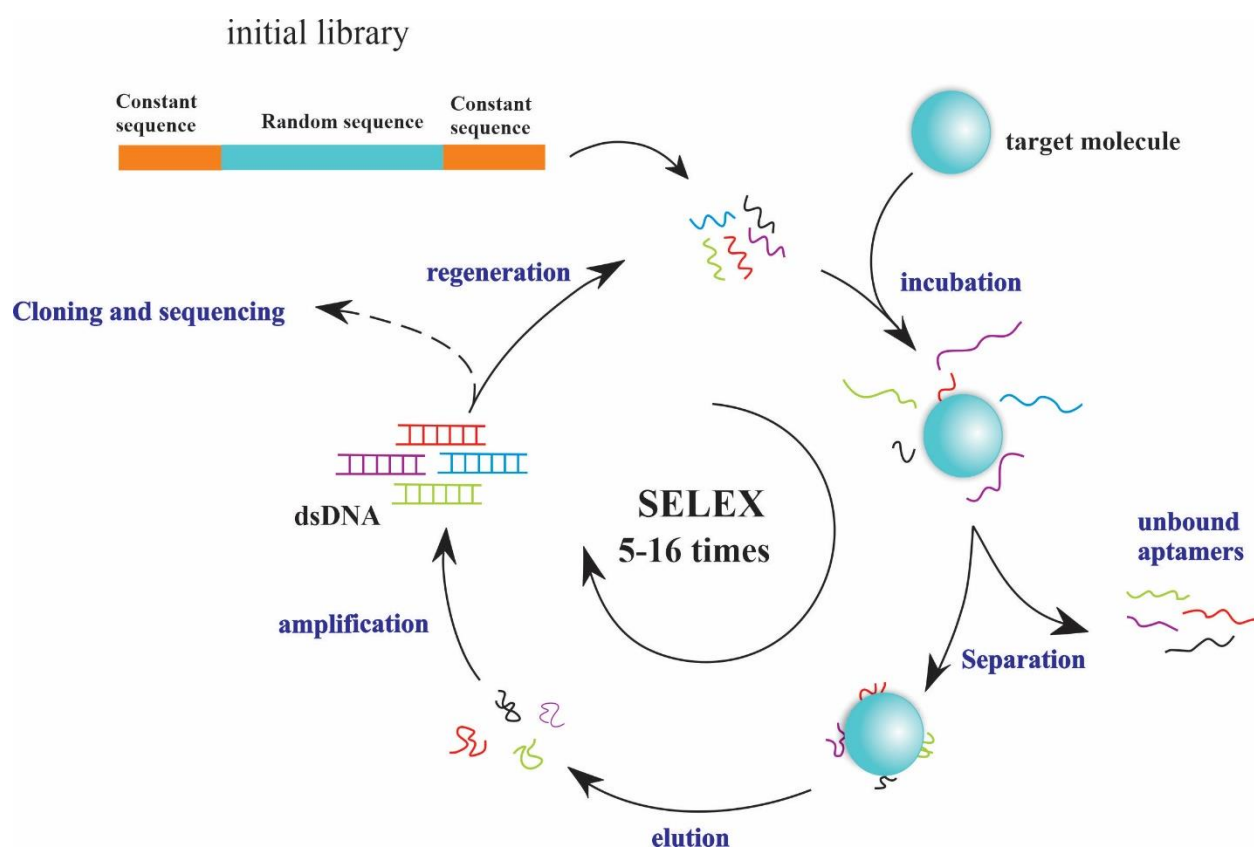


Figure S1. Chemical structure of Rivaroxaban



Scheme S1. A schematic outline of the SELEX procedure using modified RIV-Sepharose toward aptamer discovery.

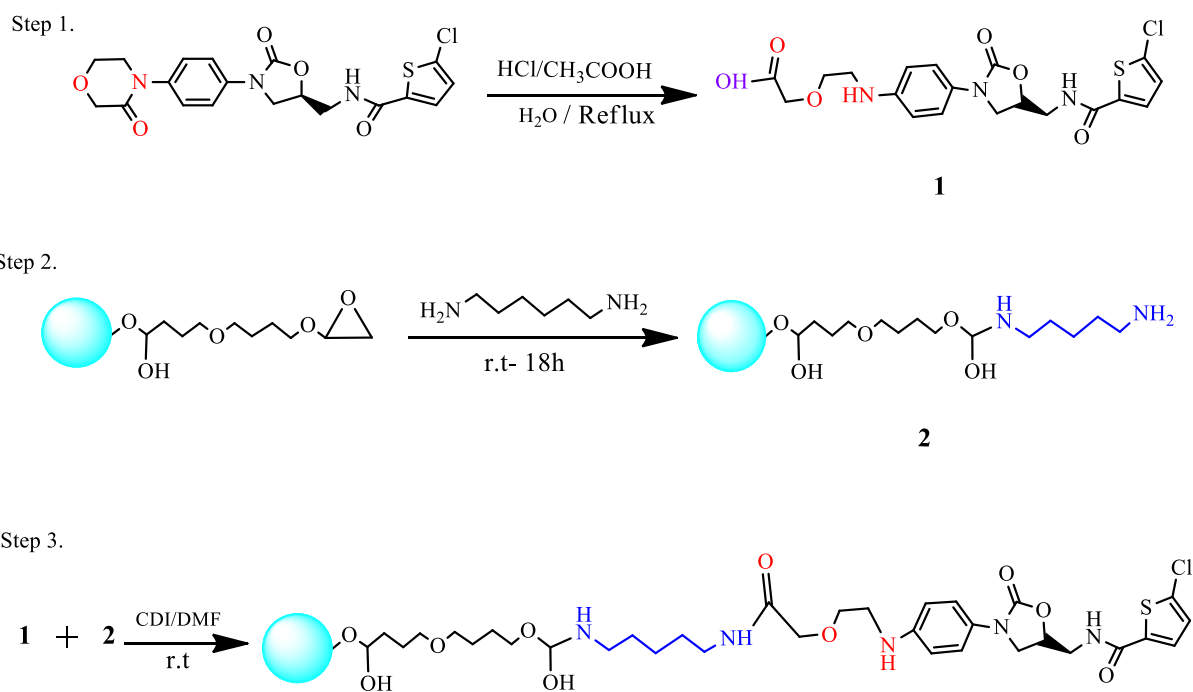


Figure S2. Coupling reaction of RIV with epoxy-activated Sepharose bead.

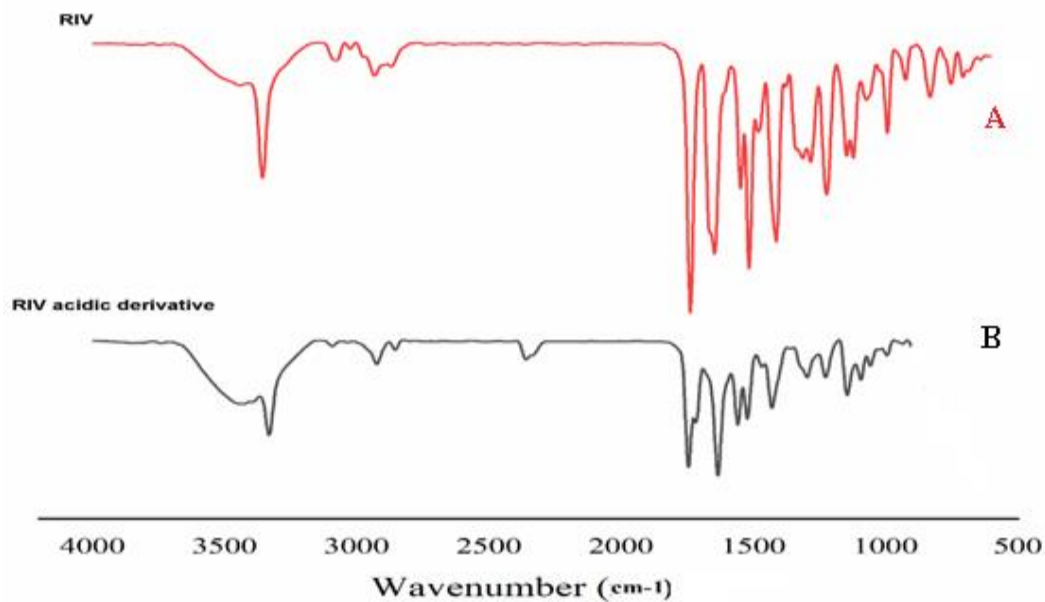


Figure S3. The FT-IR spectra of A, RIV, B, RIV-acidic derivative

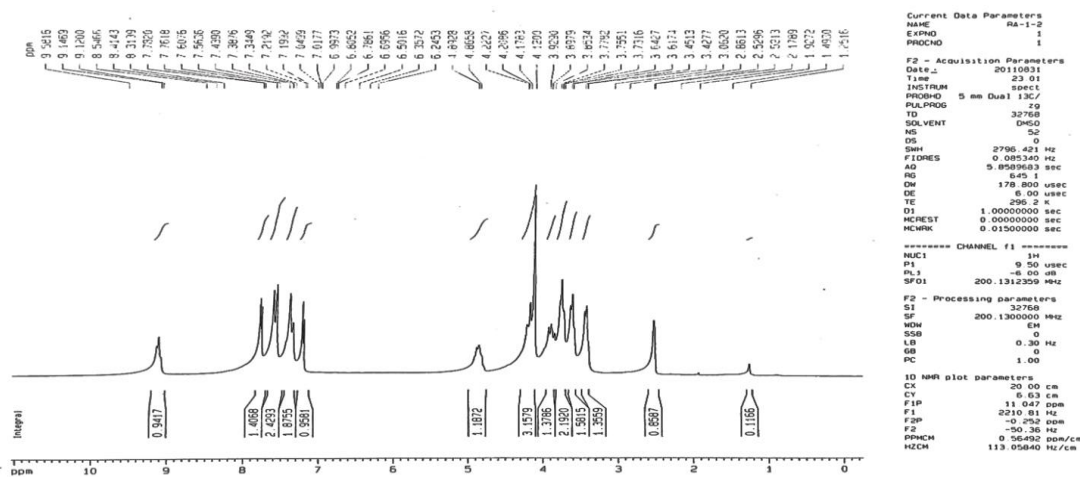


Figure S4. ^1H NMR spectra of compound (1)

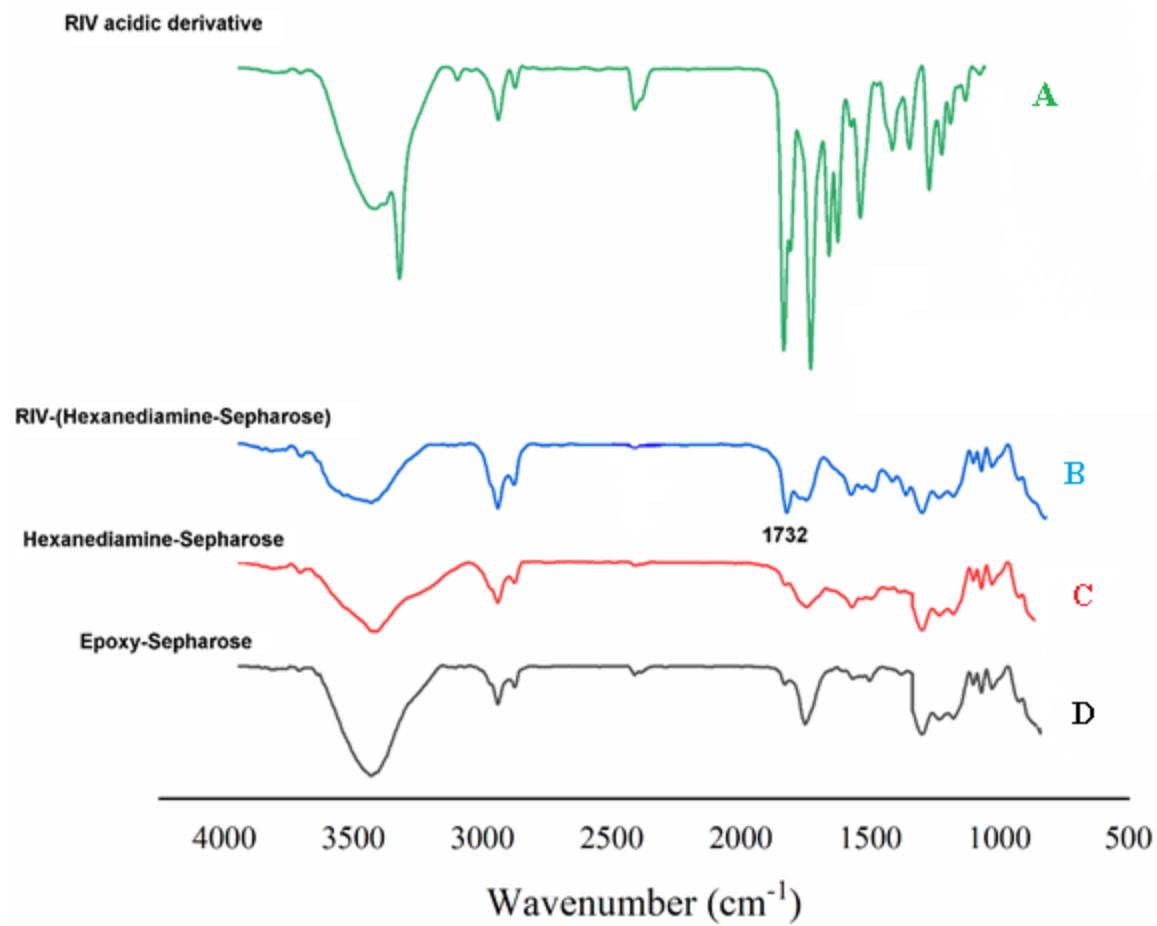


Figure S5. FT-IR spectrum of **A**, RIV acidic derivative, **B**, RIV-hexandiamine-Sepharose, **C**, hexandiamine-Sepharose, **D**, epoxy-Sepharose

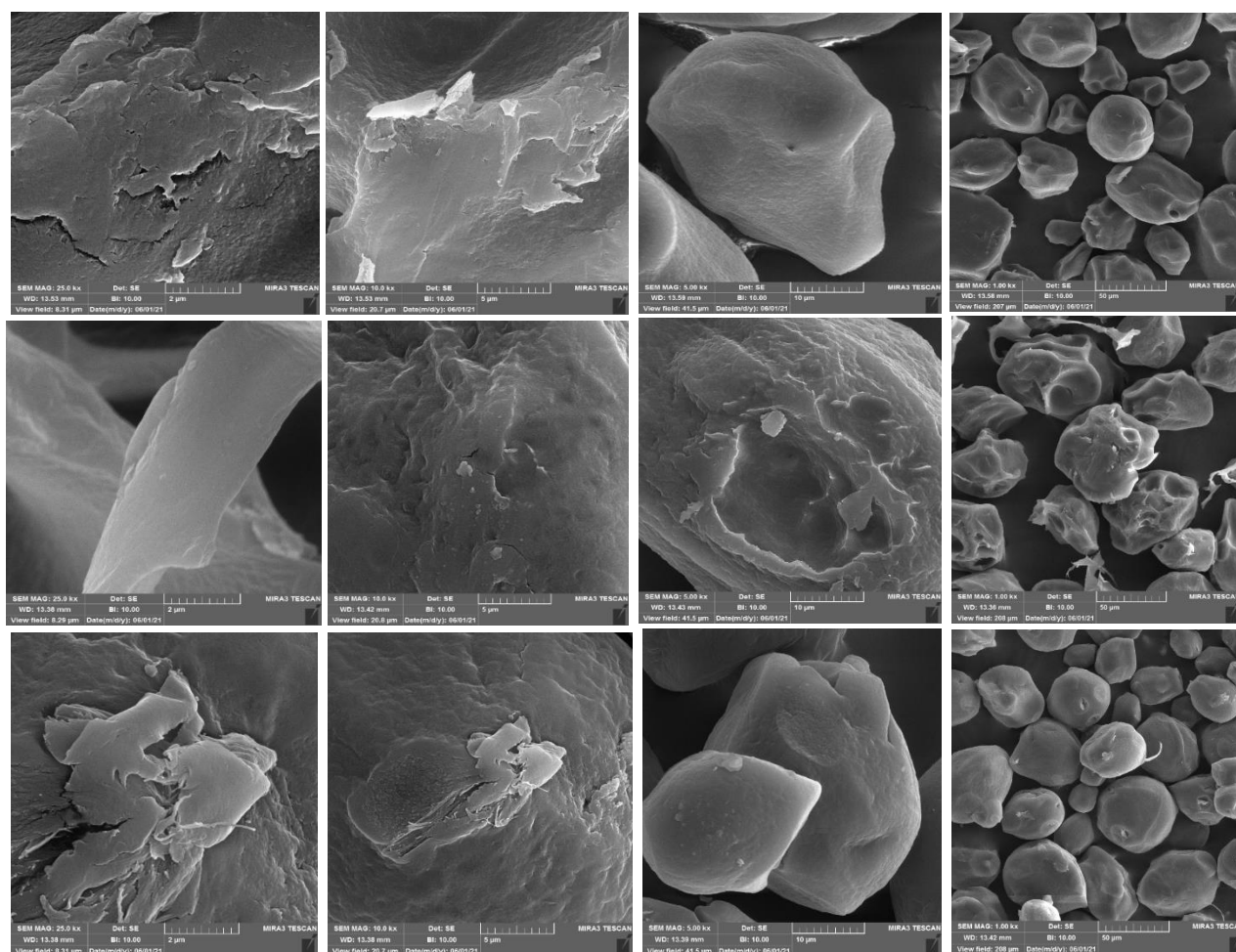


Figure S6. FE-SEM images of Sepharose beads at different magnification. For first row: epoxy-activated Sepharose 6B, second row: Sepharose-hexanediamine, and third row: Riv-(Sepharose-hexanediamine) matrix.

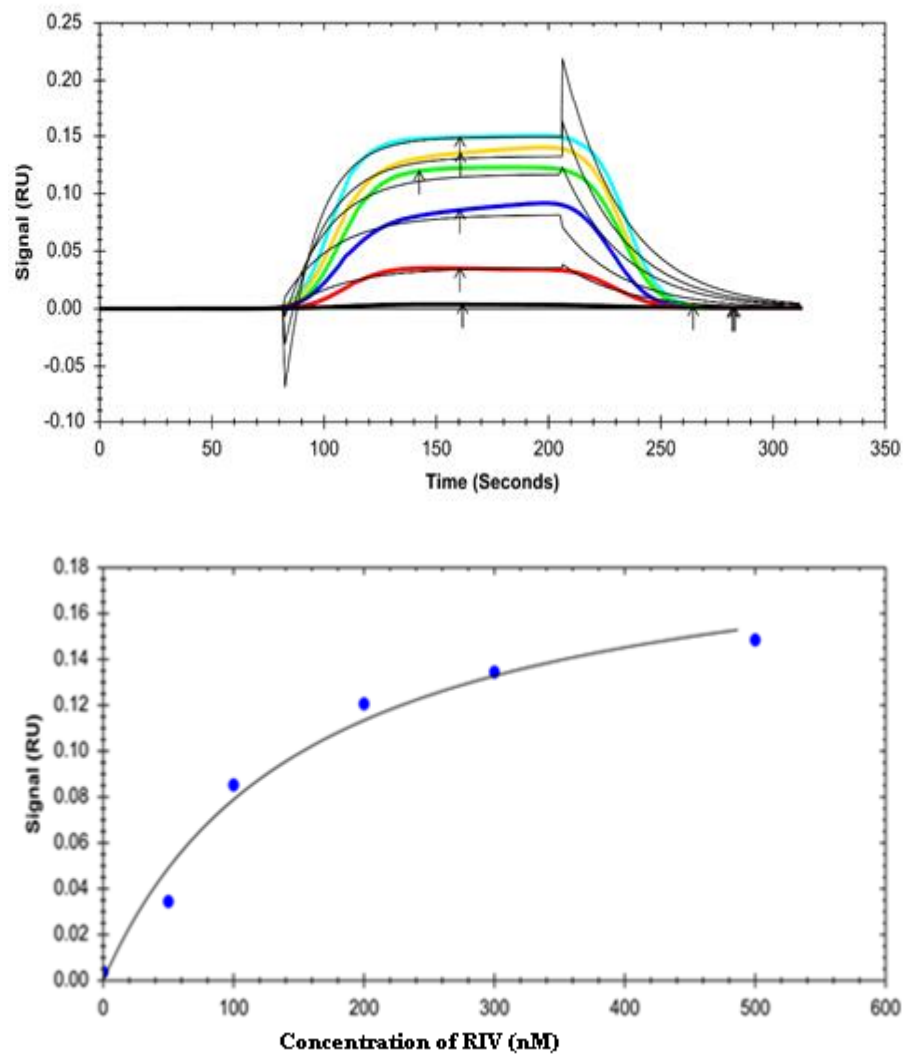


Figure S7. SPR sensor response for determination of the dissociation constants (k_d) of thiolated aptamer in combination with a concentration series of the target RIV (20–500 nM). On the basis of the obtained curves, non-linear regression analysis was used to compute K_d values.

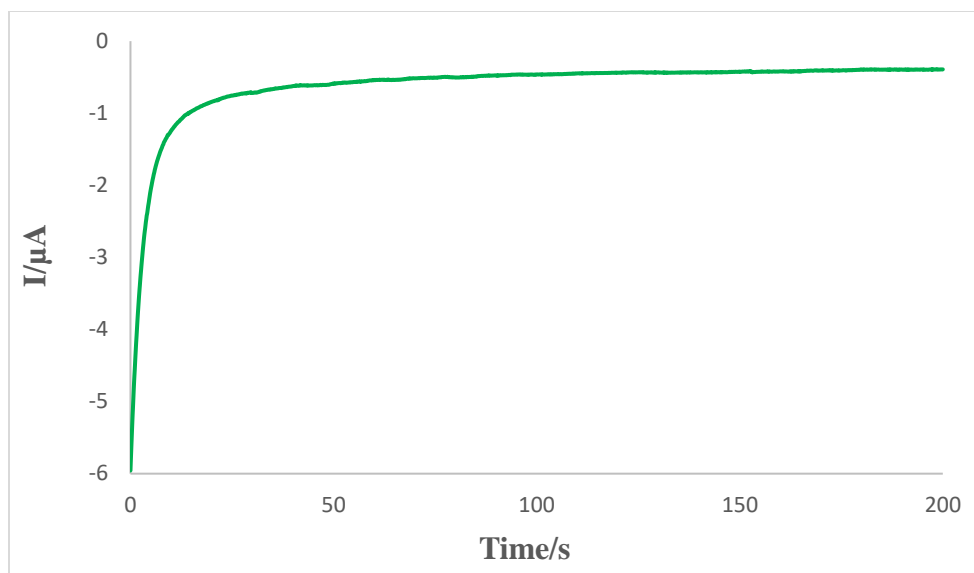


Figure S8. ChA of electrodeposition of AuNPs on the surface of ITO-PET-OH at the potential of $E=0.0$ V, time=200 s. 0.01 M $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$ and 0.01 M CTAB and arginine (0.003 g).

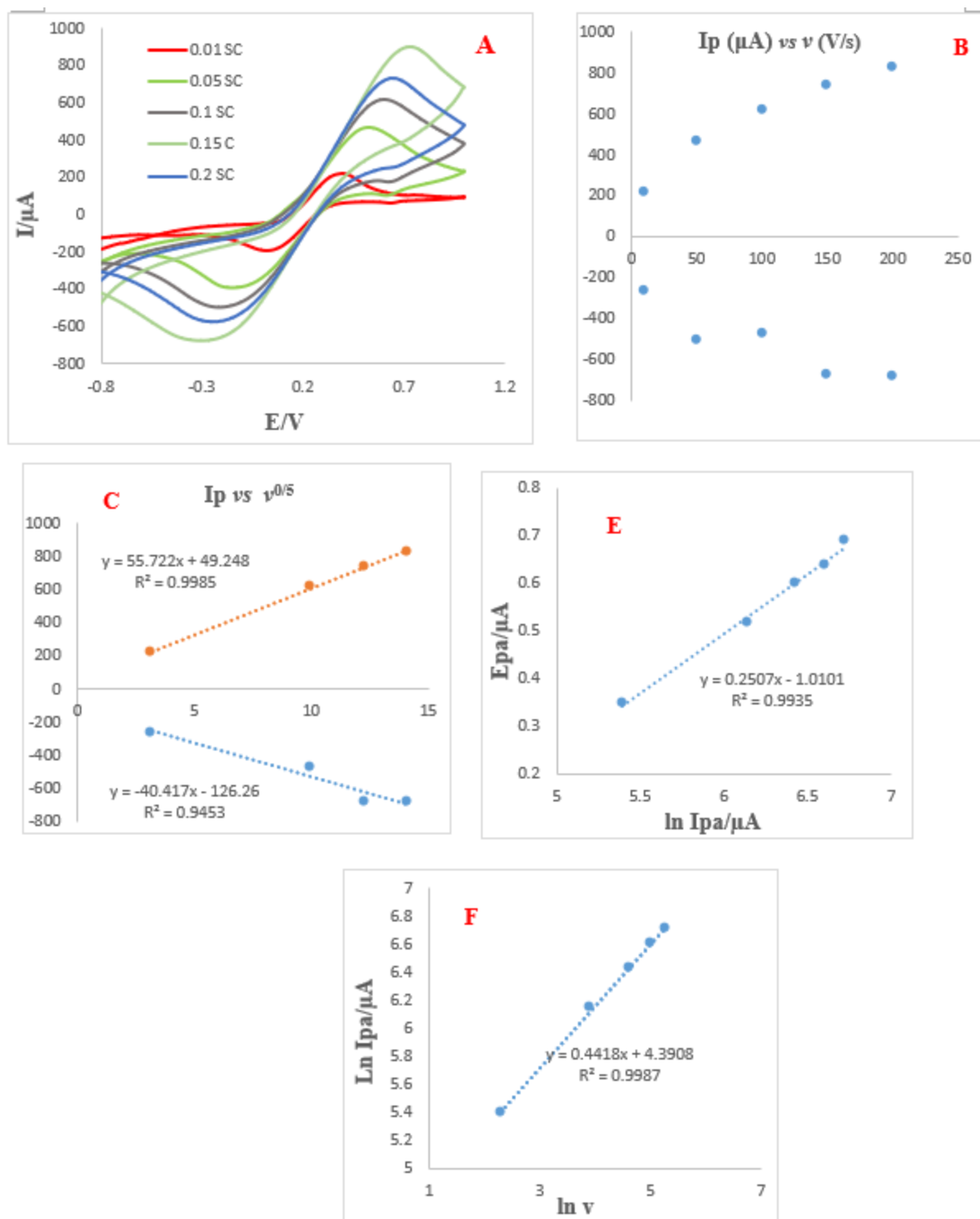


Figure S9. **A)** CVs of ITO-PET/OH/AuNPs in 0.01M $\text{Fe}(\text{CN})_6^{3-/4-}$ containing KCl, in various potential sweep rates (from inner to outer): 10, 50, 100, 150, 200, and 1000 mVs^{-1} , respectively. **B)** Dependence of anodic/cathodic peak currents vs. sweep rate. **C)** Dependence of anodic/cathodic peak currents vs. square

root of potential sweep rate. **E)** Tafel plot (variation of peak potential *versus* $\ln I_{pa}$. **F)** Dependency of $\ln I_{pa}$ *versus* $\ln v$.

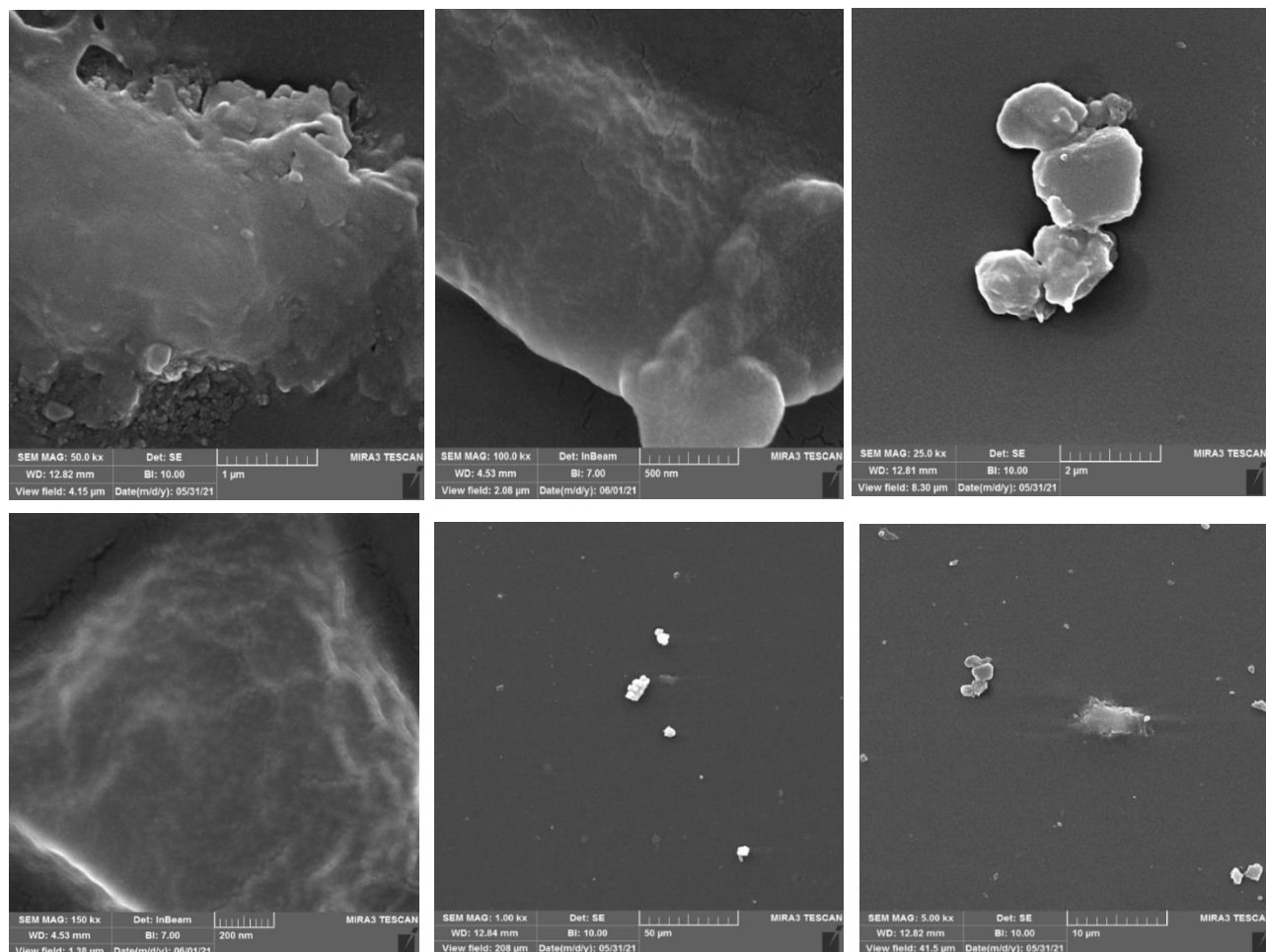


Figure S10A. FESEM images ITO PET-OH electrode in different magnifications.

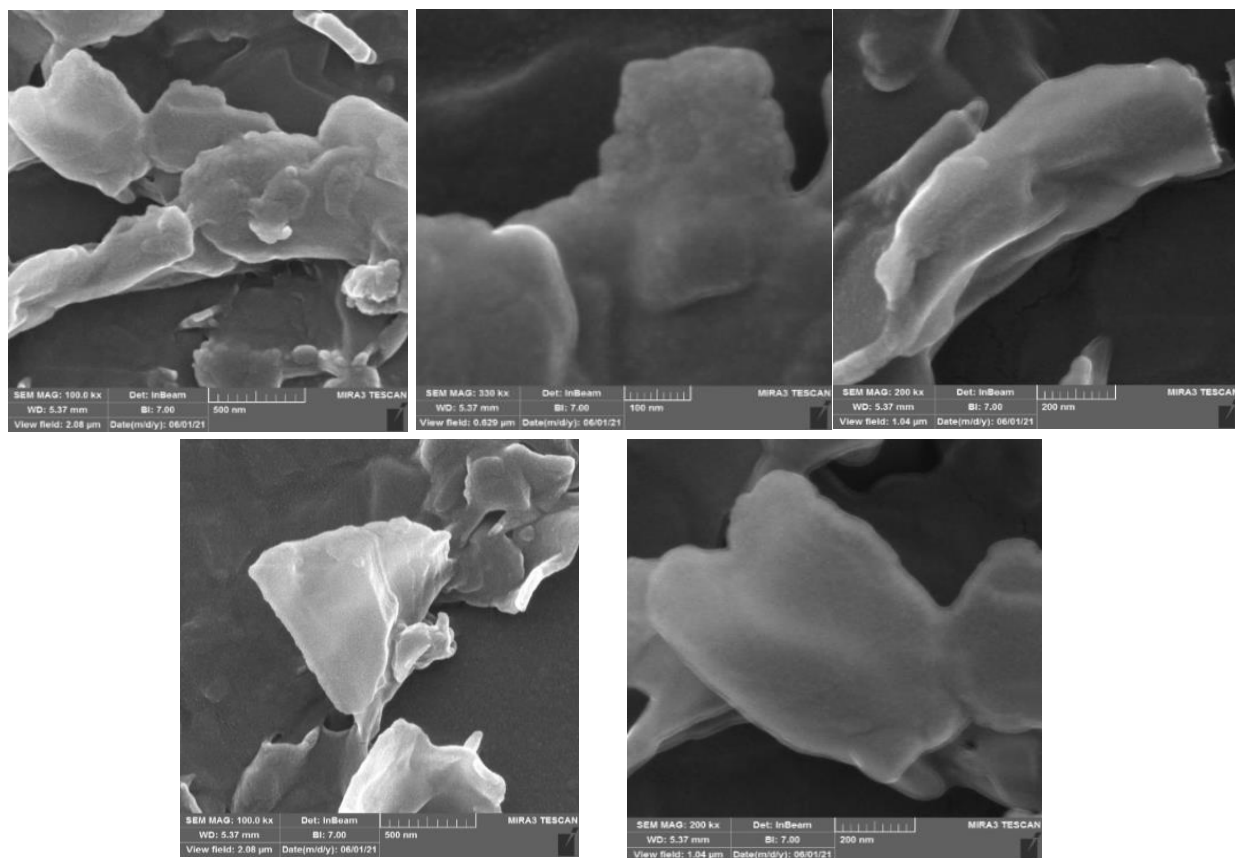


Figure S10B. FE-SEM images of ITO PET-OH modified Au electrode at various magnifications.

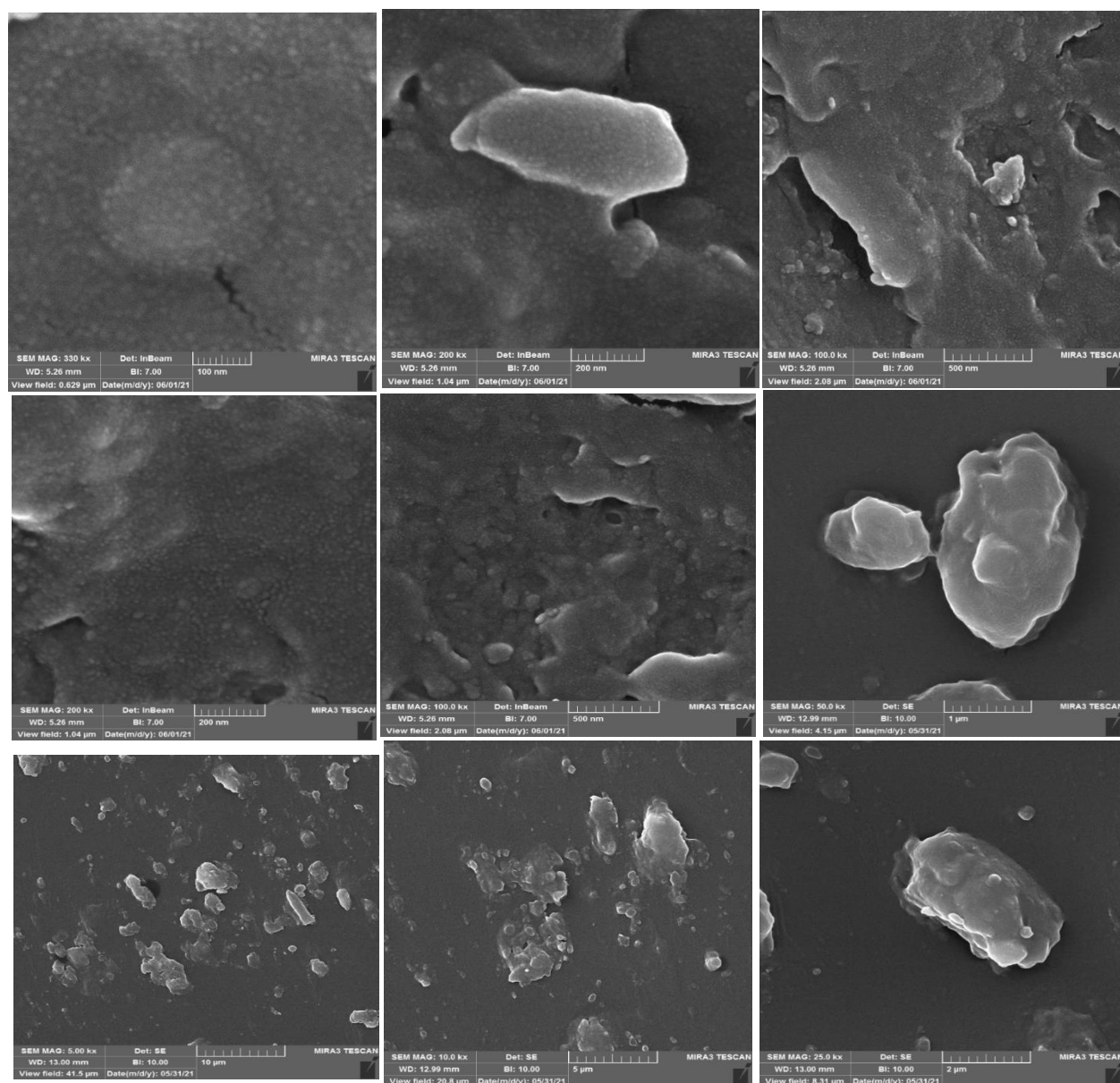


Figure S10C. FE-SEM of ITO PET-OH-AuNPs-aptamer in different magnifications.

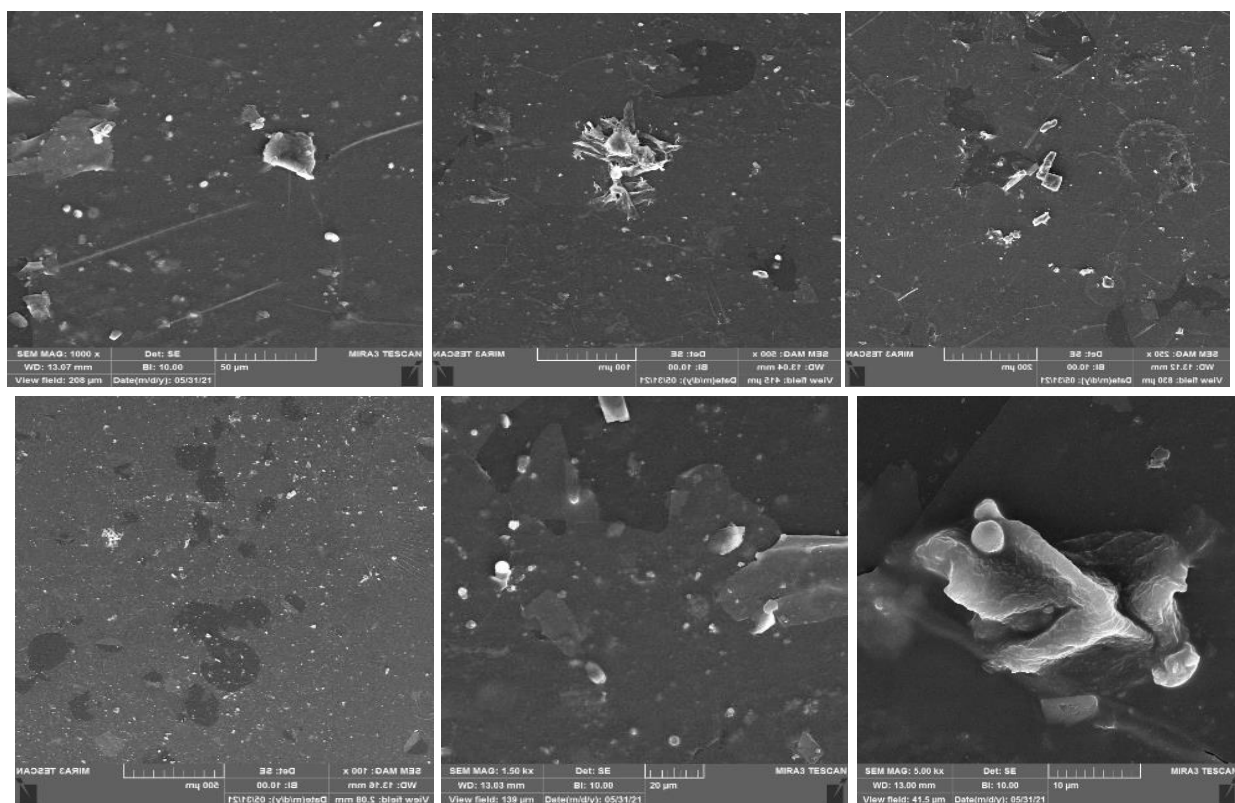
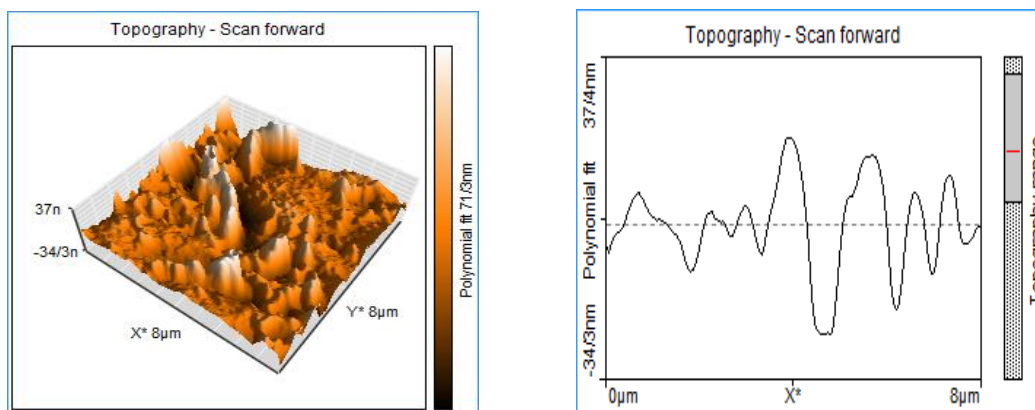
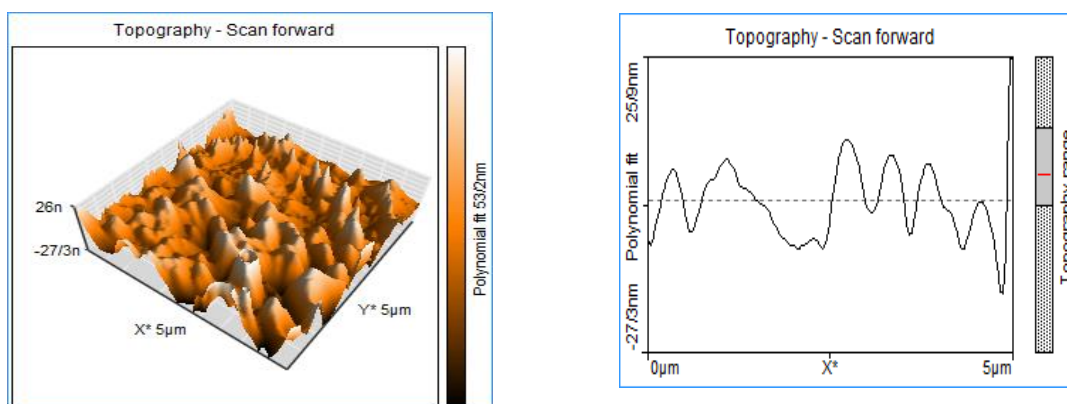


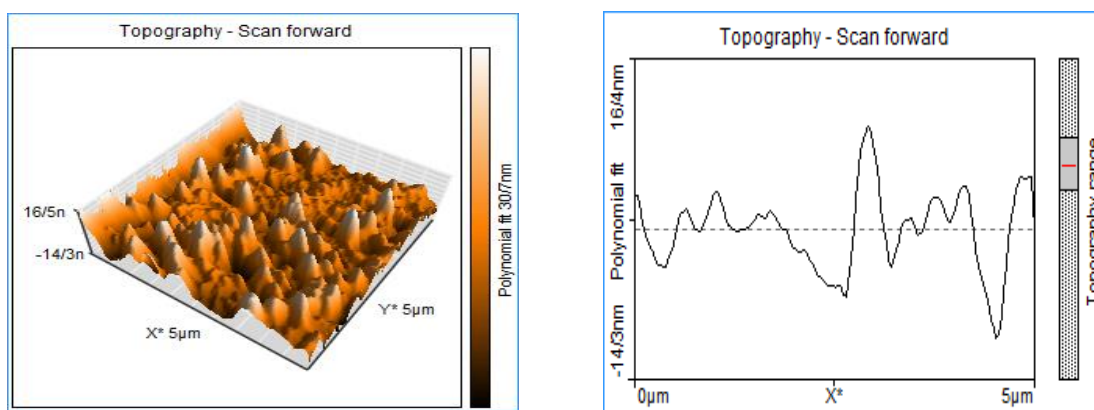
Figure S10D. FE-SEM image of ITO PET- OH-AuNPs-aptamer after incubation with RIV in different magnification.



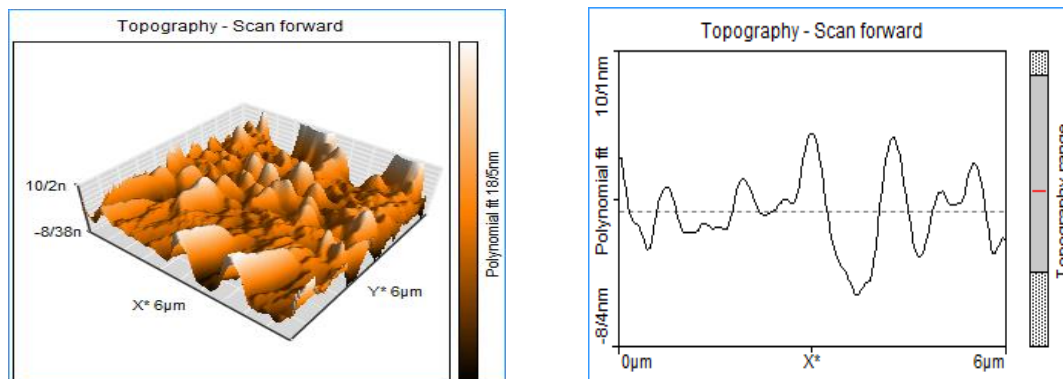
(A). ITO-PET-OH



(B). ITO-PET-OH-AuNPs

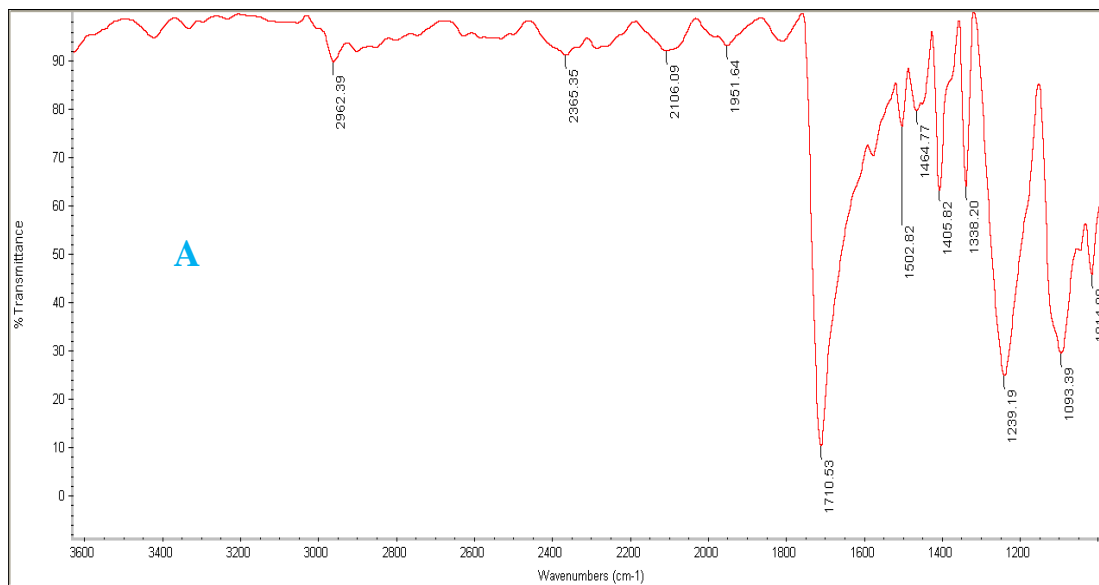


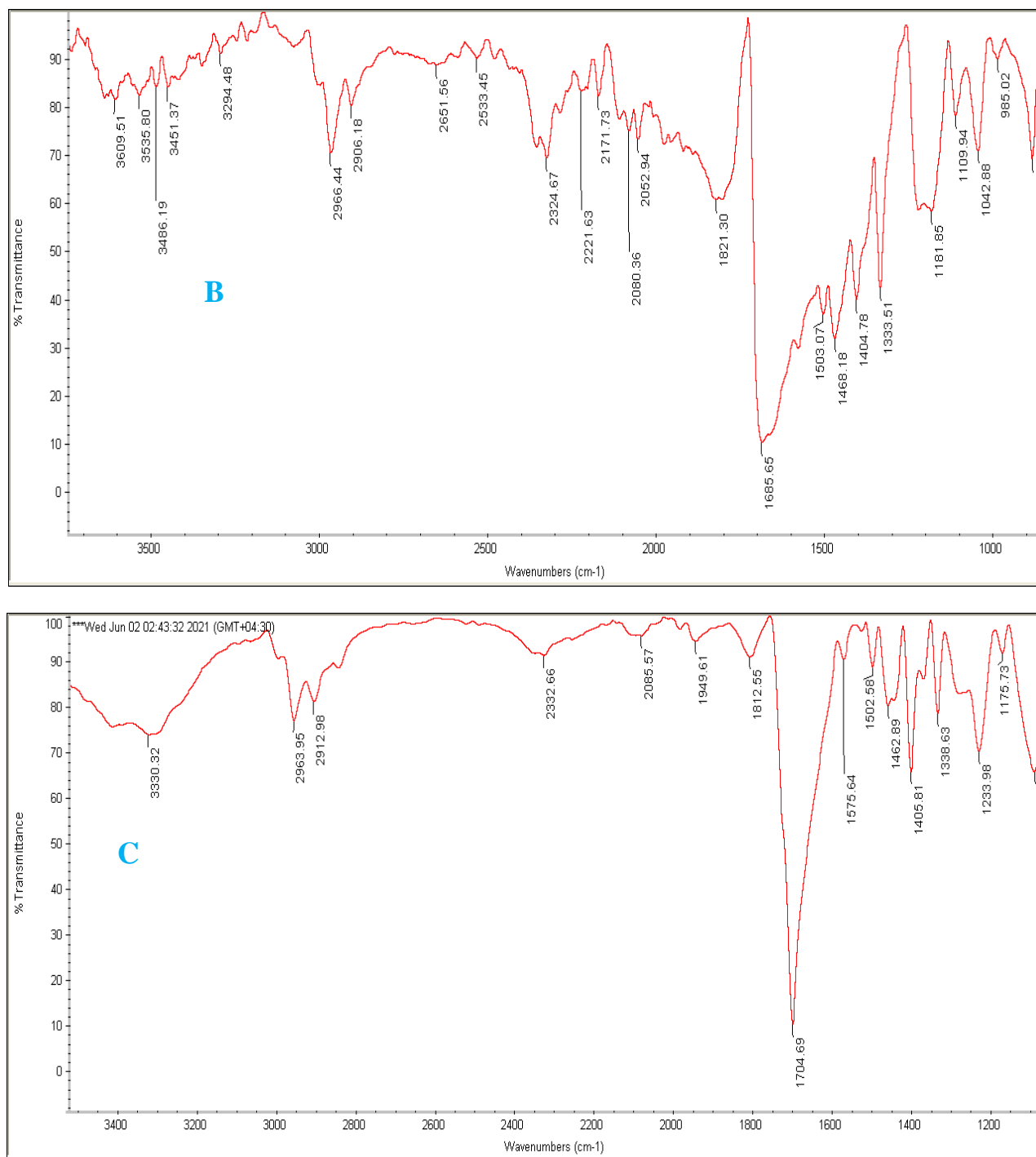
(C). ITO-PET-OH-AuNPs-aptamer



(D). ITO-PET-OH-AuNPs-aptamer/RIV

Figure S11. Topographical AFM images of (A) ITO film on PET, (B) ITO-PET/OH, (C) ITO-PET/OH/AuNPs, (D) electrode substrates after thiol aptamer assembly and planar ITO-PET/OH/AuNPs/aptamer/RIV substrates





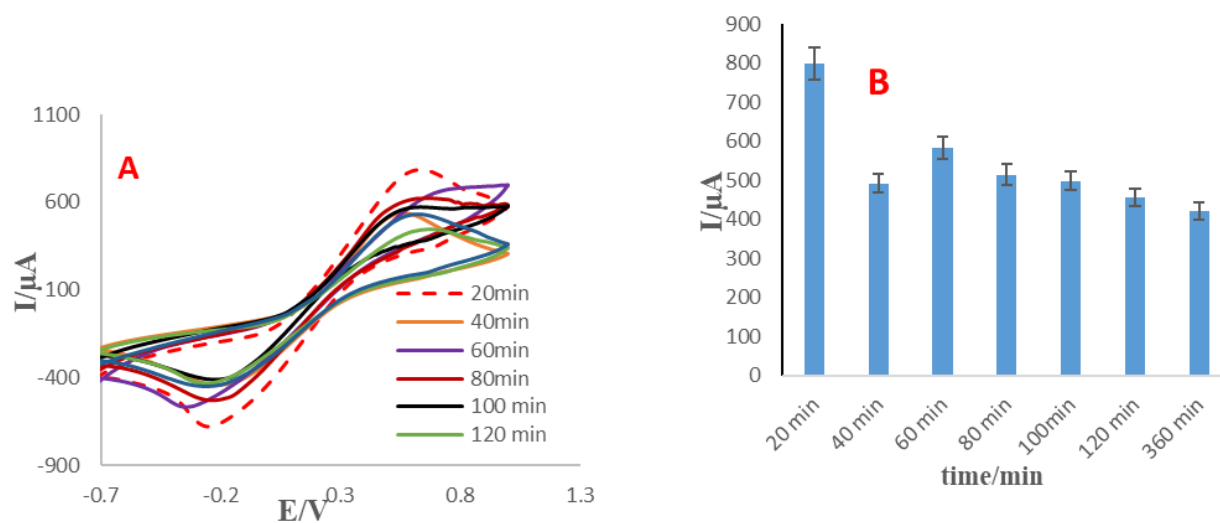


Figure S13. Optimization of various incubation time (20, 40, 60, 80, 100, 120 and 360 min) for aptamer immobilization (A) CVs of ITO PET-OH-Au NPs-aptamer in ferrocyanide/ferricyanide/KCl (0.01 M) at the potential range of -1 to +1 V, sweep rate: 100 mv/s (B) Histogram of peak current *versus* time of incubation. (n=3)

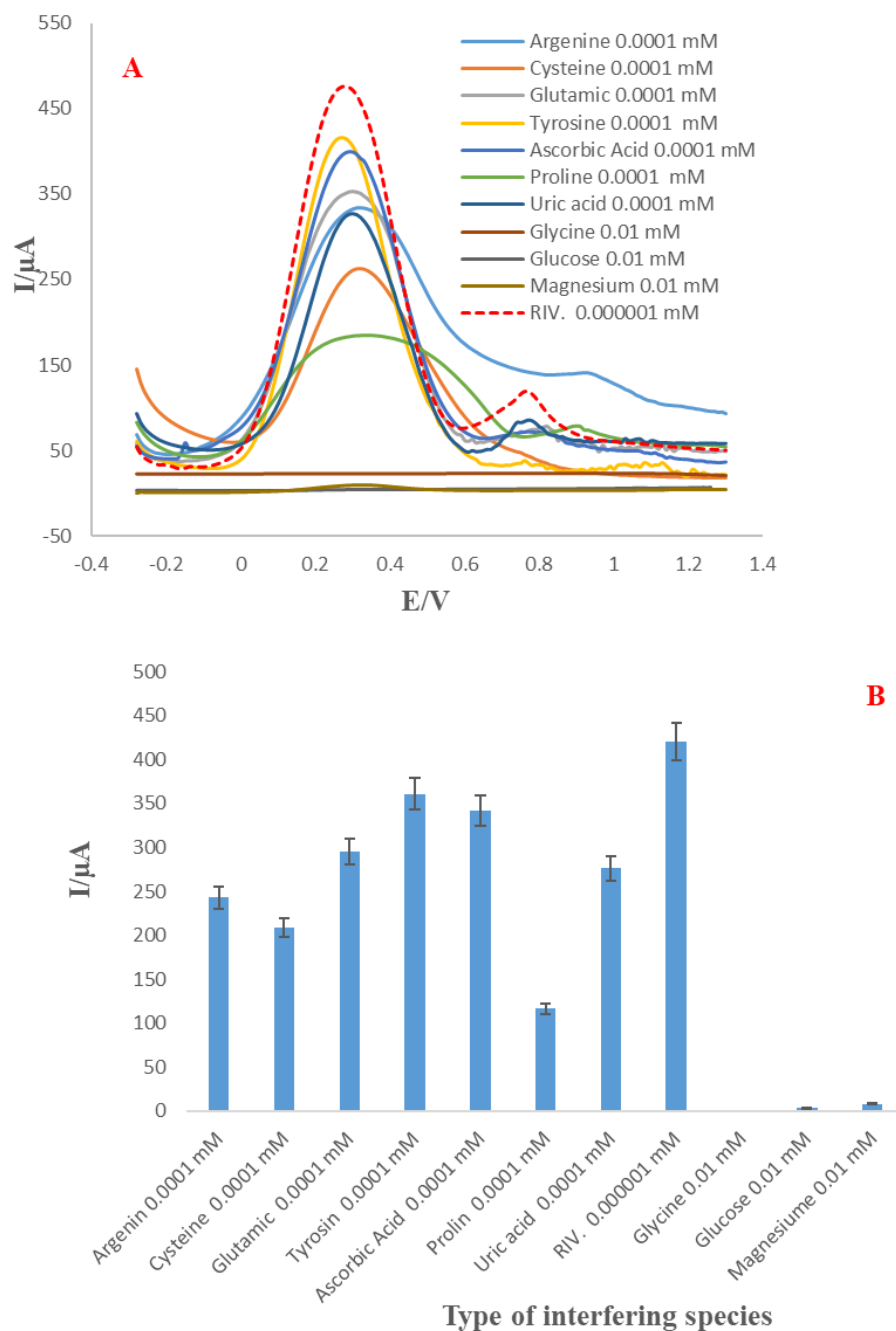
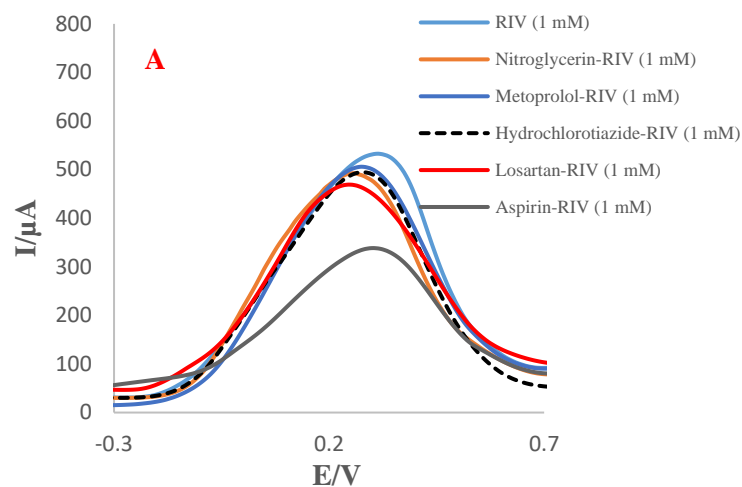


Figure S14. **A)** SWV of aptasensor in the presence of different interfering agents. **B)** Variation of peak currents *versus* type of interfering species. Supporting electrolyte is 0.01 M $[\text{Fe}(\text{CN})_6]^{3-/4-}$, SWV parameters: potential range of -1 V to +1 V with a step potential of 0.01 mV, amplitude 0.25 mV and frequency 1 Hz.



B

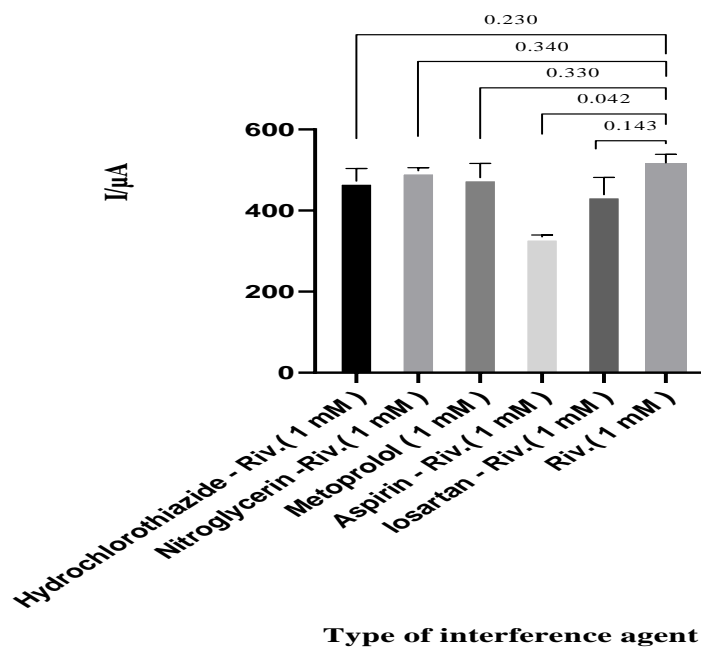


Figure S15. A) SWV of aptasensor in the presence of different species (hydrochlorothiazide, metoprolol, nitroglycerin, atorvastatin, aspirin, and losartan). B) Dependency of peak currents of aptasensor *versus* the type of interfering agents. The concentrations of analogs were 1 mM. Supporting electrolyte is 0.01 M $[\text{Fe}(\text{CN})_6]^{3-/4-}$, SWV parameters: potential range of -1 V to +1 V with a step potential of 0.01 mV, amplitude 0.25 mV, and frequency 1 Hz. Statistical significance was determined by Paired t-test. The aspirin shows a significantly lower response than other drugs ($p < 0.05$) indicating significant differences relative to displayed bars as determined.

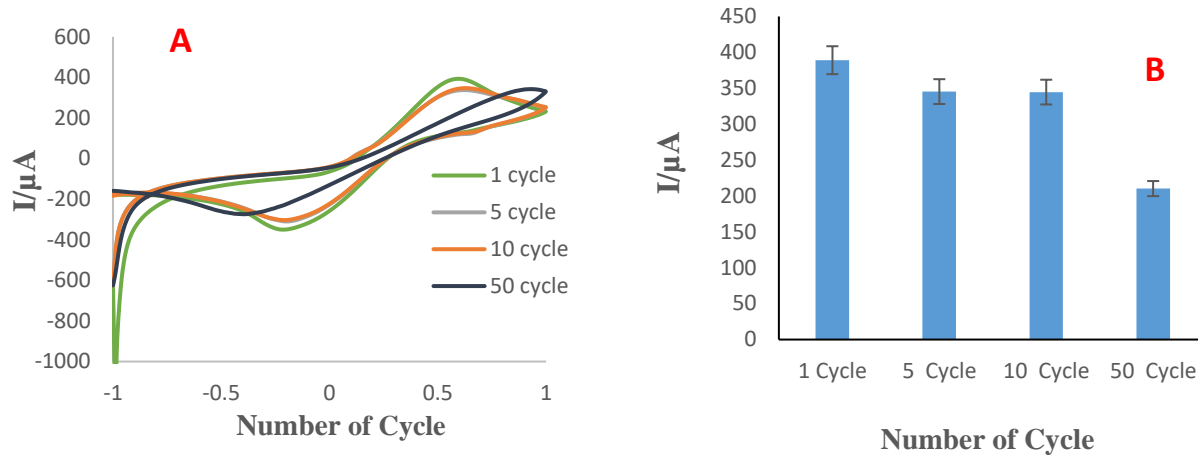


Figure S16. A) CVs of ITO-PET-OH-AuNPs in different cycle number (1-50 Cycle). **B)** Histogram of peak current *versus* number of cycles. Sweep rate is 100 mV/s. supporting electrode is $[\text{Fe}(\text{CN})_6]^{3-/4-}/\text{KCl}$ (0.01 M)]. (n=3).

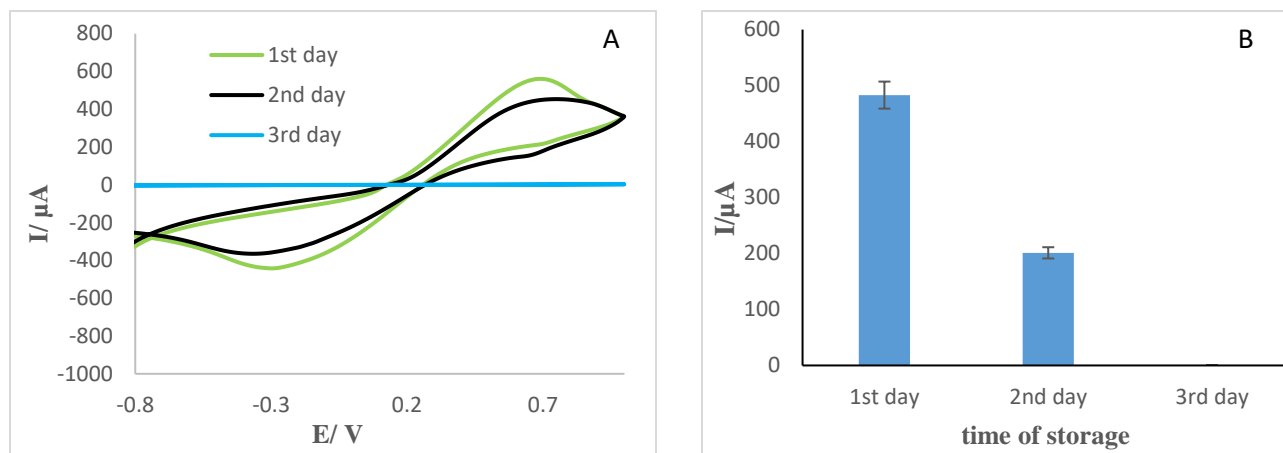


Figure S17. A, CVs of ITO-PET-OH-AuNPs/Aptamer in the various storage time (24, 48, and 72 h) in supporting electrolyte $[\text{Fe}(\text{CN})_6]^{3-/4-}/\text{KCl}$ (0.01 M) sweep rate is 100 mv/s. (1–3 days). (n=3). B) Histogram of peak current *versus* time of Storage.

Table S1. Validation data of proposed method for quantification of RIV in exhaled breath condensate (EBC) and human plasma samples (n=3).

Parameters	EBC	Plasma
Linear range (nM)	10-600	10-600
Slope	0.1609	0.196
Intercept	90.163	38.356
Correlation coefficient	0.9952	0.9941
Number of data points	7	8
LOD (nM)	6.03	14.08
LOQ (nM)	19.9	46.47

Table S2. Recovery of RIV from exhaled breath condensate (EBC) and plasma samples (n=3). Data are presented as mean \pm relative standard deviation (RSD)

plasma and EBC samples	Added RIV (nM)	EBC			Plasma		
		Found (nM)	Recovery (%)	RSD (%) (n=3)	Found (nM)	Recovery (%)	RSD (%) (n=3)
1	10	7.6 (\pm 3.7)	88.7	6.1	22 (\pm 2.1)	107.7	4.9
2	50	57.9 (\pm 2.2)	101.8	2.3	46.5 (\pm 2.5)	108.1	5.0
3	100	118.5 (\pm 0.5)	104.3	0.4	95.6 (\pm 1.3)	99.5	2.3
4	300	289 (\pm 1.9)	98.9	1.4	280.3 (\pm 1.3)	96.2	2.0
5	500	494.7 (\pm 2.2)	99.5	1.3	498.7 (\pm 2.2)	99.7	1.6

Table S3. Comparison of the recovery of RIV from exhaled breath condensate (EBC) and plasma samples (n=3), and the relative standard deviations (RSD %) analyzed by the proposed aptasensor and LC-MS/MS method

Sample (nM)	EBC Recoveries (%)		Plasma Recoveries (%)	
	Aptasensor	LC-MS/MS	Aptasensor	LC-MS/MS
10	88.7 (\pm 6.1)	107.0 (\pm 0.21)	107.7 (\pm 4.9)	103.3 (\pm 2.1)
50	101.8 (\pm 2.3)	105.7 (\pm 0.2)	108.1 (\pm 5.0)	105.4 (\pm 0.8)
100	104.3 (\pm 0.4)	100.2 (\pm 0.25)	99.5 (\pm 2.3)	98.9 (\pm 0.4)
300	98.9 (\pm 1.4)	96.7 (\pm 0.06)	96.2 (\pm 2.0)	99.9 (\pm 0.2)
500	99.5 (\pm 1.3)	101.0 (\pm 0.08)	99.7 (\pm 1.6)	99.7 (\pm 0.1)

Table S4. The repeatability of aptasensor

Concentration(nM)	Ip1	Ip2	SD	AVE _{STDV}
10	278	276	1.41	2.8
50	300	295	3.53	
100	321	317	2.82	
300	411	414	2.12	
500	474	480	4.24	