

Supplementary materials

Article

Ultrasensitive Photochemical Immunosensor Based on Flowerlike SnO₂/BiOI/Ag₂S Composites for Detection of Procalcitonin

Nuo Zhang ^{1,2}, **Jinhui Feng** ², **Guanhui Zhao** ², **Xiaoyi Duan** ¹, **Yaoguang Wang** ^{3,*}, **Daopeng Zhang** ^{1,*} and **Qin Wei** ²

¹ School of Chemistry and Chemical Engineering, Shandong University of Technology, Zibo 255049, China; chm_zhangn@ujn.edu.cn (N.Z.); sennuo_1225@163.com (X.D.)

² Key Laboratory of Interfacial Reaction & Sensing Analysis in Universities of Shandong, School of Chemistry and Chemical Engineering, University of Jinan, Jinan 250022, China; fengjinhui2011@163.com (J.F.); zhaoguanhui9001@163.com (G.Z.); sdjndxwq@163.com (Q.W.)

³ Shandong Provincial Key Laboratory of Molecular Engineering, School of Chemistry and Chemical Engineering, Qilu University of Technology (Shandong Academy of Sciences), Jinan 250353, China

* Correspondence: wangyaoguang@qlu.edu.cn (Y.W.); dpzhang@sdu.edu.cn (D.Z.); Tel./Fax: +86-531-8276-7872

Citation: Zhang, N.; Feng, J.; Zhao, G.; Duan, X.; Wang, Y.; Zhang, D.; Wei, Q. Ultrasensitive Photochemical Immunosensor Based on Flowerlike SnO₂/BiOI/Ag₂S Composites for Detection of Procalcitonin. *Biosensors* **2021**, *11*, 421. <https://doi.org/10.3390/bios11110421>

Received: 7 October 2021

Accepted: 25 October 2021

Published: 28 October 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).

1. Materials and apparatus

Thioglycolic acid (TGA) was obtained from Macklin Reagent Co., Ltd. (Shanghai, China). 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC) and N-hydroxysuccinimide (NHS) were obtained from Aladdin Reagent Database Inc. (Shanghai, China). Tin (IV) chloride pentahydrate (SnCl_4), thioacetamide, potassium iodide (KI), Bismuth nitrate pentahydrate ($\text{Bi}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$), sodium sulfide (Na_2S), silver nitrate (AgNO_3), ascorbic acid (AA), absolute ethanol, isopropyl alcohol and acetone were purchased from Sinopharm Chemical Reagent Co., Ltd. (Beijing, China). Phosphate buffered solution (PBS, $1/15 \text{ mol} \cdot \text{L}^{-1} \text{ KH}_2\text{PO}_4$ and $1/15 \text{ mol} \cdot \text{L}^{-1} \text{ Na}_2\text{HPO}_4$) containing AA was used as an electrolyte for the PEC measurements. All other chemicals in the experiment were analytical grade and were used as received without further purification.

2. Apparatus

Scanning electron microscope (SEM) images and energy dispersive spectroscopy (EDS) were obtained by using a field-emission SEM (Zeiss, Gemini 300, Germany). Electrochemical impedance spectroscopy (EIS) analysis was performed with an RST5200F electrochemical workstation (Zhengzhou Shiruisi Technology Co., Ltd, China) with a three-electrode system in a $5.0 \text{ mmol} \cdot \text{L}^{-1} [\text{Fe}(\text{CN})_6]^{3-/4-}$ solution containing $0.10 \text{ mol} \cdot \text{L}^{-1} \text{ KCl}$. UV-vis diffuse reflectance spectrum measurements were performed with a Shimadzu UV-3101PC spectrometer (Japan). All PEC experiments were measured on a CHI760E electrochemical workstation (Chenhua Instrument Shanghai Co., Ltd, China) by using a conventional three-electrode system comprising of a saturated calomel electrode as reference electrode, a platinum wire as a counter-electrode, and the as-prepared $\text{SnO}_2/\text{BiOI}/\text{Ag}_2\text{S}$ modified ITO electrode ($2.5 \times 1.0 \text{ cm}^2$) as working electrode.

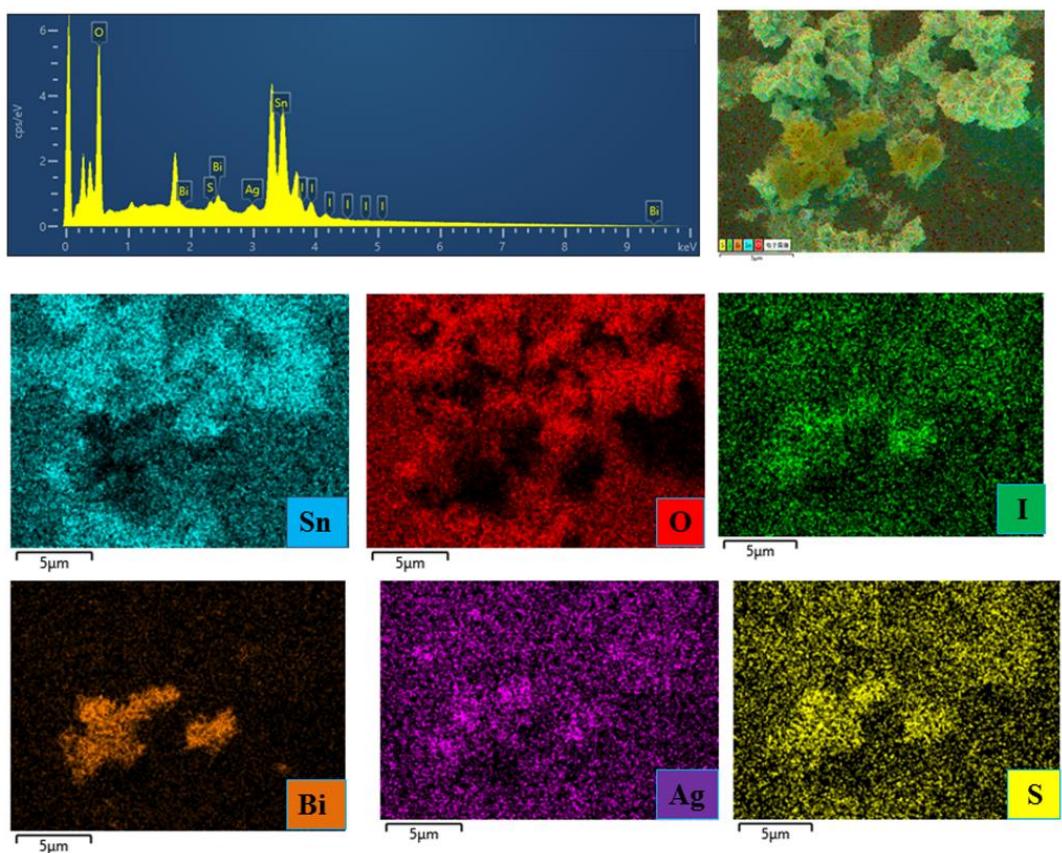


Figure S1 The EDS mapping images of SnO₂/BiOI/Ag₂S composites.

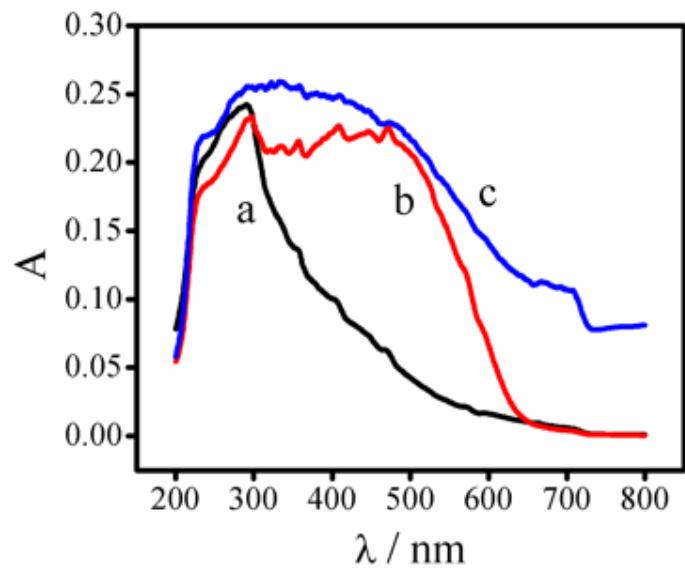


Figure S2 UV-Vis diffuse reflectance spectra of SnO₂ (a), SnO₂/BiOI (b) and SnO₂/BiOI/Ag₂S (c).

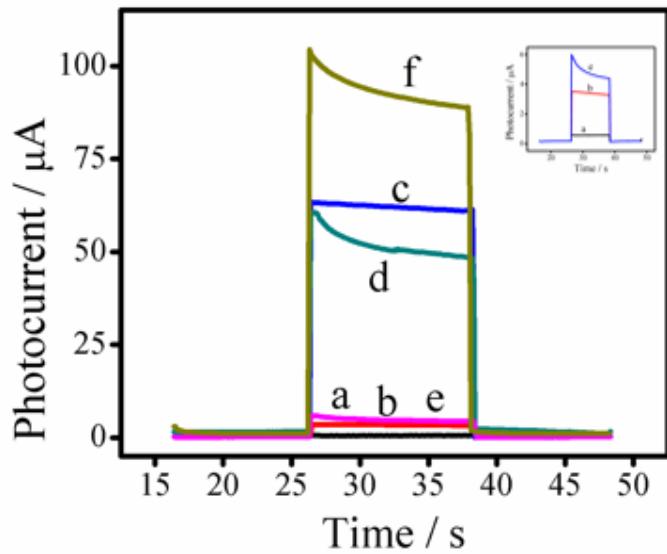


Figure S3 Time-based photocurrent response curves of SnO_2 (a), BiOI (b), SnO_2/BiOI (c), $\text{SnO}_2/\text{Ag}_2\text{S}$ (d), $\text{BiOI}/\text{Ag}_2\text{S}$ (e) and $\text{SnO}_2/\text{BiOI}/\text{Ag}_2\text{S}$ (f).

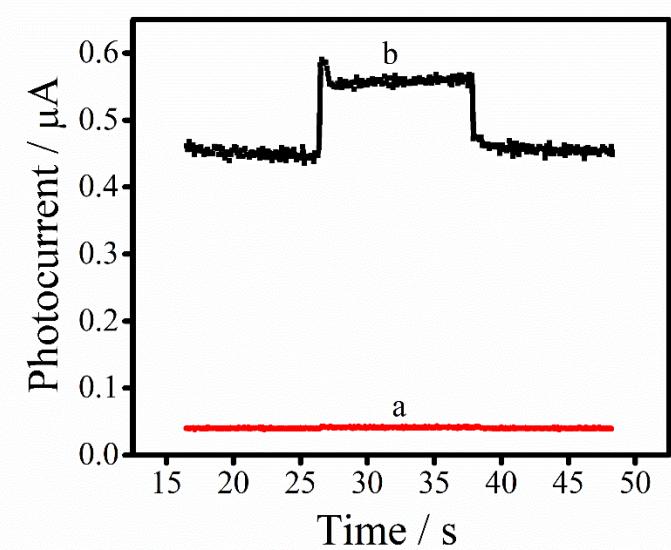


Figure S4 Time-based photocurrent response curves of ITO electrode (a) and ITO/SnO₂ (b)

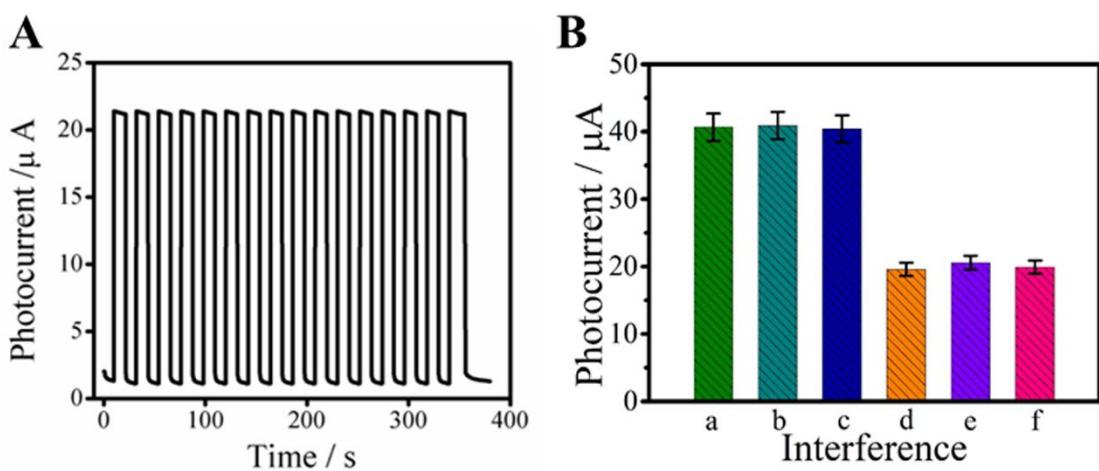


Figure 5. Stability curve of chronocurrent (A); Selectivity of the PEC immunosensor for detecting PCT (B) : (a) Blank , (b) Blank+100 ng·mL⁻¹ CEA, (c) Blank+100 ng·mL⁻¹ PSA, (d) 1.0 ng·mL⁻¹ PCT, (e) 1.0 ng·mL⁻¹ PCT+100 ng·mL⁻¹ CEA, (g) (f) 1.0 ng·mL⁻¹ PCT+100 ng·mL⁻¹ PSA. The applied potential was 0 V. Error bars = SD ($n = 5$).

Table S1 Comparison of the performance of the proposed PEC immunosensor for PCT detection
and those of other reports.

Detection method	Liner range ($\text{pg}\cdot\text{mL}^{-1}$)	Detection limit ($\text{pg}\cdot\text{mL}^{-1}$)	References
Chemiluminescence immunoassay	$44 - 1.0 \times 10^5$	44	1
Electrochemical immunosensor	$1.0 - 5.0 \times 10^4$	0.36	2
Ratiometric electrochemical immunosensor	$1.0 - 1.0 \times 10^5$	0.30	3
Immunosorbent assay	$1.0 - 1.0 \times 10^5$	0.095	4
Immunochromatographic assay	$62.5 - 4.0 \times 10^5$	62.5	5
SERS-based immunochromatographic assay	$10 - 1.0 \times 10^7$	8.017	6
Time-resolved digital immunoassay	$4.2 - 1.25 \times 10^4$	2.8	7
Electrochemical paper-based analytical device	$0.50 - 2.5 \times 10^5$	0.27	8
Lateral flow immunoassay	$12 - 1.0 \times 10^4$	31	9
Capillary immunosensor	$0.10 - 1.0 \times 10^5$	0.01	10
Electrochemiluminescence immunosensor	$0.10 - 1.0 \times 10^4$	0.0257	11
Electrochemiluminescence immunosensor	$0.10 - 1.0 \times 10^5$	0.01258	12
Electrochemiluminescence immunosensor	$0.010 - 1.0 \times 10^5$	0.00365	13
Electrochemiluminescence immunosensor	0.005 - 100	0.0021	14
Electrochemiluminescence immunosensor	$0.10 - 5.0 \times 10^4$	0.054	15
Electrochemiluminescence immunosensor	$0.05 - 1.0 \times 10^5$	0.017	16
Photoelectrochemical immunosensor	$0.10 - 5.0 \times 10^4$	0.020	17
Photoelectrochemical immunosensor	$0.10 - 1.0 \times 10^5$	0.030	18
Photoelectrochemical immunosensor	$0.50 - 1.0 \times 10^5$	0.17	19
Photoelectrochemical immunosensor	$1.0 - 1.0 \times 10^5$	0.42	20
Photoelectrochemical immunosensor	$0.50 - 1.0 \times 10^5$	0.14	This work

Table S2. The results of the PCT determination in human serum samples

Content in samples (ng·mL ⁻¹)	Added content (ng·mL ⁻¹)	Average content		
		(n=11) (ng·mL ⁻¹)	RSD (n=11, %)	Recovery (%)
0.0400	0.0100	0.0512	8.4	103%
	1.00	1.04	7.6	100%
0.800	1.00	1.79	4.7	98.8%
	10.0	10.8	2.2	100 %

References:

- [1] Enqi Huang, Dezhi Huang, Yu Wang, et al. Active droplet-array microfluidics-based chemiluminescence immunoassay for point-of-care detection of procalcitonin. *Biosensors and Bioelectronics*, 2022, 195, 113684.
- [2] Hui Ding, Lei Yang, Hongying Jia, et al. Label-free electrochemical immunosensor with palladium nanoparticles functionalized MoS₂/NiCo heterostructures for sensitive procalcitonin detection. *Sensors and Actuators B: Chemical*, 2020, 312, 127980.
- [3] Juncong Miao, Kang Du, Xuan Li, et al. Ratiometric electrochemical immunosensor for the detection of procalcitonin based on the ratios of SiO₂-Fc-COOH-Au and UiO-66-TB complexes. *Biosensors and Bioelectronics*, 2021, 171, 112713.
- [4] Chang Yue Chiang, Tze Ta Huang, Chih Hui Wang, et al. Fiber optic nanogold-linked immunosorbent assay for rapid detection of procalcitonin at femtomolar concentration level. *Biosensors and Bioelectronics*, 2020, 151, 111871.
- [5] Shu Zhou, Yuanli Peng, Jing Hu, et al. Quantum dot nanobead-based immunochromatographic assay for the quantitative detection of the procalcitonin antigen in serum samples. *Microchemical Journal*, 2020, 159, 105533.
- [6] Ji Xia, Dan Lu, Yifan Liu, et al. Prediction of premature rupture of membranes via simultaneous detection of procalcitonin and interleukin-6 by a SERS-based immunochromatographic assay. *New Journal of Chemistry*, 2020, 44, 17099.
- [7] Wenwen Jing, Yan Wang, Yunze Yang, et al. Time-resolved digital immunoassay for rapid and sensitive quantitation of procalcitonin with plasmonic imaging. *ACS Nano*, 2019, 13, 8609-8617.
- [8] Suchanat Boonkaew, Ilhoon Jang, Eka Noviana, et al. Electrochemical paper-based analytical device for multiplexed, point-of-care detection of cardiovascular disease biomarkers. *Sensors and Actuators B: Chemical*, 2021, 330, 129336.
- [9] Liang Huang, Yuxing Zhang, Tao Liao, et al. Compact magneto-fluorescent colloids by hierarchical assembly of dual-components in radial channels for sensitive point-of-care immunoassay. *Small*, 2021, 17, 2100862.
- [10] Xuexue Xu, Xingda Song, Rongbin Nie, et al. Ultra-sensitive capillary immunosensor

combining porous-layer surface modification and biotin-streptavidin nano-complex signal amplification: Application for sensing of procalcitonin in serum. *Talanta*, 2019, 205, 120089.

- [11] Cui Song, Xiaojian Li, Lihua Hu, et al. Quench-Type Electrochemiluminescence Immunosensor Based on Resonance Energy Transfer from Carbon Nanotubes and Au-Nanoparticles-Enhanced g-C₃N₄ to CuO@Polydopamine for Procalcitonin Detection. *ACS Applied Materials & Interfaces*, 2020, 12, 8006-8015.
- [12] Lihua Hu, Cui Song, Tengfei Shi, et al. Dual-quenching electrochemiluminescence resonance energy transfer system from IRMOF-3 coreaction accelerator enriched nitrogen-doped GQDs to ZnO@Au for sensitive detection of procalcitonin. *Sensors and Actuators B: Chemical*, 2021, 346, 130495.
- [13] Lu Zhao, Xianzhen Song, Xiang Ren, et al. Ultrasensitive near-infrared electrochemiluminescence biosensor derived from Eu-MOF with antenna effect and high efficiency catalysis of specific CoS₂ hollow triple shelled nanoboxes for procalcitonin. *Biosensors and Bioelectronics*, 2021, 191, 113409.
- [14] Xiaojian Li, Yu Du, Peng Xu, et al. Signal-off electrochemiluminescence immunosensor based on Mn-Eumelanin coordination nanoparticles quenching PtCo-CuFe₂O₄-reduced graphene oxide enhanced luminol. *Sensors and Actuators B: Chemical*, 2020, 323, 128702.
- [15] Lei Yang, Jingwei Xue, Yue Jia, et al. Construction of well-ordered electrochemiluminescence sensing interface using peptide-based specific antibody immobilizer and N-(aminobutyl)-N-(ethylisoluminol) functionalized ferritin as signal indicator for procalcitonin analysis. *Biosensors and Bioelectronics*, 2019, 142, 111562.
- [16] Xinrong Shao, Xianzhen Song, Xin Liu, et al. A dual signal-amplified electrochemiluminescence immunosensor based on core-shell CeO₂-Au@Pt nanosphere for procalcitonin detection. *Microchimica Acta*, 2021, 188, 344.
- [17] Yanrong Qian, Jinhui Feng, Huan Wang, et al. Sandwich-type signal-off photoelectrochemical immunosensor based on dual suppression effect of PbS quantum dots/Co₃O₄ polyhedron as signal amplification for procalcitonin detection.

Sensors and Actuators B: Chemical, 2019, 300, 127001.

- [18] Chunzhu Bao, Dawei Fan, Xin Liu, et al. A signal-offfff type photoelectrochemical immunosensor for the ultrasensitive detection of procalcitonin: Ru(bpy)₃²⁺ and Bi₂S₃ co-sensitized ZnTiO₃/TiO₂ polyhedra as matrix and dual inhibition by SiO₂/PDA-Au. Biosensors and Bioelectronics, 2019, 142, 111513.
- [19] Mengdi Wang, Xuejing Liu, Hongying Jia, et al. A novel approach to photoelectrochemical immunoassay for procalcitonin on the basis of SnS₂/CdS. New Journal of Chemistry, 2020, 44, 15281.
- [20] Xin Liu, Chunzhu Bao, Xinrong Shao, et al. A procalcitonin photoelectrochemical immunosensor: NCQDs and Sb₂S₃ co-sensitized hydrangea-shaped WO₃ as a matrix through a layer-by-layer assembly. New Journal of Chemistry, 2020, 44, 2452.