

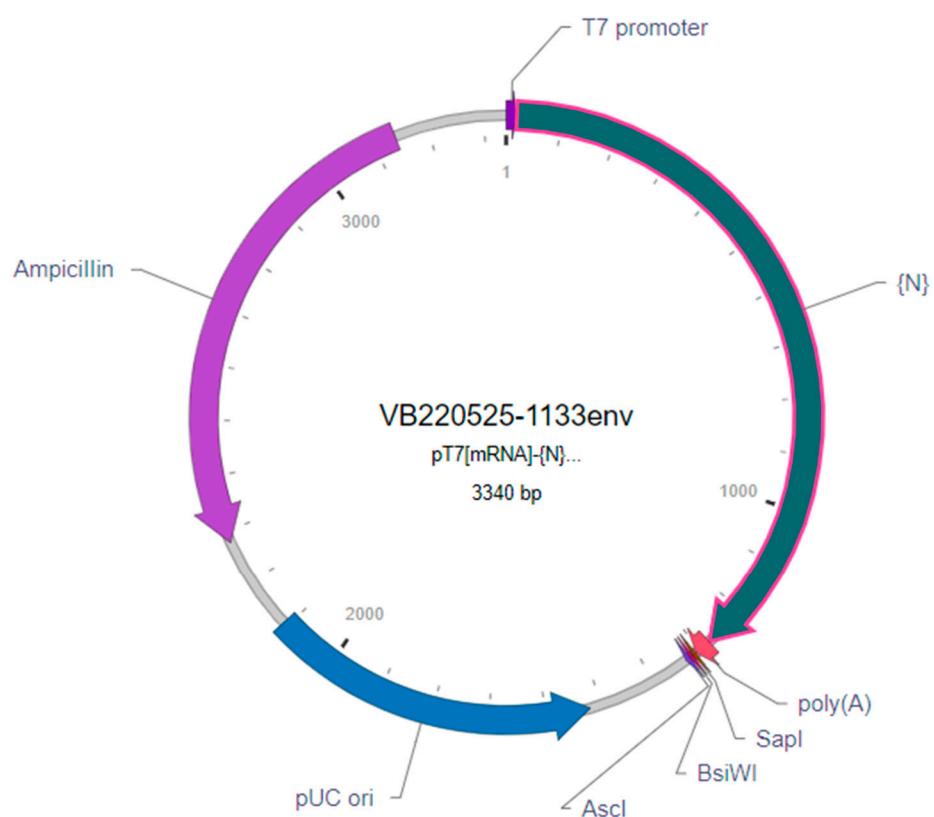
# **Simple, Visual, Point-of-Care SARS-CoV-2 Detection Incorporating Recombinase Polymerase Amplification and Target DNA–Protein Crosslinking Enhanced Chemiluminescence**

## **CONTENTS**

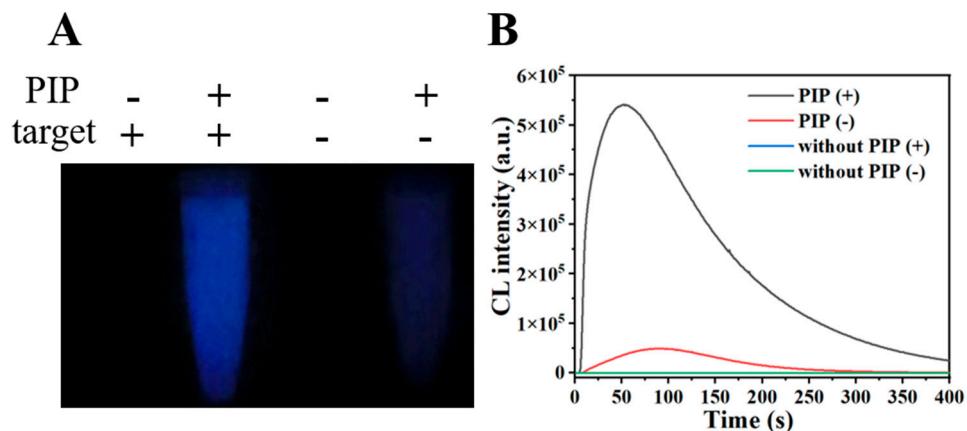
|  |    |
|--|----|
| <b>Table S1.</b> Detailed information of the oligonucleotide sequences used in this study.                                       | S2 |
| <b>Figure S1.</b> The designed SARS-CoV-2 N gene plasmid map.....  | S3 |
| <b>Figure S2.</b> Effect of the PIP enhancer on RPADPCL chemiluminescent signal strength .....                                   | S4 |
| <b>Figure S3.</b> Identification of SARS-CoV-2 N gene plasmid by 1% agarose gel electrophoresis. ....                            | S5 |
| <b>Figure S4.</b> Identification of RPA products for SARS-CoV-2 N gene plasmid detection by 1% agarose gel electrophoresis. .... | S5 |
| <b>Figure S5.</b> Identification of SARS-CoV-2 IVT RNA by 1% agarose gel electrophoresis. ....                                   | S5 |
| <b>Figure S6.</b> Identification of RPA products for IVT RNA detection by 1% agarose gel electrophoresis. ....                   | S6 |
| <b>Table S2.</b> Comparison between the proposed method and other RPA-based SARS-CoV-2 detection methods. ....                   | S7 |
| <b>Reference</b> .....   | S8 |

**Table S1.** Detailed information of the oligonucleotide sequences used in this study.

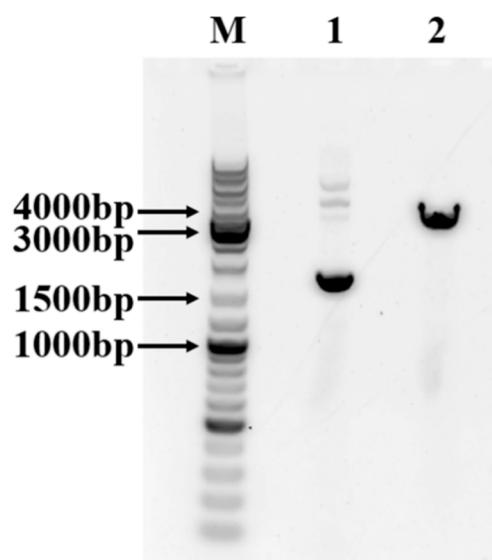
| Names                | Sequences (5'-3')   |
|----------------------|---|
| SARS-CoV-2 N<br>gene | ATGTCTGATAATGGACCCAAAATCAGCGAAATGCACCCCGCATTACGTTGGTGGAA<br>CCCTCAGATTCAACTGGCAGTAACCAGAATGGAGAACGAGTCAGTGGGCGCGATCAA<br>ACAACGTCGGCCCCAAGGTTACCAATAACTGCGTCTGGTCACCGCTCTCAC<br>TCAACATGGCAAGGAAGACCTTAAATTCCCTCGAGGACAAGGCCTCCAATTAAACA<br>CCAATAGCAGTCCAGATGACCAAATTGGCTACTACCGAAGAGCTACCAAGACGAATT<br>CGTGGTGGTGACGGTAAAATGAAAGATCTCAGTCCAAGATGGTATTCTACTACCTA<br>GGAACCTGGGCCAGAACAGCTGGACTTCCCTATGGTGCTAACAAAGACGGCATCATATG<br>GGTGCAACTGAGGGAGCCTTGAATACACAAAAGATCACATTGGCACCCGCAATC<br>CTGCTAACAAATGCTGCAATCGTCTACAACCTCCTCAAGGAACAAACATTGCCAAAAG<br>GCTTCTACGCAGAACAGGGAGCAGAGGCGGAGTCAGCCTCTCGTCCCTCATCAC<br>GTAGTCGCAACAGTTCAAGAAATTCAACTCCAGGCAGCAGTAGGGAAACTTCTCCT<br>GCTAGAAATGGCTGGCAATGGCGGTGATGCTGCTCTGCTTGCTGCTGCTGACAGA<br>TTGAACCAGCTTGAGAGCAAAATGTCGGTAAAGGCCAACAAACAACAGGCCAAAC<br>TGTCACTAAGAAATCTGCTGCTGAGGCTCTAAGAACGCTCGGAAAAACGTACTGC<br>CACTAAAGCATAACAATGTAACACAAGCTTCGGCAGACGTGGTCCAGAACAAACCC<br>AAGGAAATTGGGGACCAGGAACTAATCAGACAAGGAACGTGATTACAAACATTGG<br>CCGCAAATTGCAAAATTGCCCCCAGCGCTTCAGCGTTCTCGGAATGTCGCGCATT<br>GGCATGGAAGTCACACCTCGGGAACGTGGTTGACCTACACAGGTGCCATCAAATT<br>GGATGACAAAGATCCAATTCAAAGATCAAGTCATTGCTGAATAAGCATATTGA<br>CGCATAACAAACATTCCCACCAACAGAGCCTAAAAGGACAAAAGAAGAAGGCT<br>GATGAAACTCAAGCCTTACCGCAGAGACAGAACAGCAAACGTGACTCTCT<br>TCCTGCTGCAGATTGGATGATTCTCAAACAATTGCAACAATCCATGAGCAGTGC<br>TGACTCAACTCAGGCCTAA |
| SARS-2 FP            | CAACTTCCTCAAGGAACAAACATTGCCAAAA   |
| SARS-2 RP            | TGGAGTTGAATTCTTGAACGTGGCGACT  |
| SARS-2 FP-biotin     | biotin-CAACTTCCTCAAGGAACAAACATTGCCAAAA  |
| SARS-2 RP-biotin     | biotin-TGGAGTTGAATTCTTGAACGTGGCGACT   |



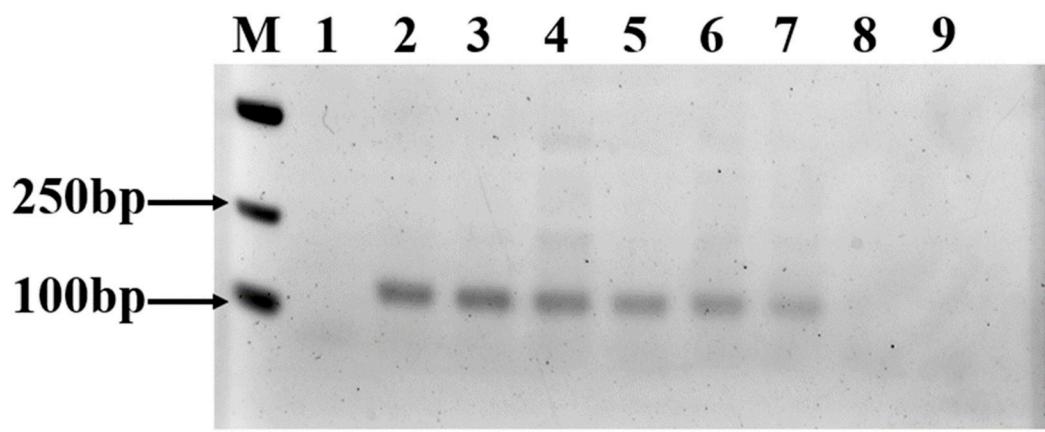
**Figure S1.** The designed SARS-CoV-2 N gene plasmid map.



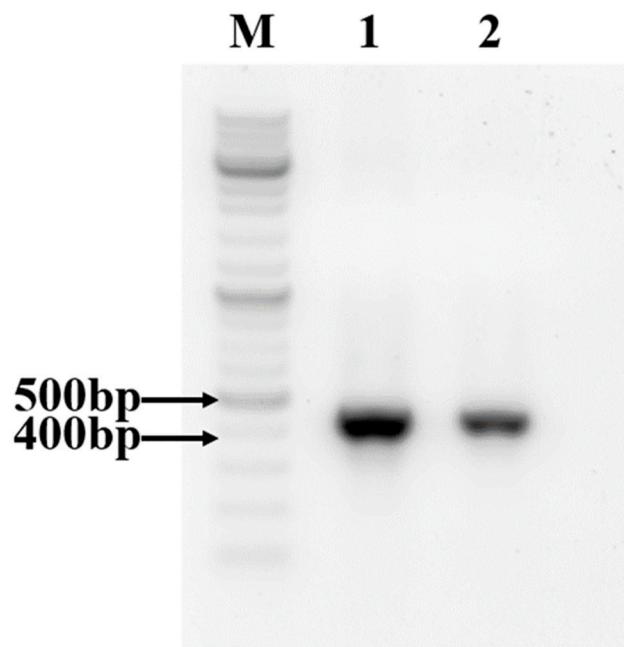
**Figure S2.** Effect of the PIP enhancer on RPADPCL chemiluminescent signal strength.



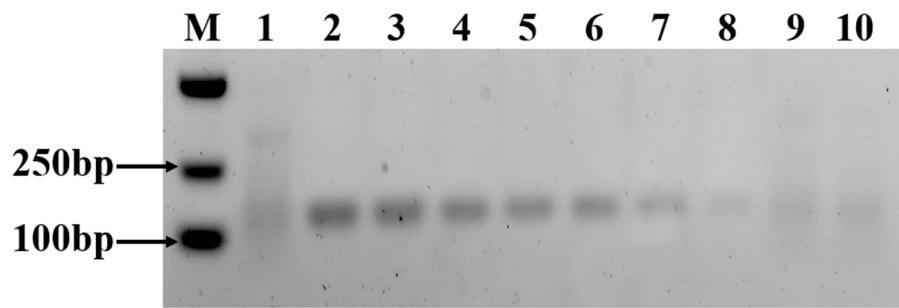
**Figure S3.** Identification of SARS-CoV-2 N gene plasmid by 1% agarose gel electrophoresis. M: DNA marker; Lane 1: The designed SARS-CoV-2 N gene plasmid. Lane 2: The plasmid after AscI enzyme digestion.



**Figure S4.** Identification of RPA products for SARS-CoV-2 N gene plasmid detection by 1% agarose gel electrophoresis. M: DNA marker; Lane 1: negative control; Lane 2-9: The concentration of SARS-CoV-2 N gene plasmid was 1000, 500, 300, 200, 100, 50, 10, 1 copies, respectively.



**Figure S5.** Identification of SARS-CoV-2 IVT RNA by 1% agarose gel electrophoresis. M: DNA marker; Lane 1: The IVT RNA synthetized by RiboMAX Large Scale RNA Production System Kit. Lane 2: The IVT RNA synthetized by T7 Quick High Yield RNA Transcription Kit.



**Figure S6.** Identification of RPA products for IVT RNA detection by 1% agarose gel electrophoresis.

M: DNA marker; Lane 1: negative control; Lane 2-10: The concentration of SARS-CoV-2 N gene plasmid was 2000, 1000, 500, 300, 200, 100, 50, 10, 1 copies, respectively.

**Table S2.** Comparison between the proposed method and other RPA-based SARS-CoV-2 detection methods.

| No. | Methods                          | Target genes  | Sensitivity<br>(LOD) | Quantification | Visualization | Detection time | Ref.      |
|-----|----------------------------------|---------------|----------------------|----------------|---------------|----------------|-----------|
| 1   | Strip                            | N/ORF1ab gene | 10 copies            | ×              | √             | 60 min         | [1]       |
| 2   | CRISPR/Cas9-mediated strip       | E/ORF1ab gene | 100 copies           | ×              | √             | 58 min         | [2]       |
| 3   | Strip                            | N/S gene      | 10 copies            | ×              | √             | 45 min         | [3]       |
| 4   | Microfluidic-integrated strip    | N gene        | 30 copies            | ×              | √             | 20 min         | [4]       |
| 5   | Strip                            | N gene        | 35.4 copies          | ×              | √             | 45 min         | [5]       |
| 6   | CRISPR/Cas fluorometry           | N/ORF1ab gene | 2 copies             | √              | ×             | 50 min         | [6]       |
| 7   | Colorimetric CRISPR/Cas12a assay | N/ORF1ab gene | 1 copy               | √              | ×             | 240 min        | [7]       |
| 8   | Real-time RPA                    | N gene        | 10 copies            | √              | ×             | 27 min         | [8]       |
| 9   | Real-time RPA                    | ORF1ab/S gene | 10 copies            | √              | ×             | 24 min         | [9]       |
| 10  | Real-time RPA                    | N/E/RdRP gene | 15 copies            | √              | ×             | 15 min         | [10]      |
| 11  | Fluorescent strip                | E/RdRP gene   | 9.5 copies           | √              | ×             | 30 min         | [11]      |
| 12  | CRISPR/Cas-based lab-on-paper    | N/S gene      | 100 copies           | √              | ×             | 60 min         | [12]      |
| 13  | Chemiluminometry                 | N gene        | 15 copies            | √              | √             | 60 min         | This work |

## Reference

1. Sun, Y.; Qin, P.; He, J.; Li, W.; Shi, Y.; Xu, J.; Wu, Q.; Chen, Q.; Li, W.; Wang, X., Rapid and simultaneous visual screening of SARS-CoV-2 and influenza viruses with customized isothermal amplification integrated lateral flow strip. *Biosens. Bioelectron.* **2022**, 197, 113771.
2. Xiong, E.; Jiang, L.; Tian, T.; Hu, M.; Yue, H.; Huang, M.; Lin, W.; Jiang, Y.; Zhu, D.; Zhou, X., Simultaneous dual-gene diagnosis of SARS-CoV-2 based on CRISPR/Cas9-mediated lateral flow assay. *Angew. Chem. Int. Edit.* **2021**, 60, (10), 5307-5315.
3. Qian, J.; Boswell, S. A.; Chidley, C.; Lu, Z.-x.; Pettit, M. E.; Gaudio, B. L.; Fajnzylber, J. M.; Ingram, R. T.; Ward, R. H.; Li, J. Z., An enhanced isothermal amplification assay for viral detection. *Nat. Commun.* **2020**, 11, (1), 5920.
4. Liu, D.; Shen, H.; Zhang, Y.; Shen, D.; Zhu, M.; Song, Y.; Zhu, Z.; Yang, C., A microfluidic-integrated lateral flow recombinase polymerase amplification (MI-IF-RPA) assay for rapid COVID-19 detection. *Lab Chip* **2021**, 21, (10), 2019-2026.
5. Shelite, T. R.; Uscanga-Palomeque, A. C.; Castellanos-Gonzalez, A.; Melby, P. C.; Travi, B. L., Isothermal recombinase polymerase amplification-lateral flow detection of SARS-CoV-2, the etiological agent of COVID-19. *J. Virol. Methods* **2021**, 296, 114227.
6. Huang, Z.; Tian, D.; Liu, Y.; Lin, Z.; Lyon, C. J.; Lai, W.; Fusco, D.; Drouin, A.; Yin, X.; Hu, T., Ultra-sensitive and high-throughput CRISPR-powered COVID-19 diagnosis. *Biosens. Bioelectron.* **2020**, 164, 112316.
7. Zhang, W. S.; Pan, J.; Li, F.; Zhu, M.; Xu, M.; Zhu, H.; Yu, Y.; Su, G., Reverse transcription recombinase polymerase amplification coupled with CRISPR-Cas12a for facile and highly sensitive colorimetric SARS-CoV-2 detection. *Anal. Chem.* **2021**, 93, (8), 4126-4133.
8. Wu, T.; Ge, Y.; Zhao, K.; Zhu, X.; Chen, Y.; Wu, B.; Zhu, F.; Zhu, B.; Cui, L., A reverse-transcription recombinase-aided amplification assay for the rapid detection of N gene of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). *Virology* **2020**, 549, 1-4.
9. Xue, G.; Li, S.; Zhang, W.; Du, B.; Cui, J.; Yan, C.; Huang, L.; Chen, L.; Zhao, L.; Sun, Y., Reverse-transcription recombinase-aided amplification assay for rapid detection of the 2019 novel coronavirus (SARS-CoV-2). *Anal. Chem.* **2020**, 92, (14), 9699-9705.
10. El Wahed, A. A.; Patel, P.; Maier, M.; Pietsch, C.; Rüster, D.; Böhlken-Fascher, S.; Kissenkötter,

- J.; Behrmann, O.; Frimpong, M.; Diagne, M. M., Suitcase lab for rapid detection of SARS-CoV-2 based on recombinase polymerase amplification assay. *Anal. Chem.* **2021**, 93, (4), 2627-2634.
11. Cherkaoui, D.; Huang, D.; Miller, B. S.; Turbé, V.; McKendry, R. A., Harnessing recombinase polymerase amplification for rapid multi-gene detection of SARS-CoV-2 in resource-limited settings. *Biosens. Bioelectron.* **2021**, 189, 113328.
12. Yin, K.; Ding, X.; Li, Z.; Sfeir, M. M.; Ballesteros, E.; Liu, C., Autonomous lab-on-paper for multiplexed, CRISPR-based diagnostics of SARS-CoV-2. *Lab Chip* **2021**, 21, (14), 2730-2737.