



Nanotechnology in the Restoration of Polluted Soil

Vishnu D. Rajput ^{1,*}, Tatiana Minkina ¹, Sudhir K. Upadhyay ², Arpna Kumari ¹, Anuj Ranjan ¹, Saglara Mandzhieva ¹, Svetlana Sushkova ¹, Rupesh Kumar Singh ³ and Krishan K. Verma ⁴

- ¹ Academy of Biology and Biotechnology, Southern Federal University, 344090 Rostov-on-Don, Russia; tminkina@mail.ru (T.M.); kumari@sfedu.ru (A.K.); randzhan@sfedu.ru (A.R.); msaglara@mail.ru (S.M.); terra_rossa@mail.ru (S.S.)
- ² Department of Environmental Science, V.B.S. Purvanhal University, Jaunpur 222003, India; sku.env.lko@gmail.com
- ³ InnovPlantProtect Collaborative Laboratory, Department of Protection of Specific Crops, 7350–999 Elvas, Portugal; rupeshbio702@gmail.com
- ⁴ Guangxi Academy of Agricultural Sciences, Nanning 530007, China; drvermakishan@gmail.com
- * Correspondence: rajput.vishnu@gmail.com; Tel.: +7-918-589-00-93

Abstract: The advancements in nanoparticles (NPs) may be lighting the sustainable and eco-friendly path to accelerate the removal of toxic compounds from contaminated soils. Many efforts have been made to increase the efficiency of phytoremediation, such as the inclusion of chemical additives, the application of rhizobacteria, genetic engineering, etc. In this context, the integration of nanotechnology with bioremediation has introduced new dimensions for revamping the remediation methods. Hence, advanced remediation approaches combine nanotechnological and biological remediation methods in which the nanoscale process regulation supports the adsorption and deterioration of pollutants. Nanoparticles absorb/adsorb a large variety of contaminants and also catalyze reactions by lowering the energy required to break them down, owing to their unique surface properties. As a result, this remediation process reduces the accumulation of pollutants while limiting their spread from one medium to another. Therefore, this review article deals with all possibilities for the application of NPs for the remediation of contaminated soils and associated environmental concerns.

Keywords: pollution; heavy metals and metalloids; phytoremediation potential; phytorestoration strategy; nanotechnology

1. Introduction

Rapidly increasing anthropogenic/technogenic activities are adding potentially toxic metals, agrochemicals, and an excess of nutrients to the soil [1]. Soil is a basis of crop production as it supports plants to uptake nutrients [2]. In fact, agriculture sustains and defines human lives; however, it is often disruptive of natural ecosystems. Humans' voracious appetites for getting the benefits from natural resources grow in tandem with population growth. The conflict between the benefits and the sustainable management of agricultural land and its conservation has been reported in the literature for a long time [3,4]. Land pollution is a threat to livelihoods, quality of life, and sustainable development [5]. Thus, conserving soil is the utmost requirement for the current era owing to the pressures of increasing population and the shrinking of arable lands by technogenic activities.

The advancements in nanotechnology open a window globally to remediate or restore polluted soil in an effective way [6,7]. It has been claimed that nanotechnology has great potential as an environmentally cleaner technology, including by alleviation of the toxicities of various metals/metalloids [8,9]. Besides, nanotechnology has been recognized as a potential method for the remediation of pollutants in a variety of environmental matrices, including soils [6]. In this context, soil remediation is one of the main domains where nanotechnological approaches have been widely used. The uses of NPs have been

Citation: Rajput, V.D.; Minkina, T.; Upadhyay, S.K.; Kumari, A.; Ranjan, A.; Mandzhieva, S.; Sushkova, S.; Singh, R.K.; Verma, K.K. Nanotechnology in the Restoration of Polluted Soil. *Nanomaterials* **2022**, *12*, 769. https://doi.org/10.3390/ nano12050769

Academic Editor: Andreu Cabot

Received: 29 December 2021 Accepted: 23 February 2022 Published: 24 February 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/). explored lately to remove contaminants in a variety of ways, including by adsorption, redox reactions, precipitation, and co-precipitation, all of which are aided by their enormous specific surface area [9].

With the help of NPs, hyperaccumulators and indigenous soil microbes could enhance biodegradation processes, thereby increasing the potential extent of remediation. This could be called nano-phytoremediation and microbial-mediated nano-remediation. The use of NPs with bioremediation approaches may result in a lot of benefits as these particles are small (1–100 nm) with a larger surface area and reactivity [10]. The broad range of studies indicated that the foliar use of NPs alleviates metal-pollutant toxicity and enhances plant growth, resulting in a high accumulation of elemental toxic content in plant tissues [11–13]. The degree of contamination, the bioavailability, and the accumulation of metals by the plants are decisive for the efficiency of nano-phytoremediation as a way of removing heavy metals (HMs) from contaminated sites [14,15].

The utilization of NPs based on a metal–organic framework (MOF) has received a lot of important attention lately, but it is primarily employed for drinking water and wastewater treatment [16–18]. Thus, there is a massive opportunity for scientists to envisage potential uses of MOF in soil remediation. However, concerns related to the safe use of NPs to remediate polluted soils, and their release into ecosystems, are still less explored, and this becomes a matter of concern [19,20]

The present review unbinds the possibilities for the elimination of contaminants, with particular emphasis on microbe-mediated remediation, hyperaccumulator plants, and NPs, as well as the approaches to restore HM-contaminated soils, the benefits, and the potential risks associated with nano-bioremediation technologies. The gaps and future perspectives are comprehensively elaborated.

2. An Appraisal of Nanobioremediation-Based Removal of Pollutants; Special Emphasis on Microbe-Mediated Remediation

Soil rich in vital nutrients and micronutrients is believed to best support the optimum health of the plant and its growth [21]. Human-made activities have currently polluted the soil with a variety of persistent organic compounds, viz., polyaromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), volatile organic compounds (VOCs), HMs (Hg and Pb), agrochemicals (pesticides, fungicides, and fertilizers), and also sometimes with excess nutrients [22]. At the same time, urbanization and industrialization have also added solid wastes, varieties of chemicals, and solvents to the environment and agricultural soil [23].

Nanobioremediation is a cost-effective technique of utilizing plants and microbes for the breakdown of pollutant compounds, ultimately improving soil quality and reducing pollution. By breaking down contaminants in the soil, the process may be able to eradicate, retain, or reduce the amount of pollutants present [24,25]. The efficiency of bioremediation has been studied and enhanced in the past using chemical additives or biotechnology [26] but nanotechnology further improved the process with a newer aspect [27]. A summary of different NP-mediated removal of pollutants from contaminated media is elaborated in Table 1.

Nanobioremediation implies both nanotechnology and bioremediation together, where the process is executed at the nanoscale. The target pollutants are adsorbed, degraded, or modified owing to the unique physicochemical properties of the NPs, which also act as catalysts and help to reduce the activation energy required for breaking down the compounds [28]. The nanobioremediation process has been explored and studied, and the most exploited NPs are carbon- and metal-based [29,30]. Polymeric NPs in the form of nanocapsules or nanospheres are also exceptional in the elimination of persistent pesticide compounds and long-chain hydrocarbons [31]. However, in the case of HMs, the challenge is entirely different as they are non-biodegradable, as well as very prone to entering biological systems and food chains [32]. Biosorption and bioaccumulation using plants and microbes are traditional methods to remove HMs from polluted soils. However, recent pieces of evidence have reported the use of NPs in HM remediation with remarkable outcomes [33]. Nanoparticles are reported to have been applied in combination simultaneously or sequentially with specific microbes and the results have been convincing [34]. They could help to speed up the elimination of HMs by acting as nanocarriers of microbes or microbial biosorbents [35]. A pictorial diagram that represents the process of nanobioremediation, especially for biogenic NPs, is depicted in the Figure 1.

The synergy of NPs and bacterial degradation has also gained attention; however, the availability of a handful of published papers would not permit the making of a categorical review. Many authors have given their efforts to nanobioremediation, and yet it is still too insignificant for making any conclusive decision [10].



Figure 1. An overview of the processes of nanobioremediation using biogenic nanoparticles.

Integration of NPs with microbes for bioremediation is a two-phasic process that involves overlapping abiotic and biotic processes (Figure 1) [36]. In the first phase, after the entry of NPs into the system, pollutants undergo varieties of physicochemical processes and modifications depicting abiotic processes such as absorption, adsorption, dissolution, and chemical catalysis of photocatalytic reactions [37]. The second phase includes biotic processes such as biocides, bioaccumulation, biostimulation, and biotransformation [38,39]. These biotic processes play a crucial role in the removal of pollutants from the system.

2.1. Nanobioremediation of Heavy Metals

The existence of HMs in the environment is largely due to increased anthropogenic activities. However, disturbed biogeochemical cycles are also responsible for their release into the environment as pollutants. Elements like As, Cd, Cr, Hg, and Pb have no biological functions to perform in the biological system. Heavy metals comprise major inorganic pollutants as they exhibit substantial toxic impacts on biota even at the lowest concentrations [40,41]. The toxicity of HMs also rests on their bioavailability and absorption [42].

Acidic environments instigate the toxicity of HMs, especially if the soil structure is poor and has low nutrients (e.g., mining areas) [43].

Table 1. Summary of different nanoparticles-mediated removal of different pollutants from contaminated media.

Nanoparticles	Remediated Contami- nant(s)	Operational Conditions and Removal Efficiency	References
Polyvinylpyrrolidone (PVP) coated iron oxide nanoparticles	Cd and Pb	NPs applications were integrated with the process of bi- oremediation mediated by <i>Halomonas</i> sp. In the removal setup of Cd and Pb, <i>Halomonas</i> sp. was inoculated for 48 h at 180 rpm, 28 °C. The 100% removal was recorded after 24 h, while for Cd, it was observed after 48 h.	[44]
Zero-valent iron (nZVI) commercial suspension at two doses (1% and 10%)	As	pH was set at 12.2 ± 0.1 of the nZVI suspension. To avoid the aggregation of nZVI in the suspension, polyacrylic acid was used as a stabilizer. Maximal immobilization of As in brownfield soil was recorded at 10% of nZVI. Applications of nZVI and nGOx to the polluted soils	[45]
Graphene oxide nano- particles (nGOx) and nZVI	Metals, viz., Cd, Pb, Zn, Cu, and As in the As- Metals polluted soil	Cu, Pb, and Cd were immobilized by nGOx, while mo- bilized As and P. In a turn of nZVI, it immobilized the effectively As and Pb, and poorly Cd but enhanced availability of Cu. This study revealed that both NPs applications might be act as strategies for the immobilization and stabiliza-	[46]
Titanium oxide nano- particles-bonded-chi- tosan nanolayer (NTiO2- NCh)	Cd and Cu	 tion that can later be utilized for phytoremediation. The pH was set at 7.0 during the experimentation. The removal was assisted by 60–70 s heating by using microwave–enforced sorption approach. Application of NTiO₂-NCh was found to eliminate Cu and Cd by 88.01% and 70.67%, respectively. 	[47]
Palladium (Pd), Pd NPs	Cr	It was found that Pd NPs completely reduced Cr ⁶⁺ in 12 h. 6.3 mg of PdNPs was used to reduce 5.0 umol of Cr ⁶⁺	[48]
Magnetic iron oxide na- noparticles (Fe ₃ O ₄ NPs) treated with <i>Staphylococ-</i> <i>cus aureus</i> , and surface encapsulated with phthalic acid (n-Fe ₃ O ₄ - Phth-S <i>aureus</i>)	Cu, Ni, Pb	n-Fe ₃ O ₄ -Phth-S was found to remediate 83.0–89.5%, for Cu ²⁺ , 99.4–100%, for Pb ²⁺ , and 92.6–7.5% for Ni ²⁺ . The study also identified n-Fe ₃ O ₄ -Phth- <i>S. aureus</i> as an excellent biosorbent for the removal of divalent ions from an aqueous medium.	[49]
ZnO NPs	Cu, Cd, Cr, and Pb	The applications of ZnO-NPs at 5 mgL ⁻¹ with <i>Bacillus cereus</i> and <i>Lysinibacillus macroides</i> showed the maximal removal of Cr, Cu, and Pb which was 60%, 70%, and 85%, respectively. The optimal pH for efficient removal was 8.0. The removal was less in the case of bacteria-mediated remediation which was found to be 83 and 70% in <i>B. cereus</i> and 60 and 65% in <i>L. macroides</i> .	[50]

Heavy metals primarily affect the plants and lower soil organisms by inducing the generation of reactive oxygen species (ROS), which further results in the damage of macromolecules such as proteins and nucleic acids [41]. The existence of HMs in the soil affects crops and vegetation, their nutritional quality, and the ecological aspects associated with them. The effect of HMs on crops varies depending on the crop species, soil physicochemical characteristics, and HM type [51]. The general mechanism of toxicity exerted by HMs on crop plants includes ROS generation, which affects the cell organelles, macromolecules such as proteins and nucleic acids, and other components of the plant's structure and function [51,52]. It has also been reported to affect respiration and photosynthesis, reduce enzyme activities, elevate oxidative stress, reduce biomass, diminish crop yield, and affect the abundance, activity, diversity, and genetic makeups of useful soil microflora [53,54].

One of the key methods for the elimination of HMs includes site stabilization that immobilizes them at a specific site to decreases mobility and availability in the soil, and stops them from leaching across the sites [55]. The use of various NPs, including biogenic, has been gaining a lot of attention for the removal of HMs [56]. Biogenic NPs are those that are synthesized using biological organisms. The commonly known biogenic NPs, such as Ag NPs, are formed by *Morganella psychrotolerans* [57,58].

Nanoparticles of FeO coated with polyvinylpyrrolidone (PVP) have been successfully used for improving the bioremediation of the soil contaminated with Pb and Cd by a Gram-negative bacteria, *Halomonas* sp. This approach has significantly removed nearly 100% of Pb after 24 h, and Cd after 48 h, as compared to removal by bacteria or only NPs [59]. A biosorbent of magnetic Fe₃O₄ NPs treated with *S. aureus*, with a surface encapsulated with phthalic acid (as a n-Fe₃O₄-Phth-S complex), was used for the removal of Cu, Ni, and Pb, and the adsorptive removal of 795, 1355, and 985 µmol g⁻¹ for Cu, Pb, and Ni was achieved, respectively. In terms of percentage, the recovery rates of 83.0–89.5% for Cu²⁺, 99.4–100% for Pb²⁺, and 92.6–7.5% for Ni²⁺ were observed. The comparative study with dried *S. aureus* and n-Fe₃O₄-Phth-S for HM removal inferred that the n-Fe₃O₄-Phth-S core of the NPs, as well as the functional groups present on the microbial surface, played a key role in the removal of HMs [49]. Thus, this work revealed that the core of the NPs, as well as functional groups present on the microbial surface, had an important impact on the elimination of the contaminants.

A recent study on the removal of Cu, Cd, Cr, and Pb using HM-resistant bacteria such as *B. cereus* (PMBL-3) and *L. macroides* (PMBL-7) evidently confirmed that ZnO NPs at 5 mg L⁻¹ synergistically removes the Cr by 60%, the Cu by 70%, and the Pb by 85%, as compared to *B. cereus* (80 and 60%) and *L. macroides* (55 and 50%) at neutral pH, respectively [46]. At neutral pH the surface of ZnO NPs exhibit negative charges that promote electrostatic interactions with metal cations; however, at lower pH, the HMs get precipitated as hydroxides and then hydrogen ions compete for binding with adsorbents [60]. The strain XMCr-6 of *B. cereus* has also been reported to reduce the Cr⁶⁺ through an enzyme-mediated process. The reduced Cr³⁺ was observed to have a binding affinity to cells using coordination bonds with the functional group present on the surface of the bacterial cell wall. The formation of Cr₂O₃ NPs was found on the cell surface as a by-product [61].

The use of probiotic bacteria (*L. casei* and *L. fermentum*) to absorb Cd from water in association with Se⁵⁺ and Se NPs was also investigated. The higher absorption of Cd by *L. casei* with Se⁴⁺ ions (65%), compared to Se NPs (55.90%), was discovered in this study, and it was correlated to the higher solubility of Se⁵⁺ compared to Se NPs. When comparing *L. fermentum* and *L. casei*, the efficiency of Cd absorption was significantly higher in *L. fermentum* (50.87%) than *L. casei* (43.78%). The percentage of Cd adsorption by *L. casei* when used in conjunction with Se NPs shows no significant change. However, with increased Se NPs ratio percentages, Cd absorption was slightly increased from 5.49 to 16.54 in the presence of *L. casei* with Se NPs, compared to *L. casei* [62].

A threefold approach is now gaining popularity as the HM pollutants can be used by selective microbes to synthesize biogenic NPs (resource recovery), thereby removing them from the environment (remediation) and yielding value for the waste (effective waste

utilization). A study using *Enterococcus faecalis* for biorecovery of Pd as Pd NPs reported the synthesis of intra- and extra-cellular (membrane-bound). The range of Pd NPs was observed as 10 nm by transmission electron microscopy; however, the size of the Pd NPs was dependent on environmental conditions such as temperature, pH, and biomass. The obtained Pd NPs have great use as a bionanocatalyst that shows good catalytic efficiency (6.3 mg Pd NPs completely reduced 5.0 μ mol Cr⁶⁺ in 12 h) and the application is potentially useful to treat industrial effluents [44].

A similar study produced Te NPs from anaerobic sludge based upon supplementation with riboflavin [63]. It formed insoluble elemental tellurium (Te⁰ NPs) using pollutant tellurite Te⁴⁺ oxyanions present in the wastewater. It has been reported that 2-Hydroxy-1,4-naphthoquinone promotes the reduction of Te⁴⁺ and the quantity of Te⁰NPs synthesis [64]. The process is supported by *Rhodobacter capsulatus*, where malate is the electron-donating substrate [65], and riboflavin speeds up the rate of Te⁴⁺ reduction by anaerobic methanogenic granular sludge [66].

2.2. Degradation of Persistent Organic Pollutants

The pollution posed by POPs has been shown to have a negative influence on both the environment and human health, as certain POPs have been found to bioaccumulate in adipose tissue and to have the potential to act as carcinogens. Therefore, their remediation is a major challenge and is obligatory. A Gram-negative bacterial strain (NM05 of *Sphingomonas*) was earlier reported to degrade the pesticide hexachlorocyclohexane (HCH) [67] upon treatment with Pd/Fe0 bimetallic NPs (CMC-Pd/nFe0), showing the synergistic effect on the degradation of HCH that was enhanced by nearly 1.7–2.1-fold compared to the controls that had the *Sphingomonas* sp. strain NM05 or CMC-Pd/nFe0 alone [60]. The degradation process was found to be affected by experimental conditions (pH, temperature, HCH concentrations, etc.) [68].

The perovskite (LaFeO₃) NPs and biochar from water caltrop (*Trapa natans*) shells studied on marine sediment reported enhanced degradation of PAHs. The study used lignocellulosic fiber-reinforced biodegradable composites (LFBC) at 0.75 g L⁻¹ and pH 6.0 to activate the peroxymonosulfate (3×10^{-4} M) that helped in the oxidation of oxidizing PAHs in the sediments.

Up to 90% of total degradation was achieved; however, individually 2-ring PAHs 52%, 3-ring PAHs 61%, 4-ring PAHs 66%, 5-ring PAHs 56%, and 6-ring PAHs 29% were observed [69]. The process also reported improved microbial diversity of sediment and the major phylum *Proteobacteria* was observed initially, but after the process, *Hyphomonas* was predominantly observed [70]. In a continuous-flow experiments system for the degradation of naphthalene in the groundwater, 400 mg L⁻¹ of synthesized CaO₂ NPs degraded the naphthalene of optimum concentration 20 mg L⁻¹. This study highlights complete remediation of naphthalene in the presence of CaO₂ NPs and microbes (an abundance of *Coccobacilli*) from column effluent within 50 days [71].

In the case of soil, improving the microbial community by application of NPs is another way to reduce/remove the toxic pollutant loads from it. Si NPs have been reported to improve microbial colonization and biomass, including the rhizospheric microbes that are helpful for improving soil health [72,73]. However, prolonged exposure and accumulation of these NPs in soil may affect the nutrient and organic matter content.

3. Remediation of Contaminated Soils with Heavy Metals Using Hyperaccumulator Plants and Nanoparticles

Both from an ecological standpoint and one of restoring degraded areas, the remediation of HM-contaminated soils is a pressing issue that needs to be addressed immediately [74]. This section discusses the application of hyperaccumulation systems based on plants and NPs for the cleanup of various HMs from contaminated sites. The kinds of soil found at a contaminated site, as well as the percentage of metal contamination present, influence the rate at which hyperaccumulating plants can be utilized to remediate the site [75]. The bioavailability of metals in the rhizospheric region rests on soil pH, the gradient in elemental concentration, microbial population change, redox potential, the ratio of CO₂ and O₂, etc. [76]. The rhizospheric environment also directly rests on plant species in terms of root exudates and root architecture [75].

A few plant species, such as *Pedioplanis burchelli*, *Amaranthus spinosus*, and *Alternanthera pungens*, have survived at an optimal level of HMs by rhizofiltration and demonstrated avoidance mechanisms for HM uptake in their environment [70]. Beyond the optimum concentration, HMs pose an adverse impact on plant growth and human health [77]. However, hyperaccumulating plant species ingest metals in large quantities from contaminated soils, then transport and accumulate them in the organs above the soil in higher concentrations than in non-hyperaccumulating species without any obvious phytotoxic effects [78,79]. The activity of plant hyperaccumulators for HMs based on their phytoremediation potentials such as phytostabilization, phytoextraction, and rhizodegradation was demonstrated [80].

The hyperaccumulator plant that has a bio-concentration factor (BCF) of HMs of more than one owes it to the mechanism of phytostabilization and phytoextraction [76]. A BCF and TF (translocation factor) of more than one shows the characteristic traits of phytostabilization [81]. Similarly, Kisku et al. [81] demonstrated both phytostabilization and phytoextraction activities could be found in *Parthenium hysterophorus, Sacrum munja,* and *Ipomoea carnea,* and the authors observed more than one BCF and TF for Cr, Ni, Cd, and Pb, which revealed a phytostabilization mechanism, while more than one BCF and less than one TF for Zn and Mn showed a phytoextraction process for HMs [82]. Schematic representations of hyperaccumulator plant mechanistic supplemented with NPs for the elimination of toxic elements from contaminated soil are presented in Figure 2.



Figure 2. Schematic representation of hyperaccumulator plant mechanistic supplemented with nanoparticles for removal of heavy metals from contaminated soil.

Rhizodegradation is the process through which pollutants are deposited in the rhizospheric area of soil by microbial activity, where bacteria metabolize these contaminants for energy and nutrition. In this mechanism, microbes are capable of breaking down hazardous pollutants into nontoxic and harmless products [83]. Plant roots release natural carbon-containing substances, such as sugar, alcohol, and acid, and thereby provide the microorganisms with additional nutrients, which further stimulates rhizodegradation activities [84].

A wide range of treatment strategies, including physicochemical and biological methods, have been used to decontaminate sites polluted with HMs. The mechanisms of these methods are based on redox reactions, adsorption, ion-exchange, bioremediation, and phytoremediation [85]. All these methods have their own deserves and demerits, and out of these methods' bioremediation have come the most suitable eco-friendly techniques to achieve the sustainable goals [86,87]. Phytoremediation is a commonly explored technique and its potential application in contaminated land can be manipulated by added material such as NPs. Out of numerous mechanisms, the adsorbent mechanism plays a crucial role in the elimination of a broad range of HMs from contaminated soil in a short time.

Recently, adsorbent materials such as activated carbon, biochar, and NPs became commonly available; these adsorbent materials exhibit rapid adsorption capacity, cover large surface areas, provide more interplay sites for HMs, and have low price value [88]. Hence, using hyperaccumulator plants supplemented with good adsorbent material can be a promising approach for the elimination of HMs from contaminated soil. According to the existing state of knowledge, NPs have considerable potential for the remediation of soils contaminated with HMs. Nanophytoremediation is a process for the remediation of pollutants, i.e., pollutants that use synthetic NPs from plants [89].

For the remediation of soil contaminated with HMs, carbon or non-carbon NPs have primarily been employed; CNTs (carbon nanotubes), nZVI (nano zero-valent iron), TiO₂ NPs, and Ag NPs have all been extensively investigated for their potential for soil remediation [90]. The HM-removal efficiency of CNTs also relies on the pH and temperature of the adjacent surrounding environment, contact time, and the dose of CNTs. The CNTs (0.05 g) remove 99.9% of Zn²⁺ at 10.0 pH [84] and a high percent of Cd²⁺ are removed at 3.0 pH [77]; FeS NPs removed 99.65% of Cr⁶⁺ at 6.0 pH, while at high pH (>10.0) the FeS NPs elimination rate was decreased [91]. The equilibrium of the adsorption ability of NPs rests on temperature fluctuations. The adsorption of Pb2+ by Fe3O4 NPs rises from 298 to 328K [92], while chitosan-alginate NPs remove Hg²⁺ at 30 °C [93]. Hg²⁺ removal efficacy by FeS NPs (stabilized by sodium carboxymethyl cellulose) achieves its highest elimination percent at 30 min of contact time [94]. The nZVI-NPs are effective for the elimination of HMs from contaminated sites, nZVI contains shells of zerovalent iron, Fe²⁺ and Fe³⁺, which develop the excellent potential of nZVI for the elimination of HMs. Removal of Zn²⁺ (109.7 mg g^{-1}), Cu²⁺ (161.9 mg g^{-1}), and Pb²⁺ (195 mg g^{-1}) was achieved by utilization of nZVI after 6 h [95].

Several HMs (Cr, Zn, Pb, As, U, V, etc.) are effectively removed by the application of nZVI at the highest rate from contaminated sites [96]. The nZVI @ BC (8 g kg⁻¹) was utilized in the pot experiment, and it was observed that the immobilization efficiency of Cr⁶⁺ from the soil was increased by 100% after 15 days [77]. The elimination of Cd, Pb, and Zn from contaminated soils was increased by the application of the nanomaterial OA-nZVI (0.4 g kg⁻¹) by 46.66%, 48.88%, and 47.01%, respectively, reported [97]. The low concentration of nZVI substantially improved seedlings, root length, and leaf area in white willow (*Salix alba* L.) with an increased bio-concentration factor for Cd, while a high concentration of nZVI showed an adverse effect on plant growth and BCF for Cu and Pb [98]. The elimination of Zn, Pb, and Cd was investigated by Mitzia et al., [99] using nZVI with biochar and they observed that both combinations significantly affect the immobilization of Zn, Pb, and Cd. The nZVI (sodium carboxymethyl cellulose stabilized) showed a significantly increased immobilization of Cr in edible rapeseed (*Brassica napus*) and Chinese cabbage

(*B. rapa* subsp. pekinensis) [100]. The use of NPs decreased the bioavailability and bioaccumulation of Cr in both plants.

It is observed that TiO₂ NPs have high reactivity and photo-catalytic activity, which enables contaminates to get adsorbed on its surface area; this trait of TiO₂ NPs serves to minimize the toxic behavior of HMs, as well as maintain their mobility. It absorbed 88.01% of Cu²⁺ and 70.67% of Cd²⁺ effectively at 7.0 pH [47]. Other NPs such as modified carbon substantially improved the plant growth of *Suaeda salsa* and decreased the uptake of Cd and Ni compared to the control [101]. Ag NPs can effectively reduce Ni, Pb, Na, Zn, and Cu uptake in maize (*Zea mays*) plants and promote plant growth by optimizing the gibberellin, abscisic content in *Z. mays* leaves and PGPR interaction [102]. In combination with *B. cereus* (LPR2), Ag NPs significantly induce the growth of *Z. mays* [103]. Hence, perfect strategies are required for the elimination of HMs from contaminated soil, such as the selection of hyperaccumulator plants with compatible NPs, which could be a promising technique for the restoration of contaminated soil.

4. Nanotechnological Approaches for Restoring Metalloid-Contaminated Soils

Metalloids are elements with a plurality of attributes, i.e., they have the physicochemical features of both metals and nonmetals. They are naturally found, but the introduction of industrialization has resulted in concentrations that exceed the allowable limit [104]. Antimony (Sb), arsenic (As), boron (B), germanium (Ge), tellurium (Te), and silicon (Si) are the six elements of the periodic table that are generally known as metalloids [105]. Metalloid pollution of soil is a global issue owing to the deposition of these chemicals in the environment, affecting human health, plants, and animals. Also, they are hazardous at any concentration, even at extremely low levels [9,106]. Besides, HMs/metalloids can have a major effect on the abundance, structure, and diversity of soil microbial communities, which can lead to a shift in ecosystem functioning [107,108].

In previous years, several remediation methods have been introduced for the reclamation of these metalloids from soil. However, successful remediation of sites contaminated by metalloids is hindered by the fact that these pollutants do not decompose on their own and that it is not always possible to extract all of the contaminated soil [9]. Therefore, chemical stabilization of metalloids in soils, through adsorption, surface precipitation, structural integration, or ion exchange, is a promising solution for such sites since it immobilizes contaminants in the soils, limiting their mobility, bioavailability, and bio-accessibility [109]. Furthermore, it is evident that the initial phase in phytoremediation, the lowering of their toxicity, is critical for the creation of plant cover on contaminated soils [80].

Among NPs, iron oxides NPs are considered to remediate contaminants like HMs from soils owing to their capacity to absorb these pollutants [109]. In this context, in a study, Zhang et al. [110] revealed the effectiveness of iron-based NPs, viz., nZVI, FeS, and Fe₃O₄ particles for immobilizing As in contaminated soils. Among these, Fe₃O₄ NPs were reported to be more effective than other NPs for immobilizing As. Besides iron oxides, other NPs are also described as remediating soils polluted with different metals and metalloids [111]. Apart from magnetite, other famous iron NPs are nZVI, which are utilized for remediation of metal/metalloid-polluted water. In a study, the photolytic system was used for the decrease in reducing the toxic form of chromium Cr⁶⁺ to non-toxic Cr³⁺ in aqueous [112]. In the same study, the Cr⁶⁺ reduction efficiency was recorded up to 90%. Furthermore, in some cases, NPs were also employed to immobilize the trace elements in contaminated soil. For example, nanostructured TiO₂, MgO, and ZnO were found to be efficient adsorbents for the elimination of Cr ions from leather factory waste treated soil [113]. Hence, these aspects can be further explored and investigated to envisage the potential of such NPs for the elimination of metalloids from polluted soils. The nanoscale amorphous MnO was also evaluated for its capacity to remediate the soils polluted with HMs such as Cd, Cu, Zn, As, and Pb [114,115]. Although different kinds of NPs are less generally used for metalloid remediation, but they can still be applied in this context because some NPs are utilized to remove HMs from other contaminated media, such as water.

5. Nanobioremediation: Environment Concerns and Fate of Nanoparticles

Despite the numerous roles of and advancements in nanotechnology, there is still concern about its presence in environmental spheres, its fate, and the consequent toxicological impacts. Therefore, the main focus of this section is to distinctly provide an overview on the fate and the emerging environmental challenges of the deliberate emission of NPs, owing to their enhanced applications, especially in remediation processes. Recently, some reports documented that the increasing applications of NPs in agriculture have imposed serious implications. For example, the deliberate application of NPs may result in their accumulation or in an increase in the concentration of their constituents in the soil, thus affecting the soil's properties [8,20]. The presence of NPs in soils is reported to alter the soil pH, which is one of the most important parameters that influences soil nutrient availability, microbial dynamics, overall soil health, and plant growth and development [116].

A study conducted by Cullen reported that modification of nanoscale zero-valent iron (nZVI) in the soil can cause a significant rise in the pH of a soil solution [117]. A similar report was claimed by a study of CuO NPs on soil pH where CuO NPs utilize H⁺ from the soil to yield Cu ions and Cu(OH)+ ionic complex. Furthermore, this process is stated to be more enhanced in acidic soil [118]. Also, NPs of Ag, Au, Ti, and Zn have been reported to affect soil pH and their presence has been associated with adverse effects on beneficial soil microorganisms and nematodes [119]. The extent of the induced detrimental effects of the presence of NPs in soil is influenced by their concentration and type, soil type, and the enzymatic activity of the soil [120]. In addition, an elevated concentration of NPs is associated with decreased dehydrogenase activity, which disrupts the equilibrium of the soil nutrient and fertility levels [120,121]. Furthermore, the absorption and internalization of such NPs by microbes significantly affects mycelium and damages their normal cell functioning [122].

In soil, there are three main procedures, viz., surface complexation, hydrophobic partitioning, and ion exchange for the sorption of NPs, which are also involved in the colloidfacilitated transport of pollutants [123]. Mobilization processes mediated by colloids are also responsible for the likelihood of assisted transfer. It is possible for nanoscale particles to have a significant impact on trace metal transport, either by slowing it down when they are trapped in the matrix or by increasing the speed while they are moving [124]. The presence of NPs has been reported in water [125,126], in soil [127,128], and also in the air [129,130]. Continuous NP emissions into the environment would very probably result in their ubiquitous presence, with NPs likely to infiltrate the food chain at various trophic levels and exert toxicological effects on a variety of aquatic and terrestrial animals, as well as on human health [131,132]. The ecotoxicity of NPs is strongly related to their negative effect on the environment, where they might interact with biological systems because of their unique physicochemical properties [133,134].

Once NPs enter into water bodies, they act as pollutants and are ingested by lower aquatic biota. Metal NPs were tested in three different animal models representing different trophic levels, including the *Danio rero* (zebrafish), *Daphnia pulex* (daphnia), and *Pseudokirchneriella subcapitata* (microalgae), and they caused acute toxicity and filter-feeding, according to the findings of the study. It was noted that the NPs were more hazardous to daphnia and microalgae than to zebrafish, which is understandable because daphnids are particle filter feeders and, hence, are more susceptible to being exposed to NPs [135]. Thus, after entering the biological system, NPs have an impact on biochemical processes at the molecular and tissue-organ levels [136,137]. DNA damage, ROS-induced oxidative stress, protein folding disruption, and cell death are the most common toxicological effects [138–140]. Some studies also documented that SiO₂ NPs and TiO₂ NPs induced immunomodulatory and immunosuppressive effects in biological systems [141,142]. Thus, before the

implementation of NPs for nanobioremediation, these environmental concerns should be considered as of the utmost importance and NPs should be designed in ways that promote sustainable applications.

6. Conclusion and Future of Nanoremediation

In this modern period, there is a rapid release of pollutants into the environment, posing a substantial hazard to both human and the environmental health. As a result, having effective techniques for removing them from various environmental media will be critical in preventing their negative impacts. Because many conventional procedures are incapable of efficiently removing different groups of contaminants, novel approaches are therefore required to eliminate pollutants to the maximum extent possible. Meanwhile, a very likely method for developing remedies for polluted media is nanotechnology, according to the observations from a profusion of studies in this respect. A favorable result is the filling of the data gap on real-world remediation, which is aided by closer collaboration between the research and the industry. Thus, it will be useful for driving future soil cleanup efforts, which are sometimes necessary and cannot wait for experimental results to be obtained, utilizing methods to be attained in the meanwhile. Besides, the majority of the currently available research on nanobioremediation is limited to laboratory experiments and computational modelling. Therefore, to alleviate soils contaminated with a varied range of contaminants in the field, it is necessary to utilize multidisciplinary approaches. However, the environmental fate of NPs should be carefully taken into consideration before widening the application of nanobioremediation approaches.

Author Contributions: Conceptualization, V.D.R. and T.M.; methodology, software, V.D.R., S.K.U., A.K. and T.M.; validation, V.D.R., K.K.V. and T.M.; formal analysis, S.S., S.M., A.K. and A.R.; investigation, resources.; data curation, writing—original draft preparation, V.D.R., S.K.U., A.K. and A.R.; visualization, T.M. and S.M.; supervision T.M.; project administration, T.M., S.S. and S.M.; funding acquisition, T.M., V.D.R., R.K.S. and S.M. All authors have read and agreed to the published version of the manuscript.

Funding: Not applicable.

Institutional Review Board Statement: Not Applicable.

Informed Consent Statement: Not Applicable.

Data Availability Statement: Not Applicable.

Acknowledgments: The research was financially supported by the Russian Science Foundation, project no. 21-77-20089.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Midhat, L.; Ouazzani, N.; Hejjaj, A.; Ouhammou, A.; Mandi, L. Accumulation of heavy metals in metallophytes from three mining sites (Southern Centre Morocco) and evaluation of their phytoremediation potential. *Ecotoxicol. Environ. Saf.* 2019, 169, 150–160, https://doi.org/10.1016/j.ecoenv.2018.11.009.
- 2. Parikh, S.J.; James, B.R. Soil: The Foundation of Agriculture. Nat. Educ. Knowl. 2012, 3, 2.
- Struik, P.C.; Kuyper, T.W. Sustainable intensification in agriculture: The richer shade of green. A review. *Agron. Sustain. Dev.* 2017, 37, 39, https://doi.org/10.1007/s13593-017-0445-7.
- Pretty, J.; Bharucha, Z.P. Sustainable intensification in agricultural systems. Ann. Bot. 2014, 114, 1571–1596, https://doi.org/10.1093/aob/mcu205.
- Martin, J.-L.; Maris, V.; Simberloff, D.S. The need to respect nature and its limits challenges society and conservation science. Proc. Natl. Acad. Sci. USA 2016, 113, 6105, https://doi.org/10.1073/pnas.1525003113.
- Rajput, V.; Minkina, T.; Kumari, A.; Shende, S.; Ranjan, A.; Faizan, M.; Barakvov, A.; Gromovik, A.; Gorbunova, N.; Rajput, P.; et al. A review on nanobioremediation approaches for restoration of contaminated soil. *Eurasian J. Soil Sci.* 2021, *11*, 43–60, https://doi.org/10.18393/ejss.990605.
- Medina-Pérez, G.; Fernández-Luqueño, F.; Vazquez-Nuñez, E.; López-Valdez, F.; Prieto-Mendez, J.; Madariaga-Navarrete, A.; Miranda-Arámbula, M. Remediating polluted soils using nanotechnologies: Environmental benefits and risks. *Pol. J. Environ. Stud.* 2019, 28, 1013–1030, https://doi.org/10.15244/pjoes/87099.

- Kumari, A.; Kumari, P.; Rajput, V.D.; Sushkova, S.N.; Minkina, T. Metal(loid) nanosorbents in restoration of polluted soils: Geochemical, ecotoxicological, and remediation perspectives. *Environ. Geochem. Health* 2021, 44, 235–246, https://doi.org/10.1007/s10653-021-00996-x.
- 9. Raffa, C.M.; Chiampo, F.; Shanthakumar, S. Remediation of metal/metalloid-polluted soils: A short review. *Appl. Sci.* 2021, *11*, 4134, https://doi.org/10.3390/app11094134.
- Alazaiza, M.Y.D.; Albahnasawi, A.; Ali, G.A.M.; Bashir, M.J.K.; Copty, N.K.; Amr, S.S.A.; Abushammala, M.F.M.; Al Maskari, T. Recent advances of nanoremediation technologies for soil and groundwater remediation: A review. *Water* 2021, 13, 2186, https://doi.org/10.3390/w13162186.
- Ghani, M.I.; Saleem, S.; Rather, S.A.; Rehmani, M.S.; Alamri, S.; Rajput, V.D.; Kalaji, H.M.; Saleem, N.; Sial, T.A.; Liu, M. Foliar application of zinc oxide nanoparticles: An effective strategy to mitigate drought stress in cucumber seedling by modulating antioxidant defense system and osmolytes accumulation. *Chemosphere* 2022, 289, 133202, https://doi.org/10.1016/j.chemosphere.2021.133202.
- 12. Faizan, M.; Sehar, S.; Rajput, V.D.; Faraz, A.; Afzal, S.; Minkina, T.; Sushkova, S.; Adil, M.F.; Yu, F.; Alatar, A.A.; et al. Modulation of cellular redox status and antioxidant defense system after synergistic application of zinc oxide nanoparticles and salicylic acid in rice (*Oryza sativa*) plant under arsenic stress. *Plants* **2021**, *10*, 2254, https://doi.org/10.3390/plants10112254.
- Faizan, M.; Rajput, V.D.; Al-Khuraif, A.A.; Arshad, M.; Minkina, T.; Sushkova, S.; Yu, F. Effect of foliar fertigation of chitosan nanoparticles on cadmium accumulation and toxicity in *Solanum lycopersicum*. *Biology* 2021, 10, 666, https://doi.org/10.3390/biology10070666.
- 14. Rajput, V.; Minkina, T.; Semenkov, I.; Klink, G.; Tarigholizadeh, S.; Sushkova, S. Phylogenetic analysis of hyperaccumulator plant species for heavy metals and polycyclic aromatic hydrocarbons. *Environ. Geochem. Health* **2020**, *16*, 68–75, https://doi.org/10.1007/s10653-020-00527-0.
- Ghazaryan, K.A.; Movsesyan, H.S.; Khachatryan, H.E.; Ghazaryan, N.P.; Minkina, T.M.; Sushkova, S.N.; Mandzhieva, S.S.; Rajput, V.D. Copper phytoextraction and phytostabilization potential of wild plant species growing in the mine polluted areas of Armenia. *Environ. Geochem. Health* 2018, 19, 155–163, https://doi.org/10.1144/geochem2018-035.
- Haldar, D.; Duarah, P.; Purkait, M.K. MOFs for the treatment of arsenic, fluoride and iron contaminated drinking water: A review. *Chemosphere* 2020, 251, 126388, https://doi.org/10.1016/j.chemosphere.2020.126388.
- 17. Younis, S.A.; Serp, P.; Nassar, H.N. Photocatalytic and biocidal activities of ZnTiO₂ oxynitride heterojunction with MOF-5 and g-C₃N₄: A case study for textile wastewater treatment under direct sunlight. *J. Hazard. Mater.* **2021**, *410*, 124562, https://doi.org/10.1016/j.jhazmat.2020.124562.
- Yang, G.; Zhang, D.; Zhu, G.; Zhou, T.; Song, M.; Qu, L.; Xiong, K.; Li, H. A Sm-MOF/GO nanocomposite membrane for efficient organic dye removal from wastewater. RSC Adv. 2020, 10, 8540–8547, https://doi.org/10.1039/D0RA01110J.
- Moameri, M.; Khalaki, M.A. Chapter 5–Toxicity/risk assessment of nanomaterials when used in soil treatment. In *Nanomaterials for Soil Remediation*; Amrane, A., Mohan, D., Nguyen, T.A., Assadi, A.A., Yasin, G., Eds.; Elsevier: Amsterdam, The Netherlands, 2021; pp. 87–100.
- Rajput, V.; Minkina, T.; Mazarji, M.; Shende, S.; Sushkova, S.; Mandzhieva, S.; Burachevskaya, M.; Chaplygin, V.; Singh, A.; Jatav, H. Accumulation of nanoparticles in the soil-plant systems and their effects on human health. *Ann. Agric. Sci.* 2020, 65, 137–143, https://doi.org/10.1016/j.aoas.2020.08.001.
- McGrath, J.M.; Spargo, J.; Penn, C.J. Soil Fertility and Plant Nutrition. In *Encyclopedia of Agriculture and Food Systems*; Van Alfen, N.K., Ed.; Academic Press: Oxford, UK, 2014; pp. 166–184.
- Gorovtsov, A.; Demin, K.; Sushkova, S.; Minkina, T.; Grigoryeva, T.; Dudnikova, T.; Barbashev, A.; Semenkov, I.; Romanova, V.; Laikov, A.; et al. The effect of combined pollution by PAHs and heavy metals on the topsoil microbial communities of Spolic Technosols of the lake Atamanskoe, Southern Russia. *Environ. Geochem. Health* 2021, 1–17, https://doi.org/10.1007/s10653-021-01059-x.
- 23. Rajput, V.D.; Yadav, A.N.; Jatav, H.S.; Singh, S.K.; Minkina, T. Sustainable Management and Utilization of Sewage Sludge; Springer: Cham, Switzerland, 2022.
- 24. Ashraf, S.; Siddiqa, A.; Shahida, S.; Qaisar, S. Titanium-based nanocomposite materials for arsenic removal from water: A review. *Heliyon* **2019**, *5*, e01577, https://doi.org/10.1016/j.heliyon.2019.e01577.
- Chandra, R.; Kumar, V.; Tripathi, S.; Sharma, P. Phytoremediation of industrial pollutants and life cycle assessment. In *Phytore-mediation of Environmental Pollutants*; CRC Press: Boca Raton, FL, USA, 2017; pp. 441–470.
- 26. Gerhardt, K.E.; Gerwing, P.D.; Greenberg, B.M. Opinion: Taking phytoremediation from proven technology to accepted practice. *Plant Sci.* **2017**, 256, 170–185.
- 27. Gong, X.; Huang, D.; Liu, Y.; Peng, Z.; Zeng, G.; Xu, P.; Cheng, M.; Wang, R.; Wan, J. Remediation of contaminated soils by biotechnology with nanomaterials: Bio-behavior, applications, and perspectives. *Crit. Rev. Biotechnol.* **2018**, *38*, 455–468.
- 28. Mehndiratta, P.; Jain, A.; Srivastava, S.; Gupta, N. Environmental pollution and nanotechnology. Environ. Pollut. 2013, 2, 49.
- 29. Gong, J.-L.; Wang, B.; Zeng, G.-M.; Yang, C.-P.; Niu, C.-G.; Niu, Q.-Y.; Zhou, W.-J.; Liang, Y. Removal of cationic dyes from aqueous solution using magnetic multi-wall carbon nanotube nanocomposite as adsorbent. *J. Hazard. Mater.* **2009**, *164*, 1517–1522.
- Chen, C.; Tsyusko, O.V.; McNear, D.H., Jr.; Judy, J.; Lewis, R.W.; Unrine, J.M. Effects of biosolids from a wastewater treatment plant receiving manufactured nanomaterials on *Medicago truncatula* and associated soil microbial communities at low nanomaterial concentrations. *Sci. Total Environ.* 2017, 609, 799–806, https://doi.org/10.1016/j.scitotenv.2017.07.188.

- Chauhan, R.; Yadav, H.O.S.; Sehrawat, N. Nanobioremediation: A new and a versatile tool for sustainable environmental clean up-Overview. J. Mater. Environ. Sci. 2020, 11, 564–573.
- 32. Briffa, J.; Sinagra, E.; Blundell, R. Heavy metal pollution in the environment and their toxicological effects on humans. *Heliyon* **2020**, *6*, e04691, https://doi.org/10.1016/j.heliyon.2020.e04691.
- Misra, M.; Ghosh Sachan, S. Nanobioremediation of heavy metals: Perspectives and challenges. J. Basic Microbiol. 2021, 1–16, https://doi.org/10.1002/jobm.202100384.
- Abdi, O.; Kazemi, M. A review study of biosorption of heavy metals and comparison between different biosorbents. J. Mater. Environ. Sci. 2015, 6, 1386–1399.
- Ayangbenro, A.S.; Babalola, O.O. A new strategy for heavy metal polluted environments: A review of microbial biosorbents. *Int. J. Environ. Res. Public Health* 2017, 14, 94.
- Usman, M.; Farooq, M.; Wakeel, A.; Nawaz, A.; Cheema, S.A.; Rehman, H.; Ashraf, I.; Sanaullah, M. Nanotechnology in agriculture: Current status, challenges and future opportunities. *Sci. Total Environ.* 2020, 721, 137778.
- 37. Abebe, B.; Murthy, H.C.A.; Amare, E. Summary on adsorption and photocatalysis for pollutant remediation: Mini review. *J. Encapsulation Adsorpt. Sci.* **2018**, *8*, 225–255.
- 38. Desiante, W.L.; Minas, N.S.; Fenner, K. Micropollutant biotransformation and bioaccumulation in natural stream biofilms. *Water Res.* 2021, 193, 116846.
- Filote, C.; Roşca, M.; Hlihor, R.M.; Cozma, P.; Simion, I.M.; Apostol, M.; Gavrilescu, M. Sustainable application of biosorption and bioaccumulation of persistent pollutants in wastewater treatment: Current practice. *Processes* 2021, *9*, 1696.
- Alissa, E.M.; Ferns, G.A. Heavy metal poisoning and cardiovascular disease. J. Toxicol. 2011, 2011, 870125, https://doi.org/10.1155/2011/870125.
- 41. Jaishankar, M.; Tseten, T.; Anbalagan, N.; Mathew, B.B.; Beeregowda, K.N. Toxicity, mechanism and health effects of some heavy metals. *Interdiscip. Toxicol.* 2014, 7, 60.
- Rasmussen, L.D.; Sørensen, S.J.; Turner, R.R.; Barkay, T. Application of a mer-lux biosensor for estimating bioavailable mercury in soil. Soil Biol. Biochem. 2000, 32, 639–646.
- 43. Mukhopadhyay, S.; Maiti, S.K. Phytoremediation of metal mine waste. Appl. Ecol. Environ. Res. 2010, 8, 207–222.
- 44. Cao, X.; Alabresm, A.; Chen, Y.P.; Decho, A.W.; Lead, J. Improved metal remediation using a combined bacterial and nanoscience approach. *Sci. Total Environ.* **2020**, *704*, 135378.
- Gil-Díaz, M.; Diez-Pascual, S.; González, A.; Alonso, J.; Rodríguez-Valdés, E.; Gallego, J.R.; Lobo, M.C. A nanoremediation strategy for the recovery of an As-polluted soil. *Chemosphere* 2016, 149, 137–145, https://doi.org/10.1016/j.chemosphere.2016.01.106.
- 46. Baragaño, D.; Forján, R.; Welte, L.; Gallego, J.L.R. Nanoremediation of As and metals polluted soils by means of graphene oxide nanoparticles. *Sci. Rep.* 2020, *10*, 1896, https://doi.org/10.1038/s41598-020-58852-4.
- 47. Mahmoud, M.E.; Abou Ali, S.A.A.; Elweshahy, S.M.T. Microwave functionalization of titanium oxide nanoparticles with chitosan nanolayer for instantaneous microwave sorption of Cu(II) and Cd(II) from water. *Int. J. Biol. Macromol.* 2018, 111, 393–399, https://doi.org/10.1016/j.ijbiomac.2018.01.014.
- 48. Ha, C.; Zhu, N.; Shang, R.; Shi, C.; Cui, J.; Sohoo, I.; Wu, P.; Cao, Y. Biorecovery of palladium as nanoparticles by Enterococcus faecalis and its catalysis for chromate reduction. *Chem. Eng. J.* **2016**, *288*, 246–254.
- Mahmoud, M.E.; Abdou, A.E.H.; Mohamed, S.M.S.; Osman, M.M. Engineered staphylococcus aureus via immobilization on magnetic Fe₃O₄-phthalate nanoparticles for biosorption of divalent ions from aqueous solutions. *J. Environ. Chem. Eng.* 2016, 4, 3810–3824, https://doi.org/10.1016/j.jece.2016.08.022.
- Akhtar, N.; Khan, S.; Rehman, S.U.; Rehman, Z.U.; Khatoon, A.; Rha, E.S.; Jamil, M. Synergistic effects of zinc oxide nanoparticles and bacteria reduce heavy metals toxicity in rice (*Oryza sativa* L.) Plant. *Toxics* 2021, 9, 113.
- Chaplygin, V.A.; Minkina, T.M.; Mandzhieva, S.S.; Nazarenko, O.G.; Zimulina, I.V.; Bauer, T.V.; Litvinov, Y.A.; Rajput, V. Heavy metals in agricultural crops of Rostov region through the example of soft wheat (*Triticum aestivum*). *IOP Conf. Ser. Earth Environ. Sci.* 2021, 624, 012204, https://doi.org/10.1088/1755-1315/624/1/012204.
- 52. Huang, H.; Ullah, F.; Zhou, D.-X.; Yi, M.; Zhao, Y. Mechanisms of ROS Regulation of Plant Development and Stress Responses. *Front. Plant Sci.* 2019, 10, 193–198, https://doi.org/10.3389/fpls.2019.00800.
- Enez, A.; Hudek, L.; Bräu, L. Reduction in trace element mediated oxidative stress towards cropped plants via beneficial microbes in irrigated cropping systems: A review. *Appl. Sci.* 2018, *8*, 1953, https://doi.org/10.3390/app8101953.
- 54. Arif, N.; Yadav, V.; Singh, S.; Singh, S.; Ahmad, P.; Mishra, R.K.; Sharma, S.; Tripathi, D.K.; Dubey, N.K.; Chauhan, D.K. Influence of high and low levels of plant-beneficial heavy metal ions on plant growth and development. *Front. Environ. Sci.* **2016**, *4*, 69, https://doi.org/10.3389/fenvs.2016.00069.
- 55. Suman, J.; Uhlik, O.; Viktorova, J.; Macek, T. Phytoextraction of heavy metals: A promising tool for clean-up of polluted environment? *Front. Plant Sci.* 2018, 9, 1476.
- Rajput, V.D.; Minkina, T.; Kimber, R.L.; Singh, V.K.; Shende, S.; Behal, A.; Sushkova, S.; Mandzhieva, S.; Lloyd, J.R.; Semrau, J.D. Insights into the Biosynthesis of Nanoparticles by the Genus Shewanella. *Appl. Environ. Microbiol.* 2021, *87*, e01390-21, https://doi.org/10.1128/AEM.01390-21.
- Capeness, M.J.; Echavarri-Bravo, V.; Horsfall, L.E. Production of biogenic nanoparticles for the reduction of 4-Nitrophenol and oxidative laccase-like reactions. *Front. Microbiol.* 2019, 10, 997, https://doi.org/10.3389/FMICB.2019.00997/BIBTEX.

- Sharma, D.; Kanchi, S.; Bisetty, K. Biogenic synthesis of nanoparticles: A review. Arab. J. Chem. 2019, 12, 3576–3600, https://doi.org/10.1016/J.ARABJC.2015.11.002.
- Alabresm, A.; Chen, Y.P.; Decho, A.W.; Lead, J. A novel method for the synergistic remediation of oil-water mixtures using nanoparticles and oil-degrading bacteria. *Sci. Total Environ.* 2018, 630, 1292–1297, https://doi.org/10.1016/j.scitotenv.2018.02.277.
- Xie, Y.; He, Y.; Irwin, P.L.; Jin, T.; Shi, X. Antibacterial activity and mechanism of action of zinc oxide nanoparticles against Campylobacter jejuni. *Appl. Environ. Microbiol.* 2011, 77, 2325–2331.
- 61. Laslo, V.; Pinzaru, S.C.; Zaguła, G.; Kluz, M.; Vicas, S.I.; Cavalu, S. Synergic effect of selenium nanoparticles and lactic acid bacteria in reduction cadmium toxicity. *J. Mol. Struct.* **2022**, *1247*, 131325, https://doi.org/10.1016/j.molstruc.2021.131325.
- 62. Dong, G.; Wang, Y.; Gong, L.; Wang, M.; Wang, H.; He, N.; Zheng, Y.; Li, Q. Formation of soluble Cr (III) end-products and nanoparticles during Cr (VI) reduction by *Bacillus cereus* strain XMCr-6. *Biochem. Eng. J.* **2013**, *70*, 166–172.
- 63. Ramos-Ruiz, A.; Sesma-Martin, J.; Sierra-Alvarez, R.; Field, J.A. Continuous reduction of tellurite to recoverable tellurium nanoparticles using an upflow anaerobic sludge bed (UASB) reactor. *Water Res.* **2017**, *108*, 189–196, https://doi.org/10.1016/j.watres.2016.10.074.
- 64. Ramos-Ruiz, A.; Field, J.A.; Wilkening, J.V.; Sierra-Alvarez, R. Recovery of elemental tellurium nanoparticles by the reduction of tellurium oxyanions in a methanogenic microbial consortium. *Environ. Sci. Technol.* **2016**, *50*, 1492–1500.
- 65. Borghese, R.; Baccolini, C.; Francia, F.; Sabatino, P.; Turner, R.J.; Zannoni, D. Reduction of chalcogen oxyanions and generation of nanoprecipitates by the photosynthetic bacterium *Rhodobacter capsulatus*. *J. Hazard. Mater.* **2014**, *269*, 24–30.
- Manickam, N.; Reddy, M.K.; Saini, H.S.; Shanker, R. Isolation of hexachlorocyclohexane-degrading Sphingomonas sp. by dehalogenase assay and characterization of genes involved in γ-HCH degradation. J. Appl. Microbiol. 2008, 104, 952–960.
- Singh, R.; Manickam, N.; Mudiam, M.K.R.; Murthy, R.C.; Misra, V. An integrated (nano-bio) technique for degradation of γ-HCH contaminated soil. J. Hazard. Mater. 2013, 258, 35–41.
- Hung, C.-M.; Huang, C.-P.; Chen, C.-W.; Dong, C.-D. Degradation of organic contaminants in marine sediments by peroxymonosulfate over LaFeO₃ nanoparticles supported on water caltrop shell-derived biochar and the associated microbial community responses. *J. Hazard. Mater.* 2021, 420, 126553.
- 69. Gholami, F.; Shavandi, M.; Dastgheib, S.M.M.; Amoozegar, M.A. Naphthalene remediation from groundwater by calcium peroxide (CaO₂) nanoparticles in permeable reactive barrier (PRB). *Chemosphere* **2018**, *212*, 105–113.
- Rizwan, M.; Ali, S.; ur Rehman, M.Z.; Adrees, M.; Arshad, M.; Qayyum, M.F.; Ali, L.; Hussain, A.; Chatha, S.A.S.; Imran, M. Alleviation of cadmium accumulation in maize (*Zea mays* L.) by foliar spray of zinc oxide nanoparticles and biochar to contaminated soil. *Environ. Pollut.* 2019, 248, 358–367.
- Rajput, V.D.; Minkina, T.; Feizi, M.; Kumari, A.; Khan, M.; Mandzhieva, S.; Sushkova, S.; El-Ramady, H.; Verma, K.K.; Singh, A. Effects of silicon and silicon-based nanoparticles on rhizosphere microbiome, plant stress and growth. *Biology* 2021, 10, 791.
- Srivastava, S.; Shukla, A.; Rajput, V.D.; Kumar, K.; Minkina, T.; Mandzhieva, S.; Shmaraeva, A.; Suprasanna, P. Arsenic Remediation through Sustainable Phytoremediation Approaches. *Minerals* 2021, *11*, 936, https://doi.org/10.3390/min11090936.
- 73. Gajić, G.; Djurdjević, L.; Kostić, O.; Jarić, S.; Mitrović, M.; Pavlović, P. Ecological Potential of Plants for Phytoremediation and Ecorestoration of Fly Ash Deposits and Mine Wastes. *Front. Environ. Sci.* **2018**, *6*, 124, https://doi.org/10.3389/fenvs.2018.00124.
- Palansooriya, K.N.; Shaheen, S.M.; Chen, S.S.; Tsang, D.C.W.; Hashimoto, Y.; Hou, D.; Bolan, N.S.; Rinklebe, J.; Ok, Y.S. Soil amendments for immobilization of potentially toxic elements in contaminated soils: A critical review. *Environ. Int.* 2020, 134, 105046, https://doi.org/10.1016/j.envint.2019.105046.
- Ahkami, A.H.; Allen White, R.; Handakumbura, P.P.; Jansson, C. Rhizosphere engineering: Enhancing sustainable plant ecosystem productivity. *Rhizosphere* 2017, *3*, 233–243, https://doi.org/10.1016/j.rhisph.2017.04.012.
- 76. Upadhyay, S.K.; Ahmad, M.; Srivastava, A.K.; Abhilash, P.C.; Sharma, B. Optimization of eco-friendly novel amendments for sustainable utilization of Fly ash based on growth performance, hormones, antioxidant, and heavy metal translocation in chickpea (*Cicer arietinum* L.) plant. *Chemosphere* 2021, 267, 129216, https://doi.org/10.1016/j.chemosphere.2020.129216.
- Sun, T.Y.; Bornhoft, N.A.; Hungerbuhler, K.; Nowack, B. Dynamic probabilistic modeling of environmental emissions of engineered nanomaterials. *Environ. Sci. Technol.* 2016, 50, 4701–4711, https://doi.org/10.1021/acs.est.5b05828.
- Jabeen, R.; Ahmad, A.; Iqbal, M. Phytoremediation of heavy metals: Physiological and molecular mechanisms. *Bot. Rev.* 2009, 75, 339–364, https://doi.org/10.1007/s12229-009-9036-x.
- 79. Rascio, N.; Navari-Izzo, F. Heavy metal hyperaccumulating plants: How and why do they do it? And what makes them so interesting? *Plant Sci.* **2011**, *180*, 169–181, https://doi.org/10.1016/j.plantsci.2010.08.016.
- Yan, A.; Wang, Y.; Tan, S.N.; Mohd Yusof, M.L.; Ghosh, S.; Chen, Z. Phytoremediation: A promising approach for revegetation of heavy metal-polluted land. *Front. Plant Sci.* 2020, *11*, 359, https://doi.org/10.3389/fpls.2020.00359.
- Usman, K.; Al-Ghouti, M.A.; Abu-Dieyeh, M.H. The assessment of cadmium, chromium, copper, and nickel tolerance and bioaccumulation by shrub plant *Tetraena qataranse*. Sci. Rep. 2019, 9, 5658, https://doi.org/10.1038/s41598-019-42029-9.
- Kisku, G.C.; Kumar, V.; Sahu, P.; Kumar, P.; Kumar, N. Characterization of coal fly ash and use of plants growing in ash pond for phytoremediation of metals from contaminated agricultural land. *Int. J. Phytoremediation* 2018, 20, 330–337, https://doi.org/10.1080/15226514.2017.1381942.
- Ojuederie, O.B.; Babalola, O.O. Microbial and plant-assisted bioremediation of heavy metal polluted environments: A review. Int. J. Environ. Res. Public Health 2017, 14, 1504, https://doi.org/10.3390/ijerph14121504.

- Rai, G.K.; Bhat, B.A.; Mushtaq, M.; Tariq, L.; Rai, P.K.; Basu, U.; Dar, A.A.; Islam, S.T.; Dar, T.U.H.; Bhat, J.A. Insights into decontamination of soils by phytoremediation: A detailed account on heavy metal toxicity and mitigation strategies. *Physiol. Plant* 2021, *173*, 287–304, https://doi.org/10.1111/ppl.13433.
- 85. Yu, G.; Wang, X.; Liu, J.; Jiang, P.; You, S.; Ding, N.; Guo, Q.; Lin, F. Applications of nanomaterials for heavy metal removal from water and soil: A review. *Sustainability* **2021**, *13*, 713, https://doi.org/10.3390/su13020713.
- Mazarji, M.; Bayero, M.T.; Minkina, T.; Sushkova, S.; Mandzhieva, S.; Tereshchenko, A.; Timofeeva, A.; Bauer, T.; Burachevskaya, M.; Kızılkaya, R.; et al. Realizing united nations sustainable development goals for greener remediation of heavy metals-contaminated soils by biochar: Emerging trends and future directions. *Sustainability* 2021, 13, 3825, https://doi.org/10.3390/su132413825.
- Kumar, V.; Shahi, S.K.; Singh, S. Bioremediation: An Eco-sustainable Approach for Restoration of Contaminated Sites. In *Microbial Bioprospecting for Sustainable Development*; Singh, J., Sharma, D., Kumar, G., Sharma, N.R., Eds.; Springer: Singapore, 2018; pp. 115–136.
- Ihsanullah; Abbas, A.; Al-Amer, A.M.; Laoui, T.; Al-Marri, M.J.; Nasser, M.S.; Khraisheh, M.; Atieh, M.A. Heavy metal removal from aqueous solution by advanced carbon nanotubes: Critical review of adsorption applications. *Sep. Purif. Technol.* 2016, 157, 141–161, https://doi.org/10.1016/j.seppur.2015.11.039.
- Guerra, F.D.; Attia, M.F.; Whitehead, D.C.; Alexis, F. Nanotechnology for environmental remediation: Materials and applications. *Molecules* 2018, 23, 1760, https://doi.org/10.3390/molecules23071760.
- Mubarak, N.M.; Sahu, J.N.; Abdullah, E.C.; Jayakumar, N.S.; Ganesan, P. Microwave-assisted synthesis of multi-walled carbon nanotubes for enhanced removal of Zn(II) from wastewater. *Res. Chem. Intermed.* 2016, 42, 3257–3281, https://doi.org/10.1007/s11164-015-2209-9.
- Wu, X.; Hu, J.; Wu, F.; Zhang, X.; Wang, B.; Yang, Y.; Shen, G.; Liu, J.; Tao, S.; Wang, X. Application of TiO₂ nanoparticles to reduce bioaccumulation of arsenic in rice seedlings (*Oryza sativa* L.): A mechanistic study. *J. Hazard. Mater.* 2021, 405, 124047, https://doi.org/10.1016/j.jhazmat.2020.124047.
- 92. Nassar, N.N. Rapid removal and recovery of Pb(II) from wastewater by magnetic nanoadsorbents. *J. Hazard. Mater.* **2010**, *184*, 538–546, https://doi.org/10.1016/j.jhazmat.2010.08.069.
- 93. Dubey, R.; Bajpai, J.; Bajpai, A.K. Chitosan-alginate nanoparticles (CANPs) as potential nanosorbent for removal of Hg (II) ions. *Environ. Nanotechnol. Monit. Manag.* 2016, *6*, 32–44, https://doi.org/10.1016/j.enmm.2016.06.008.
- Gong, Y.; Liu, Y.; Xiong, Z.; Zhao, D. Immobilization of mercury by carboxymethyl cellulose stabilized iron sulfide nanoparticles: Reaction mechanisms and effects of stabilizer and water chemistry. *Environ. Sci. Technol.* 2014, 48, 3986–3994, https://doi.org/10.1021/es404418a.
- Yang, F.; Zhang, S.; Sun, Y.; Cheng, K.; Li, J.; Tsang, D.C.W. Fabrication and characterization of hydrophilic corn stalk biocharsupported nanoscale zero-valent iron composites for efficient metal removal. *Bioresour. Technol.* 2018, 265, 490–497, https://doi.org/10.1016/j.biortech.2018.06.029.
- Klimkova, S.; Cernik, M.; Lacinova, L.; Filip, J.; Jancik, D.; Zboril, R. Zero-valent iron nanoparticles in treatment of acid mine water from in situ uranium leaching. *Chemosphere* 2011, *82*, 1178–1184, https://doi.org/10.1016/j.chemosphere.2010.11.075.
- Cao, Y.; Zhang, S.; Zhong, Q.; Wang, G.; Xu, X.; Li, T.; Wang, L.; Jia, Y.; Li, Y. Feasibility of nanoscale zero-valent iron to enhance the removal efficiencies of heavy metals from polluted soils by organic acids. *Ecotoxicol. Environ. Saf.* 2018, 162, 464–473, https://doi.org/10.1016/j.ecoenv.2018.07.036.
- Mokarram-Kashtiban, S.; Hosseini, S.M.; Tabari Kouchaksaraei, M.; Younesi, H. The impact of nanoparticles zero-valent iron (nZVI) and rhizosphere microorganisms on the phytoremediation ability of white willow and its response. *Environ. Sci. Pollut. Res. Int.* 2019, 26, 10776–10789, https://doi.org/10.1007/s11356-019-04411-y.
- Mitzia, A.; Vítková, M.; Komárek, M. Assessment of biochar and/or nano zero-valent iron for the stabilisation of Zn, Pb and Cd: A temporal study of solid phase geochemistry under changing soil conditions. *Chemosphere* 2020, 242, 125248, https://doi.org/10.1016/j.chemosphere.2019.125248.
- Wang, Y.; Fang, Z.; Kang, Y.; Tsang, E.P. Immobilization and phytotoxicity of chromium in contaminated soil remediated by CMC-stabilized nZVI. J. Hazard. Mater. 2014, 275, 230–237, https://doi.org/10.1016/j.jhazmat.2014.04.056.
- 101. Cheng, J.; Sun, Z.; Yu, Y.; Li, X.; Li, T. Effects of modified carbon black nanoparticles on plant-microbe remediation of petroleum and heavy metal co-contaminated soils. *Int. J. Phytoremediation* **2019**, *21*, 634–642, https://doi.org/10.1080/15226514.2018.1556581.
- 102. Khan, N.; Bano, A. Modulation of phytoremediation and plant growth by the treatment with PGPR, Ag nanoparticle and untreated municipal wastewater. *Int. J. Phytoremediation* **2016**, *18*, 1258–1269, https://doi.org/10.1080/15226514.2016.1203287.
- 103. Kumar, P.; Pahal, V.; Gupta, A.; Vadhan, R.; Chandra, H.; Dubey, R.C. Effect of silver nanoparticles and Bacillus cereus LPR2 on the growth of *Zea mays. Sci. Rep.* **2020**, *10*, 20409, https://doi.org/10.1038/s41598-020-77460-w.
- 104. Fordyce, F.M.; Everett, P.A.; Bearcock, J.M.; Lister, T.R. Soil metal/metalloid concentrations in the Clyde Basin, Scotland, UK: quality. 108, Implications for Earth Environ. Sci. Trans. R. Soc. Edinb. 2018, land 191-216. https://doi.org/10.1017/S1755691018000282.
- 105. Yazdi, M.; Sepehrizadeh, Z.; Mahdavi, M.; Shahverdi, A.R.; Faramarzi, M. Metal, metalloid, and oxide nanoparticles for therapeutic and diagnostic oncology. *Nano Biomed. Eng.* 2016, *8*, 246–267, https://doi.org/10.5101/nbe.v8i4.p246-267.
- Martínez-Alcalá, I.; Bernal, M.P. Environmental Impact of Metals, Metalloids, and Their Toxicity. In *Metalloids in Plants*; John and Wiley and Sons: Hoboken, NJ, USA, 2020; pp. 451–488.

- 107. Huang, C.-C.; Liang, C.-M.; Yang, T.-I.; Chen, J.-L.; Wang, W.-K. Shift of bacterial communities in heavy metal-contaminated agricultural land during a remediation process. *PLoS ONE* **2021**, *16*, e0255137, https://doi.org/10.1371/journal.pone.0255137.
- Crowley, D. Impacts of metals and metalloids on soil microbial diversity and ecosystem function. *Rev. Cienc. Suelo Nutr. Vegl.* 2008, *8*, 6–11.
- Kumpiene, J.; Lagerkvist, A.; Maurice, C. Stabilization of As, Cr, Cu, Pb and Zn in soil using amendments–A review. Waste Manage. 2008, 28, 215–225, https://doi.org/10.1016/j.wasman.2006.12.012.
- Zhang, M.; Wang, Y.; Zhao, D.; Pan, G. Immobilization of arsenic in soils by stabilized nanoscale zero-valent iron, iron sulfide (FeS), and magnetite (Fe₃O₄) particles. *Sci. Bull.* 2010, *55*, 365–372, https://doi.org/10.1007/s11434-009-0703-4.
- Martínez-Fernández, D.; Vítková, M.; Michálková, Z.; Komárek, M. Engineered Nanomaterials for Phytoremediation of Metal/Metalloid-Contaminated Soils: Implications for Plant Physiology. In *Phytoremediation: Management of Environmental Contaminants*; Ansari, A.A., Gill, S.S., Gill, R., Lanza, G.R., Newman, L., Eds.; Springer: Cham, Switzerland, 2017; Volume 5, pp. 369-403.
- 112. Kim, Y.; Joo, H.; Her, N.; Yoon, Y.; Lee, C.-H.; Yoon, J. Self-rotating photocatalytic system for aqueous Cr(VI) reduction on TiO₂ nanotube/Ti mesh substrate. *Chem. Eng. J.* **2013**, 229, 66–71, https://doi.org/10.1016/j.cej.2013.05.116.
- 113. Taghipour, M.; Jalali, M. Effect of clay minerals and nanoparticles on chromium fractionation in soil contaminated with leather factory waste. J. Hazard Mater. 2015, 297, 127–133, https://doi.org/10.1016/j.jhazmat.2015.04.067.
- 114. Della Puppa, L.; Komárek, M.; Bordas, F.; Bollinger, J.-C.; Joussein, E. Adsorption of copper, cadmium, lead and zinc onto a synthetic manganese oxide. *J. Colloid Interface Sci.* 2013, 399, 99–106, https://doi.org/10.1016/j.jcis.2013.02.029.
- 115. Michálková, Z.; Komárek, M.; Šillerová, H.; Della Puppa, L.; Joussein, E.; Bordas, F.; Vaněk, A.; Vaněk, O.; Ettler, V. Evaluating the potential of three Fe- and Mn-(nano)oxides for the stabilization of Cd, Cu and Pb in contaminated soils. *J. Environ. Manage.* 2014, 146, 226–234, https://doi.org/10.1016/j.jenvman.2014.08.004.
- 116. Fernández, F.G.; Hoeft, R.G. Managing soil pH and crop nutrients. Illinois Agron. Handb. 2009, 24, 91-112.
- 117. Cullen, L.G.; Tilston, E.L.; Mitchell, G.R.; Collins, C.D.; Shaw, L.J. Assessing the impact of nano-and micro-scale zerovalent iron particles on soil microbial activities: Particle reactivity interferes with assay conditions and interpretation of genuine microbial effects. *Chemosphere* **2011**, *82*, 1675–1682.
- 118. Shi, J.; Ye, J.; Fang, H.; Zhang, S.; Xu, C. Effects of copper oxide nanoparticles on paddy soil properties and components. *Nanomaterials* **2018**, *8*, 839.
- García-Gómez, C.; Fernández, M.D.; García, S.; Obrador, A.F.; Letón, M.; Babín, M. Soil pH effects on the toxicity of zinc oxide nanoparticles to soil microbial community. Environ. Sci. Pollut. Res. 2018, 25, 28140–28152.
- 120. Shin, Y.-J.; Kwak, J.I.; An, Y.-J. Evidence for the inhibitory effects of silver nanoparticles on the activities of soil exoenzymes. *Chemosphere* **2012**, *88*, 524–529.
- 121. Jośko, I.; Oleszczuk, P.; Futa, B. The effect of inorganic nanoparticles (ZnO, Cr₂O₃, CuO and Ni) and their bulk counterparts on enzyme activities in different soils. *Geoderma* **2014**, 232, 528–537.
- 122. Ameen, F.; Alsamhary, K.; Alabdullatif, J.A.; ALNadhari, S. A review on metal-based nanoparticles and their toxicity to beneficial soil bacteria and fungi. *Ecotoxicol. Environ. Saf.* 2021, 213, 112027.
- Gao, J.; Wang, Y.; Du, Y.; Zhou, L.; He, Y.; Ma, L.; Yin, L.; Kong, W.; Jiang, Y. Construction of biocatalytic colloidosome using lipase-containing dendritic mesoporous silica nanospheres for enhanced enzyme catalysis. *Chem. Eng. J.* 2017, 317, 175–186.
- 124. Tiede, K.; Boxall, A.B.A.; Tear, S.P.; Lewis, J.; David, H.; Hassellöv, M. Detection and characterization of engineered nanoparticles in food and the environment. Food Addit. Contam.–Part A Chem. Anal. *Control. Expo. Risk Assess.* 2008, 25, 795–821, https://doi.org/10.1080/02652030802007553.
- 125. Blinova, I.; Ivask, A.; Heinlaan, M.; Mortimer, M.; Kahru, A. Ecotoxicity of nanoparticles of CuO and ZnO in natural water. *Environ. Pollut.* **2010**, *158*, 41–47.
- 126. Gondikas, A.P.; von der Kammer, F.; Reed, R.B.; Wagner, S.; Ranville, J.F.; Hofmann, T. Release of TiO₂ nanoparticles from sunscreens into surface waters: A one-year survey at the old Danube recreational Lake. *Environ. Sci. Technol.* 2014, 48, 5415– 5422.
- 127. Stuart, E.J.E.; Compton, R.G. Nanoparticles-emerging contaminants. In *Environmental Analysis by Electrochemical Sensors and Biosensors*; Springer: Berlin/Heidelberg, Germany, 2015; pp. 855–878.
- 128. Antisari, L.V.; Carbone, S.; Gatti, A.; Vianello, G.; Nannipieri, P. Toxicity of metal oxide (CeO₂, Fe₃O₄, SnO₂) engineered nanoparticles on soil microbial biomass and their distribution in soil. *Soil Biol. Biochem.* **2013**, *60*, 87–94.
- Maher, B.A.; González-Maciel, A.; Reynoso-Robles, R.; Torres-Jardón, R.; Calderón-Garcidueñas, L. Iron-rich air pollution nanoparticles: An unrecognised environmental risk factor for myocardial mitochondrial dysfunction and cardiac oxidative stress. *Environ. Res.* 2020, 188, 109816.
- 130. Liu, N.M.; Miyashita, L.; Maher, B.A.; McPhail, G.; Jones, C.J.P.; Barratt, B.; Thangaratinam, S.; Karloukovski, V.; Ahmed, I.A.; Aslam, Z. Evidence for the presence of air pollution nanoparticles in placental tissue cells. *Sci. Total Environ.* **2021**, *751*, 142235.
- 131. Rico, C.M.; Majumdar, S.; Duarte-Gardea, M.; Peralta-Videa, J.R.; Gardea-Torresdey, J.L. Interaction of nanoparticles with edible plants and their possible implications in the food chain. *J. Agric. Food Chem.* **2011**, *59*, 3485–3498.
- Cedervall, T.; Hansson, L.-A.; Lard, M.; Frohm, B.; Linse, S. Food chain transport of nanoparticles affects behaviour and fat metabolism in fish. *PLoS ONE* 2012, 7, e32254.
- 133. Zhu, X.; Wang, J.; Zhang, X.; Chang, Y.; Chen, Y. Trophic transfer of TiO₂ nanoparticles from daphnia to zebrafish in a simplified freshwater food chain. *Chemosphere* **2010**, *79*, 928–933.

- 135. Dietz, K.-J.; Herth, S. Plant nanotoxicology. Trends Plant Sci. 2011, 16, 582-589.
- 136. Liu, J.; Fan, D.; Wang, L.; Shi, L.; Ding, J.; Chen, Y.; Shen, S. Effects of ZnO, CuO, Au, and TiO₂ nanoparticles on Daphnia magna and early life stages of zebrafish Danio rerio. *Environ. Prot. Eng.* **2014**, *40*, 139–149.
- Monteiro-Riviere, N.A.; Inman, A.O.; Zhang, L.W. Limitations and relative utility of screening assays to assess engineered nanoparticle toxicity in a human cell line. *Toxicol. Appl. Pharmacol.* 2009, 234, 222–235.
- 138. Nel, A.; Xia, T.; Mädler, L.; Li, N. Toxic potential of materials at the nanolevel. *Science* 2006, 311, 622–627.
- Yang, W.; Wang, L.; Mettenbrink, E.M.; DeAngelis, P.L.; Wilhelm, S. Nanoparticle Toxicology. *Annu. Rev. Pharmacol. Toxicol.* 2021, 61, 269–289, https://doi.org/10.1146/annurev-pharmtox-032320-110338.
- Olaru, D.; Olaru, A.; Hussein Kassem, G.; Popescu-Drigă, M.V.; Pinoşanu, L.R.; Dumitraşcu, D.I.; Popescu, E.L.; Hermann, D.M.; Popa-Wagner, A. Toxicity and health impact of nanoparticles. Basic biology and clinical perspective. *Rom. J. Morphol. Embryol.* 2019, 60, 787–792.
- 141. Maurer-Jones, M.A.; Lin, Y.-S.; Haynes, C.L. Functional assessment of metal oxide nanoparticle toxicity in immune cells. *ACS Nano* **2010**, *4*, 3363–3373.
- 142. Maurer-Jones, M.A.; Christenson, J.R.; Haynes, C.L. TiO₂ nanoparticle-induced ROS correlates with modulated immune cell function. *J. Nanoparticle Res.* **2012**, *14*, 1291.