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EXPERIMENTAL SECTION

Chemicals and Materials. 3,3',5,5'-tetramethylbenzidine (TMB), ascorbic acid (AA), glutathione (GSH), tryptophan (Trp), arginine (Arg), and glutamic acid (Glu) were obtained from Sigma-Aldrich (Shanghai, China). Papain (Pap), pepsin (Pep), lysozyme (Lys), deoxyribonuclease (DNase), and acetylcholinesterase (AChE) were obtained from Beyotime Biotechnology (Shanghai, China). Ferrocene (Fc), and zinc nitrate hexahydrate ($\text{Zn}(\text{NO}_3)_2$, 99.5%), were purchased from Alfa Aesar (Shanghai, China). 2-Methylimidazole (98%), 30% hydrogen peroxide (H_2O_2), acetic acid (HAc), sodium acetate (NaAc), and sulfuric acid (H_2SO_4) (98.08%) were purchased from Adamas-beta (Shanghai, China). Methanol (99.5%) and ethanol (99.7%) were purchased from General-reagent (Shanghai, China). Ultrapure water was used in the experiments involving aqueous solution. All chemicals were used without further purification. Ultrapure water with a resistivity of $18 \text{ M}\Omega \cdot \text{cm}^{-1}$ was used in the experiments.

Apparatus. UV-vis spectra were recorded by using UV-2600 spectrometer (Shimadzu, Japan). The pore parameters such as Brunauer-Emmett-Teller (BET) and Barrett-Joyner-Halenda (BJH) pore size distribution were measured using N_2 adsorption/desorption at 393 K on a Quantachrome Autosorb IQ3 (USA). Transmission electron microscopy (TEM) image and high-angle annular dark-field scanning transmission electron microscopy (HAADF-STEM) image were obtained on Talos F200S G2 transmission electron microscope (Thermo Fisher Scientific, America). Scanning electron microscopy (SEM) image was photographed by TESCAN MIRA3 LM field emission scanning electron microscope (TESCAN, Czech). Powder X-ray diffraction (XRD) patterns were measured by X'Pert3 Powder X-ray diffractometer (PANalytical, Netherlands). X-ray Photoelectron Spectroscopy (XPS) spectra were obtained by K-Alpha X-ray photoelectron spectrometer (Thermo Fisher Scientific, America).

Pretreatment of serum samples. Firstly, 0.3 μL serum was incubated with 0.2 μL N-Ethylmaleimide NEM (100 mM) for 2 h at room temperature. Then, centrifugated the mixed solution for 10 min. Finally, transferred the supernatant as pretreated serum to prepare for the next experiment.

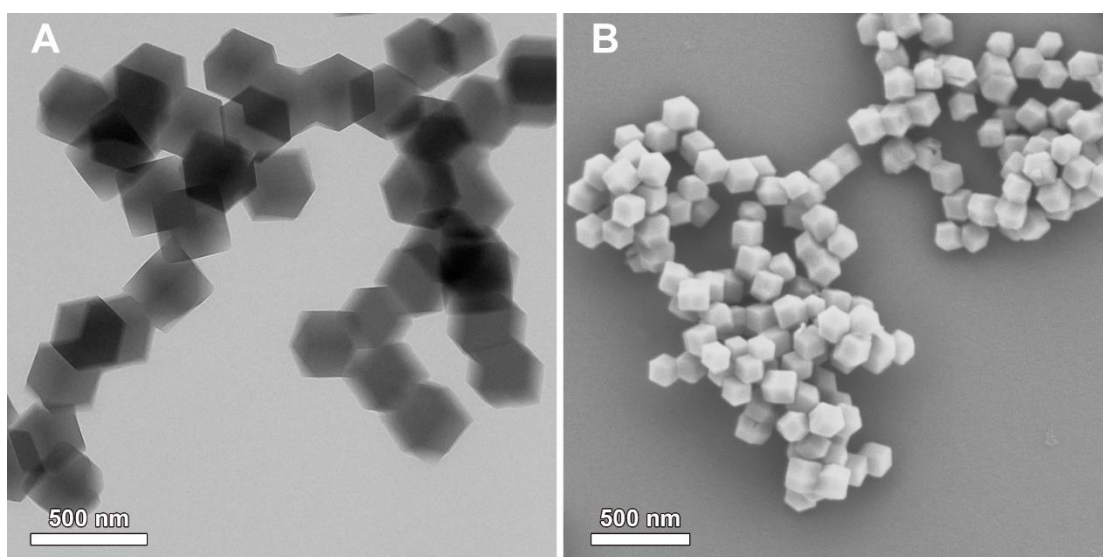


Figure S1. TEM (A) and SEM (B) images of ZIF-8.

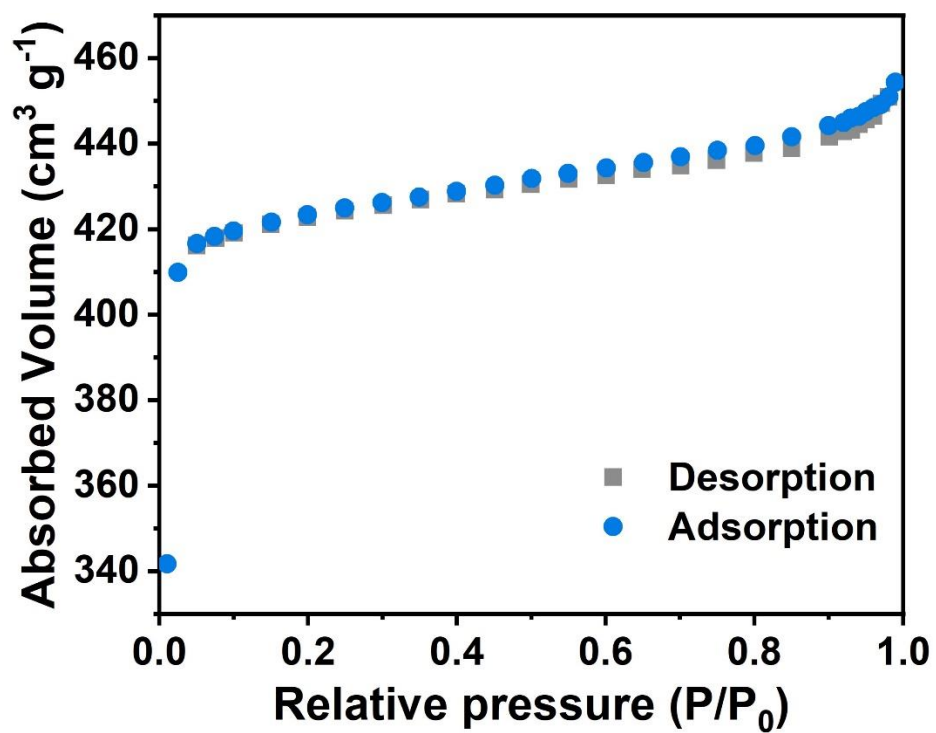


Figure S2. N₂ adsorption/desorption isotherms of ZIF-8.

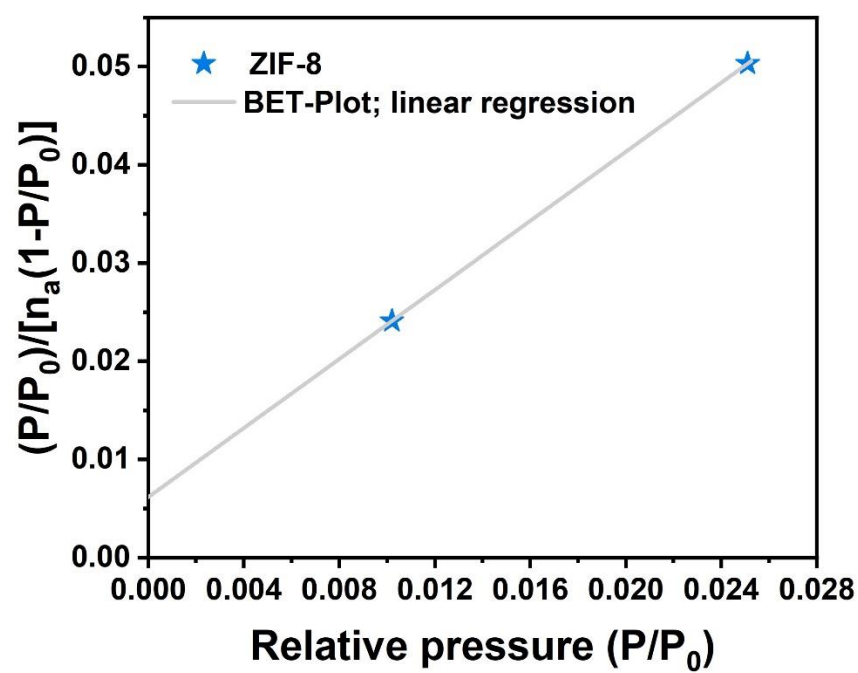


Figure S3. BET-Plot with linear regression for ZIF-8.

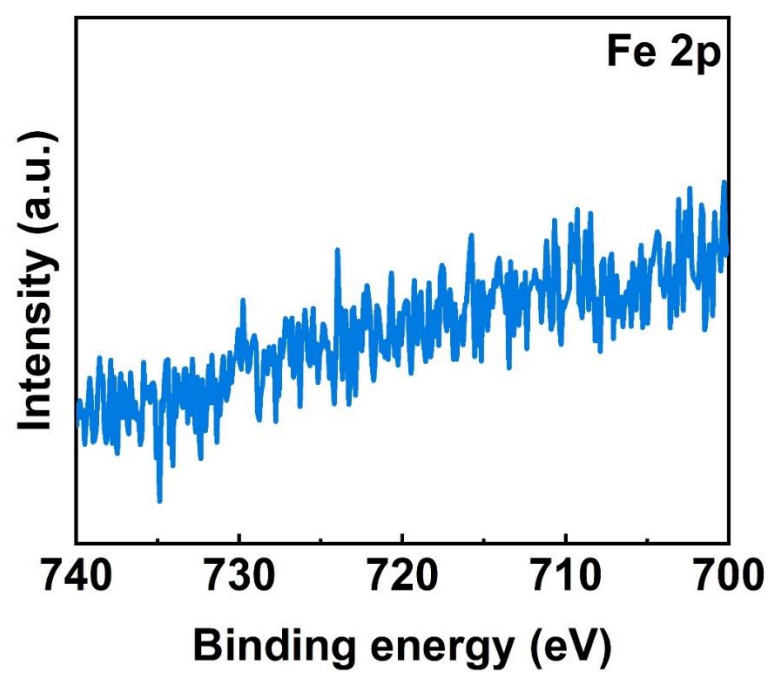


Figure S4. XPS spectrum for Fe 2p.

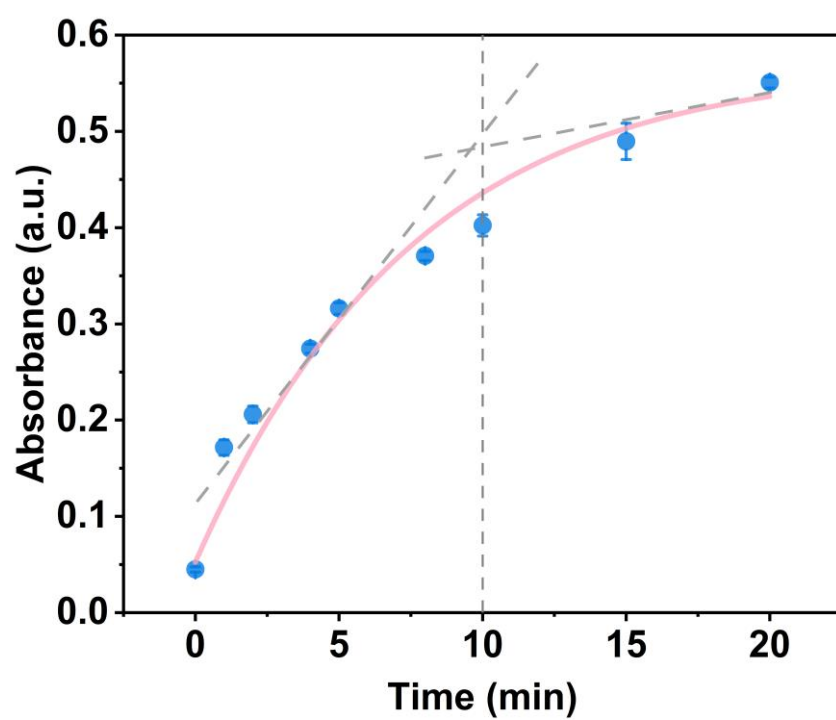


Figure S5. Effect of reaction time on oxidase-like activity of Fe-N-C.

Table S1. Atomic percentage of the Fe-N-C.

Sample	Fe 2p (%)	N 1s (%)	C 1s (%)	O 1s (%)
Fe-N-C	0.07	7.23	86.45	6.25

Table S2. Comparison of kinetic parameters for Fe-N-C and other oxidase mimics.

Catalyst	K_m (mM)	V_{max} (10^{-8} M s $^{-1}$)	Ref.
^a Fe-N/C-CNT (Fe-N ₃)	0.13	2.25	[1]
^b Fe-N-C SAC	1.81	0.601	[2]
^c Fe-N-C-400	0.269	33.80	[3]
Co-SAC	0.578	34.5	[4]
Ni-N/C-CNTs	0.284	0.245	[5]
Fe-N/C-CNTs	0.62	52.6	
Co-N/C-CNTs	1.17	20.8	
FeBi-NC	0.21	0.7	[6]
Glucose oxidase	12.1	60.1	[7]
Fe-N-C	0.253	4.136	This work

^a Fe-N/C-CNT (Fe-N₃): ferrum-nitrogen-carbon carbon nanotube (with Fe-N₃ units);

^b Fe-N-C SAC: Fe-N-C single-atom catalyst;

^c Fe-N-C-400: Fe-N-C under carbonization at 400 °C.

Table S3. Comparison of the sensing performance of our proposed method with other reported AA sensors.

Nanomaterial	Method	Linear range	LOD	Ref
CuO/Pt	Colorimetry	1 μ M–0.6 mM	0.796 μ M	[8]
MOF-808	Colorimetry	30 μ M–1 mM	15 μ M	[9]
^a CL/TiN/GCE	Electrochemistry	50–1500 μ M	1.52 μ M	[10]
^b ITO/rGO/AuNPs	Electrochemistry	20–150 μ M	9.4 μ M	[11]
^c PPy@C/GCE	Electrochemistry	10–150 μ M	0.75 μ M	[12]
Cu-Ag/rGO nanocomposite	Colorimetry	5–30 μ M	3.6 μ M	[13]
^d PPYNTs/Co ₃ O ₄	Electrochemistry	5–80 μ M	0.23 μ M	[14]
^e PMo ₁₂ -PANI/ β -CD/ITO	Electrochemistry	0.01–3000 μ M	0.015 μ M	[15]
Fe-N-C	Colorimetry	0.1–2 μ M	0.123 μ M	This work

^a CL/TiN/GCE: glassy carbon electrode modified with chrysanthemum-like titanium nitride nanostructures;

^b ITO/rGO/AuNPs: indium tin oxide-based electrodes with both reduced graphene oxide and Au nanoparticles;

^c PPy@C/GCE: glassy carbon electrode modified with polypyrrole grafted cellulose;

^d PPYNTs/Co₃O₄: polypyrrole nanotubes with Co₃O₄ nano powder;

^e PMo₁₂-PANI/ β -CD/ITO: polyaniline (PANI) and phosphomolybdic acid (PMo₁₂) co-polymerized on the film of β -cyclodextrin (β -CD) modified ITO.

Table S4. Comparison of the sensing performance of our proposed method with other reported GSH sensors.

Nanomaterial	Method	Linear range	LOD	Ref
Gold nanoclusters	Colorimetry	2–25 μM	0.42 μM	[16]
^a COF-300-AR	Colorimetry	1–15 μM	1.0 μM	[17]
^b N, S-CQDs	Fluorescence	0–100 μM	6.7 μM	[18]
^c MnO ₂ -induced PDA-NPs	Fluorescence	0–350 μM	1.5 μM	[19]
^d PDI-C ₄ SH	Electrochemistry	50–5000 μM	17 μM	[20]
^e Receptor 1	Colorimetry	60–120 μM	5.86 μM	[21]
^f Rho-GSH	Fluorescence	0–5000 μM	49.6 μM	[22]
Fe-N-C	Colorimetry	1–10 μM	1.31 μM	This work

^a COF-300-AR: reductive product of COF-300;

^b N, S-CQDs: nitrogen and sulfur co-doped carbon quantum dots;

^c MnO₂-induced PDA-NPs: MnO₂-induced polydopamine nanoparticles;

^d PDI-C₄SH: *N, N'*-2-octyloxy-1-methyl-2-oxoethyl)-(-2,4-thiolbutyloxy)-1-methyl-2-oxoethyl)-3,4:9,10-perylenetetracarboxyldiimide;

^e Receptor 1: (E)-5-(diethylamino)-2-(((2-(methylthio) phenyl)imino)methyl)phenol;

^f Rho-GSH: 3', 6'-bis(ethylamino)-2', 7'-dimethyl-2-(4-nitrophenoxy) spiro [isoindoline-1, 9' -xanthen]-3-one.

Table S5. Results for the determination of AA in normal human serum samples.

Samples	AA spiked (μM)	Total detected (μM)	Recovery (%)	RSD (% , n = 3)
Serum 1	0.5	0.54	108.4	5.3
	1	1.10	109.6	5.4
	1.5	1.44	96.1	6.3
Serum 2	0.5	0.49	97.5	1.6
	1	1.12	111.9	0.7
	1.5	1.38	97.9	0.6
Serum 3	0.5	0.61	110.0	0.4
	1	1.11	111.5	1.9
	1.5	1.49	99.1	1.1

Table S6. Results for the determination of GSH in normal human serum samples.

Samples	GSH spiked (μM)	Total detected (μM)	Recovery (%)	RSD (% , n = 3)
Serum 1	0	1.23	-	0.8
	2	3.18	98.4	1.5
	4	5.11	97.8	1.9
	8	9.29	100.7	2.5
Serum 2	0	1.25	-	1.4
	2	3.14	96.7	2.3
	4	5.02	95.7	0.8
	8	9.41	101.7	2.9
Serum 3	0	1.55	-	0.7
	2	3.64	102.7	1.9
	4	5.54	99.8	0.2
	8	9.49	99.4	1.3

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