



Supplementary Materials

Covalent Organic Frameworks-TpPa-1 as an Emerging Platform for Electrochemical Sensing

Gang Li, Baiqing Yuan *, Sidi Chen, Liju Gan and Chunying Xu

School of Chemistry and Materials Science, Ludong University, Yantai 264025, China

* Correspondence: baiqingyuan1981@126.com or bqyuan@ldu.edu.cn

The electroactive surface areas of these electrodes could be calculated by the Randles–Sevcik equation using the CV responses of the corresponding electrodes in 0.1 M KCl solution in the presence of 5 mM $\text{K}_3\text{Fe}(\text{CN})_6$ at different scan rates at 298 K (Hosseini, H., Ahmar, H., Dehghani, A., Bagheri, A., Tadjarodi, A., Fakhari, A.R., 2013. Bio-sens. Bioelectron. 42, 426–429.):

$$I_p = 2.69 \times 10^5 \times A \times n^{3/2} \times D^{1/2} \times C \times v^{1/2}$$

where, I_p (A) is the peak current, A (cm^2) is the electroactive surface area, n is the number of electron transfer which equals 1, $D = 7.6 \times 10^{-6}$ (cm^2/s), v is the scan rate (V/s), C is the concentration of $\text{K}_3\text{Fe}(\text{CN})_6$ (mol/mL).

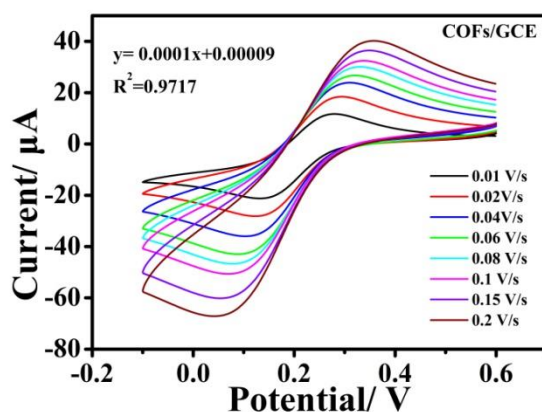


Figure S1. The CVs of COFs/GCE in 0.1 M KCl solution in the presence of 5 mM $\text{K}_3\text{Fe}(\text{CN})_6$ at different scan rates.

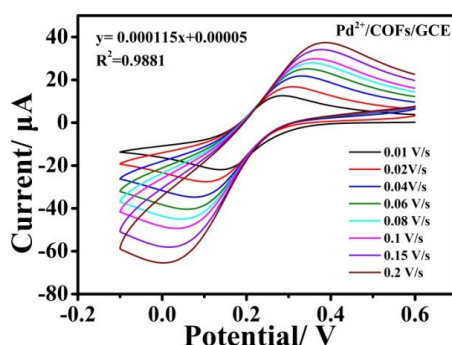
COF/ GCE: $A = 0.027 \text{ cm}^2$ 

Figure S2. The CVs of Pd^{2+} /COFs/GCE in 0.1 M KCl solution in the presence of 5 mM $\text{K}_3\text{Fe}(\text{CN})_6$ at different scan rates.

Pd/ COF/ GCE: $A=0.041 \text{ cm}^2$

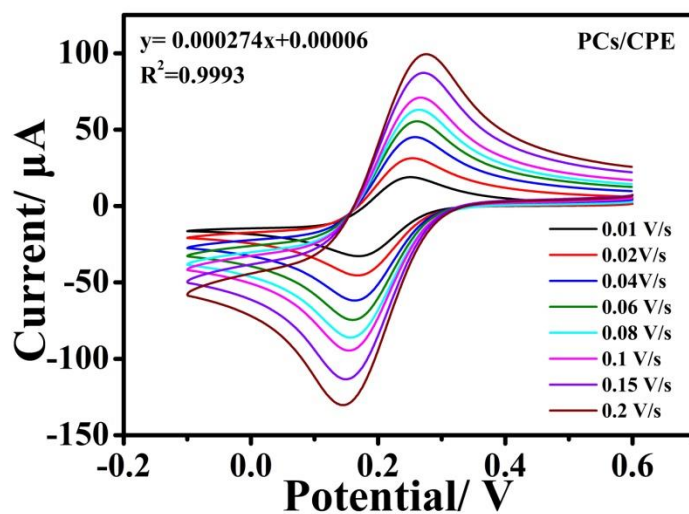


Figure S3. The CVs of PCs/GCE in 0.1 M KCl solution in the presence of 5 mM $\text{K}_3\text{Fe}(\text{CN})_6$ at different scan rates.

PCs: $A=0.074 \text{ cm}^2$

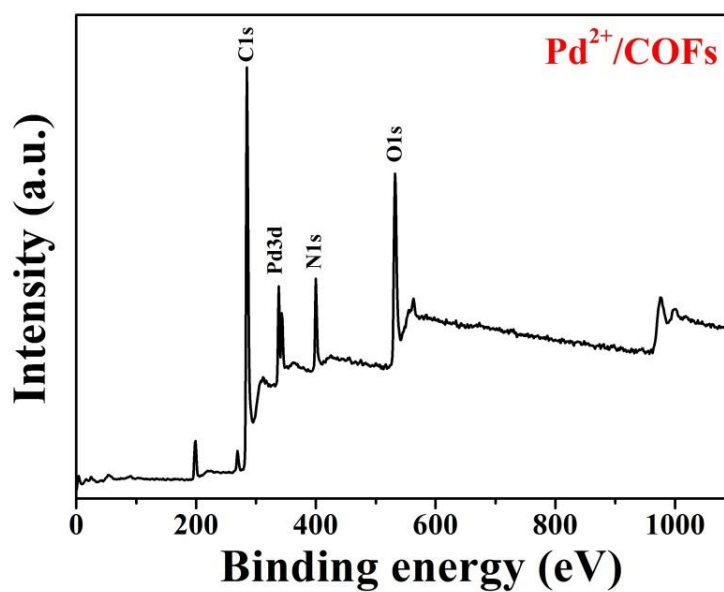


Figure S4. XPS spectra of $\text{Pd}^{2+}/\text{COFs}$.

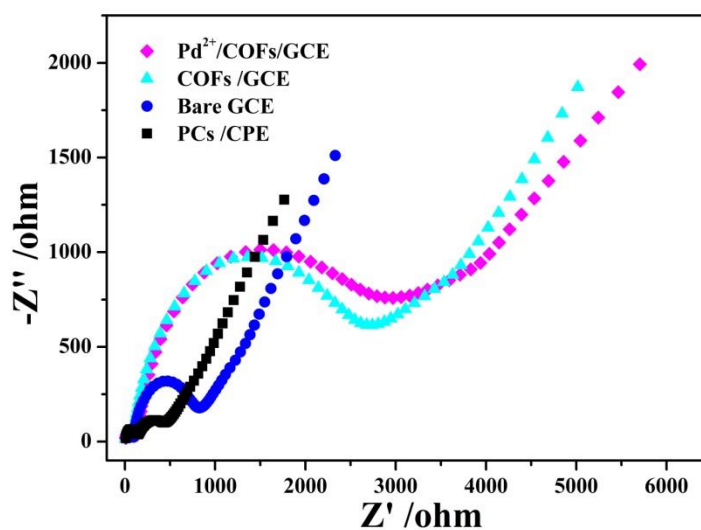


Figure S5. EIS of PCs /GCE, Bare GCE ,COFs/GCE, and Pd²⁺/COFs/GCE in 0.1 M KCl solution in the presence of 5 mM K₃Fe(CN)₆.

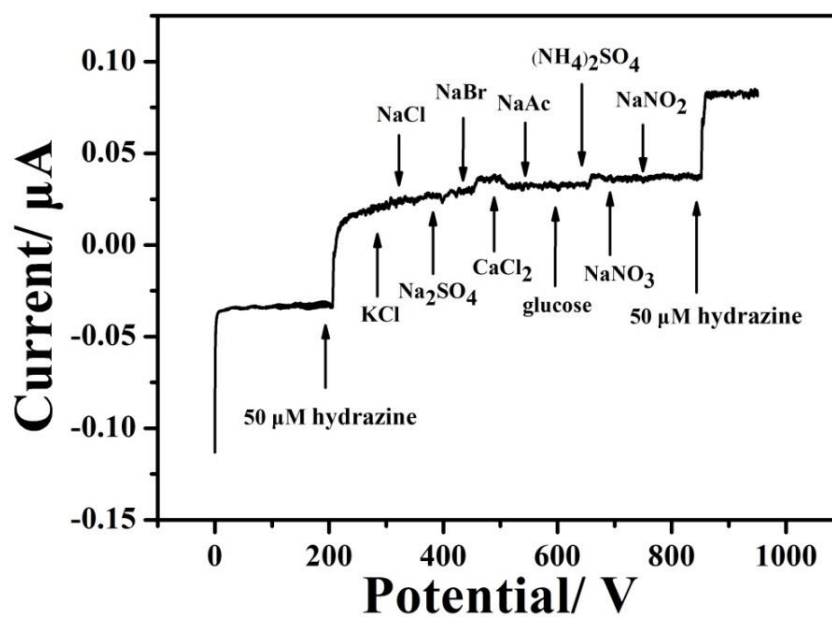


Figure S6. Amperometric curve of Pd²⁺/COFs/GCE with 50 μ M hydrazine and 500 μ M of interferences such as KCl, NaCl, Na₂SO₄, NaBr, CaCl₂, NaAc, glucose, (NH₄)₂SO₄, NaNO₃, and NaNO₂. (Electrolyte: 0.1 M NaOH).

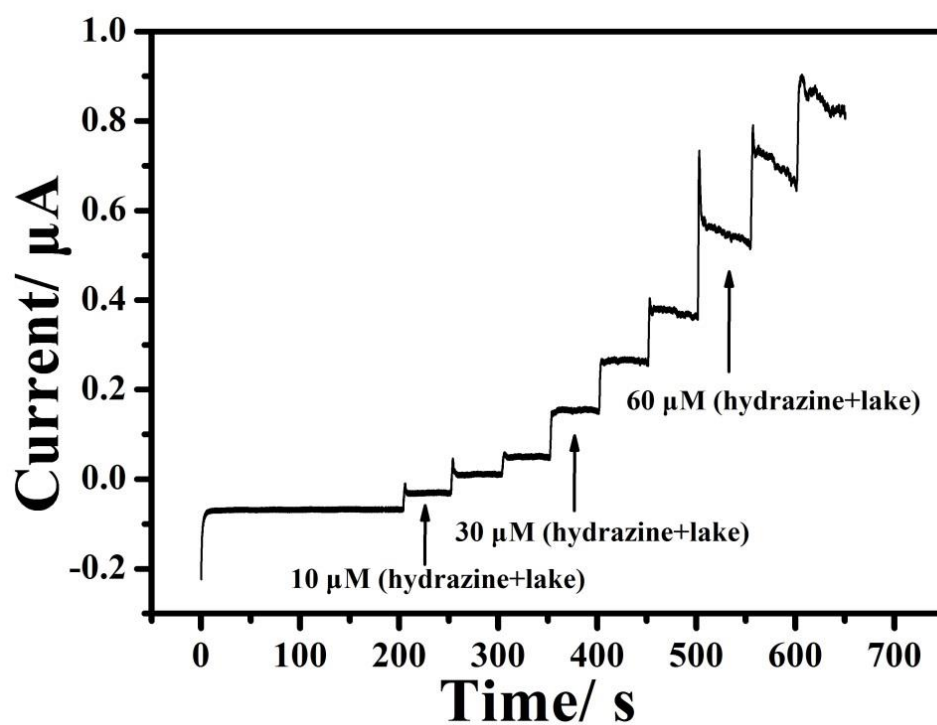


Figure S7. Amperometric curve of $\text{Pd}^{2+}/\text{COFs}/\text{GCE}$ for the detection of hydrazine spiked in water samples at -0.1 V. (Electrolyte: 0.1 M NaOH).

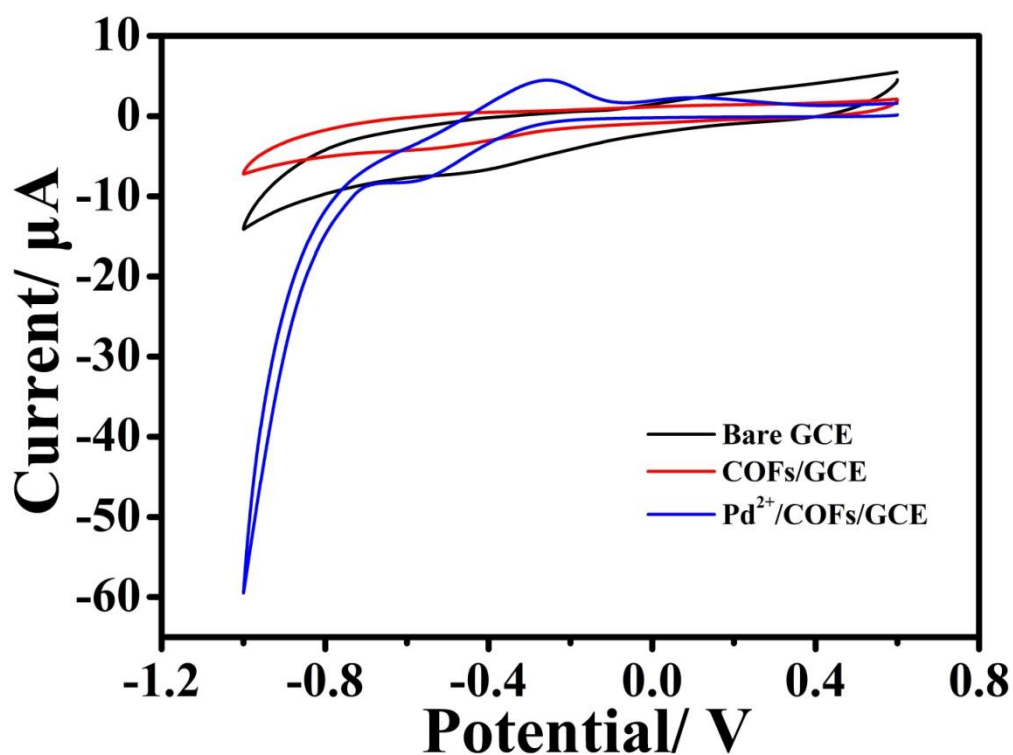


Figure S8. CVs of Bare GCE, COFs/GCE, and $\text{Pd}^{2+}/\text{COFs}/\text{GCE}$ in 0.1 M PBS at a scan rate of 50 mV s^{-1} (pH=7.0).

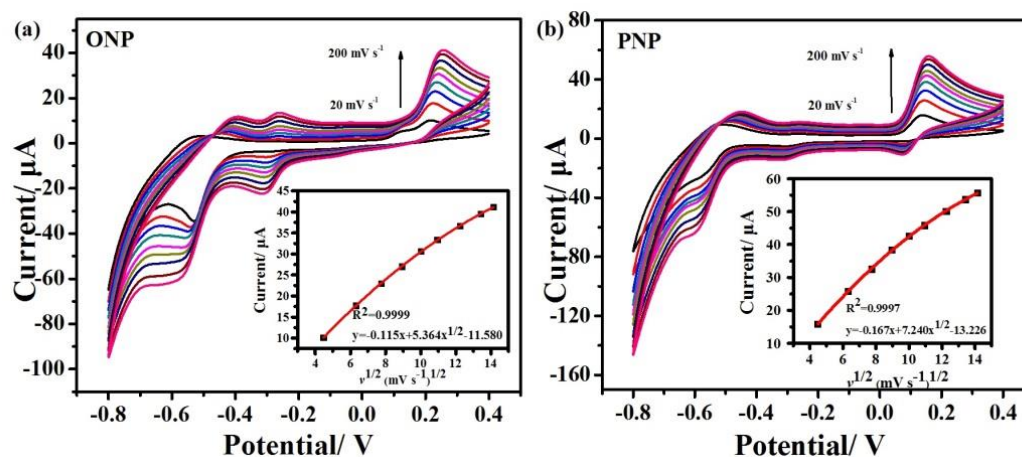


Figure S9. CVs of 500 μM of (a) ONP and (b) PNP on $\text{Pd}^{2+}/\text{COFs}/\text{GCE}$ with different scan rates (20–200 mV s^{-1}) in 0.1 M PBS (pH=7.0). (Inset: the corresponding calibration plot.).

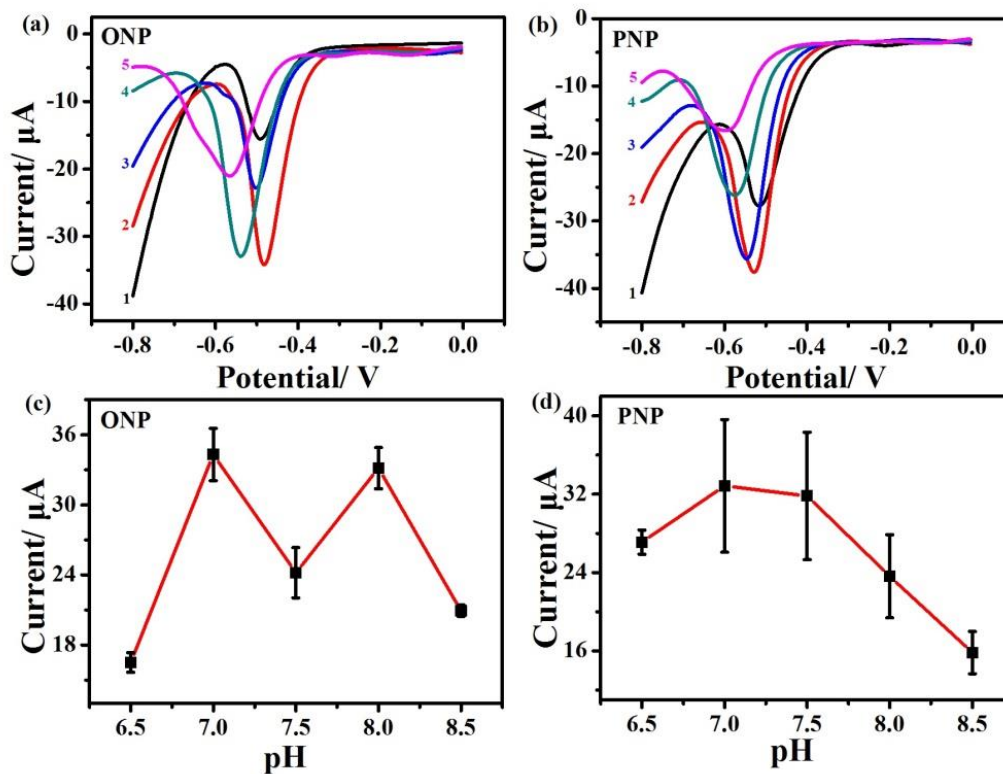


Figure S10. DPV curves of 500 μM of (a) ONP and (b) PNP on $\text{Pd}^{2+}/\text{COFs}/\text{GCE}$ in 0.1 M PBS (pH=7.0) with different pH values (1–5: 6.5, 7.0, 7.5, 8.0, and 8.5). Plots of I_{pc} vs. pH for (c) ONP and (d) PNP.

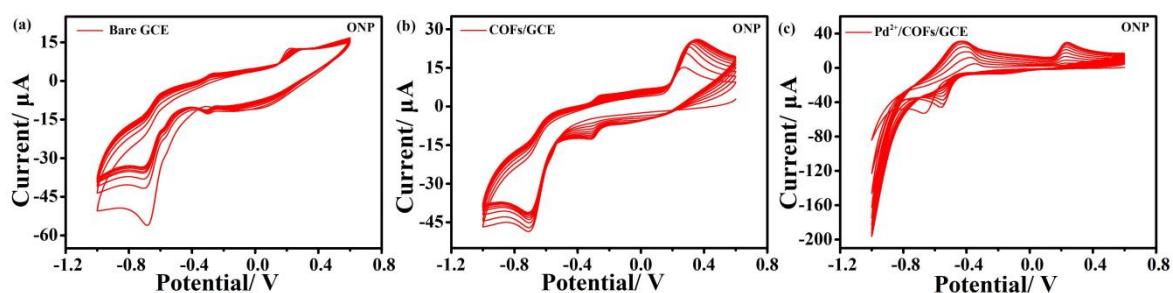


Figure S11. Successive CVs of Bare GCE (a), COFs/GCE (b), and Pd²⁺/COFs/GCE (c) in the presence of 500 µM nitrophenol ONP at a scan rate of 50 mV s⁻¹ in 0.1 M PBS (pH=7).

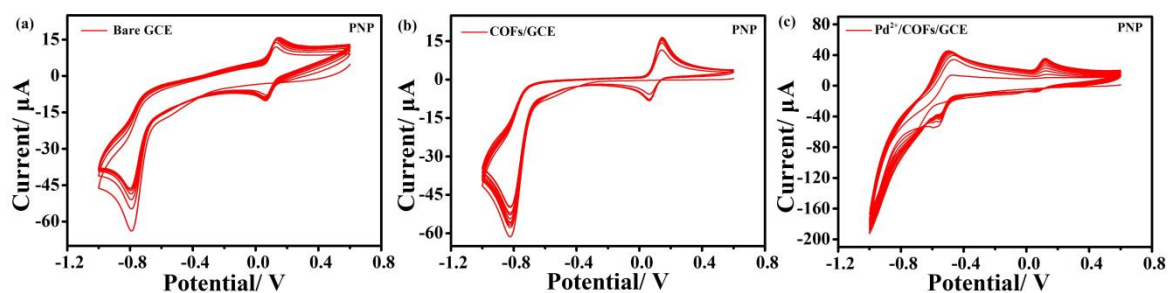


Figure S12. Successive CVs of Bare GCE (a), COFs/GCE (b), and Pd²⁺/COFs/GCE (c) in the presence of 500 µM nitrophenol PNP at a scan rate of 50 mV s⁻¹ in 0.1 M PBS (pH=7.0).

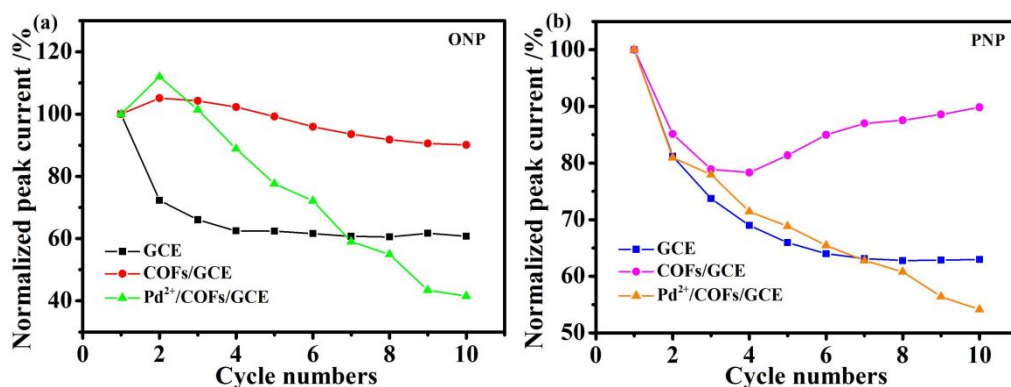


Figure S13. Normalized peak current of bare GCE, COFs/GCE, and Pd²⁺/COFs/GCE in the presence of 500 µM nitrophenol ((a): ONP, (b): PNP) at a scan rate of 50 mV s⁻¹ in 0.1 M PBS (pH=7.0) for ten cycles.

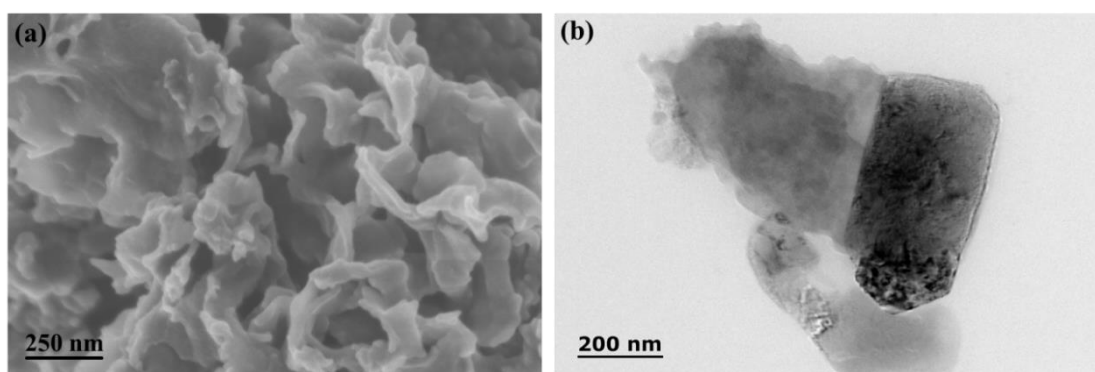


Figure S14. SEM (a) and TEM (b) images of PCs.

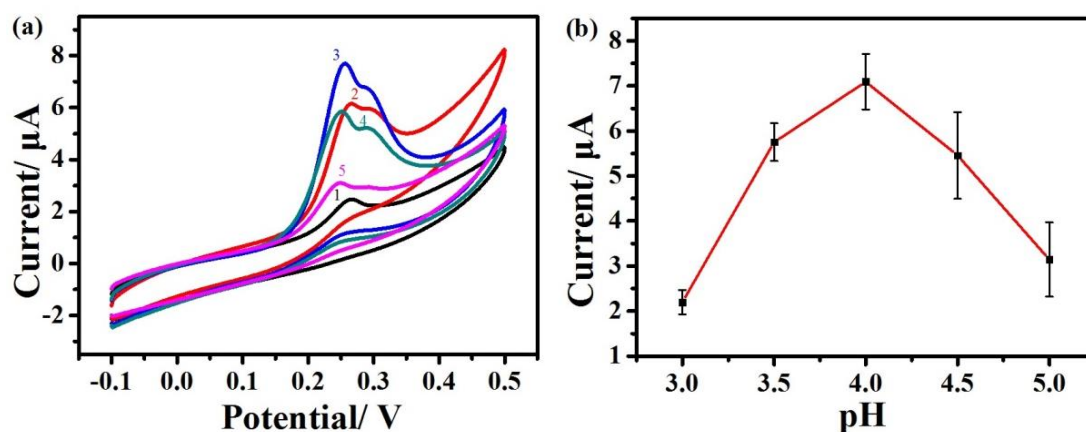


Figure S15. CVs of 5 mM of GSH on (a) PCs/CPE in 0.1 M PBS with different pH values (1-5: 3.0, 3.5, 4.0, 4.5 and 5.0). Plots of I_{pc} vs. pH for (b) GSH.

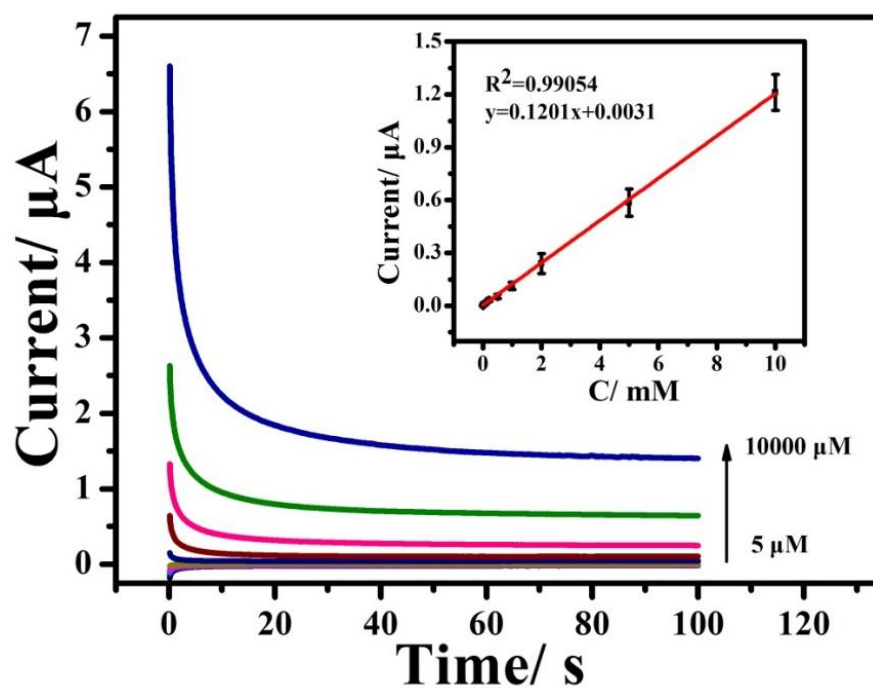


Figure S16. Amperometric responses of PCs/CPE to additions of various concentrations of GSH in 0.1 M PBS (pH=4.0) at 0.25 V. (Inset: the corresponding calibration plot.).

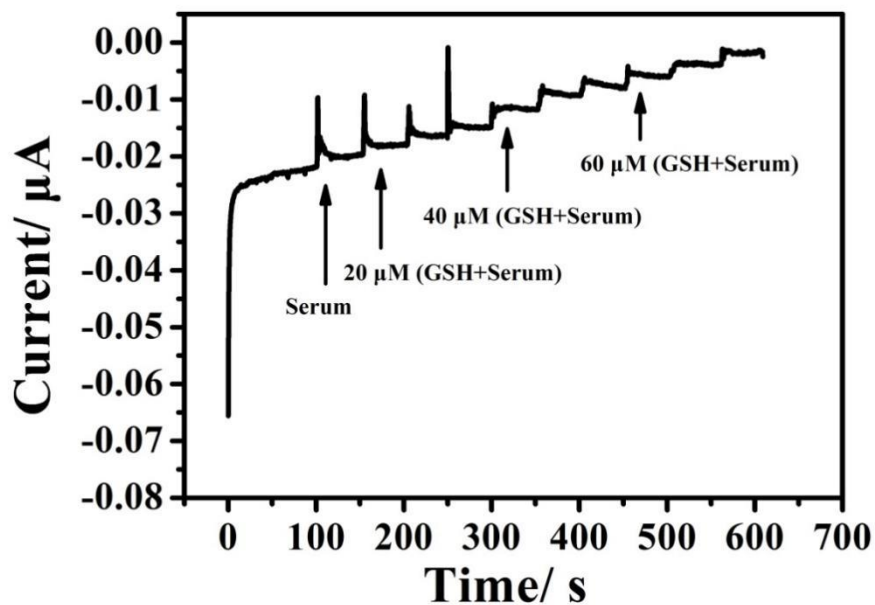


Figure S17. Amperometric responses of PCs/CPE to successive additions of various concentrations of GSH spiked in serum in 0.1 M PBS (pH=4.0) at 0.25 V.

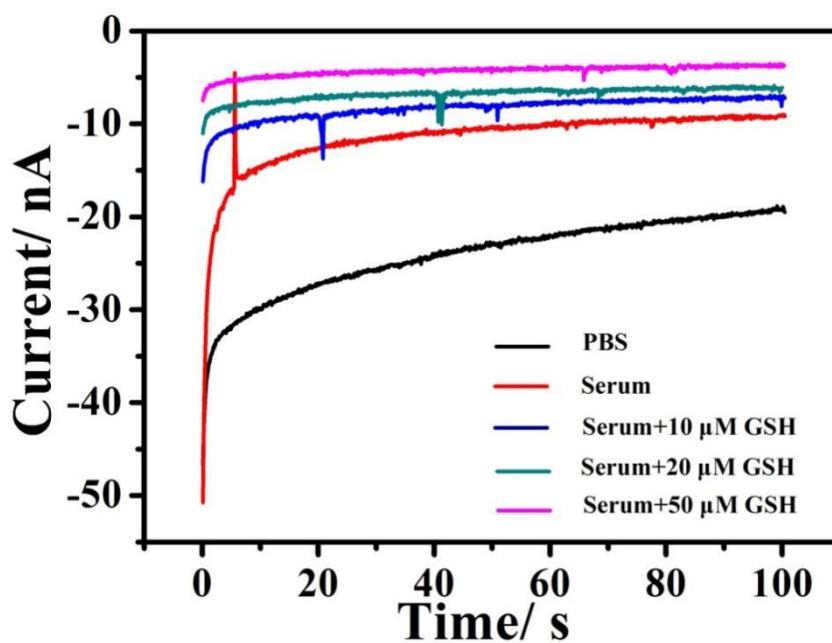


Figure S18. Amperometric responses of PCs/CPE for the detection of GSH spiked in human serum in 0.1 M PBS (pH=4.0) at 0.25 V.

Table S1. Comparison of the Pd²⁺/COFs/GCE for different parameters with previously reported hydrazine electrodes.

Electrode Material	Electrochemical Method	Potential (V)	Linear Range(μM)	Detection Range (μM)	Ref.
Cu-MOFs/OMC/GCE	Amperometry	0.4	0.5-771	0.35	[1]
CuO/OMC/GCE	Amperometry	0.31	1-2110	0.887	[2]
MSRG/Au/GCE	Amperometry	0.29	2-30	0.5	[3]
MXene/ZIF-8/GCE	Amperometry	0.35	10-7700	5.1	[4]
Au@porous P-MWCNT/rGO/GCE	Amperometry	0.4	3000-55000	0.31	[5]
CNT/NiHCF/GCE	Amperometry	0.36	20-200	0.8	[6]
Cu NP/C	Amperometry	0.45	100-2400	1.4	[7]
Ag/ZIF-67/CPE	Amperometry	-0.05	4-326 326-4700	1.45	[8]
NPs/CPE	Amperometry	0.5	1-1300	0.28	[9]
NiCo ₂ S ₄ /GCE	Amperometry	0.3	1.7-7800	0.6	[10]
Pt-Pd/ERGO/GCE	Amperometry	-0.71	7-5500	1.7	[11]
PAYR/Ag@C/GCE	Amperometry	0.4	1-1320	0.25	[12]
rGO-Co ₃ O ₄ @Au/GCE	Amperometry	0.079	10-620	0.443	[13]
ZnO/Co ₃ O ₄ /GCE	Amperometry	-0.2	2-1500 1500-5500	0.52	[14]
Pd²⁺/COFs/GCE	Amperometry	-0.1	0.5-1588.5	0.2	This work

Table S2. Determination of hydrazine spiked in water samples.

Added (μM)	Found (μM)	Recovery (%)	RSD (%)
10	9.5	95	7.3
30	26.7	89	3.9
60	51.3	85.3	1.4

Table S3. Comparison of different modified GCE for detection of ONP and PNP.

Electrode Material	Electrochemical Method	Linear Range (μM)		Detection Limit (μM)		References
		ONP	PNP	ONP	PNP	
BSO-gCN/GCE	LSV	/	1.6-50	/	1	[15]
AcSCD-AuNPs-MC/GCE	DPV	/	0.1-10 10-350	/	3.63	[16]
CeO ₂ :Ag/GCE	CV	/	7.81-1000	/	1.2	[17]
5-sulfosalicylic acid doped polyaniline/GCE	DPV	/	6.7-112.1	/	3.2	[18]
BaO NRs/GCE	DPV	5-640	/	0.5	/	[19]
Mg/Fe-LDH /GCE	Amperometry	1-700	/	4	/	[20]
SDS-HTLC/GCE	DPV	1-600	/	0.5	/	[21]
MnFe-PBA/GCE	DPV	1-700	/	0.59	/	[22]
nano-Au/GCE	CV	10-1000	10-1000	8	8	[23]
meso-ZnCo ₂ O ₄ /GCE	DPV	1-4000	1-4000	0.3	0.3	[24]
PEDOT:PSS/GCE	LSV	10-3000	10-3000	4.55	4.51	[25]
Pd²⁺/COFs/GCE	DPV	5-2000	5-2000	1.75	0.91	This work

Table S4. Comparison of different modified electrodes for electrochemical sensing of GSH.

Electrode Material	Electrochemical Method	pH	Linear Range (μM)	Detection Limit (μM)	Ref.
GO/GCE	Amperometry	5	5-875 875-4080	5	[26]
CoPcS-NWE	Amperometry	4	10-20000	8.3	[27]
N-doped ZnO/C hollow rhombic dodecahedral/GCE	CV	7.4	1-500	8	[28]
MWCNTs/SPE	CV	7.3	5-20	2	[29]
CNTs/SPE	CV	7	10-60	3	[30]
Co-MOCP/CPE	Amperometry	5.5	2.5-950	2.5	[31]
Pt/rGMnO	Amperometry	6.5	1-10 10-100	0.9	[32]
Co(II)TAPc-TA/Au	DPV	7.4	10-100	0.28	[33]
N-G/CoPc/GCE	Amperometry	0.1 M NaOH	1-8000	0.6	[34]
RuHCF/rGO/PIGE	SWV	5	5.12-25.58	1.7	[35]
PCs/CPE	Amperometry	4	5-10000	1	This work

Table S5. Determination of GSH spiked in human serum by successive additions.

Added (μM)	Expected (μM)	Recovery (%)	RSD (%)
20	19.2	96	4.2
40	39.6	99	2.5
60	54.9	91.5	1.7

Table S6. Determination of GSH spiked in human serum by a non-successive addition method.

Added (μM)	Detected (μM)	Recovery (%)	RSD (%)
10	10.7	107	3.0
20	18.9	94.5	2.4
50	46	92	1.1

References

1. L. Wang, Q.Q. Teng, X.T. Sun, Y.T. Chen, Y.M. Wang, H. Wang, Y.F. Zhang, Facile synthesis of metal-organic frameworks/ordered mesoporous carbon composites with enhanced electrocatalytic ability for hydrazine, *J. Colloid. Interf. Sci.* 512(2018) 127-133.
2. L. Wang, T.J. Meng, H.X. Jia, Y. Feng, T. Gong, H. Wang, Y.F. Zhang, Electrochemical study of hydrazine oxidation by leaf-shaped copper oxide loaded on highly ordered mesoporous carbon composite, *J. Colloid Interface Sci.* 549(2019) 98-104.
3. M. Gharani, A. Bahari, S. Ghasemi, Preparation of MoS₂-reduced graphene oxide/Au nanohybrid for electrochemical sensing of hydrazine, *J. Mater. Sci. Mater. Electron.* 32(2021) 7765-7777.
4. Y.Q. Yao, X.H. Han, X.H. Yang, J. Zhao, C.P. Chai, Detection of Hydrazine at MXene/ZIF-8 Nanocomposite Modified Electrode, *Chin. J. Chem.* 39(2021) 330-336.
5. X.J. Zhang, J.B. Zheng, Amperometric hydrazine sensor based on the use of a gold nanoparticle-modified nanocomposite consisting of porous polydopamine, multiwalled carbon nanotubes and reduced graphene oxide, *Mikrochim. Acta.* 187(2020) 89.
6. N. Vishnu, A.S. Kumar, S. Badhulika, Selective in-situ derivatization of intrinsic nickel to nickel hexacyanoferrate on carbon nanotube and its application for electrochemical sensing of hydrazine, *J. Electroanal. Chem.* 837(2019) 60-66.
7. S. Li, W.S. Feng, X.H. Gao, A.M. Guo, H.J. Li, Copper-based materials derived from metal-organic frameworks for electrochemical sensing of hydrazine, *Micro Nano Lett.* 16(2021) 478-483.
8. F. Asadi, S.N. Azizi, S. Ghasemi, Preparation of Ag nanoparticles on nano cobalt-based metal organic framework (ZIF-67) as catalyst support for electrochemical determination of hydrazine, *J. Mater. Sci. Mater. Electron.* 30(2019) 5410-5420.
9. A. Avanes, M. Hasanazadeh-Karamjavan, G. Shokri-Jarcheloo, Electrocatalytic oxidation and amperometric determination of hydrazine using a carbon paste electrode modified with beta-nickel hydroxide nanoplatelets, *Mikrochim. Acta.* 186(2019) 441.

10. C.Q. Duan, Y.H. Dong, Q.L. Sheng, J.B. Zheng, A high-performance non-enzymatic electrochemical hydrazine sensor based on NiCo_2S_4 porous sphere, *Talanta*. 198(2019) 23-29.
11. S. Ghasemi, S.R. Hosseini, F. Hasanpoor, S. Nabipour, Amperometric hydrazine sensor based on the use of Pt-Pd nanoparticles placed on reduced graphene oxide nanosheets, *Mikrochim. Acta*. 186(2019) 601.
12. A. Maleki, R. Rezaee, H. Daraei, B. Shahmoradi, N. Amini, Fabrication of a sensitive electrochemical sensor to environmental pollutant of hydrazine in real water samples based on synergistic catalysis of Ag@C core-shell and polyalizarin yellow R, *J. Alloys Compd.* 763(2018) 997-1004.
13. M.M. Shahid, P. Rameshkumar, W.J. Basirunc, U. Wijayantha, W.S. Chiu, P.S. Khiew, N.M. Huang, An electrochemical sensing platform of cobalt oxide@gold nanocubes interleaved reduced graphene oxide for the selective determination of hydrazine, *Electrochim. Acta*. 259(2018) 606-616.
14. A. Mousavi-Majd, S. Ghasemi, S.R. Hosseini, Zeolitic imidazolate framework derived porous $\text{ZnO}/\text{Co}_3\text{O}_4$ incorporated with gold nanoparticles as ternary nanohybrid for determination of hydrazine, *J. Alloys Compd.* 896(2022).
15. S. Vinoth, P. Sampathkumar, K. Giribabu, A. Pandikumar, Ultrasonically assisted synthesis of barium stannate incorporated graphitic carbon nitride nanocomposite and its analytical performance in electrochemical sensing of 4-nitrophenol, *Ultrasonics Sonochemistry*, *Ultrason. Sonochem.* 62(2020) 104855.
16. Y.Y. Zhou, J. Zhao, S.H. Li, M.J. Guo, Z. Fan, An electrochemical sensor for the detection of p-nitrophenol based on a cyclodextrin-decorated gold nanoparticle-mesoporous carbon hybrid, *Analyst*. 144(2019) 4400-4406.
17. A.A. Ansari, M. Alam, M.A. Ali, Nanostructured CeO_2 : Ag platform for electrochemically sensitive detection of nitrophenol, *Colloids Surf. A*. 613(2021) 126116.
18. R. Suresh, K. Giribabu, R. Manigandan, S. Praveen Kumar, S. Munusamy, S. Muthamizh, V. Narayanan, Polyaniline Nanorods: Synthesis, Characterization, and Application for the Determination of para-Nitrophenol, *Anal. Lett.* 49(2015) 269-281.
19. M.M. Alam, A.M. Asiri, M.M. Rahman, Electrochemical detection of 2-Nitrophenol using a glassy carbon electrode modified with BaO Nanorods, *Chem. Asian. J.* 16(2021) 1475-1485.
20. K. Nejati, K. Asadpour-Zeynali, Z. Rezvani, R. Peyghami, Determination of 2-nitrophenol by electrochemical synthesized Mg/Fe layered double hydroxide sensor, *Int. J. Electrochem. Sci.* 9(2014) 5222-5234.
21. H.S. Yin, Y.L. Zhou, S.Y. Ai, L. Cui, L.S. Zhu, Electrochemical determination of 2-Nitrophenol in Water Samples using Mg-Al-SDS Hydrotalcite-Like Clay modified glassy carbon electrode, *Electroanalysis*. 22(2010) 1136-1142.
22. J.H. Li, L.Z. He, J.B. Jiang, Z.F. Xu, M.Q. Liu, X. Liu, H.X. Tong, Z. Liu, D. Qian, Facile syntheses of bimetallic Prussian blue analogues ($\text{K}_x\text{M}[\text{Fe}(\text{CN})_6] \cdot n\text{H}_2\text{O}$, $\text{M}=\text{Ni}$, Co , and Mn) for electrochemical determination of toxic 2-nitrophenol, *Electrochim. Acta*, 353(2020) 136579.
23. L. Chu, L. Han, X.L. Zhang, Electrochemical simultaneous determination of nitrophenol isomers at nano-gold modified glassy carbon electrode, *J. Appl. Electrochem.* 41(2011) 687-694.
24. J.J. Zhang, S.Q. Cui, Y.P. Ding, X.X. Yang, K. Guo, J.T. Zhao, Bioelectronics, Two-dimensional mesoporous ZnCo_2O_4 nanosheets as a novel electrocatalyst for detection of o-nitrophenol and p-nitrophenol, *Biosens. Bioelectron.* 112(2018) 177-185.
25. B.M. Hryniewicz, E.S. Orth, M. Vidotti, Enzymeless PEDOT-based electrochemical sensor for the detection of nitrophenols and organophosphates, *Sens. Actuators B Chem.* 257(2018) 570-578.
26. B.Q. Yuan, X.Y. Zeng, C.Y. Xu, L. Liu, Y.H. Ma, D.J. Zhang, Y. Fan, Electrochemical modification of graphene oxide bearing different types of oxygen functional species for the electro-catalytic oxidation of reduced glutathione, *Sens. Actuators B Chem.* 184(2013) 15-20.
27. W.T. Wu, X. Chen, Y.T. Jiao, W.T. Fan, Y.L. Liu, W.H. Huang, Versatile Construction of Biomimetic Nanosensors for Electrochemical Monitoring of Intracellular Glutathione, *Electroanal. Chem.* 134(2022) e202115820.
28. L.Z. Zhao, L. Zhao, Y. Miao, C.X. Zhang, Selective electrochemical determination of glutathione from the leakage of intracellular GSH contents in HeLa cells following doxorubicin-induced cell apoptosis, *Electrochim. Acta*. 206(2016) 86-98.
29. P. Lee, R.G. Compton, Selective electrochemical detection of thiol biomarkers in saliva using multiwalled carbon nanotube screen-printed electrodes, *Sens. Actuators B Chem.* 209(2015) 983-988.
30. P.T. Lee, L.M. Goncalves, R.G. Compton, Electrochemical determination of free and total glutathione in human saliva samples, *Sens. Actuators B Chem.* 221(2015) 962-968.
31. B.Q. Yuan, R.C. Zhang, X.X. Jiao, J. Li, H.Z. Shi, D.J. Zhang, Amperometric determination of reduced glutathione with a new Co-based metal-organic coordination polymer modified electrode, *Electrochem. Commun.* 40(2014) 92-95.
32. S. Kannappan, L. Ramachandra Bhat, N. Nesakumar, K.J. Babu, A.J. Kulandaisamy, J.B.B. Rayappan, Design and development of a non-enzymatic electrochemical biosensor for the detection of Glutathione, *Electroanalysis*. 34(2022)1-12.
33. M.N. Abbas, A.A. Saeed, M.B. Ali, A. Errachid, N. Zine, A. Baraket, B. Singh, Biosensor for the oxidative stress biomarker glutathione based on SAM of cobalt phthalocyanine on a thioctic acid modified gold electrode, *J. Solid State Electrochem.* 23(2019) 1129-1144.
34. H.Y. Xu, J.J. Xiao, B. Liu, S. Griveau, F. Bedioui, Enhanced electrochemical sensing of thiols based on cobalt phthalocyanine immobilized on nitrogen-doped graphene, *Biosens. Bioelectron.* 66(2015) 438-444.
35. S. Saranya, B. Geetha, P.N. Deepa, Simultaneous detection of glutathione, threonine, and glycine at electrodeposited RuHCF/rGO -modified electrode, *Ionics*. 25(2019) 5537-5550.