



Supplementary material

Regulation of Antimicrobial Effect of Hemicyanine-based photosensitizer via Supramolecular Assembly

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Materials and Measurements

All chemicals were purchased from the commercial sources (Sigma-Aldrich Shanghai China, and Alfa-Aesar Shanghai China). All organic solvents were obtained from Beijing Chemical Works. UV-Vis absorption spectra were got from a JASCO V-550 spectrophotometer. Fluorescence spectra were measured on a Hitachi F-4500 fluorometer. Three microorganisms (Amp^r *E. coli*, *S. aureus*, *C. albicans*) were selected as representative strains, which were purchased from China General Microbiological Culture Collection Center. The concentration of microbial suspensions (OD₆₀₀) tested by a UV-Vis spectrophotometer (JASCO V-550) at 600 nm. Bacterial culture related consumables are used after sterilization. Photographs of bright-field and fluorescence-field were taken with a fluorescence microscope (Ti2-U, Nikon, Tokyo, Japan). Zeta potentials were measured on a Nano ZS (ZEN3600) (Malvern, Malvern, UK) system. The ¹H NMR spectra were collected by Bruker Avance III 400 HD nuclear magnetic resonance spectrometer.

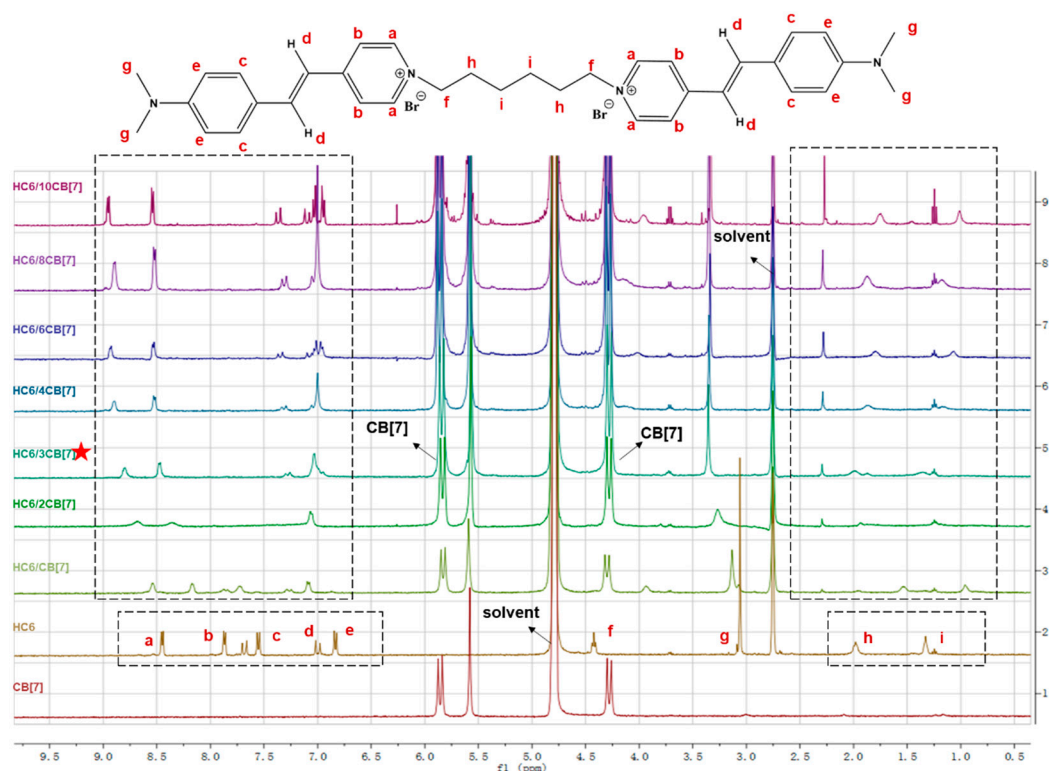


Figure S1. ^1H NMR (400MHz, D_2O with 0.17% $\text{DMSO-}d_6$) spectra of HC6 before and after adding CB[7]. The chemical shifts of protons in aromatic and dimethyl amino groups gradually move to downfield, indicating the assembly complexes HC6/CB[7] form. This is because that the formation of HC6/CB[7] complexes can reduce the aggregation degree of HC6, resulting in the decrease of electron density in aromatic and dimethyl amino groups which lead to a higher ppm value in NMR spectra.

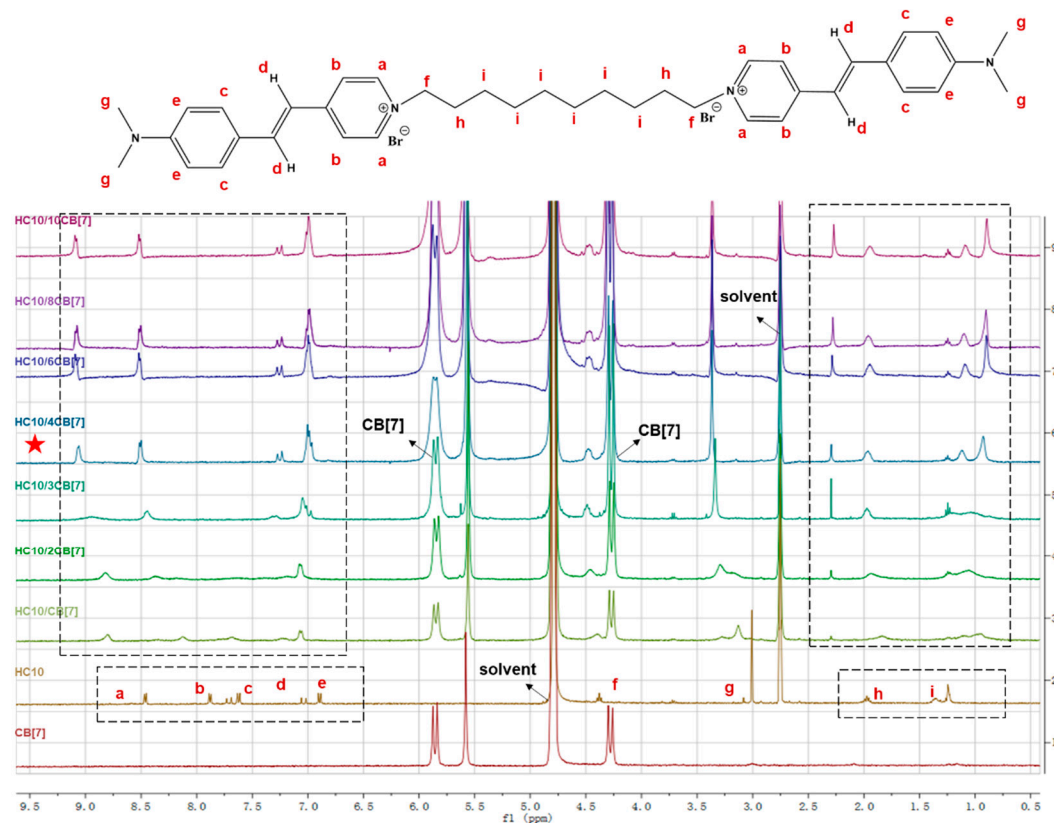


Figure S2. ^1H NMR (400MHz, D_2O with 0.17% $\text{DMSO-}d_6$) spectra of HC10 before and after adding CB[7].

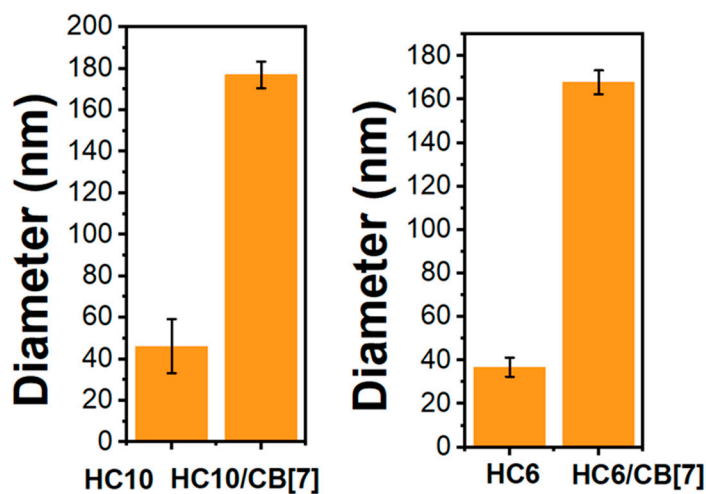


Figure S3. Size histograms of HC6 or HC10 and its supramolecular complex HCs/CB[7].

Table S1. Zeta potential data of Amp^r *E. coli*, *S. aureus* and *C. albicans* before and after binding with HCs or supramolecular conjugate HCs/CB[7] complex, respectively.

Samples		Zeta potentials (ζ , mV)
Amp ^r <i>E. coli</i>	Blank	-44.5 ± 0.2
	HC6	-41.6 ± 0.4
	HC6/CB[7]	-45.4 ± 0.3
	HC10	-39.1 ± 0.5
	HC10/CB[7]	-44.5 ± 0.1
<i>S. aureus</i>	Blank	-26.2 ± 0.1
	HC6	-23.4 ± 0.4
	HC6/CB[7]	-24.1 ± 0.1
	HC10	-19.2 ± 1.0
	HC10/CB[7]	-26.1 ± 0.6
<i>C. albicans</i>	Blank	-18.0 ± 0.1
	HC6	-17.1 ± 0.3
	HC6/CB[7]	-17.3 ± 0.3
	HC10	-18.2 ± 0.9
	HC10/CB[7]	-18.3 ± 1.1

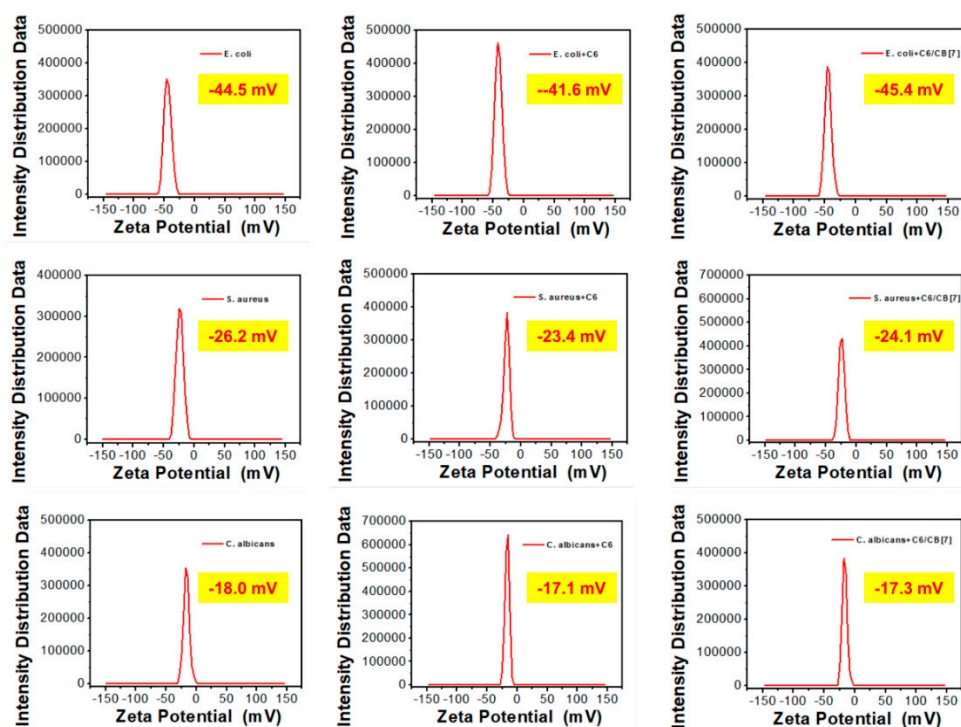


Figure S4. Zeta potentials of the three pathogens before and after binding with HC6 or supramolecular conjugate complex HC6/CB[7].

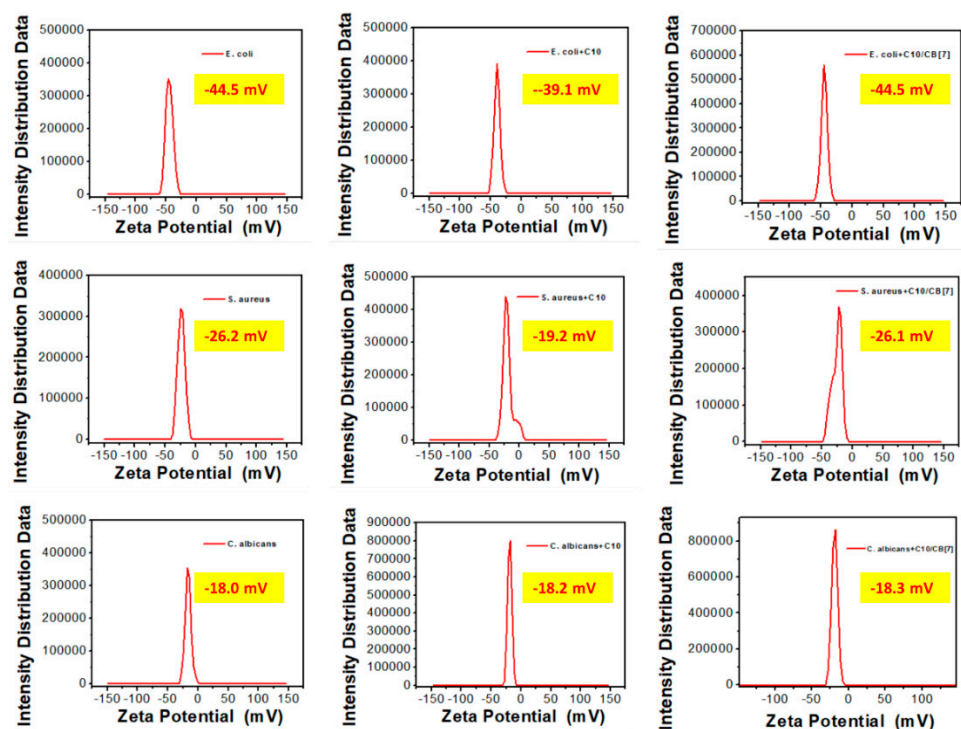


Figure S5. Zeta potentials of the three pathogens before and after binding with HC10 or supramolecular conjugate complex HC10/CB[7].

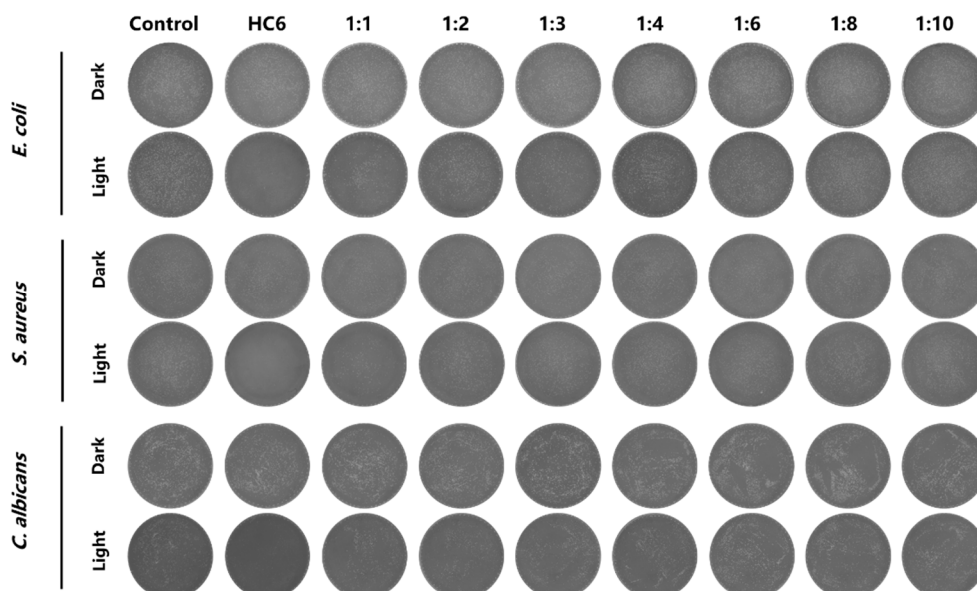


Figure S6. Plate photographs of *E. coli*, *S. aureus* and *C. albicans* acting with HC6 and its supramolecular assemblers in different proportions. [HC6] = 25 μ M, [CB[7]] = 0, 25, 50, 75, 100, 150, 200, 250 μ M.

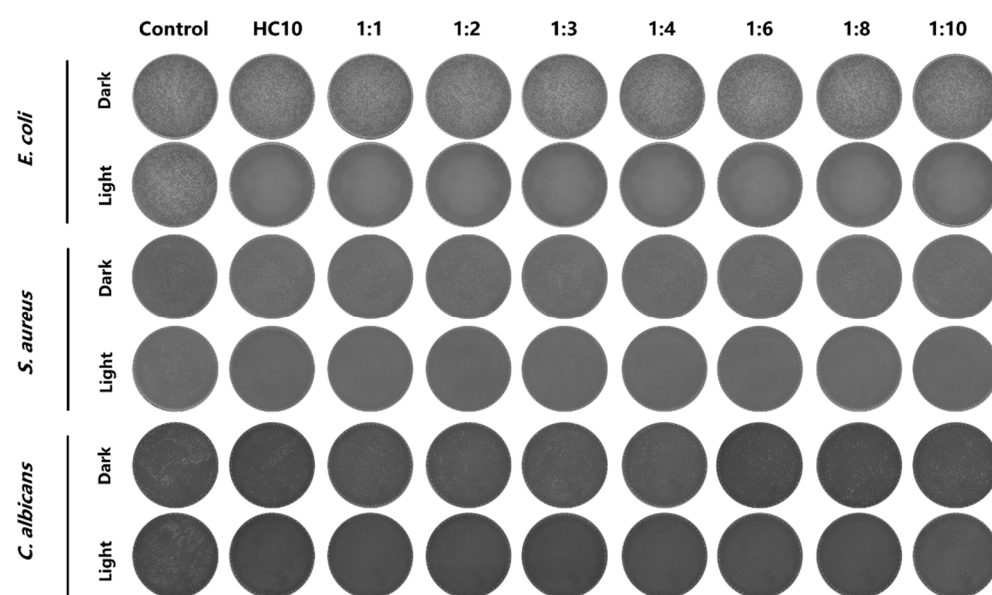


Figure S7. Plate photographs of *E. coli*, *S. aureus* and *C. albicans* acting with HC10 and its supramolecular assemblers in different proportions. [HC10] = 25 μ M, [CB[7]] = 0, 25, 50, 75, 100, 150, 200, 250 μ M.

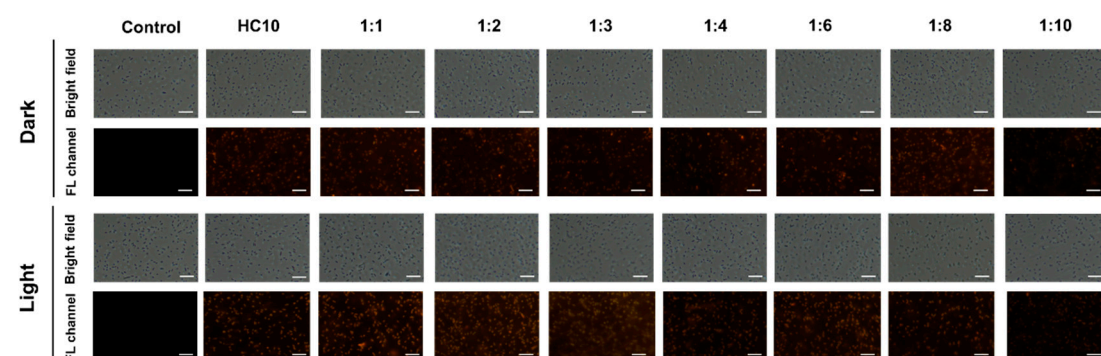


Figure S8. Fluorescence microscope images of *E. coli* before and after binding with HC10 and HC10/CB[7] in different proportions under light (65 mW/cm²) and darkness, [HC10] = 25 μ M, [CB[7]] = 0, 25, 50, 75, 100, 150, 200, 250 μ M, the scale is 20 μ m.

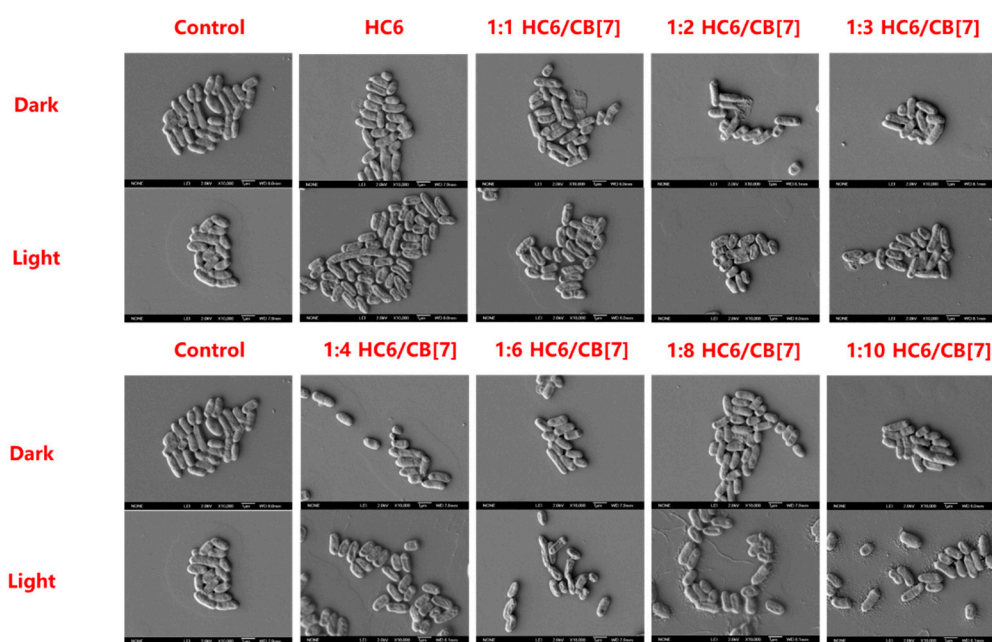


Figure S9. SEM images of Amp^r *E. coli* with HC6 and HC6/CB[7] before and after the interaction under dark or light (65 mW/cm²), the scale is 1 μm.