aeruginosa*, MDR-*Acinetobacter baumannii*, or any bacteria non-susceptible to ≥ 1 agent in ≥ 3 antimicrobial categories [4,28–30].

Following collection, total beta-lactam plasma concentrations were quantified using liquid chromatography with tandem mass spectrometry assays at the Infectious Disease Pharmacokinetics Laboratory at UF. The calibration range was 2–100 mg/L with an interand intra-day accuracy and precision < 10%. Values below the limit of quantification were assigned a value of "0" for analysis. Free drug concentrations were estimated using previously published values (80% for cefepime, 98% for meropenem, and 70% for piperacillin) [31–33].

Posterior predictions were generated using the nonparametric adaptive grid (NPAG) in Pmetrics v1.9.7 (Laboratory of Applied Pharmacokinetics and Bioinformatics, Los Angeles, CA, USA) with previously published cefepime, meropenem, and piperacillin models, drug doses, drug concentrations, and covariates, including renal function (CrCl or SCr), weight, and age [34–36]. Beta-lactam exposure was generated from initiation of antibiotics up until therapy discontinuation or the time of resistant bacteria culture collection, whichever was reported first. Beta-lactam posterior predictions were imported to Phoenix WinNonlin v8.3.4 (Certara, Princeton, NJ, USA) to calculate the free area under the time concentration curve (*f*AUC) and *f*T > MIC and *f*T > 4× MIC for 0–24 h, 0–7 days, and the duration of beta-lactam therapy or up to the day of resistant bacterial culture. Calculated *f*AUC was used to estimate mean daily *f*AUC to MIC (*f*AUC/MIC) ratios. If patients received beta-lactam therapy for less than or had a resistant bacteria culture in less than 7 days, PK/PD calculations were stopped after the last dose of beta-lactam.

4.2. Statistical Analysis

Statistical analysis was performed on JMP Pro v17 (SAS Institute, Cary, NC, USA). Continuous data were presented as the median and interquartile range (IQR) and categorical data as count and percentages. Covariates including age, BMI, RRT, days on antibiotics, days between first and last culture, mechanical ventilation, hospital and ICU LOS, diabetes, liver disease, COPD, heart failure, and SOFA score were tested individually for association with resistance emergence (defined as both any increase in MIC or $\geq 2 \times$ increase in MIC) in a univariate analysis. Significant covariates identified in the univariate analysis were included and controlled for in the final multiple regression models, and PK/PD parameters were tested individually for associations with bacterial resistance. PK/PD parameters tested in the multiple regression models included the mean daily fAUC/MIC and fT > MICand $fT > 4 \times$ MIC for 0 to 24 h, 0 to 7 days, and the duration of therapy. Classification and regression tree (CART) analysis was used to test PK/PD parameters including fT > MIC and fAUC/MIC for breakpoint values of significance. These breakpoints were then tested in the multiple regression models for associations with bacterial resistance. A subgroup analysis was also conducted to test PK/PD relationships with resistance emergence when only using cultures of the same source. The same significant covariates were included in the final multiple regression model. Kaplan–Meier estimators were reported for associations between PK/PD parameters and the time on beta-lactam therapy. A *p*-value of less than 0.05 was considered statistically significant for all analyses.

5. Conclusions

Daily $fAUC/MIC \ge 494$ may be associated with a decreased risk of Gram-negative resistance emergence and a reduced duration of antibiotics. These results could be a potential PK/PD target for future investigations, such as interventional prospective studies with a larger sample size. *P. aeruginosa* was significantly associated with an increase in resistance emergence. No associations between resistance emergence and fT > MIC and $fT > 4 \times MIC$ were found.

Author Contributions: N.F.M. performed data collection, data analysis, and wrote the manuscript. J.W. performed data collection and assisted with manuscript writing. M.H.A. was involved in the study design, analysis, and contributed a model for Pmetrics. T.W.F. contributed a model for Pmetrics. C.A.P. assisted with the study design and oversaw the quantification of all beta-lactams measured in the study. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: This study was reviewed and approved as exempt by the University of Florida IRB (IRB # 201902910).

Informed Consent Statement: Patient consent was waived due to the retrospective, noninterventional design of the study. The confidentiality of all data obtained during the research was guaranteed.

Data Availability Statement: Data supporting the reported results are available upon request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Centers for Disease Control and Prevention. About Antimicrobial Resistance. 2022. Available online: https://www.cdc.gov/ drugresistance/about.html (accessed on 23 March 2023).
- 2. World Health Organization. Antimicrobial Resistance: Global Report on Surveillance. 1 April 2014. Available online: https://www.who. int/publications/i/item/9789241564748 (accessed on 23 March 2023).
- 3. Murray, C.J.L.; Ikuta, K.S.; Sharara, F.; Swetschinski, L.; Aguilar, G.R.; Gray, A.; Han, C.; Bisignano, C.; Rao, P.; Wool, E.; et al. Global burden of bacterial antimicrobial resistance in 2019: A systematic analysis. *Lancet* **2022**, *399*, 629–655. [CrossRef]
- Magiorakos, A.-P.; Srinivasan, A.; Carey, R.B.; Carmeli, Y.; Falagas, M.E.; Giske, C.G.; Harbarth, S.; Hindler, J.F.; Kahlmeter, G.; Olsson-Liljequist, B.; et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: An international expert proposal for interim standard definitions for acquired resistance. *Clin. Microbiol. Infect.* 2012, *18*, 268–281. [CrossRef] [PubMed]
- Fernandes, P. Antibacterial discovery and development—The failure of success? *Nat. Biotechnol.* 2006, 24, 1497–1503. [CrossRef] [PubMed]
- MacVane, S.H. Antimicrobial Resistance in the Intensive Care Unit: A Focus on Gram-Negative Bacterial Infections. J. Intensiv. Care Med. 2016, 32, 25–37. [CrossRef] [PubMed]
- Sumi, C.D.; Heffernan, A.J.; Lipman, J.; Roberts, J.A.; Sime, F.B. What Antibiotic Exposures Are Required to Suppress the Emergence of Resistance for Gram-Negative Bacteria? A Systematic Review. *Clin. Pharmacokinet.* 2019, 58, 1407–1443. [CrossRef]
- Heffernan, A.J.; Sime, F.B.; Lipman, J.; Roberts, J.A. Individualising Therapy to Minimize Bacterial Multidrug Resistance. *Drugs* 2018, 78, 621–641. [CrossRef]
- 9. Adembri, C.; Novelli, A.; Nobili, S. Some Suggestions from PK/PD Principles to Contain Resistance in the Clinical Setting—Focus on ICU Patients and Gram-Negative Strains. *Antibiotics* 2020, *9*, 676. [CrossRef]
- 10. Rybak, M.J. Pharmacodynamics: Relation to antimicrobial resistance. *Am. J. Infect. Control* **2006**, *34*, S38–S45; discussion S64–S73. [CrossRef]
- Alshaer, M.H.; Maranchick, N.; Bai, C.; Maguigan, K.L.; Shoulders, B.; Felton, T.W.; Mathew, S.K.; Mardini, M.T.; Peloquin, C.A. Using Machine Learning to Define the Impact of Beta-Lactam Early and Cumulative Target Attainment on Outcomes in Intensive Care Unit Patients with Hospital-Acquired and Ventilator-Associated Pneumonia. *Antimicrob. Agents Chemother.* 2022, 66, e0056322. [CrossRef]
- Roberts, J.A.; Paul, S.K.; Akova, M.; Bassetti, M.; De Waele, J.J.; Dimopoulos, G.; Kaukonen, K.-M.; Koulenti, D.; Martin, C.; Montravers, P.; et al. DALI: Defining Antibiotic Levels in Intensive Care Unit Patients: Are Current β-Lactam Antibiotic Doses Sufficient for Critically III Patients? *Clin. Infect. Dis.* 2014, *58*, 1072–1083. [CrossRef]
- Guilhaumou, R.; Benaboud, S.; Bennis, Y.; Dahyot-Fizelier, C.; Dailly, E.; Gandia, P.; Goutelle, S.; Lefeuvre, S.; Mongardon, N.; Roger, C.; et al. Optimization of the treatment with beta-lactam antibiotics in critically ill patients—Guidelines from the French Society of Pharmacology and Therapeutics (Société Française de Pharmacologie et Thérapeutique—SFPT) and the French Society of Anaesthesia and Intensive Care Medicine (Société Française d'Anesthésie et Réanimation—SFAR). *Crit. Care* 2019, 23, 104. [CrossRef]
- Gatti, M.; Cojutti, P.G.; Pascale, R.; Tonetti, T.; Laici, C.; Dell'olio, A.; Siniscalchi, A.; Giannella, M.; Viale, P.; Pea, F. Assessment of a PK/PD Target of Continuous Infusion Beta-Lactams Useful for Preventing Microbiological Failure and/or Resistance Development in Critically Ill Patients Affected by Documented Gram-Negative Infections. *Antibiotics* 2021, 10, 1311. [CrossRef]
- Drusano, G.L.; Louie, A.; MacGowan, A.; Hope, W. Suppression of Emergence of Resistance in Pathogenic Bacteria: Keeping Our Powder Dry, Part 1. Antimicrob. Agents Chemother. 2016, 60, 1183–1193. [CrossRef] [PubMed]
- McKinnon, P.S.; Paladino, J.A.; Schentag, J.J. Evaluation of area under the inhibitory curve (AUIC) and time above the minimum inhibitory concentration (T>MIC) as predictors of outcome for cefepime and ceftazidime in serious bacterial infections. *Int. J. Antimicrob. Agents* 2008, 31, 345–351. [CrossRef] [PubMed]
- Schentag, J.J.; Nix, D.E.; Adelman, M.H. Mathematical Examination of Dual Individualization Principles (I): Relationships between AUC above MIC and Area under the Inhibitory Curve for Cefmenoxime, Ciprofloxacin, and Tobramycin. *DICP* 1991, 25, 1050–1057. [CrossRef] [PubMed]
- Craig, W.A. State-of-the-Art Clinical Article: Pharmacokinetic/Pharmacodynamic Parameters: Rationale for Antibacterial Dosing of Mice and Men. *Clin. Infect. Dis.* 1998, 26, 1–10. [CrossRef] [PubMed]
- Felton, T.W.; Goodwin, J.; O'Connor, L.; Sharp, A.; Gregson, L.; Livermore, J.; Howard, S.J.; Neely, M.N.; Hope, W.W. Impact of Bolus Dosing versus Continuous Infusion of Piperacillin and Tazobactam on the Development of Antimicrobial Resistance in Pseudomonas aeruginosa. *Antimicrob. Agents Chemother.* 2013, 57, 5811–5819. [CrossRef] [PubMed]
- Taccone, F.S.; Laterre, P.-F.; Dugernier, T.; Spapen, H.; Delattre, I.; Witebolle, X.; De Backer, D.; Layeux, B.; Wallemacq, P.; Vincent, J.-L.; et al. Insufficient β-lactam concentrations in the early phase of severe sepsis and septic shock. *Crit. Care* 2010, 14, R126. [CrossRef] [PubMed]
- Abdulla, A.; Dijkstra, A.; Hunfeld, N.G.M.; Endeman, H.; Bahmany, S.; Ewoldt, T.M.J.; Muller, A.E.; van Gelder, T.; Gommers, D.; Koch, B.C.P. Failure of target attainment of beta-lactam antibiotics in critically ill patients and associated risk factors: A two-center prospective study (EXPAT). *Crit. Care* 2020, 24, 558. [CrossRef]
- 22. Doern, G.V.; Brecher, S.M. The Clinical Predictive Value (or Lack Thereof) of the Results of In Vitro Antimicrobial Susceptibility Tests. *J. Clin. Microbiol.* 2011, 49, S11–S14. [CrossRef]

- Venugopalan, V.; Hamza, M.; Santevecchi, B.; DeSear, K.; Cherabuddi, K.; Peloquin, C.A.; Alshaer, M.H. Implementation of a β-lactam therapeutic drug monitoring program: Experience from a large academic medical center. *Am. J. Health Pharm.* 2022, *79*, 1586–1591. [CrossRef] [PubMed]
- Hayakawa, K.; Gattu, S.; Marchaim, D.; Bhargava, A.; Palla, M.; Alshabani, K.; Gudur, U.M.; Pulluru, H.; Bathina, P.; Sundaragiri, P.R.; et al. Epidemiology and Risk Factors for Isolation of Escherichia coli Producing CTX-M-Type Extended-Spectrum β-Lactamase in a Large U.S. Medical Center. *Antimicrob. Agents Chemother.* 2013, 57, 4010–4018. [CrossRef]
- Ben-Ami, R.; Rodríguez-Baño, J.; Arslan, H.; Pitout, J.D.D.; Quentin, C.; Calbo, E.S.; Azap, K.; Arpin, C.; Pascual, A.; Livermore, D.M.; et al. A Multinational Survey of Risk Factors for Infection with Extended-Spectrum β-Lactamase–Producing Enterobacteriaceae in Nonhospitalized Patients. *Clin. Infect. Dis.* 2009, *49*, 682–690. [CrossRef] [PubMed]
- 26. Maina, J.W.; Onyambu, F.G.; Kibet, P.S.; Musyoki, A.M. Multidrug-resistant Gram-negative bacterial infections and associated factors in a Kenyan intensive care unit: A cross-sectional study. *Ann. Clin. Microbiol. Antimicrob.* 2023, 22, 85. [CrossRef] [PubMed]
- Al Hamdan, A.S.; Alghamdi, A.; Alyousif, G.F.; Hamza, F.; Shafey, M.M.; AlAmri, A.M.; Sunki, A.A. Evaluating the Prevalence and the Risk Factors of Gram-Negative Multi-Drug Resistant Bacteria in Eastern Saudi Arabia. *Infect. Drug Resist.* 2022, 15, 475–490. [CrossRef]
- Tamma, P.D.; Aitken, S.L.; Bonomo, R.A.; Mathers, A.J.; van Duin, D.; Clancy, C.J. Infectious Diseases Society of America 2023 Guidance on the Treatment of Antimicrobial Resistant Gram-Negative Infections. *Clin. Infect. Dis.* 2023, 2023, ciad428. [CrossRef] [PubMed]
- 29. Wang, M.; Wei, H.; Zhao, Y.; Shang, L.; Di, L.; Lyu, C.; Liu, J. Analysis of multidrug-resistant bacteria in 3223 patients with hospital-acquired infections (HAI) from a tertiary general hospital in China. *Bosn. J. Basic Med. Sci.* 2019, 19, 86–93. [CrossRef]
- Balkhair, A.; Al-Farsi, Y.M.; Al-Muharrmi, Z.; Al-Rashdi, R.; Al-Jabri, M.; Neilson, F.; Al-Adawi, S.S.; El-Beeli, M.; Al-Adawi, S. Epidemiology of Multi-Drug Resistant Organisms in a Teaching Hospital in Oman: A One-Year Hospital-Based Study. *Sci. World J.* 2014, 2014, 157102. [CrossRef]
- Wong, G.; Briscoe, S.; Adnan, S.; McWhinney, B.; Ungerer, J.; Lipman, J.; Roberts, J.A. Protein Binding of β-Lactam Antibiotics in Critically Ill Patients: Can We Successfully Predict Unbound Concentrations? *Antimicrob. Agents Chemother.* 2013, 57, 6165–6170. [CrossRef]
- 32. Craig, W.A. The Pharmacology of Meropenem, A New Carbapenem Antibiotic. Clin. Infect. Dis. 1997, 24, S266–S275. [CrossRef]
- Adnan, S.; Paterson, D.L.; Lipman, J.; Kumar, S.; Li, J.; Rudd, M.; Roberts, J.A. Pharmacokinetics of Beta-Lactam Antibiotics in Patients with Intra-Abdominal Disease: A Structured Review. *Surg. Infect.* 2012, 13, 9–17. [CrossRef] [PubMed]
- Alshaer, M.H.; Goutelle, S.; Santevecchi, B.A.; Shoulders, B.R.; Venugopalan, V.; Cherabuddi, K.; Liu, J.; Kiel, P.J.; Roberts, J.A.; Sime, F.B.; et al. Cefepime Precision Dosing Tool: From Standard to Precise Dose Using Nonparametric Population Pharmacokinetics. *Antimicrob. Agents Chemother.* 2022, 66, e0204621. [CrossRef] [PubMed]
- Mathew, S.K.; Mathew, B.S.; Neely, M.N.; Naik, G.S.; Prabha, R.; Jacob, G.G.; Subramani, K.; Fleming, D.H. A Nonparametric Pharmacokinetic Approach to Determine the Optimal Dosing Regimen for 30-Minute and 3-Hour Meropenem Infusions in Critically Ill Patients. *Ther. Drug Monit.* 2016, *38*, 593–599. [CrossRef] [PubMed]
- Felton, T.W.; Roberts, J.A.; Lodise, T.P.; Van Guilder, M.; Boselli, E.; Neely, M.N.; Hope, W.W. Individualization of Piperacillin Dosing for Critically Ill Patients: Dosing Software to Optimize Antimicrobial Therapy. *Antimicrob. Agents Chemother.* 2014, 58, 4094–4102. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.