

Article

The Biology of Mesopelagic Fishes and Their Catches (1950–2018) by Commercial and Experimental Fisheries

Daniel Pauly ¹, Chiara Piroddi ², Lincoln Hood ³, Nicolas Bailly ¹, Elaine Chu ¹, Vicky Lam ¹,
Evgeny A. Pakhomov ^{4,5}, Leonid K. Pshenichnov ⁶, Vladimir I. Radchenko ⁷ and Maria Lourdes D. Palomares ^{1,*}

¹ *Sea Around Us*, Institute of Oceans and Fisheries, University of British Columbia, Vancouver, BC, V6T 1Z4 Canada; d.pauly@oceans.ubc.ca (D.P.); n.bailly@oceans.ubc.ca (N.B.); e.chu@oceans.ubc.ca (E.C.); v.lam@oceans.ubc.ca (V.L.)

² European Commission, Joint Research Centre, Ispra, 21027 Italy; chiara.piroddi@ec.europa.eu

³ *Sea Around Us* – Indian Ocean, Marine Futures Lab, School of Biological Sciences, University of Western Australia, Crawley, WA, 6012, Australia; lincoln.hood@research.uwa.edu.au

⁴ Earth, Ocean and Atmospheric Sciences Department and the Institute for the Oceans and Fisheries, University of British Columbia, Vancouver, BC, V6T 1Z4, Canada; epakhomov@eoas.ubc.ca

⁵ Hakai Institute, Heriot Bay, BC, V0P 1H0, Canada

⁶ Institute of Fisheries and Ecology of the Sea (IFES) 8, Konsulskaya Str., Berdyansk, 71118, Ukraine; lkpikentnet@gmail.com

⁷ North Pacific Anadromous Fish Commission, Suite 502, 889 West Pender Street, Vancouver, BC V6C 3B2, Canada; vlrad@npafc.org

* Correspondence: m.palomares@oceans.ubc.ca

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Abstract: Following a brief review of their biology, this contribution is an attempt to provide a global overview of the catches of mesopelagic fishes (of which 2.68 million tonnes were officially reported to the FAO) throughout the world ocean from 1950 to 2018, to serve as a baseline to a future development of these fisheries. The overview is based on a thorough scanning of the literature dealing with commercial or experimental fisheries for mesopelagics and their catches, and/or the mesopelagic bycatch of other fisheries. All commercial (industrial and artisanal) fisheries for mesopelagic fishes were included, as well as experimental fisheries of which we were aware, while catches performed only to obtain scientific samples were omitted. The processes of generating bycatch and causing discards are discussed, with emphasis on Russian fisheries. From peer-reviewed and gray literature, we lifted information on mesopelagic fisheries and assembled it into one document (see Online Supplementary Material), which we then summarized into two text tables with catch data, one by country/region, the other by species or species groups.

Keywords: Myctophiformes; reconstructed fisheries catch; *Sea Around Us*; bycatch; discards; growth

Table S1. Species of fish in FishBase belonging to the Myctophiformes (n=254), Neoscolecidae (n=6) and Myctophidae (n=248), and considered to contribute the bulk of mesopelagic fishes. Where available, the depth range (or a single depth of occurrence), maximum recorded length and trophic level are provided (see www.fishbase.org). Note: Lmax is the maximum length in standard length (SL).

No.	Species (Families, Subfamilies)	Depth range (m)	Lmax (SL, cm)	Trophic level
	Family Neoscopelidae	entry 1	data	data
1	<i>Neoscopelus macrolepidotus</i>	300–1180	25.0	4.2
2	<i>Neoscopelus microchir</i>	250–700	30.5	3.2
3	<i>Neoscopelus porosus</i>	454–642	18.3	3.6
4	<i>Scopelengys clarkei</i>	0–1000	--	3.2
5	<i>Scopelengys tristis</i>	400–1830	20.0	3.1
6	<i>Solivomer arenidens</i>	1241–2022	--	3.2
	Family Myctophidae			
	Subfamily Diaphinae			
7	<i>Diaphus adenomus</i>	180–600	18.0	3.2
8	<i>Diaphus aliciae</i>	489	6.0	3.1
9	<i>Diaphus anderseni</i>	100–560	3.2	3.1
10	<i>Diaphus antonbruuni</i>	500	5.5	3.1
11	<i>Diaphus arabicus</i>	0–468	--	3.1
12	<i>Diaphus basileusi</i>	120	16.4	3.2
13	<i>Diaphus bertelseni</i>	0–300	9.1	3.1
14	<i>Diaphus brachycephalus</i>	200–600	6.0	3.1
15	<i>Diaphus burtoni</i>	312	--	3.1
16	<i>Diaphus chrysorhynchus</i>	213–587	1.1	3.0
17	<i>Diaphus coeruleus</i>	457–549	13.7	3.9
18	<i>Diaphus confusus</i>	562	--	3.1
19	<i>Diaphus dahlgreni</i>	320	--	3.1
20	<i>Diaphus danae</i>	350	12.6	3.3
21	<i>Diaphus dehaveni</i>	247	--	3.1
22	<i>Diaphus diadematus</i>	350	4.2	3.1
23	<i>Diaphus diademophilus</i>	0–1808	4.9	3.1
24	<i>Diaphus drachmanni</i>	300	--	3.1
25	<i>Diaphus dumerilii</i>	0–805	8.7	3.0
26	<i>Diaphus effulgens</i>	0–6000	15.0	3.0
27	<i>Diaphus ehrhorni</i>	382	--	3.1
28	<i>Diaphus faustinoi</i>	540	--	3.1
29	<i>Diaphus fragilis</i>	15–1313	12.3	3.1
30	<i>Diaphus fulgens</i>	85–1000	4.5	3.1
31	<i>Diaphus garmani</i>	0–2091	6.0	3.1
32	<i>Diaphus gigas</i>	100–839	--	3.1
33	<i>Diaphus handi</i>	774	--	3.1
34	<i>Diaphus holti</i>	40–777	7.0	3.1
35	<i>Diaphus hudsoni</i>	0–840	8.4	3.3
36	<i>Diaphus impostor</i>	0–140	--	3.1
37	<i>Diaphus jenseni</i>	350–1389	5.0	3.1
38	<i>Diaphus kapalae</i>	0–290	--	3.1
39	<i>Diaphus knappi</i>	122–664	17.3	3.2
40	<i>Diaphus kora</i>	0–387	--	3.1
41	<i>Diaphus kuroshio</i>	100–1537	6.3	3.1
42	<i>Diaphus lobatus</i>	--	--	3.1
43	<i>Diaphus lucidus</i>	0–2999	11.8	3.0
44	<i>Diaphus lucifrons</i>	564	--	3.1
45	<i>Diaphus luetkeni</i>	40–750	6.0	3.8
46	<i>Diaphus malayanus</i>	1000–2000	4.5	3.1

No.	Species (Families, Subfamilies)	Depth range (m)	Lmax (SL, cm)	Trophic level
47	<i>Diaphus mascarensis</i>	237–800	14.4	3.2
48	<i>Diaphus meadi</i>	250	5.4	3.0
49	<i>Diaphus megalops</i>	1–528	8.5	3.1
50	<i>Diaphus metopoclampus</i>	90–1085	7.5	3.3
51	<i>Diaphus minax</i>	476	--	3.1
52	<i>Diaphus mollis</i>	50–600	6.6	3.0
53	<i>Diaphus nielseni</i>	--	4.0	3.1
54	<i>Diaphus ostentfeldi</i>	350	12.0	3.2
55	<i>Diaphus pacificus</i>	--	--	3.1
56	<i>Diaphus pallidus</i>	310	--	3.1
57	<i>Diaphus parini</i>	320	--	3.1
58	<i>Diaphus parri</i>	350–1071	6.5	3.1
59	<i>Diaphus perspicillatus</i>	0–1500	7.1	3.1
60	<i>Diaphus phillipsi</i>	588–1330	7.7	3.1
61	<i>Diaphus problematicus</i>	40–820	10.5	3.0
62	<i>Diaphus rafinesquii</i>	40–2173	9.0	3.4
63	<i>Diaphus regani</i>	750	1.4	3.0
64	<i>Diaphus richardsoni</i>	350–1000	6.0	3.1
65	<i>Diaphus rivatoni</i>	0–152	9.0	3.1
66	<i>Diaphus roei</i>	558	--	3.1
67	<i>Diaphus sagamiensis</i>	549	--	3.1
68	<i>Diaphus schmidtii</i>	100–1400	5.3	3.2
69	<i>Diaphus signatus</i>	1270	4.0	3.1
70	<i>Diaphus similis</i>	0–631	7.2	3.1
71	<i>Diaphus splendidus</i>	0–8000	9.0	3.0
72	<i>Diaphus suborbitalis</i>	387–1537	7.3	3.1
73	<i>Diaphus subtilis</i>	40–750	8.5	3.1
74	<i>Diaphus taaningi</i>	40–475	7.0	3.3
75	<i>Diaphus termophilus</i>	40–850	8.0	3.1
76	<i>Diaphus theta</i>	10–3400	9.3	3.2
77	<i>Diaphus thiollierei</i>	--	10.0	3.3
78	<i>Diaphus trachops</i>	100–686	6.3	3.1
79	<i>Diaphus umbroculus</i>	311	--	3.1
80	<i>Diaphus vanhoeffeni</i>	40–750	4.2	3.1
81	<i>Diaphus watasei</i>	100–2005	17.0	3.2
82	<i>Diaphus whitleyi</i>	311	--	3.1
83	<i>Diaphus wisneri</i>	50–375	--	3.1
84	<i>Idiolychnus urolampus</i>	124–582	11.0	3.2
85	<i>Lobianchia dofleini</i>	0–4000	5.0	3.0
86	<i>Lobianchia gemellarii</i>	25–800	6.0	3.0
Subfamily Gymnoscopelinae				
87	<i>Gymnoscopelus bolini</i>	4200	28.0	3.3
88	<i>Gymnoscopelus braueri</i>	2700	13.2	3.2
89	<i>Gymnoscopelus fraseri</i>	50–250	8.8	3.2
90	<i>Gymnoscopelus hintonoides</i>	2200–2350	14.0	3.2
91	<i>Gymnoscopelus microlampas</i>	200–500	11.7	3.0
92	<i>Gymnoscopelus nicholsi</i>	300	15.7	3.4
93	<i>Gymnoscopelus opisthopterus</i>	550–900	16.2	3.3
94	<i>Gymnoscopelus piabilis</i>	--	14.6	3.2

No.	Species (Families, Subfamilies)	Depth range (m)	Lmax (SL, cm)	Trophic level
95	<i>Hintonia candens</i>	--	13.0	3.2
96	<i>Lampanyctodes hectoris</i>	--	7.0	3.2
97	<i>Lampichthys procerus</i>	1–2000	8.2	3.1
98	<i>Notoscopelus bolini</i>	1–1300	10.2	3.1
99	<i>Notoscopelus caudispinosus</i>	1–360	14.0	3.2
100	<i>Notoscopelus elongatus</i>	45–1000	14.2	3.4
101	<i>Notoscopelus japonicus</i>	391–794	13.3	3.2
102	<i>Notoscopelus kroyeri</i>	0–1000	14.3	3.2
103	<i>Notoscopelus resplendens</i>	777–2121	9.5	3.0
104	<i>Scopelopsis multipunctatus</i>	3–2000	8.1	3.0
Subfamily Lampanyctinae				
105	<i>Bolinichthys distofax</i>	100–690	9.0	3.1
106	<i>Bolinichthys indicus</i>	25–900	4.5	3.1
107	<i>Bolinichthys longipes</i>	50–1021	5.0	3.1
108	<i>Bolinichthys nikolayi</i>	25–1760	4.1	3.0
109	<i>Bolinichthys photothorax</i>	40–750	7.3	3.0
110	<i>Bolinichthys pyrsobolus</i>	60–778	9.2	3.1
111	<i>Bolinichthys supralateralis</i>	40–850	11.7	3.1
112	<i>Ceratoscopelus maderensis</i>	51–1480	8.1	3.3
113	<i>Ceratoscopelus townsendi</i>	100–500	15.1	3.5
114	<i>Ceratoscopelus warmingii</i>	391–2056	8.1	3.4
115	<i>Lampadena anomala</i>	330–2000	18.0	3.2
116	<i>Lampadena atlantica</i>	60–1000	20.0	3.2
117	<i>Lampadena chavesi</i>	40–800	8.0	3.1
118	<i>Lampadena dea</i>	1500–2390	8.9	3.1
119	<i>Lampadena luminosa</i>	50–1021	20.0	3.2
120	<i>Lampadena notialis</i>	1–800	13.9	3.2
121	<i>Lampadena pontifex</i>	1–750	11.0	3.1
122	<i>Lampadena speculigera</i>	1–1000	15.3	3.2
123	<i>Lampadena urophaos</i>	50–1000	20.0	3.2
124	<i>Lampadena yaquinae</i>	100–2056	13.0	3.2
125	<i>Lampanyctus acanthurus</i>	930–1537	13.0	3.3
126	<i>Lampanyctus achirus</i>	--	16.2	3.2
127	<i>Lampanyctus alatus</i>	40–1500	6.1	3.2
128	<i>Lampanyctus ater</i>	60–1100	14.0	3.2
129	<i>Lampanyctus australis</i>	--	13.1	3.3
130	<i>Lampanyctus bristori</i>	--	14.2	3.2
131	<i>Lampanyctus crocodilus</i>	1–1200	30.0	3.2
132	<i>Lampanyctus crypticus</i>	--	9.8	3.2
133	<i>Lampanyctus cuprarius</i>	40–1000	7.9	3.3
134	<i>Lampanyctus fernae</i>	1–750	9.1	3.2
135	<i>Lampanyctus festivus</i>	40–1052	13.8	3.3
136	<i>Lampanyctus gibbsi</i>	--	12.2	3.2
137	<i>Lampanyctus hawaiiensis</i>	300–850	8.1	3.1
138	<i>Lampanyctus hubbsi</i>	1–2500	3.0	3.1
139	<i>Lampanyctus idostigma</i>	100–500	9.6	3.2
140	<i>Lampanyctus indicus</i>	--	8.0	3.1
141	<i>Lampanyctus intricarius</i>	40–750	20.0	3.4
142	<i>Lampanyctus isaacsi</i>	0–2300	13.3	3.2

No.	Species (Families, Subfamilies)	Depth range (m)	Lmax (SL, cm)	Trophic level
143	<i>Lampanyctus iselinoides</i>	64	--	3.2
144	<i>Lampanyctus jordani</i>	588–3400	14.0	3.3
145	<i>Lampanyctus lepidolychnus</i>	312–332	11.9	3.2
146	<i>Lampanyctus lineatus</i>	60–1150	23.7	3.0
147	<i>Lampanyctus macdonaldi</i>	60–1464	16.0	3.1
148	<i>Lampanyctus macropterus</i>	0–2091	6.8	3.2
149	<i>Lampanyctus niger</i>	100–1015	11.1	3.1
150	<i>Lampanyctus nobilis</i>	100–1000	12.4	3.1
151	<i>Lampanyctus omostigma</i>	3000	2.6	3.1
152	<i>Lampanyctus parvicauda</i>	100–500	--	3.2
153	<i>Lampanyctus photonotus</i>	40–1100	8.5	3.2
154	<i>Lampanyctus phyllisae</i>	--	15.1	3.2
155	<i>Lampanyctus pusillus</i>	40–850	4.3	3.4
156	<i>Lampanyctus regalis</i>	772–3400	17.2	3.2
157	<i>Lampanyctus ritteri</i>	20–1095	12.0	3.4
158	<i>Lampanyctus simulator</i>	0–500	9.3	3.2
159	<i>Lampanyctus steinbecki</i>	80–100	3.8	3.1
160	<i>Lampanyctus tenuiformis</i>	1537	15.3	3.3
161	<i>Lampanyctus turneri</i>	1757	7.0	3.2
162	<i>Lampanyctus vadulus</i>	0–370	9.9	3.2
163	<i>Lampanyctus wisneri</i>	600–650	8.8	3.1
164	<i>Lepidophanes gaussi</i>	0–850	5.0	3.1
165	<i>Lepidophanes guentheri</i>	40–750	7.8	3.0
166	<i>Parvilux boschmai</i>	--	--	3.2
167	<i>Parvilux ingens</i>	100–500	16.4	3.1
168	<i>Stenobranchius leucopsarus</i>	31–3400	10.7	3.2
169	<i>Stenobranchius nannochir</i>	441–3400	11.0	3.0
170	<i>Taaningichthys bathyphilus</i>	400–1550	8.0	3.1
171	<i>Taaningichthys minimus</i>	90–800	6.5	3.1
172	<i>Taaningichthys paurolychnus</i>	900–2000	9.5	3.2
173	<i>Triphoturus mexicanus</i>	25	5.7	3.3
174	<i>Triphoturus nigrescens</i>	100–1000	8.1	3.1
175	<i>Triphoturus oculeum</i>	770–3243	--	3.2
Subfamily Myctophinae				
176	<i>Benthosema fibulatum</i>	1–2000	8.0	3.2
177	<i>Benthosema glaciale</i>	1–1407	10.3	3.1
178	<i>Benthosema panamense</i>	--	4.5	3.1
179	<i>Benthosema pterotum</i>	10–300	5.7	3.1
180	<i>Benthosema suborbitale</i>	50–2500	3.9	3.4
181	<i>Centrobranchus andreae</i>	650	6.5	3.4
182	<i>Centrobranchus brevirostris</i>	--	4.0	3.3
183	<i>C. choerocephalus</i>	1050	4.0	3.3
184	<i>Centrobranchus nigroocellatus</i>	1–700	5.0	3.4
185	<i>Ctenoscopelus phengodes</i>	--	9.3	3.4
186	<i>Dasyscopelus asper</i>	244–1948	6.5	3.7
187	<i>Dasyscopelus obtusirostris</i>	1–700	7.8	3.4
188	<i>Dasyscopelus selenops</i>	40–500	6.4	3.3
189	<i>Dasyscopelus spinosus</i>	1–700	9.0	3.5
190	<i>Diogenichthys atlanticus</i>	1–1050	2.9	3.1

No.	Species (Families, Subfamilies)	Depth range (m)	Lmax (SL, cm)	Trophic level
191	<i>Diogenichthys laternatus</i>	1–2091	4.0	3.2
192	<i>Diogenichthys panurgus</i>	366	2.3	3.1
193	<i>Electrona antarctica</i>	1–1010	11.5	3.2
194	<i>Electrona carlsbergi</i>	1–1008	11.2	3.3
195	<i>Electrona paucirastra</i>	--	7.0	3.3
196	<i>Electrona risso</i>	90–1485	8.2	3.4
197	<i>Electrona subaspera</i>	--	12.7	3.3
198	<i>Gonichthys barnesi</i>	1–1000	5.0	3.2
199	<i>Gonichthys cocco</i>	1–1450	6.0	3.2
200	<i>Gonichthys tenuiculus</i>	--	4.1	3.2
201	<i>Gonichthys venetus</i>	--	--	3.2
202	<i>Hygophum atratum</i>	600–3132	4.9	3.2
203	<i>Hygophum benoiti</i>	51–700	5.5	3.0
204	<i>Hygophum bruuni</i>	--	--	3.2
205	<i>Hygophum hanseni</i>	57–728	6.7	3.2
206	<i>Hygophum hygomii</i>	1–1485	6.8	3.0
207	<i>Hygophum macrochir</i>	1–750	6.0	3.2
208	<i>Hygophum proximum</i>	1–1000	5.0	3.2
209	<i>Hygophum reinhardtii</i>	1–1050	6.0	3.2
210	<i>Hygophum taaningi</i>	250–1000	6.1	3.2
211	<i>Krefflichthys anderssoni</i>	2700	7.1	3.1
212	<i>Loweina interrupta</i>	60–800	3.9	3.2
213	<i>Loweina rara</i>	1–1050	4.5	3.2
214	<i>Loweina terminata</i>	1–825	3.0	3.1
215	<i>Metelectrona ahlstromi</i>	1–2000	--	3.3
216	<i>Metelectrona herwigi</i>	98	5.5	3.2
217	<i>Metelectrona ventralis</i>	0–426	10.7	3.3
218	<i>Myctophum affine</i>	0–600	7.9	3.0
219	<i>Myctophum aurolaternatum</i>	--	11.0	3.5
220	<i>Myctophum brachygnathum</i>	--	--	3.4
221	<i>Myctophum fissunovi</i>	--	7.0	3.4
222	<i>Myctophum indicum</i>	--	--	3.4
223	<i>Myctophum lunatum</i>	--	5.7	3.3
224	<i>Myctophum lychnobium</i>	1–1000	3.8	3.2
225	<i>Myctophum nitidulum</i>	412–1537	8.3	3.4
226	<i>Myctophum orientale</i>	--	--	3.4
227	<i>Myctophum ovcharovi</i>	40–90	7.2	3.4
228	<i>Myctophum punctatum</i>	1–1000	11.0	3.4
229	<i>Protomyctophum andriashevi</i>	50–332	6.0	3.4
230	<i>Protomyctophum arcticum</i>	90–1600	6.0	3.1
231	<i>Protomyctophum beckeri</i>	1–2100	3.5	3.2
232	<i>Protomyctophum bolini</i>	364–728	6.7	3.0
233	<i>Protomyctophum chilense</i>	1–400	--	3.3
234	<i>Protomyctophum choriodon</i>	--	9.5	4.2
235	<i>Protomyctophum crockeri</i>	100–500	3.7	3.2
236	<i>Protomyctophum gemmatum</i>	2000	8.6	3.4
237	<i>Protomyctophum luciferum</i>	2000	6.1	3.5
238	<i>Protomyctophum mcginnisi</i>	--	--	3.3
239	<i>Protomyctophum normani</i>	--	5.6	3.3

No.	Species (Families, Subfamilies)	Depth range (m)	Lmax (SL, cm)	Trophic level
240	<i>Protomyctophum parallelum</i>	2500	5.0	3.3
241	<i>P. subparallelum</i>	350	3.6	3.2
242	<i>Protomyctophum tenisoni</i>	96	5.4	3.3
243	<i>Protomyctophum thompsoni</i>	785–1500	5.2	3.3
244	<i>Symbolophorus barnardi</i>	100–800	11.6	3.1
245	<i>Symbolophorus boops</i>	0–500	13.1	3.5
246	<i>Symbolophorus californiensis</i>	557–1497	11.0	3.1
247	<i>Symbolophorus evermanni</i>	100–500	8.0	3.4
248	<i>Symbolophorus krefftii</i>	1–150	11.2	3.2
249	<i>Symbolophorus reversus</i>	--	8.9	3.2
250	<i>Symbolophorus rufinus</i>	0–850	9.4	3.2
251	<i>Symbolophorus veranyi</i>	0–800	12.0	3.3
252	<i>Tarletonbeania crenularis</i>	0–710	10.4	3.1
253	<i>Tarletonbeania taylori</i>	0–1500	7.0	3.3
Subfamily Notolychninae				
254	<i>Notolychnus valdiviae</i>	25–700	5.2	3.1

Quotes. The following consists of quotes with diverse information of mesopelagic fisheries and their catches.

“During the late 1970s and early 1980s, the severe depletion of demersal fish stocks (most notably *Nothotenia rossii*) was followed in the second half of the latter decade by harvesting of benthopelagic species such as toothfish species with variable year class strengths (*C. gunnari*) and mesopelagic species such as *E. carlsbergi*.[...] Economic considerations effectively ended the *E. carlsbergi* fishery at the end of the 1991/92 season [1], while other fishing grounds, such as the Ob and Lena Seamounts, were effectively closed from the mid-1990s onwards” [2].

“Recently a fortuitous fishery for the lanternfish *Lampanyctodes hectoris* has developed incidental to the anchovy/pilchard fishery off the western coast of South Africa [3]. Annual landings of lanternfishes (mostly *L. hectoris*) were 1,134 metric tons or 0.3 percent of the pelagic fishery catch in this region in 1969 and increased to 42,560 metric tons or 10.45 percent of the catch in 1973” [4].

“There are reports of fishery for mesopelagics especially myctophids, the most well-known is the purse seine fishery for *Lampanyctodes hectoris* off South Africa [5] and also in erstwhile USSR where they fish off West Africa and off Southern Australia. Due to its high lipid (wax esters) content most of the myctophids are unpalatable for consumption and is used for the production of fish meal, fish oil and fish silage. But some species (*Diaphus coeruleus* and *Gymnoscopelus nicholski*) have been fished for human consumption [6,7]. During the 70’s *Gymnoscopelus bolini* and *G. nicholski*, caught as bycatch in the Antarctic marbled rock cod fishery has been smoked for human consumption. In India, however there have been no reports of a myctophid fishery and its use for human consumption” [8].

“Commercial lanternfish fisheries include limited operations off South Africa, in the sub-Antarctic and in the Gulf of Oman [9–12]. But majority of the myctophids are not used for direct human consumption owing to their high lipid or wax ester content, therefore they are used as predator fish feed, poultry feed, animal feed and crop fertilizers [8,13,14]. Exceptions to this are *Diaphus coeruleus*, *Gymnoscopelus nicholski* and *G. bolini* which were considered edible in the Southwest Indian Ocean and Southern Atlantic in the late 1970s [8,15–17]. There are no reports of human consumption of myctophids in India [8,17]. Lekshmy et al. [13] have carried out various methods for processing and utilization of *Benthosema pterotum*. They have also carried out nutritional evaluation of fish meal, dry fish and fish hydrolysate using casein protein as reference on rats for palatability. However, one cannot ignore the processing difficulties on a large scale. An industrial fishery for

Lampanyctodes hectoris in South African waters closed in the mid-1980s due to processing difficulties caused by the high oil content of the fish [17]. Interestingly, in eastern South Atlantic, this particular species accounted for around 42,560 tonnes (10.45%) of pelagic catch in 1973 [16] [18].

“A single haul off Argentina yielded 30 tonnes (33 tons) of *Diaphus dumerilii* in one hour. [...]. Limited commercial exploitation occurs off South Africa, where annual purse seine landings (mainly of *Lampanyctodes hectoris*) have fluctuated between 100 and 42,400 tonnes (110 to 46,725 tons). The lanternfishes are reduced to fish meal and fish oil. Because of lanternfishes' high oil content, processing plants are forced to mix them with other species to prevent clogging the machinery. Around South Georgia and Shag rocks, experimental fishing on *Electrona carlsbergi* (mainly juveniles) averaged about 20,000 tonnes (22,000 tons) per year between 1988 and 1990, but increased dramatically to 78,488 tonnes (86,494 tons) in 1991. The Commission for the Conservation of Antarctic Marine Living Resources therefore introduced a 20,000 tonne (220,400 ton) TAC (total allowable catch) for the species for the 1992 season” [19].

“During 1989–1990, 8 cruises were carried out using this vessel in the region, not only for trial fishing but also for estimating the biomass of lantern fish (myctophids) resources” [20].

“According to [21], fishermen in Suruga Bay who eat large quantities of *Diaphus* spp. sort out and discard *B. pterotum* as inedible. That does not mean that this huge production is useless; fish oil and protein have other uses than direct human consumption. Studies in India [9,14] show that meal and hydrolysate from *B. pterotum* are excellent protein supplements in fish and poultry feeds. These myctophids are readily fished; Norwegian results reached 100 tons hr⁻¹ with a sonar-guided, 750 m² (15 × 50 m) double warp trawl (which is a seriously large piece of gear)” [22].

“Pearlside fishery of 2009 landed more than 46,000t; landing in 2010 was 18,000t and decreased until 2013–2016 had 0 landings despite some trials” [23].

“The target species of the fishery are or have been marbled notothenia (*Notothenia rossii*), mackerel icefish (*Champscephalus gunnari*), grey notothenia (*Lepidonotothen* (= *Notothenia*) *squamifrons*), Günther's notothenia (*Patagonotothen guntheri*), sub-Antarctic lanternfish (indiscriminately recorded as *Electrona carlsbergi*) and Patagonian toothfish (*Dissostichus eleginoides*). [...] Owing to their small size Gunther's notothenia and lanternfish have been used for fish meal, while the other species have been fished primarily for direct human consumption [24]. [...] After the successive depletion of the demersal fish stocks, harvesting of (benthopelagic) Patagonian toothfish and (pelagic) sub-Antarctic lanternfish started in the second half of the 1980s [...] Economical considerations prompted the cessation of the fishery on lanternfish after the 1991/92 season. [...] The stock of sub-Antarctic lanternfish has yet to be properly assessed following a tentative assessment in 1991 [...], although a substantial fishery with annual catches of several tens of thousand tonnes has been conducted on the stock for a number of years” [25].

“After most of the demersal (bottom-dwelling) fish stocks were depleted, which happened before CCAMLR came into force, benthopelagic (living off the bottom) Patagonian toothfish and mesopelagic (living in oceanic midwater) sub-Antarctic lanternfish began to be harvested in the second half of the 1980s [...]. By the end of the 1980s, fishing for most species was either prohibited, as in the case of the marbled rockcod, or was limited by total allowable catches (TACs). [...] Economic considerations prompted the cessation of the fishery for lanternfish after the 1991/92 season. [...] The Soviet Union began a trawl fishery for lanternfish (reported indiscriminately as *E. carlsbergi*) in the Antarctic Polar Front in the 1980s, with annual catches initially varying between 500 and 2,500 tonnes. Catches increased from 1987/88 by 14,000 to 23,000–29,000 tonnes in the two subsequent seasons, and peaked in 1990/91 (78,000 tonnes) and 1991/92 (51,000 tonnes) [...]. The fishery lapsed in the 1992/93 season, as it was no longer considered to be economically viable” [26].

“Iceland has in the last few years collected information on mesopelagic fish in the Irminger Sea during their investigations on redfish and have also done some exploratory fishing trials. In Faroese waters Russian trawlers fishing for blue whiting have occasionally reported significant by-catches of mesopelagic fish, and the Faroese Fisheries Laboratory and the Marine Research Institute in Iceland have done some exploratory fishing, but so far without any success” [27].

“Since 2002, the Federation of Vessel owners, in cooperation with the Marine Research Institute in Reykjavík have conducted several experimental cruises. So far, none of the trials have resulted in commercially exploitable catches. The experiments were performed along the Reykjanes Ridge with commercial vessels, using a Gloria #1280 type trawl. Modifications was made on the belly part and the cod end had 9 mm mesh size. In summary there were low catch rate in all hauls, but also low acoustic recordings during the surveys, according to the fishermen. Highest catch rate during these experiments was 3 t/h of *Maurolicus muelleri*” [27].

“In the Gulf of Oman, the only myctophid present is *Benthosema pterotum* and Iranian fishers have started a commercial fishery for myctophids in their part of the Gulf of Oman [28].

“In spite of its abundance in world oceans, currently only a few commercial myctophid fisheries exist, which include limited operations off South Africa, in the sub- Antarctic, and in the Gulf of Oman [5,19,29]. Global catch of myctophids during 1970–2010, varied between a few tonnes to a maximum of 42,400 t reported during 1973 [30]. Though not commercially exploited in India, these resources have been reported as bycatch of deep-sea shrimp trawlers operating from southwest coast of India [31–33]. [It was reported that] the annual catch of myctophids during 2010–11 was 2972 t and the catch was supported mainly by five species viz., *Diaphus watasei*, *D. garmani*, *Benthosema fibulatum*, *Myctophum obtusirostre* and *Neoscopilus microchir*. Boopendranath et al. [34] reported the annual catch of myctophids, caught as bycatch in the deep-sea shrimp trawlers operating off southwest coast of India, as 3676 t, with a catch rate of 19.87 kg h⁻¹” [35].

“Myctophids are fairly abundant in Philippine waters, but are rarely caught by fishermen except when they are attracted by light at night in the open seas” [36].

“Myctophids form bycatch in deep sea shrimp trawls with an annual average catch of 2668 t during 2009–2011 in Kerala coast. Fishery occurred almost round the year with peak during November - February. [...] Along the south-west coast of India, lantern fish (Order Myctophiformes) forms a major portion (20–35%) of the bycatch in the deep-sea shrimp trawls [37]. These fishes, when landed are mostly used for fishmeal or manure production” [33].

“Fishermen in Suruga Bay, Central Japan used *Diaphus* spp. as food [21]. Commercial fishery for *Diaphus coeruleus* and *Gymnoscopelus nicholski* (edible species) in the south-west Indian Ocean and southern Atlantic began in 1977 and catch by former USSR countries reached 51,680 t in 1992, after which the fishery ceased due to decrease in catch. Despite this, the Commission for Conservation of Antarctic Marine Living Resources (CCAMLR) still permits Total Allowable Catch (TAC) of 200,000 t for this resource from the area under its jurisdiction. Industrial purse seine fishery for *Lampanyctodes hectoris* was developed in South African waters and closed in the mid-1980s due to processing difficulties caused by the high oil content in the fish [17]. Lanternfishes are harvested commercially only off South Africa and in the sub-Antarctic [19,38] [...] Catch comprised of five species viz., *Diaphus watasei* (74.23%), *Neoscopilus microchir* (20.57%), *Benthosema fibulatum* (1.94%), *Diaphus garmani* (1.69%) and *Myctophum obtusirostre* (1.58%) [...] *D. watasei* and *N. microchir* were available round the year whereas, other species occurred only seasonally. *D. watasei* was found to be dominant among the myctophids” [33].

“After a long period of high expectations, a commercial fishery for these mesopelagic fishes was initiated in the Persian side of the Oman Sea” [39].

“The federal government has prepared a draft Deep-Sea Fishing Policy for issuance of 50 Licenses for Tuna long Liners, Squid Jigger, Mesopelagic fishing to foreign flagged vessels and 6000 licenses to local fishing vessels” [40].

“Management measures: (1) TAC combined for lantern and lightfish: 50,000 t; (2) Minimum mesh size of 28 mm; (3) Sardine bycatch limitation (anchovy-directed operations); (4) Closed season from 1 November to 14 January; (5) ‘Landings monitored and estimated at factory landing sites’ [41].

“During commercial fishing trials in 1995–1998, using a pelagic trawl with cod-end mesh size of 10 mm, the average catch was between 24 and 28 t day⁻¹ in Iranian waters [...]. During trial commercial fishing in Oman waters in 1996, total monthly catches of myctophids for the months of March, April and May were 446, 1563 and 1273 t, respectively. Over 123 fishing days this gave an average catch of 20 t day⁻¹. However, catches declined during early summer and the trial was therefore discontinued” [42].

“[A] fishery for two species of myctophids which are considered edible viz., *Diaphus coeruleus* and *Gymnoscopelus nicholski* existed in the Southwest Indian Ocean and Southern Atlantic during 1977–1992 and catches up to 51,680 t has been reported in 1992. Shotton [43] has reported regarding an industrial purse seine fishery for *Lampanyctodes hectoris* in South African waters which was closed in the mid-1980s due to processing difficulties caused by the high oil content of the fish. Qeshm Fish Process Company in Iran produces fish meal and oil, mainly based on lantern fish and the plant has a nominal capacity of 3,600 tons of lanternfish per day, out of which approximately 700 tons of fish meal and 70 tons of fish oil are obtained (QFPCO 2011)” [44].

“Special attention should be paid here to numerous species from the group of Myctophidae, pelagic Gobidae and other snail-sized fish (below 10 cm in length) forming dense shoals identified as sound scattering layers. The exploitation of their stocks was begun by the Republic of South Africa (Divisions 1.4 and 1.6) when 11- and 12.7-mm mesh purse seines were introduced, although these fish inhabit the whole ICSEAF Area. At first their catches were quite substantial, equaling, for instance, 42,000 tons (mostly *L. hectoris*) for Division 1.6 in 1973. Between 1978 and 1983, the catches considerably, not exceeding 1,000 tons, with the exception of 1979 and 1981, when 10,000 tons were taken [5] (Newman, 1977)” [45].

“*Lampanyctodes hectoris* have accounted for 0.3–10.45% (1134–42,560 metric tons) of the total fish landed by South African pelagic fishing boats operating in the cold water off the west coast of south Africa during the years 1969–1973 [3]. Approximately 15 tons of another species, *Diaphus dumerilii*, were taken in a single haul at a depth of 260–265 m off Uruguay [46]” [38].

“Myctophids have been targeted by commercial fisheries in the Southern Ocean, notably in the northern Scotia Sea area where ex-Soviet Union vessels targeted *Electrona carlsbergi* at or just south of the Polar Front to the north of South Georgia [47]. Catches peaked at around 30,000 tonnes in the 1988/89 season, with the fish converted to meal, but since 1990 there has not been a targeted fishery” [48].

“An annual PUCL for mesopelagic fish of 50,000 t was introduced in 2012, following increased catches of lantern- and light fish by the experimental pelagic trawl fishery in 2011, when just over 7000 t of these species were landed. Since then, however, catches have not exceeded 1000 t. It is anticipated that catches of mesopelagic fish may again increase in 2014 with resumption of this experiment” [49].

“While under limited commercial exploitation in the southern Benguela, the mesopelagic catch has historically fluctuated between 100 and 42,400 tonnes and has accounted for some 10% of the total annual catch made by South Africa’s small pelagic fishery in some years [...] However, the fishery intermittently closed during mid-80s due to processing difficulties caused by the high wax ester content of the fish [...]. In addition to the commercial purse-seine fishery, DAFF granted two-year permits in 2010 for an experimental mid-water trawl fishery targeting mesopelagic and pelagic stocks. Of the total

catch reported for both years combined (9486.5 tonnes), 83% consisted of *L. hectoris* and 4% of *M. walvisensis*" [50].

"Some Icelandic companies are developing the maurolic fishery (*Maurolicus muelleri*) in areas south of Iceland. While he is not always successful, at the end of January, there were several successful days before main concentrations of maurolic migrated to the west.

According to the information of the First Officer and skipper of the "Faxi RE" trawler, the fishery began in the area of the Grindavík Deep, then moved south of the Eldey area. All three HB Grandi trawlers were fishing. The catches were 70 to 80 tons for long trawls. In the same area there were 12 other vessels of other companies. The "Faxi RE" used a midwater trawl with a small-mesh insert. But it seems that for a more successful harvest, a smaller mesh trawl and additional knowledge will be required. The fish is small enough. The optimal time for catching it is daytime only." [51].

References

1. Miller, D.G.M. The Southern Ocean: A global view. *Ocean Yearbook*, 2000, 14, 468–513.
2. Miller, D.G.M. Managing fishing in the sub-Antarctic. *Papers and Proceedings of the Royal Society of Tasmania*, 2007, 141(1), 121–140.
3. Centurion-Harris, O.M. The appearance of lantern-fish in commercial catches. *South African Shippings News and Fishing Industry Review*, 1974, 29, 44–45.
4. Ahlstrom, E.H.; Moser, H.G.; O'Toole, M.J. Development and distribution of larvae and early juveniles of the commercial lanternfish, *Lampanyctodes hectoris* (Günther), off the west coast of southern Africa with a discussion of phylogenetic relationships of the genus. *Bulletin Southern California Academy of Sciences*, 1976, 75, 138–152.
5. Newman, G. The living marine resources of the Southeast Atlantic. *FAO Fisheries Technical Paper 178*, FAO, Rome, 1977; 59 p.
6. Melnikova, O.M. Teknokhimisk karakteristik av *Diaphus coeruleus*. *Rybnoye Khozyaistvo*, 1973, Mosk. (5), 72.
7. Dubrovskaya, T.A.; Makarov, O.E. Chemical characteristics of some Antarctic fishes. *Trudy VNIRO*, 1969, 66, 311–317.
8. Balu, S.; Menon, N.G. Lantern fish-potential deep-sea resource. *Marine Ecosystem, ENVIS*, 2006, 5(1), 3–5.
9. Gopakumar, K.; Nair, R.K.G.; Nair, V.P.G.; Nair, L.A.; Radhakrishnan, A.G.; Nair, R.P. Studies on lantern fish (*Benthosema pterotum*) I. Biochemical and microbiological investigations. *Fishing Technology*, 1983, 20(1), 17–19.
10. Gjøsæter, J. Mesopelagic fish, a large potential resource in the Arabian Sea. *Deep-Sea Research*, 1984, 17, 215–251.
11. Moser, H.G.; Watson, W. Myctophiformes. In *Early stages of Atlantic fishes—an identification guide for the Western Central North Atlantic*, Richards, W.J. (ed); 1st Edition, Inc. Net Library, 2006; pp 461–580.
12. Valinassab, T.; Pierce, G.J.; Johannesson, K. Lantern fish (*Benthosema pterotum*) resources as a target for commercial exploitation in the Oman Sea. *Journal of Applied Ichthyology*, 2007, 23, 573–577.
13. Lekshmy, N.A.; Arul, J.M.; Mathew, P.T.; Gopakumar, K. Studies on lantern fish (*Benthosema pterotum*) II. Nutritional evaluation. *Fishing Technology*, 1983, 20(1), 17.
14. Nair, L.A.; Arul, J.M.; Mathew, P.T.; Gopakumar, K. Studies on lanternfish (*Benthosema pterotum*) II. Nutritional evaluation. *Fishing Technology*, 1983, 20(1), 20–23.
15. Nafpaktitis, B.G. Myctophidae. In: *FAO Species Identification Sheets for Fishery Purposes: Western Indian Ocean*, Fischer, W.; Bianchi, G. (eds); 3rd edition, FAO, Rome, 1982; p 1–8.
16. Hulley, P.A. Lanternfishes-Myctophidae. In *FAO species identification sheets for fishery purposes. Southern Ocean (Fishing areas 48, 58 and 88)*, Fischer W, Hureau JC (eds); FAO, Rome, 1985; Vol. 2, 316–322.
17. FAO. Review of the state of world fishery resources: Marine Fisheries. *FAO Fisheries Circular No. 920 FIRM/C920* ISSN 0429-9329, 1997; PAGES??.
18. Catul, V.; Gauns, M.; Karuppasamy, P.K. A review on mesopelagic fishes belonging to family Myctophidae. *Reviews in Fish Biology and Fisheries*, 2011, 21, 339–254.
19. Hulley, P.A. Lanternfishes. In *Encyclopedia of Fishes*, Paxton, J.R.; Eschmeyer, W.N. (eds.); Academic Press Inc., San Diego, CA, 1994; pp. 127–128.
20. Ziaee Shilat, A.; Valinassab, T. Trial fishing for lantern fishes (myctophids) in the Gulf of Oman (1989–1990). *FAO Fisheries Circular No. 935*, FAO, Rome, 1998; 76 p.
21. Kubota, T. Food of lanternfishes in Suruga Bay, central Japan. In *Proceedings of the North Pacific Aquaculture Symposium*, Anchorage, Alaska, August 18–21 1980 and Newport, Oregon August 25–27 1980; University of Alaska Sea Grant Report, 1982, 82–2, 275–283.
22. GLOBEC US. Global Ocean Ecosystems Dynamics: A Component of the U.S. Global Change Research Program, University of California, Davis, California; Report No. 9; 1993; 93 p.
23. Sigurdsson, T. The "big apple"? Mesopelagic fish: The Icelandic case. PowerPoint presentation. Marine and Freshwater Research Institute. Bergen, 2017.
24. Kock, K.H. *Antarctic fish and fisheries*. Cambridge University Press; 1992; 307 p.
25. Kock, K.H. Fishing and conservation in southern waters Karl-Hermann Kock. *Polar Record*, 1994, 30(172), 3–22.

26. Kock K.-H. Understanding CCAMLR's Approach to Management. CCAMLR, 2000; 63 p.
27. Lamhauge, S.; Jacobsen, J.A.; Jakupsstovu, H.I.; Valdemarsen, J.W.; Sigurdsson, T.; Bardarsson, B.; Filin, A. Fishery and utilisation of mesopelagic fishes and krill in the Northeast Atlantic. 526, TemaNord, Denmark, 2008.
28. Shahid, A. Personal communication, 2003.
29. Gjøsæter, J.; Kawaguchi, K. A review of the world resources of mesopelagic fish. *FAO Fisheries Technical Paper* 193, 1980; 151 p.
30. FAO. The state of world fisheries and aquaculture (SOFIA). FAO, Rome, 2012; <http://www.fao.org/docrep/016/i2727e/i2727e00.htm>.
31. Boopendranath, M.R.; Remesan, M.P.; Jose Fernandez, T.; Pradeep, K.; Vipin, P.M.; Ravi, R. Myctophids in the bycatch of deep-sea shrimp trawlers. *Fish Technology Newsletter*, 2009, 20(2), 1-2.
32. Pillai, N.G.K. Bineesh, K.K.; Sebastine, M.; Akhilesh, K.V. Lantern fishes (Myctophids): Bycatch of deep-sea shrimp trawlers operated off Kollam, south-west coast of India. *Marine Fisheries Information Service T and E Ser.*, 2009, 202, 1-4.
33. Sebastine, M.; Bineesh, K.K.; Abdussamad, M.; Pillai, N.G.K. Myctophid fishery along the Kerala coast with emphasis on population characteristics and biology of the headlight fish, *Diaphus watasei* Jordan & Starks, 1904. *Indian Journal of Fisheries*, 2013, 60(4), 7-11.
34. Boopendranath, M.R.; Vijayan, P.K.; Remesan, M.P.; Anandan, R.; Ninan, G.; Zynudheen, A.A.; Das, S.; Rajeswari, G.; Raghu Prakash, R.; Sankar, T.V.; Panda, S.K.; Mohan, C.O.; Vipin, P.M.; Fernandez, T.J.; Renju, R.; Mahato, P.K.; Pradeep, K.; Rajamoorthy, K.; Sherief, M.P.S.; Baby, L.; Abhilash, S. Final Report on CIFT Project Component on Development of Harvest and Post-harvest Technologies for Utilization of Myctophid Resources in the Arabian Sea, CIFT, Cochin, 2012; 246 p.
35. Remesan, M.P.; Prajith, K.K.; Daniel Raj, F.; Joseph, R.; Boopendranath, M.R. Investigations on aimed midwater trawling for myctophids in the Arabian Sea. *Fishery Technology*, 2016, 53, 190-196.
36. Sarnas, A.M. A revision of the Philippine Myctophidae. *The Philippine Journal of Science*, 1953, 82(4), 375-427.
37. Bineesh, K.K.; Sebastine, A.M., Akhilesh, K.V.; Rajool Shanis, C.P.; Hashim, M.; Pillai, N.G.K.; Ganga, U. Diversity of mesopelagic fishes in the deep-sea prawn trawlers from south-west coast of India with notes on myctophids. Book of Abstracts, 20th All India Congress of Zoology. Central Institute of Fisheries Education, Mumbai, 2009, p. 167.
38. Nafpaktitis, B.G.; Backus, R.H.; Craddock, J.E.; Haedrich, R.L.; Robinson, B.H.; Karnella, C. Family Myctophidae. In *Fishes of the Western North Atlantic*, Gibbs Jr., R.H. (ed.); Memoir Sears Foundation for Marine Research, New Haven, 1977; pp. 13-265.
39. Shaviklo A.R. Developing value-added products from lanternfish. INFOFISH International, 2012, 2, 42-46.
40. Anon. Sindh govt shows reservations on proposed deep-sea fishing policy. Retrieved from <https://www.thenews.com.pk/latest/151278-Sindh-govt-shows-reservations-on-proposed-deep-sea-fishing-policy>, 2016.
41. Anon. Information on fishery management in the Republic of South Africa. <http://www.fao.org/fi/old-site/FCP/en/ZAF/body.htm>, 2001.
42. Valinassab, T.; Pierce, G.J.; Johannesson, K. Lantern fish (*Benthosema pterotum*) resources as a target for commercial exploitation in the Oman Sea. *Journal of Applied Ichthyology*, 2007, 23, 573-577.
43. Shotton, R. Lanternfishes: A potential fishery in the Northern Arabian Sea? In *A review of the state of world fishery resources: Marine fisheries*. FAO fisheries circular No. 920 FIRM/C920. FAO, Rome, 1997.
44. Vipin, P.M.; Renju, R.; Fernandez, T.J.; Pradeep, K.; Boopendranath, M.R.; Remesan, M.P. Distribution myctophid resources in the Indian Ocean. *Reviews in Fish Biology and Fisheries*, 2012, 22, 423-436.
45. Wysokinski, A. The living marine resources of the Southeast Atlantic. *FAO Fisheries Technical Paper* 178, 1986; 120 p.
46. Gerhard Krefft, Walther Herwig Station. 208/1966; Institut für Seefischerei, Hamburg; personal communication, 1966.
47. Kock, K.H. The state of exploited fish stocks in the Southern-Ocean –a review. *Arch. Fischereiwiss*, 1991, 41, 1-66.
48. Collins, M.A.; Stowasser, G.; Fielding, S.; Shreeve, R. Xavier, J.C.; Venables, H.J.; Enderlein, P.; Cherel, Y.; Van de Putte, A. Latitudinal and bathymetric patterns in the distribution and abundance of mesopelagic fish in the Scotia Sea. *Deep-Sea Research II*, 2012, 59-60, 189-198.
49. Prochazaka, K. Status of the South African Marine Fishery Resources 2014. Department of Agriculture, Forestry and Fisheries, Cape Town, South Africa, 2014; 84 p.
50. Tyler, T. Examining the feeding ecology of two mesopelagic fishes (*Lampanyctodes hectoris* & *Maurollicus walvisensis*) off the west coast of South Africa using stable isotope and stomach content analyses. Master of Science thesis, University of Cape Town, Biological Sciences, Cape Town, South Africa, 2016; 78 p.
51. Anon. Novosti Otrastli: Rybnaja muka iz Mavrolika, 11 February 2009. In Russian, translated by E.A. Pakhomov, https://www.fishnet.ru/news/novosti_otrasli/11003.html.