

*Supplementary Information*

# Influence of Coating and Size of Magnetic Nanoparticles on Cellular Uptake for In Vitro MRI

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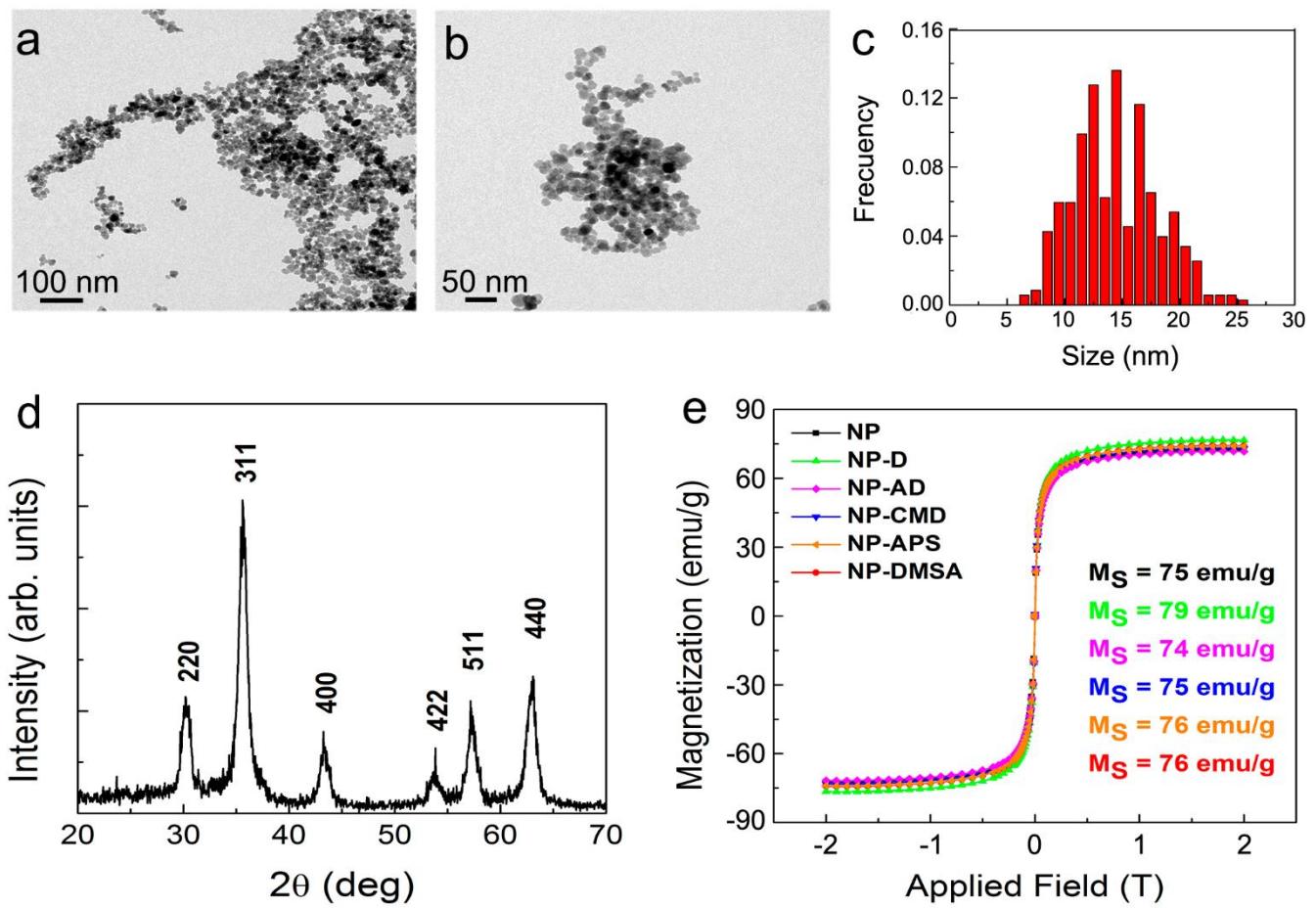
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## IONPs characterization

### XRD and VSM measurements

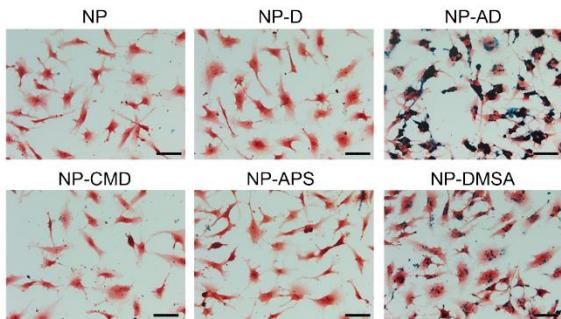
The diffraction maxima (Figure S1D) have been indexed as 220, 311, 400, 422, 511 and 440 reflections corresponding all of them to the maghemite phase (ICDD 00-039-1346). Almost identical Ms values in the range 74–79 emu/g, typical of maghemite, were measured for all samples (Figure S1E).



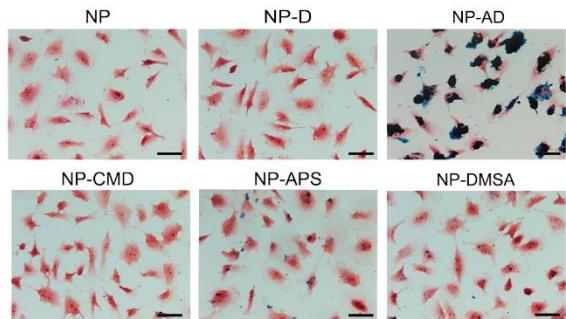
**Figure S1.** **a-c)** Selected TEM micrographs and histogram showing the size distribution of IONPs. **d)** XRD pattern and diffraction maxima from maghemite nanoparticles. **e)** VSM measurements for the different coated IONPs. Key words: NP (naked), NP-D (dextran), NP-AD (amino-dextran), NP-CMD (carboxymethyl-dextran), NP-APS (aminopropyl-triethoxy silane), and NP-DMSA (dimercaptosuccinic acid).

### IONPs uptake by the CMI method

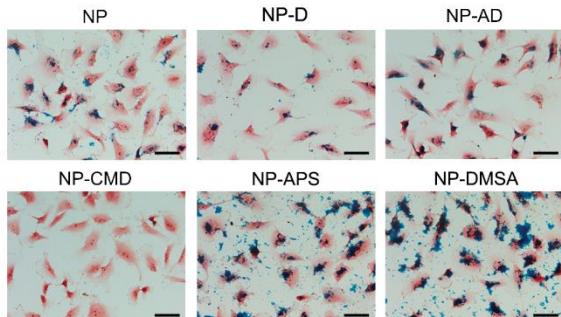
A. Filtered IONPs + DMEM 10%FBS



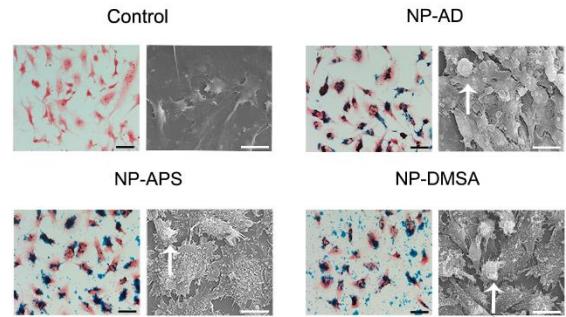
B. Unfiltered IONPs + DMEM 10%FBS



C. Unfiltered IONPs + DMEM 0%FBS



D. Prussian blue - SEM



**Figure S2.** Optical microscopy images of U373 cells after Prussian blue staining following **A)** Filtered IONPs suspended in DMEM with 10% FBS, **B)** Un-filtered IONPs suspended in DMEM with 10% of FBS, **C)** Un-filtered IONPs in DMEM at 0% of FBS. **D)** Scanning electron microscopy of a control U373 cells, filtered NP-AD on DMEM 10% FBS, un-filtered NP-APS and NP-DMSA on DMEM 0% FBS. White arrows show IONPs on the cell membrane surface. Scale bars: black = 50  $\mu$ m, white = 25  $\mu$ m.

### Colloidal properties

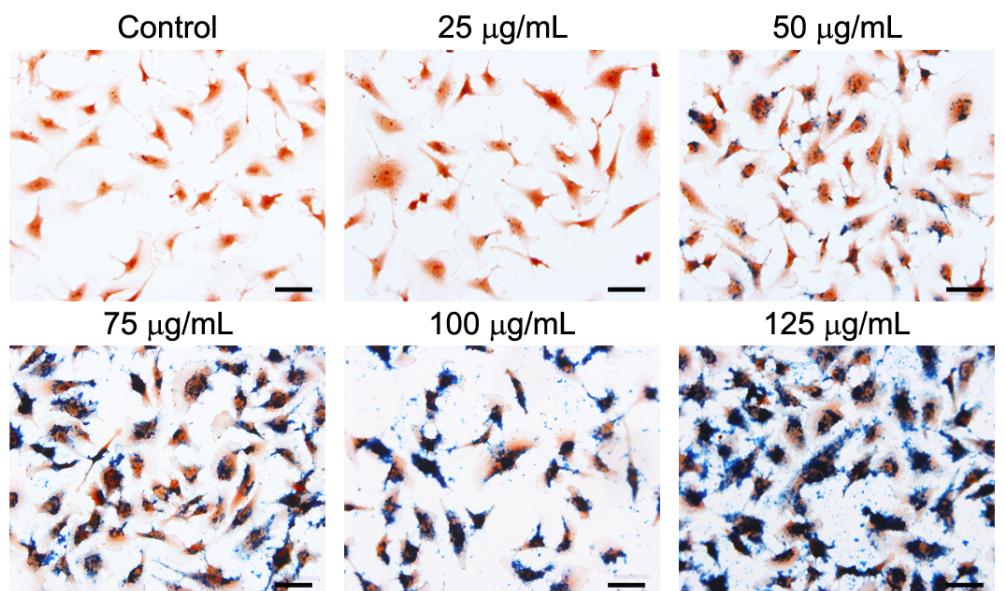
**Table S1.** Colloidal properties of IONPs Table 1, after CMI improvements.

IONPs	$D_{hyd}$ (nm)	$\zeta$ potential (mV)	Labelling (%)
NP	1347 (0.3)	-8	$97.6 \pm 8.8$
NP-D	132 (0.3)	-4	$81.6 \pm 5.7$
NP-AD	76 (0.3)	10	$97.3 \pm 2.9$
NP-CMD	84 (0.1)	-18	$2.5 \pm 0.4$
NP-APS	177 (0.2)	-1	$79.5 \pm 7.4$
NP-DMSA	335 (0.3)	-12	$98.7 \pm 1.2$

**Table S2.** Mean hydrodynamic diameters (with PDI in parentheses) and zeta potential values of IONPs with different size core suspended in water, DMEM cell culture medium and DMEM supplemented with FBS (10%).

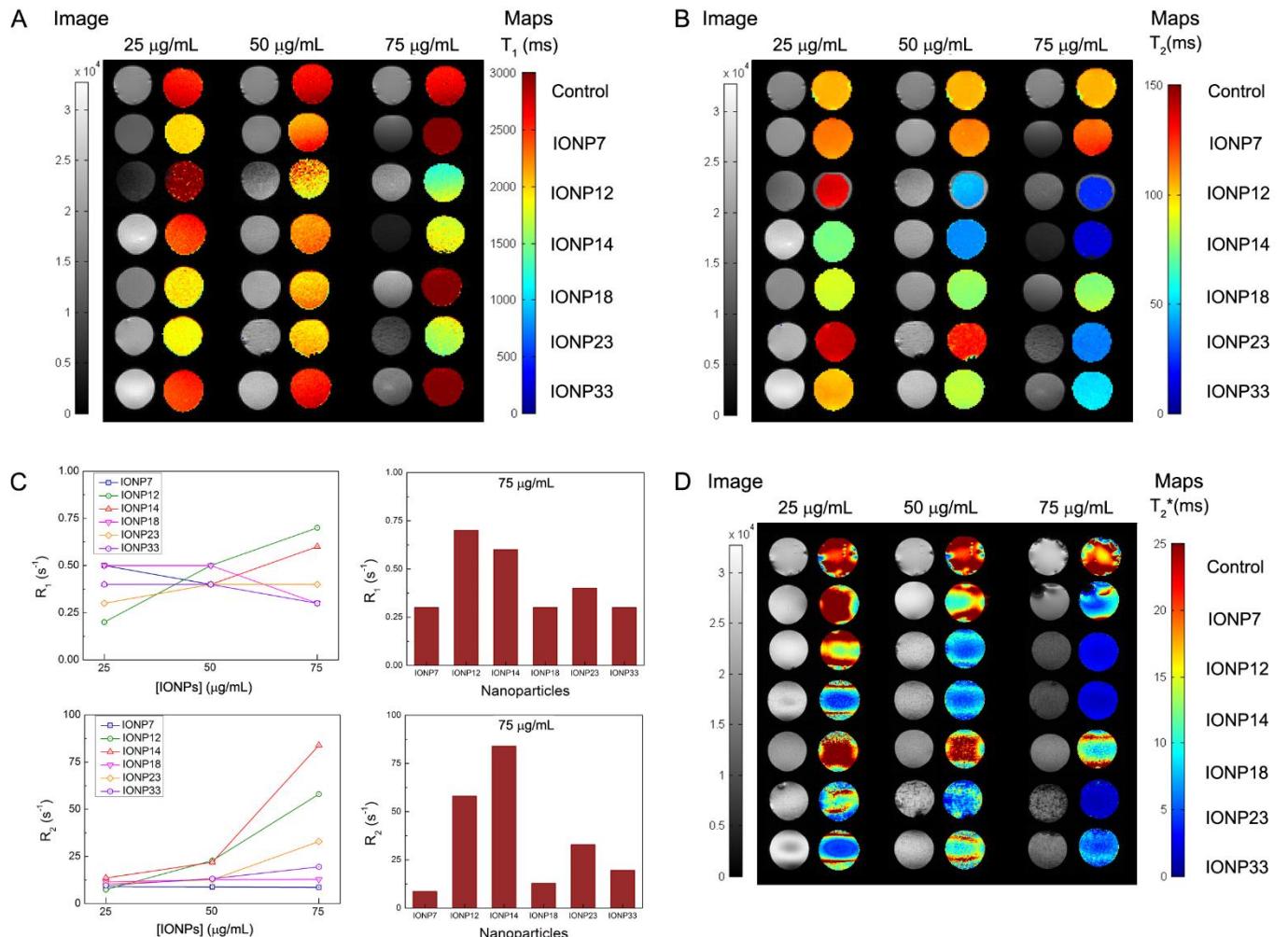
IONPs	$D_{hyd}$ (nm)			$\zeta$ potential (mV)		
	Water	DMEM	DMEM+10%FBS	Water	DMEM	DMEM+10%FBS
IONP7	53 (0.1)	958 (0.3)	376 (0.3)	-43	-15	-11
IONP12	78 (0.3)	1014 (0.2)	357 (0.3)	-41	-10	-12
IONP14	49 (0.4)	1002 (0.7)	212 (0.3)	-37	-22	-12
IONP18	44 (0.2)	820 (0.2)	223 (0.4)	-37	-22	-12
IONP23	45 (0.2)	1946 (0.2)	424 (0.4)	-43	-16	-12
IONP33	85 (0.3)	834 (0.3)	350 (0.3)	-42	-14	-11

Figure S3 shows the optical images after Prussian blue staining at different initial IONP23 concentrations. With high initial concentrations of 100 and 125 µg/mL, agglomerates appeared on the cell surface, so 25, 50 and 75 µg/mL of initial concentration were used to favour internalization.



**Figure S3.** Prussian blue optical images of IONP23 on U373 cells at different concentrations, 25, 50, 75, 100 and 125 µg/mL. Scale bar: 50 µm.

### MRI Contrast Enhancement In Vitro



**Figure S4.** Contrast images and maps of A)  $T_1$ , B)  $T_2$  at an initial concentration of 25  $\mu\text{g/mL}$ , 50  $\mu\text{g/mL}$  and 75  $\mu\text{g/mL}$  for all IONPs. C, left)  $R_1$  (up) and  $R_2$  (down) values for all IONPs at different concentrations. C, right) Values of  $R_1$  (up) and  $R_2$  (down) using 75  $\mu\text{g/mL}$  for all IONPs. D) Contrast and maps of  $T_2^*$  of all IONPs using 25  $\mu\text{g/mL}$ , 50  $\mu\text{g/mL}$  and 75  $\mu\text{g/mL}$  initial concentrations.