

Supplementary file

Article

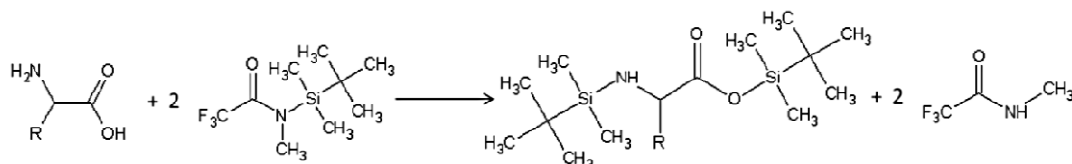
Highly Efficient Ru-Based Catalysts for Lactic Acid Conversion to Alanine

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Product analysis

Typically, the HPLC analysis of the reaction mixture containing amino acids is a complex one, performed with two different columns and a complex elution system, and requires derivatization with o-phthalaldehyde, the obtained derivatives being unstable. For this reason, the GC-MS method was chosen for the analysis of the products, which allows simultaneous separation and identification. Two commercial derivatizing agents, namely N,O-Bis(trimethylsilyl)trifluoroacetamide:trimethylchlorosilane 99:1 (BSTFA) and N-Methyl-N-tert-butyltrimethylsilyltrifluoroacetamide (MTBSTFA) were tested. During the derivatization reaction, the active hydrogen in -OH and -NH₂ groups is replaced by silane groups (for exemplification see Scheme S1). Among the two, MTBSTFA showed superior ability for alanine silylation and much better stability, which is also in agreement with the literature.



Scheme S1. Derivatization of aminoacids with MTBSTFA agent

High temperatures, often in the presence of acetonitrile, are required for complete derivatization of amino acids. Several derivatization temperatures were tested and the results are shown in Figure 1S. A complete dissolution of the reaction mixture and a higher relative abundance (%) were observed at 80 and 90 °C, respectively. However, no major differences were observed between the silylation results at 80 and 90 °C. Therefore, the derivatization was performed at 80 °C in order to avoid the formation of secondary derivatives.

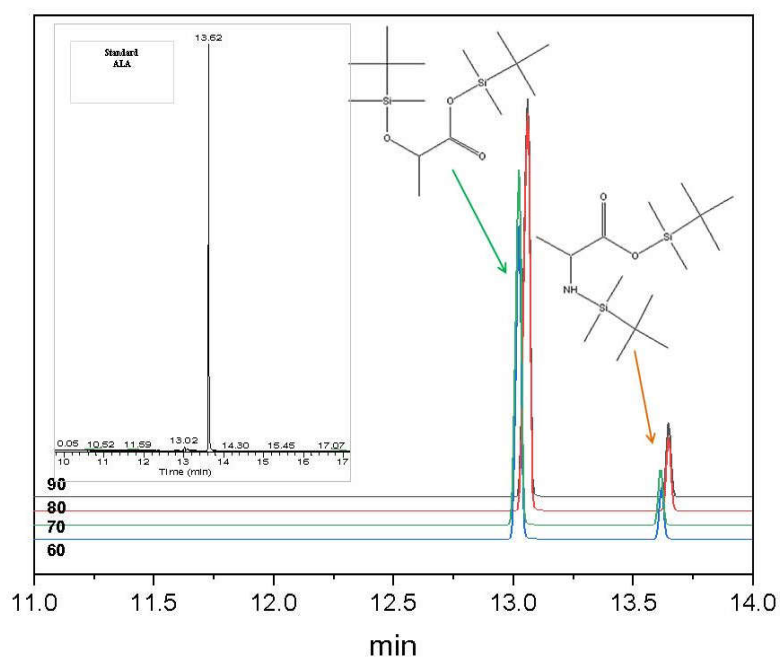


Figure S1. GC chromatograms obtained at different temperatures after derivatization with MTBSTFA for 4h. In figure is shown the peak for the derivatized standard of alanine (inset) and the derivatized alanine identified in the reaction in the following conditions: *catalyst* 5%Ru(III)/MNP - 50mg; *lactic acid*-0.35mmol; *temperature*-200°C; *NH₃-H₂O*-2.5mL; *H₂ pressure*-10atm; *reaction time*- 2h.

The mass spectra of the corresponding peaks were identified using the MS database, but also by injecting the derivatized standard compound under the same conditions. Figure S2 shows the mass spectra for the alanine peak in the reaction mixture compared to that in the database, which are identical at a probability of 93%.

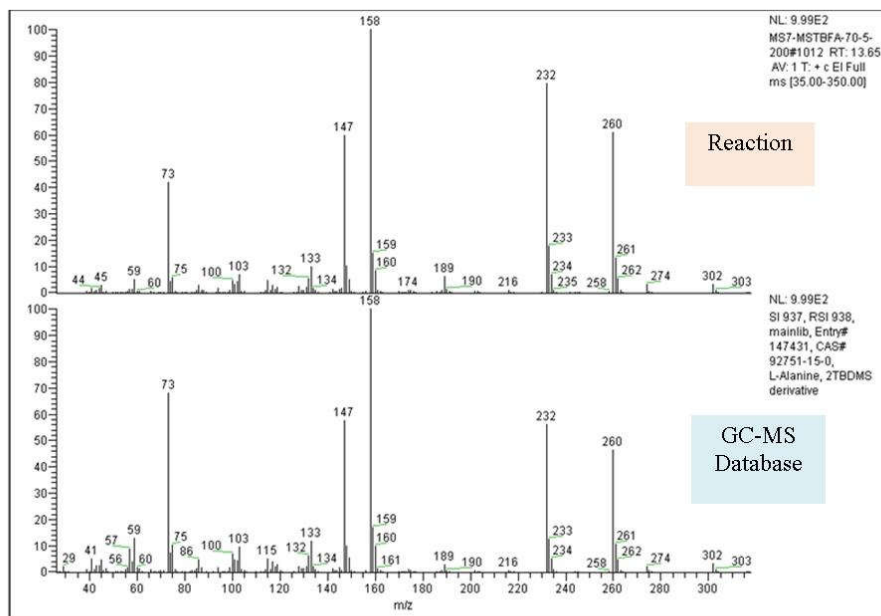


Figure S2. GC-MS identification of alanine: Mass spectra for alanine obtained in the reaction mixture compared to the database.

For additional confirmation, a UV-Vis spectrophotometric analysis of a reaction mixture was also performed using a calibration curve of alanine in $\text{NH}_3\cdot\text{H}_2\text{O}$ with ninhydrin. The calibration curve and the corresponding trendline are shown in Figure S3.

The concentration determined for sample was 0.158 mmol/mL, corresponding to 0.395 mmol in 2.5 mL and respectively a yield of 82%, comparable to that obtained by GC-MS (85%).

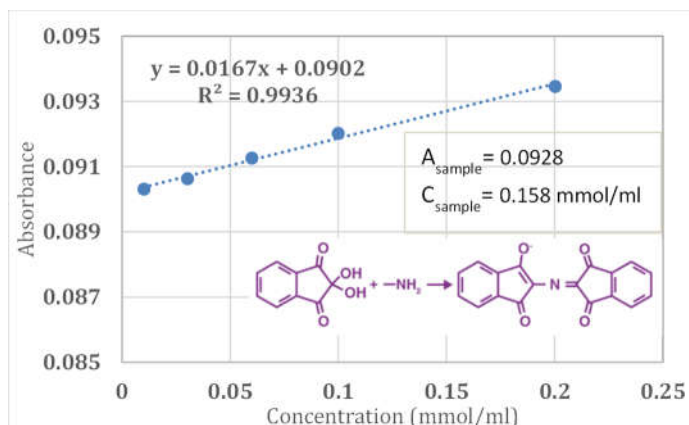


Figure S3. Calibration curve and the corresponding trendline equation for the analysis of alanine solutions in $\text{NH}_3 \cdot \text{H}_2\text{O}$ using UV-Vis spectrophotometry. The sample obtained after reaction under following reaction conditions: *catalyst 5%Ru(0)/MSN - 50mg; lactic acid-0.35mmol; temperature-200°C; $\text{NH}_3 \cdot \text{H}_2\text{O}$ -2.5mL; H_2 pressure-10atm; reaction time- 2h*

To confirm the successful separation and identification of all reaction products, a complete GC chromatogram of a reaction mixture is provided in Figure S4.

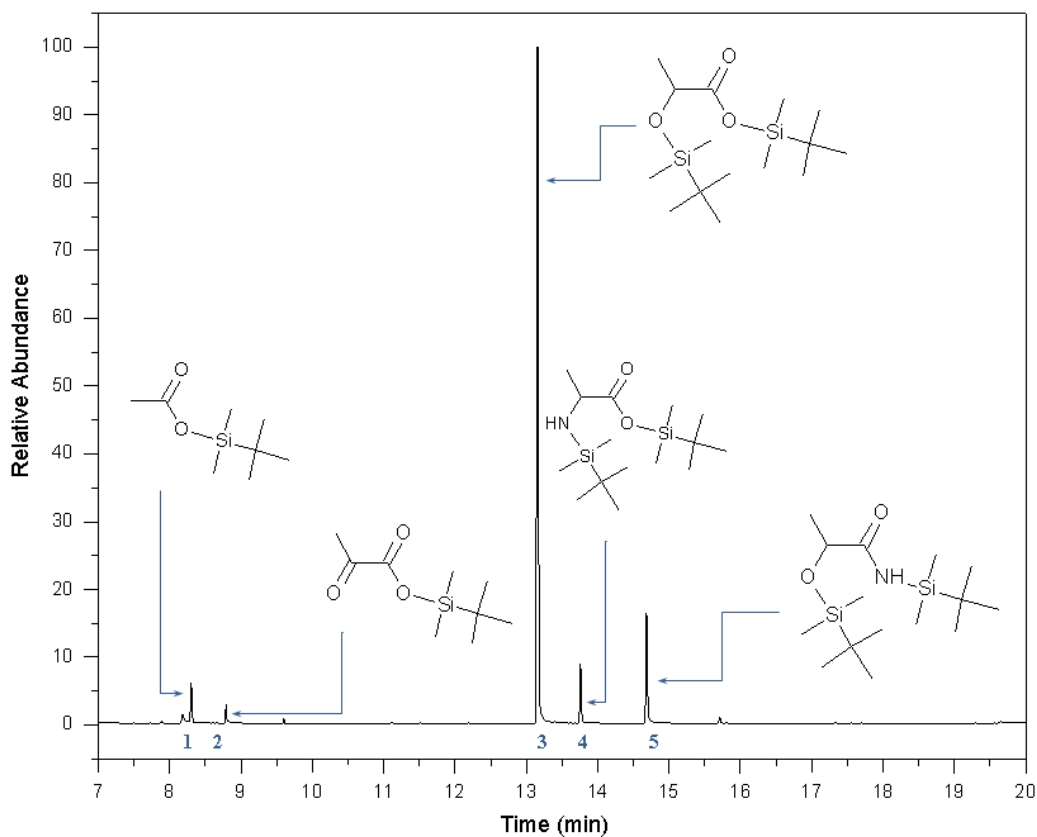


Figure S4. Representative GC chromatogram of the reaction products obtained in the following conditions: 5%Ru/CNT-25mg; lactic acid-0.7mmol; temperature-210°C; $\text{NH}_3 \cdot \text{H}_2\text{O}$ -2.5mL; H_2 pressure-10atm; reaction time- 2h. Where: 1-acetic acid (AA); 2- pyruvic acid (PA); 3 - lactic acid (LA); 4-alanine (AL); 5-lactamide (LAM).

Liquid nitrogen adsorption-desorption isotherms at -196 °C of Ru/CNT samples

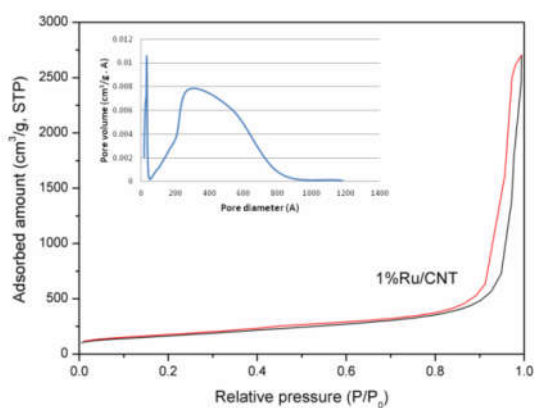


Figure S5. Nitrogen adsorption-desorption isotherm and pore size distributions (inset) for 1%Ru/CNT sample

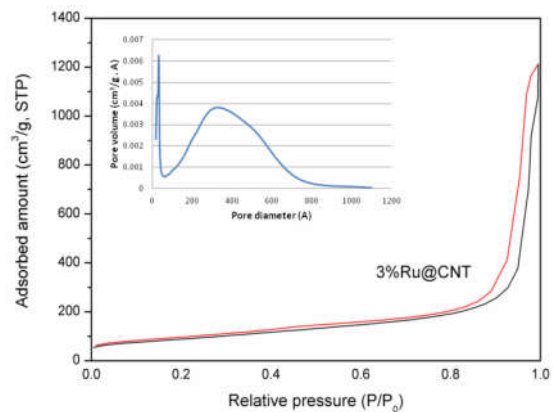


Figure S6. Nitrogen adsorption-desorption isotherm and pore size distributions (inset) for 3%Ru/CNT sample

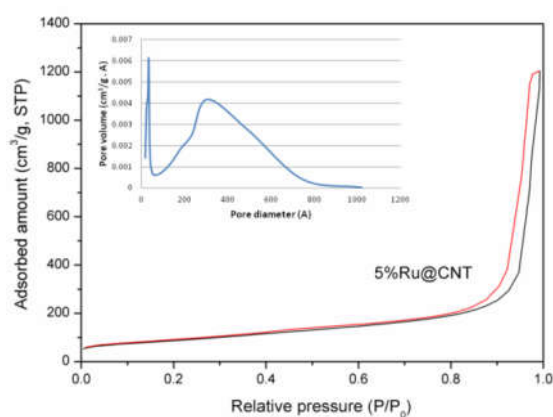


Figure S7. Nitrogen adsorption-desorption isotherm and pore size distributions (inset) for 5%Ru/CNT sample

STEM microscopy

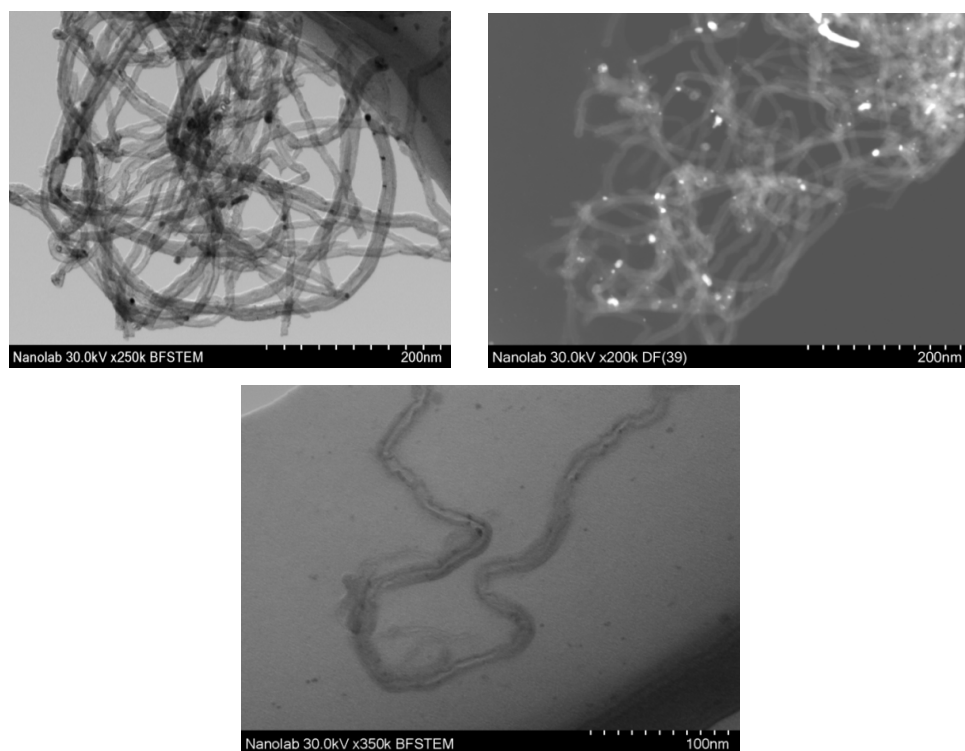
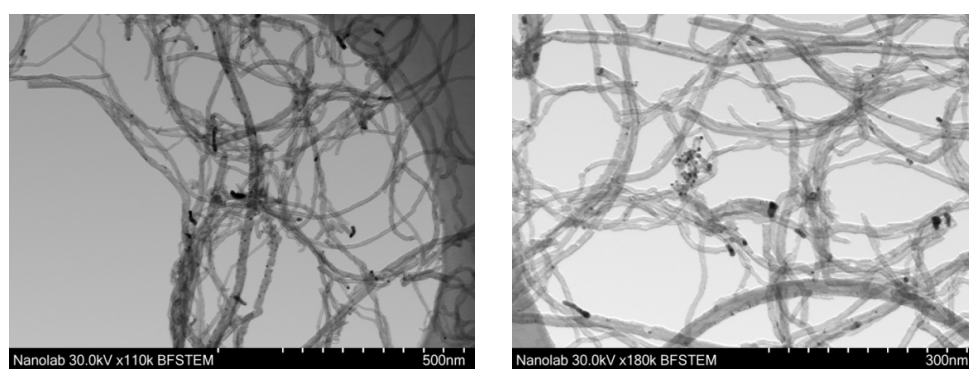


Figure S8. BF and DF-STEM images for the 3%Ru/CNT sample



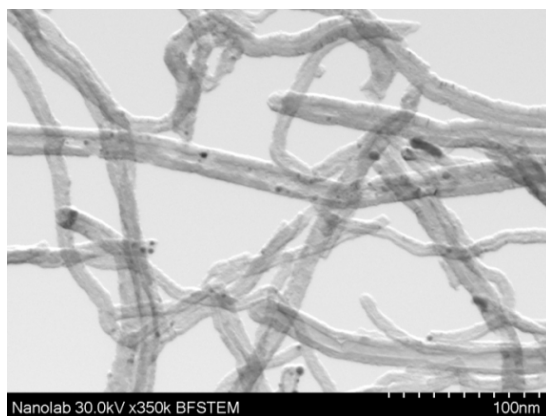


Figure S9. BF-STEM images for the 5%Ru/CNT sample

H₂-TPD

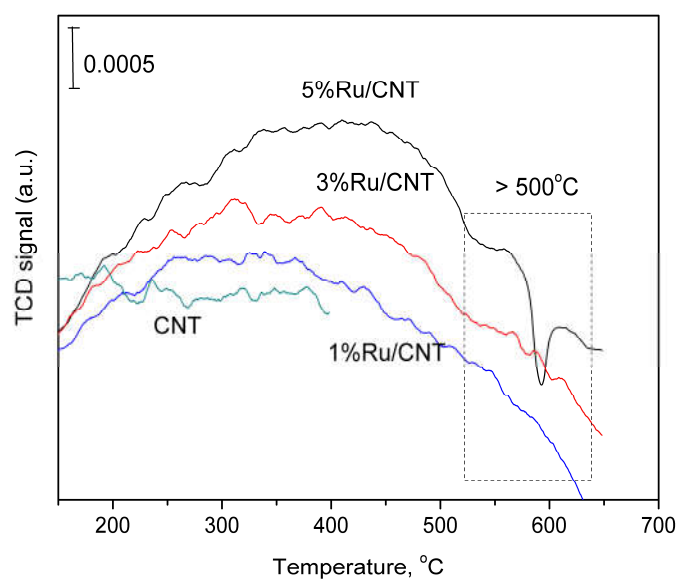


Figure S10. H₂-TPD profiles for the Ru/CNT samples

Table S1. H₂-TPD parameters for Ru/CNT samples and MWCNT carrier

Entry	Sample	Temperature at Maximum (°C)	H ₂ desorbed (mmol/g)
1	CNT	-	0

2	1%Ru/CNT	324	0.007
		338	0.002
		430	0.007
		489	0.010
		TOTAL	0.026
3	3%Ru/CNT	311	0.017
		565	0.015
		587	0.003
		610	0.003
		TOTAL	0.038
4	5%Ru/CNT	306	0.035
		561	0.005
		609	0.006
		TOTAL	0.046

NH₃- TPD results for Ru/BEA catalysts

Table S2. Acidic characteristics of the pristine BEA zeolite and 3%Ru/BEA catalyst, determined from the NH₃- TPD.

Sample	Range of temperature (°C)				Total sites conc. (μmols /g)
	50-100	100-200	200-300	300-400	
BEA zeolite	17.8	80.5	-	10.2	108.5
3%Ru/BEA	-	65.8	34.2	-	100.0

Liquid nitrogen adsorption-desorption isotherms at -196 °C of Ru/BEA samples

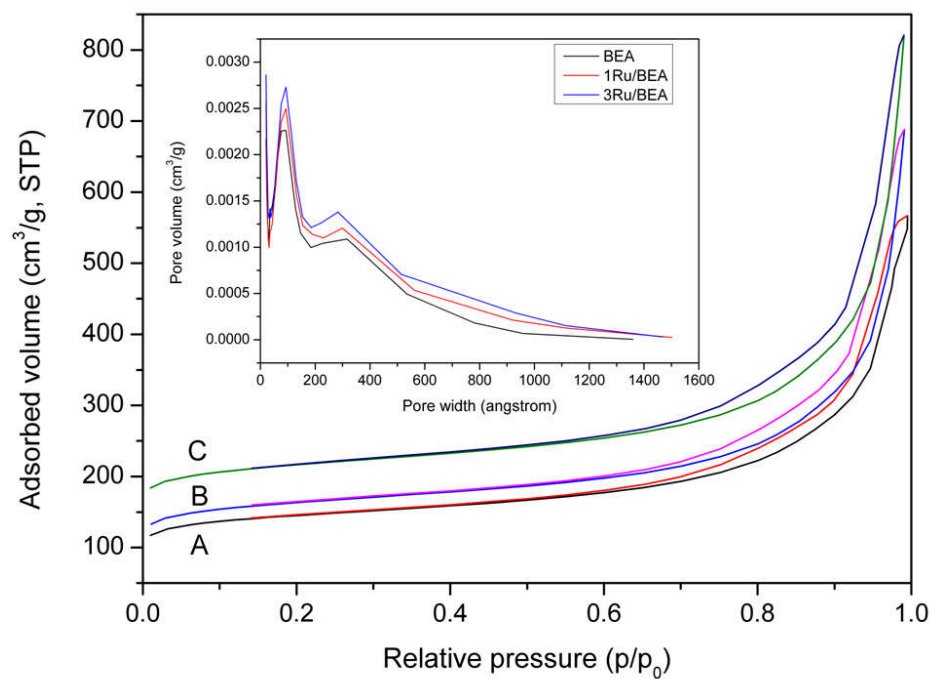


Figure S11. Adsorption-desorption isotherm of liquid nitrogen at -196°C of BEA (A), 1%Ru/BEA (B) and 3%Ru/BEA (C) sample. Inset: Pore size distribution