

Supporting Materials

Asymmetric Schottky Barrier-Generated MoS₂/WTe₂ FET Biosensor Based on a Rectified Signal

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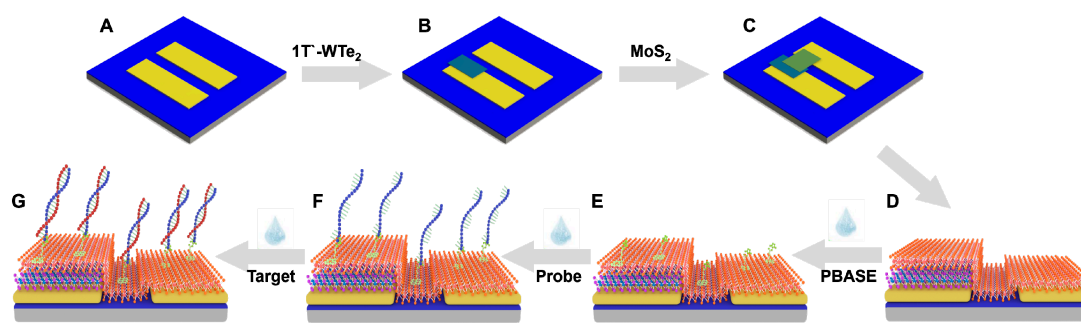


Figure S1. Fabrication process diagram of MoS₂/WTe₂ FET biosensor. (A) A clean and tidy Si/SiO₂ substrate is prepared. (B) WTe₂ is transferred by using a two-dimensional material transfer platform. (C) MoS₂ is transferred by using a two-dimensional material transfer platform. (D) A MoS₂/WTe₂ Schottky heterojunction is formed. (E) Add PBASE on the MoS₂/WTe₂ FET biosensor. (F) Add probe DNA on the MoS₂/WTe₂ FET biosensor. (G) Add target DNA on the MoS₂/WTe₂ FET biosensor.

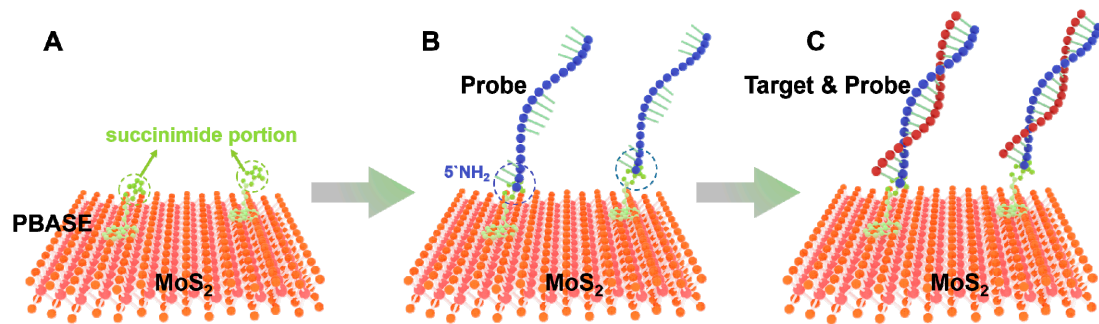


Figure S2. The functionalization and immobilization process of the device. (A) PBASE, when dissolved in DMSO, stacks its pyrene group on the surface of MoS₂, binds to MoS₂. (B) The succinimidyl ester group of PBASE is connected to the amino-modified probe DNA through a cross-linking reaction. (C) The target DNA is combined with the probe DNA through complementary base pairing.

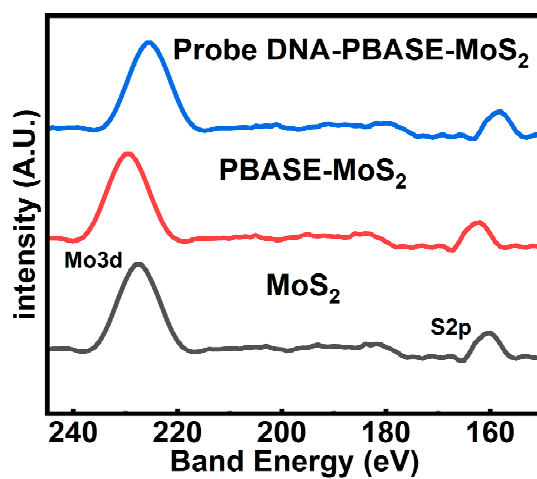


Figure S3. High-resolution XPS spectra of Mo 3d, S 2p. The movement of Mo 3d peak and S 2p peak confirmed that PBASE and probe DNA were successfully combined on the surface of MoS₂/WTe₂ FET biosensor.

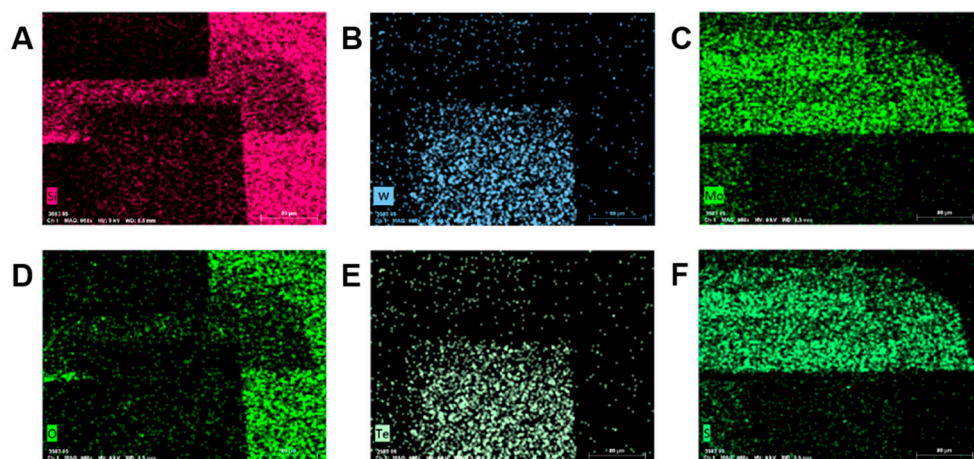


Figure S4. EDS images mentioned in Figure 3B. The above EDS images represent (A) Si, (B) W, (C) Mo, (D) O, (E) Te and (F) S elements respectively.

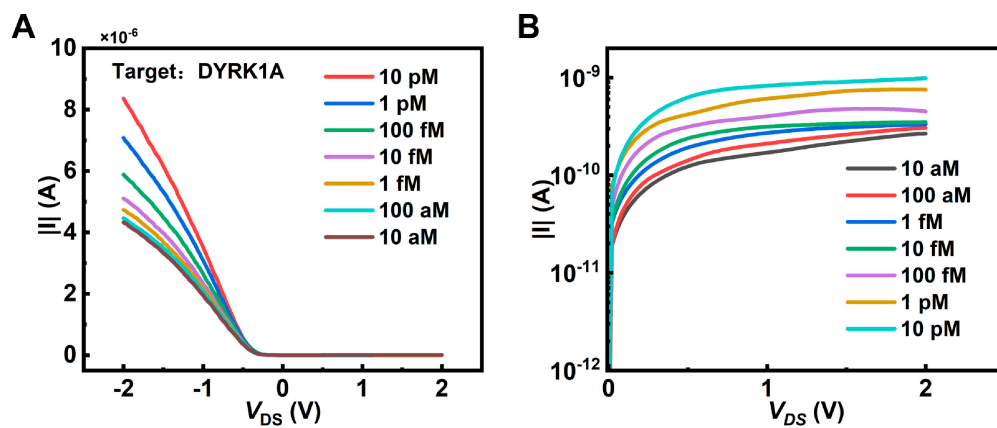


Figure S5. The current results of different concentrations of target DNA (DYRK1A) under (A) positive and (B) negative bias voltages.

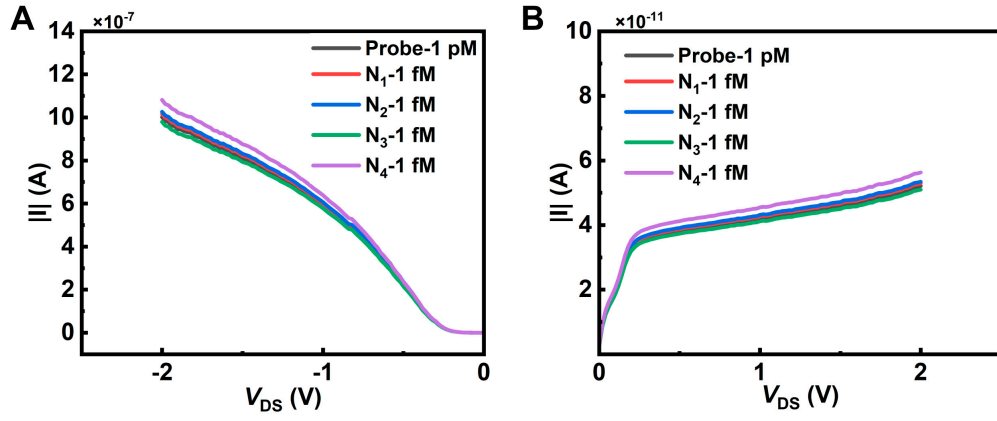


Figure S6. The complete current diagram after adding N₁-N₄ samples. Source-drain current under (A) negative bias and (B) positive bias in linear scale.

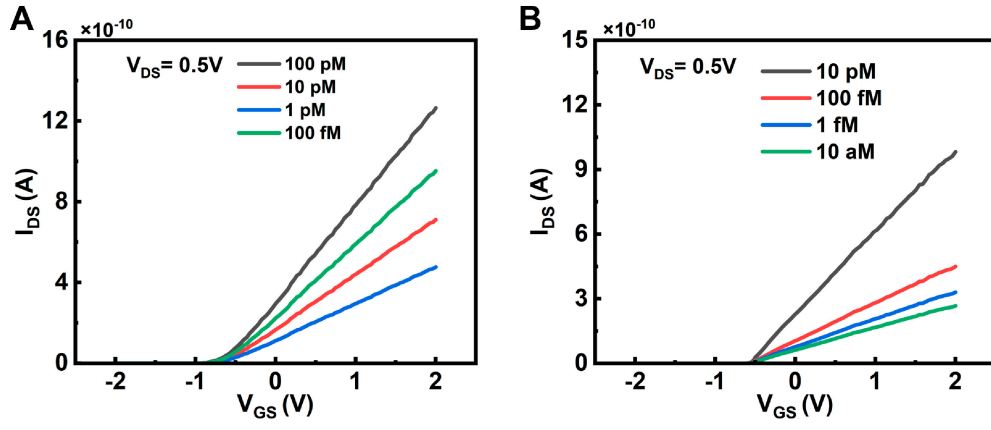


Figure S7: The transfer characteristics curves of the (A) Au/MoS₂/Au and (B) WTe₂/MoS₂/Au structures, respectively.

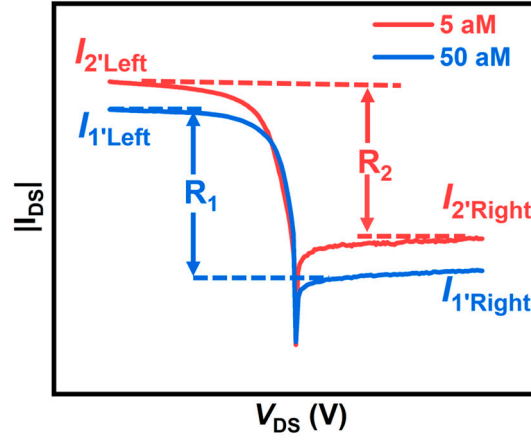


Figure S8. Output characteristics curves measured at different biomolecular concentrations in the MoS₂/WTe₂ FET biosensor. I_1 and I_2 respectively represent the source-drain current at different concentrations, I_{Left} and I_{Right} respectively represent the source-drain current dominated by different Schottky barriers, R_1 and R_2 respectively represent the rectification ratio of MoS₂/WTe₂ FET biosensor at different concentrations

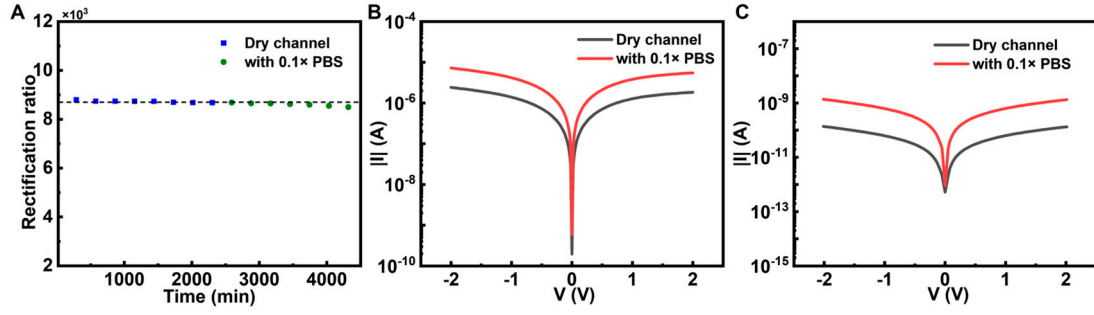


Figure S9. Stability comparison of “rectified signal” and “absolute value signal”. The output current curves of (A) $\text{WTe}_2/\text{MoS}_2/\text{Au}$ FET, (B) $\text{WTe}_2/\text{MoS}_2/\text{WTe}_2$ FET, and (C) $\text{Au}/\text{MoS}_2/\text{Au}$ FET under external factors such as water molecules, respectively. Under external factors such as water molecules, the rectification ratio signal of the $\text{WTe}_2/\text{MoS}_2/\text{Au}$ FET biosensor is stable within long times, as shown in Supplementary Figure 3A.

Table S1. The DNA sequences purchased from Sangon Biotech (Shanghai) Co., Ltd

Names	Sequences (5'→3')
Probe DNA in Figs. 1B-1C, 1H-1I, 2I, 3B-3F	NH ₂ -C ₆ -CAGAGTCACTGACTCACTA
Target DNA in Figs. 1C,1I,2I,2B-2D	TTAGTGAGTCAGTGACTCTG
RNA in Figs. 3F-3G	CAGGUGGAACCUCAUCAGGAGAUGC
T1 in Figs. 3F-3G	TTTTTTTTTTUUUUUCCCGTCCGTATGG
T2 in Figs. 3F-3G	TCTCAAGGACCACCGCATCTCTACCCATAC GGACGGG
3-base Mismatch DNA in Figs. 3F-3G	TTAGTGAGTCAGTGACTGAC

Target DNA sequence in Figs. 1C,1I,2I,2B-2D was selected from GeneBank: NG_009366.2

Table S2. Comparison of biosensing performance among various FET biosensors

Sensing materials	Functional- ization methods	Probe types	Target biomarkers	LOD	Signal types	Ref
Graphene	PBASE	ssDNA	Antigen	2.6 pM	V_{Dirac} ; I_{DS}	[1]
rGO	PBASE	Antibody	Antigen	0.1 pg/mL	V_{Dirac}	[2]
Graphene	AuNPs	ssDNA	20-mer DNA	15 aM	V_{CNP}	[3]
Mxenes/graphene	APTES	Antibody	Antigen	1 fg/mL	V_G ; I_{DS}	[4]
MoS ₂	Physically adsorb	Antibody	Antigen	1 pM	V_{th}	[5]
MoS ₂	AuNPs	ssDNA	30-mer DNA	10 aM	I_{DS}	[6]
rGO	Physically adsorb	ssDNA	48-mer DNA	5 pM	I_{DS}	[7]

Exfoliated- Graphene	EDC+NHS	Antibody	Antigen	10 fg/mL	Resist- ance	[8]
MoS ₂ /WTe ₂	PBASE	ssDNA	20-mer DNA	10 aM	Recti- fication ratio	This work

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