

Selenium Nanomaterials Enhance the Nutrients and Functional Components of

Fuding Dabai Tea

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Text S1 *HPLC-MS/MS operating parameters used in this study*

The components of theanine, catechins and alkaloids were detected by the high-performance liquid chromatography-tandem mass spectrometry (HPLC-MS/MS, Thermo Scientific, Germany). The fresh tea buds were ground into a powder. Then, samples were separated at 35 °C on a Thermo Scientific UPLC Vanquish equipped with a HSS T3 column applying the following gradient at a flow rate of 0.35 $\mu\text{L}/\text{min}$. The eluents in negative/positive mode were eluent A (0.01% v/v formic acid in water) and eluent B (0.01% formic acid in acetonitrile). The solvent gradient was set as follows: 5 % B, 0 min; 5 % B, 1 min; 95 % B, 11.0 min; 95 % B, 12min; 5% B, 12.1min; 5 % B, 14 min. The following dd-MS² product ion spectra were collected at resolution of 17500, isolation window of 3.0 m/z, and collision energy of nce: 15, 30, 40, respectively.

The metabolites in tea buds also were determined by HPLC-MS/MS. After the plant sample extraction, 10 μL samples were injected onto a Acquity HPLC HSS T3 (2.1 \times 100 mm, 1.8 μm) using an 18-min linear gradient at a flow rate of 0.35 $\text{mL}\cdot\text{min}^{-1}$. The eluents in negative/positive mode were eluent A (0.1% v/v formic acid in water) and eluent B (0.1% formic acid in acetonitrile). The solvent gradient was set as follows: 5% B, 0 min; 5% B, 1.5 min; 100% B, 10.0 min; 100% B, 11 min; 100% B, 11.5min; 5% B, 14 min. Nitrogen was used as sheath gas (35 $\text{L}\cdot\text{min}^{-1}$) and aux gas (15 $\text{L}\cdot\text{min}^{-1}$). The spray voltage was set as 3 kV (-3 kV for negative mode and 3 kV for positive mode) and capillary temperature was 320 °C. The resolution of full MS was set as 70000 and scan range was acquired between 70 to 1050 m/z in full MS-dd MS² mode. The following dd-MS² product ion spectra were collected at resolution of 17500, isolation

window of 1.5 m/z, and collision energy of nce: 20, 40, 60. The raw data files were processed using Compound Discoverer 3.1 software coupled with the mzCloud and S-7 Chem Spider libraries.

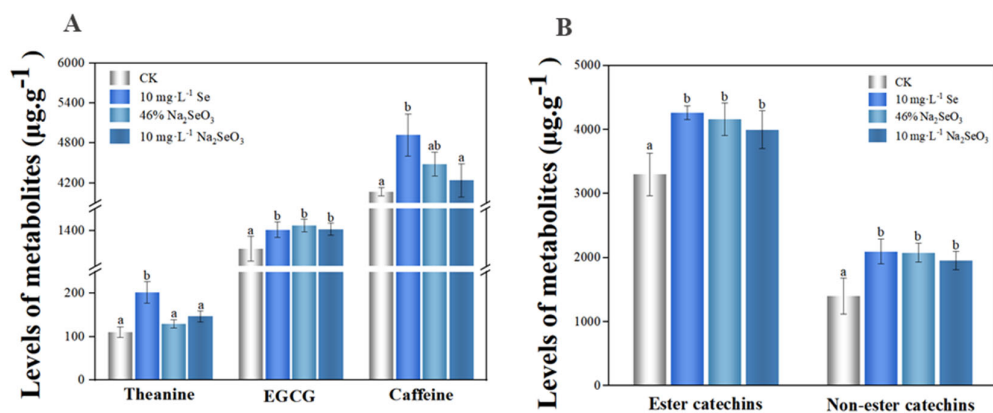


Figure S1. Changes in the content of (A) theanine, EGCG, caffeine and (B) esters, and non-ester catechins in FDDB treated with Se NMs and their equivalent ion control and fertilizer control. Letters following numbers indicate statistical significance - samples without common letters are significantly different at $p < 0.05$ according to Duncan and Toucan tests.

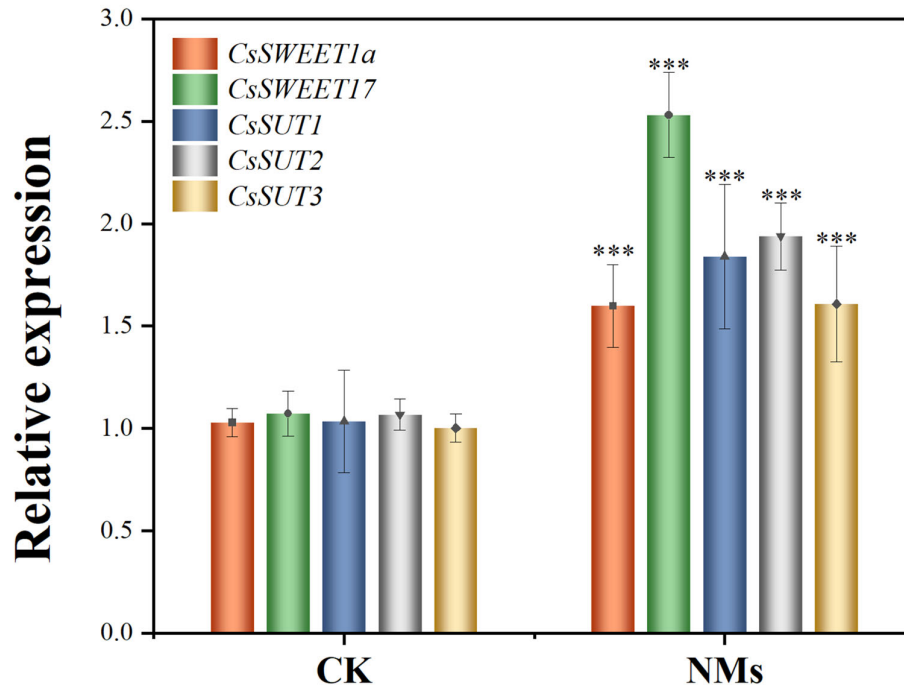


Figure S2. Relative expression of *CsSUT* and *CsSWEET* treated with Se NMs. The significant differences are marked with “***” ($p < 0.001$). The value represents the mean \pm standard error ($n = 5$).

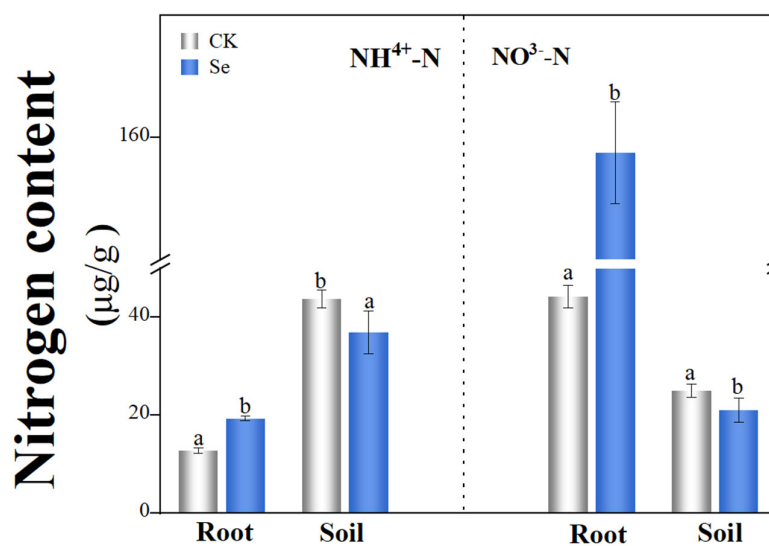


Figure S3. The content of ammonium and nitrate nitrogen in tea roots and rhizosphere soil under treatment with Se NMs. There was a significant difference between the treatment groups with different numbers ($p < 0.05$), the value represents the mean \pm standard error ($n = 5$).

Table S1 Hydrodynamic diameter and Zeta potential of Se NMs.

Name	Value	Unit
Hydrodynamic diameter	156.8 \pm 9.9	nm
Zeta potential	-24.0 \pm 1.6	mV

Table S2 *Primer sequences of several genes related to metabolism and synthesis in tea*

Gene name	Forward Primer	Reverse Primer
<i>GAPDH</i>	TTGGCATCGTTGAGGGTCT	CAGTGGAACACGGAAAGC
<i>CsTs1</i>	TCTTTCTGGACCTGTGAGTG	GCTTGAGGGTAGATAATGAGT
<i>CsGs1</i>	TAAAGTGCAACCGTCCCCTC	GTCCAAGCCAGTACCACCAA
<i>CsGOGAT</i>	CGAGGATCTTGCCCAACTGA	ACCCAGAAGCAACAGTACC
<i>CsPAL</i>	GAATGCCGGTCTTATCCACT	CGGTGAACACCTTGTCAAAC
<i>CsC4H</i>	CGAGAGGTTCTTGGAAGAGG	AGAATTGGCAGAGCAAGGAT
<i>CsCHI</i>	CACAAAGAAGATTATGGGTGAAG	CAAACCTAGAAGTTGCCAAGAGT
<i>CsANR</i>	GAATGCCGGTCTTATCCACT	CGGTGAACACCTTGTCAAAC
<i>CsLAR</i>	GAATGCCGGTCTTATCCACT	CGGTGAACACCTTGTCAAAC
<i>UGGT</i>	GGCAAGAAGCTAATAGGGTCGTT	TTGTATCATTTCGGAAGTGGTGGG
<i>CsDFR</i>	GTATTTGGACCGATGTGG	TTGTACTCCGGGTATTTT
<i>CsANS</i>	GGCCACAAGTGCCTACAATTG	CCCATGATTCACCAAATGCA
<i>CsAPP1</i>	GCTCAGTTGGGTGTTGGGTGGTTG	CGGCGAGGAGTGGAAGTGAA
<i>CsAPP2</i>	GCAGTGATTGGGTCAGGAGTGT	GCAGCGAGGAGAGTAGAAGTGT
<i>CsAPP5</i>	CTCTGTGGCTGCATGGGCTATG	TGGATGACTAGGCGGCATTGG
<i>CsAPP6</i>	TTGCGGTTGTCTTGGCTATGCT	CGGCTCTCTGCGAATGCGAATA
<i>CsAPP8</i>	CGGCACGGAATGGCTCTCAAT	TGGAGAACGGATAGGCGAAAGC
<i>CsSWEET1a</i>	AGCCAAGAAACCTCCTAATACC	TACTTGCTCGATCGCTTCTCTT
<i>CsSWEET17</i>	GGTTTCTCGGCAATACAGCATC	GGTGATCCTTGGTGCTTCCATT
<i>CsSUT1</i>	CATGCGATGCTAACTGTGCC	TTCAGCATGGCTCGACTGTT
<i>CsSUT2</i>	GGAATCGACTCCGATCTGCC	TGCAAAGCCCAACCGAATT
<i>CsSUT3</i>	TGTGGTCGGAGCTATTGCAG	CAGTGAGAAGGGGCAGAAGG