

# A point-of-care testing device utilizing graphene-enhanced fiber optic SPR sensor for real-time detection of infectious pathogens

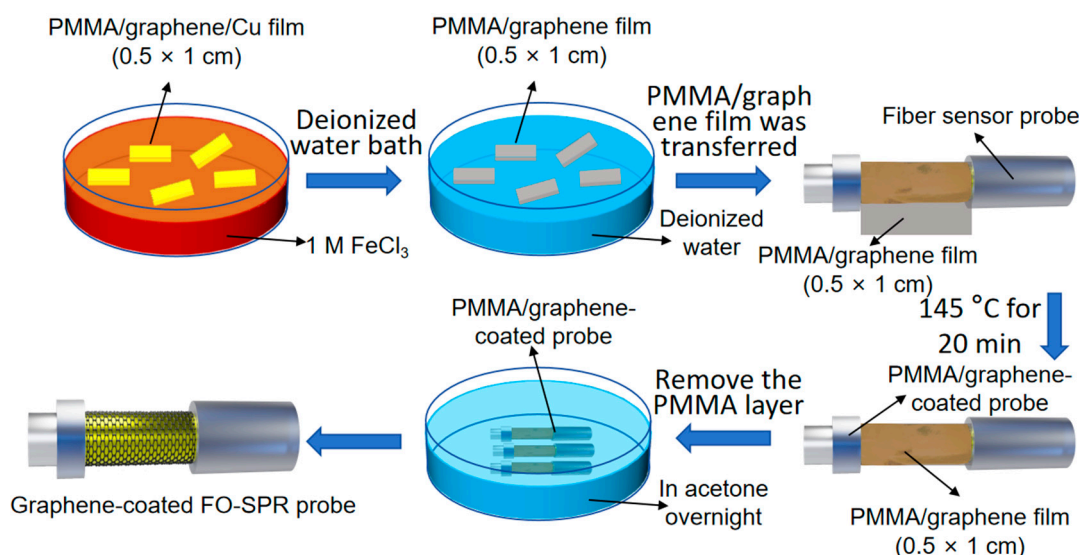
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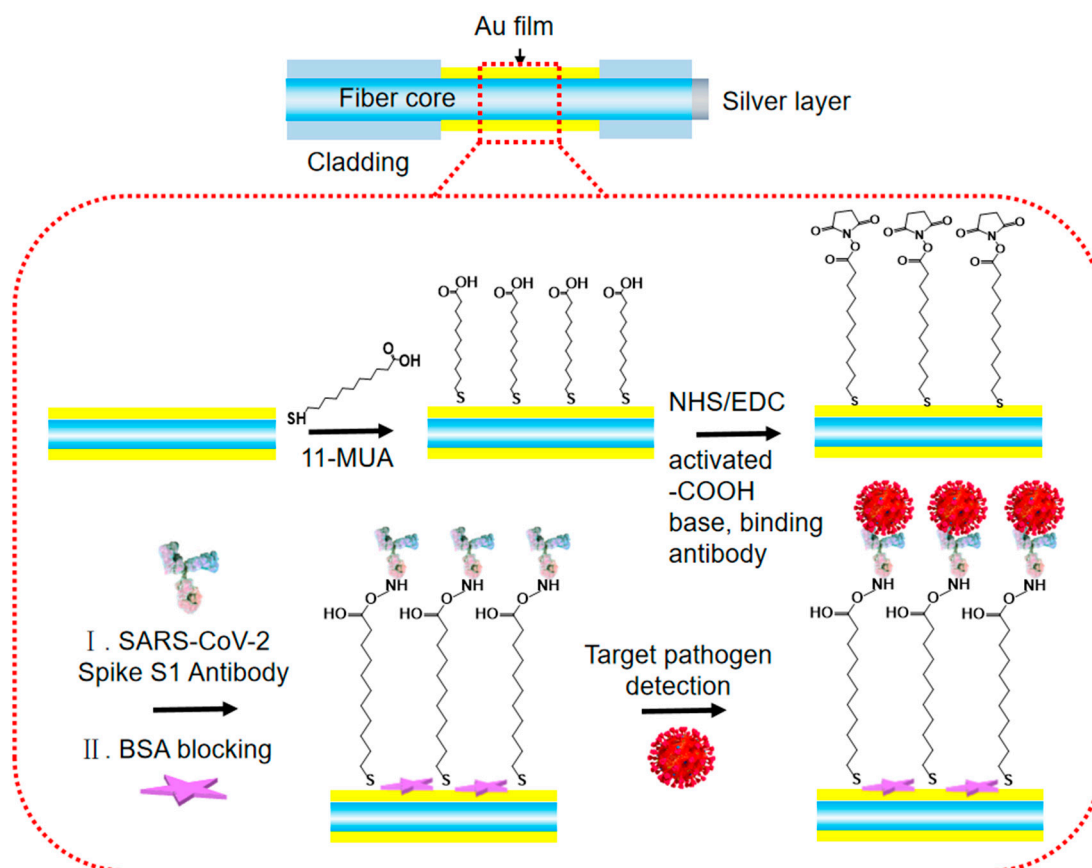
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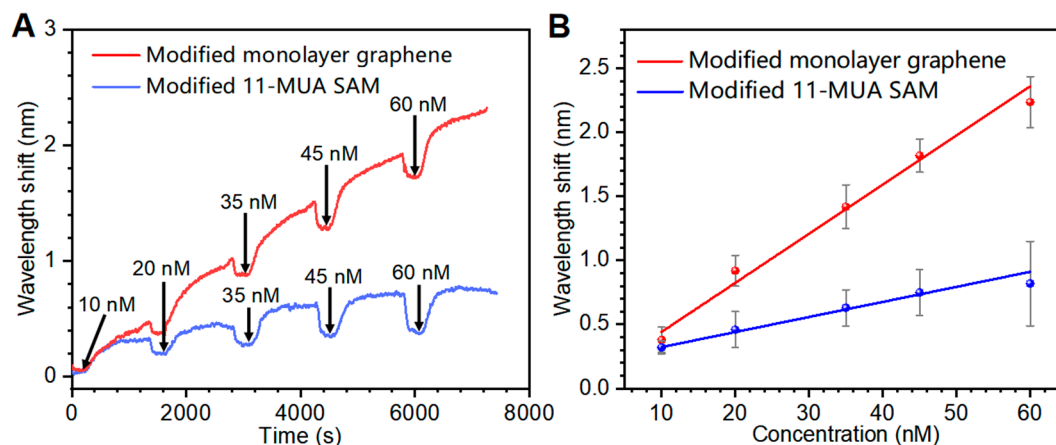
**Figure S1.** Scheme of the graphene-coated FO-SPR sensing probe.

## 11-MUA-modified FO-SPR sensing probes.

The FO-SPR sensing probe with bare gold was immersed in 1 mM 11-MUA in ethanol at room temperature overnight and alternately rinsed with ethanol and deionized water. Then, to activate the carboxyl group, the FO-SPR probe was placed in an aqueous 0.5 M NHS/EDC solution for 20 min and rinsed with deionized water. The sensing probe was immersed in 250 µg/mL SARS-CoV-2 spike S1 antibody in PBS for 45 min. Finally, the probe was put in 1 mg/mL BSA for 1 h.



**Figure S2.** Scheme of the bio-functionalized FO-SPR sensing probe.



**Figure S3.** Testing SARS-CoV-2 spike S1 protein via portable FO-SPR device. (A) The graphene-coated FO-SPR sensing probe wavelength shift curve (red) and the conventional 11-MUA-modified FO-SPR sensing probe wavelength shift curve (blue). (B) The graphene-coated FO-SPR sensing probe (red) and the conventional 11-MUA-modified FO-SPR sensing probe (blue) were linearly fitted for different protein concentrations and wavelength shifts. Error bars were determined by the standard deviation of three tests.

**Table S1.** RSD for repeatability analysis.

SARS-CoV-2 spike S1 protein concentration	Relative standard deviation (RSD)	Repeatability (100%-RSD )
10 nM	22.8	77.2
20 nM	15.5	84.5
35 nM	12.2	87.8
45 nM	8.6	91.4
60 nM	6.8	93.2

**Table S2.** Comparison of SPR methods for biosensing analysis.

Method	Type of receptor	Target analyte	Linear range	Limit of detection	Ref
SPR	Escherichia coli	Phage T4	0 PFU/mL - $2 \times 10^8$ PFU/mL	$1 \times 10^7$ PFU/mL	[1]
SPR	polyclonal antibody	Salmonella enterica	-	$10^5$ CFU/mL	[2]
LSPR	Casein	Trypsin	0.1 $\mu$ g/mL - 100 $\mu$ g/mL	4.3 nM	[3]
SPR	Hb antibody	Hb	5 $\mu$ g/mL - 250 $\mu$ g/mL	0.078 $\mu$ M	[4]
SPR	Antibody	HIV-1 p24 capsid protein	-	$4.1 \pm 0.5$ nM	[5]
FO-SPR	SPA	IgG	30 $\mu$ g/mL - 100 $\mu$ g/mL	0.5 $\mu$ g/mL	[6]
FO-SPR	DNA-aptamer	SARS-CoV-2 spike protein	25 nM - 1 $\mu$ M	37 nM	[7]
FO-SPR	Anti-ADM monoclonal antibody	ADM	-	0.25 $\mu$ g/mL	[8]
FO-SPR POCT	SARS-CoV-2 spike S1 protein antibody	SARS-CoV-2 spike S1 protein SARS-CoV-2 virus	10 nM - 60 nM $5 \times 10^3$ TCID <sub>50</sub> /mL - $6 \times 10^4$ TCID <sub>50</sub> /mL	2.5 nM $2.2 \times 10^3$ TCID <sub>50</sub> /mL	This work

LSPR: Localized surface plasmon resonance; HIV: Human immunodeficiency virus; SPA: Staphylococcal protein A; Hb: hemoglobin; IgG: Human immunoglobulin G; ADM: Adalimumab.

## References

- [1] Xiao, C.Q.; Jiang, F.L.; Zhou, B.; Li, R.; Liu, Y. Immobilization of Escherichia coli for detection of phage T4 using surface plasmon resonance. *Science China Chemistry* 2012, 55, 1931-1939. <https://doi.org/10.1007/s11426-012-4553-6>.
- [2] Makhneva, E.; Farka, Z.; Skládal, P.; Zajíčková, L. Cyclopropylamine plasma polymer surfaces for label-free SPR and QCM immunosensing of Salmonella. *Sensors and Actuators B: Chemical* 2018, 276, 447-455. <https://doi.org/10.1016/j.snb.2018.08.055>.
- [3] Svärd, A.; Neilands, J.; Palm, E.; Svensäter, G.; Bengtsson, T.; Aili, D. Protein-Functionalized Gold Nanoparticles as Refractometric Nanoplasmonic Sensors for the Detection of Proteolytic Activity of Porphyromonas gingivalis. *ACS Applied Nano Materials* 2020, 3, 10, 9822-9830. <https://doi.org/10.1021/acsanm.0c01899>.
- [4] Inci, F.; Saylan, Y.; Kojouri, A.M.; Ogut, M.G.; Denizli, A.; Demirci, U. A disposable microfluidic-integrated hand-held plasmonic platform for protein detection. *Applied Materials Today* 2020, 18, 100478. <https://doi.org/10.1016/j.apmt.2019.100478>.
- [5] Sarcina, L.; Mangiatordi, G.F.; Torricelli, F.; Bollella, P.; Gounani, Z.; Österbacka, R.; Macchia, E.; Torsi, L. Surface Plasmon Resonance Assay for Label-Free and Selective Detection of HIV-1 p24 Protein. *Biosensors* 2021, 11, 6, 180. <https://doi.org/10.3390/bios11060180>.
- [6] Wang, Q.; Jing, J.Y.; Wang, B.T. Highly Sensitive SPR Biosensor Based on Graphene Oxide and Staphylococcal Protein A Co-Modified TFBG for Human IgG Detection. *IEEE Transactions on Instrumentation and Measurement* 2019, 68, 9, 3350-3357. <https://doi.org/10.1109/TIM.2018.2875961>.
- [7] Cennamo, N.; Pasquardini, L.; Arcadio, F.; Lunelli, L.; Vanzetti, L.; Carafa, V.; Altucci, L.; Zeni, L. SARS-CoV-2 spike protein detection through a plasmonic D-shaped plastic optical fiber aptasensor. *Talanta* 2021, 233, 122532. <https://doi.org/10.1016/j.talanta.2021.122532>.
- [8] Qu, J.H.; Ordutowski, H.; Tricht, C.V.; Verbruggen, R.; Gallardo, A.B.; Bulcaen, M.; Ciwinka, M.; Cisneros, C.C.; Devriese, C.; Guluzade, S.; Janssens, X.; Kornblum, S.; Lu, Y.H.; Marolt, N.; Nanjappan, C.; Rutten, E.; Vanhauwaert, E.; Geukens, N. Thomas, D.; Dosso, F.D.; Safdar, S.; Spasic, D.; Lammertyn, J. Point-of-care therapeutic drug monitoring of adalimumab by integrating a FO-SPR biosensor in a self-powered microfluidic cartridge. *Biosensors and Bioelectronics* 2022, 206, 114125. <https://doi.org/10.1016/j.bios.2022.114125>.