

Review

# The Unexplored Potential of Edible Flowers Lipids

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**Abstract:** Edible flowers have been historically linked to traditional world cuisine and culture. They are often used as ingredients in food and beverages for medicinal or pharmaceutical purposes. However, little attention has been paid to the quality of their lipids, and therefore to their potential for oil extraction and use in the food and food supplements industries. This review summarizes the current knowledge on the lipid composition of several edible flowers, including fat content, fatty acids, vitamin E, and carotenoids profiles. Edible flower lipids were found to be rich in linoleic (C18:2) and  $\alpha$ -linolenic (C18:3) acids, which are essential fatty acids. Furthermore, most flowers are a good source of  $\alpha$ -tocopherol and xanthophylls, such as lutein and zeaxanthin. This review provides valuable information on the lipid profile of some edible flowers in order to better characterize them and to increase their popularization among the food industry and consumers, boosting agriculture demand for these products.

**Keywords:** edible flowers; lipid composition; fatty acids; tocopherols; carotenoids

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## 1. Introduction

Lipids are major and essential constituents of all plant cells, providing structural integrity and energy for various metabolic processes [1]. In plants, the compartmentation of neutral lipids is mostly associated with seed tissues, where triacylglycerols are stored [2]. As such, most of the research on the lipid composition of plants has mainly focused on the oil from their seeds [3–6]. However, some non-seed tissues, such as leaves, flowers, and fruits, also synthesize and store lipids, although until now, their formation or function in these tissues is poorly understood [2]. Indeed, lipids are among the least studied metabolites in flowers, but recently they began to be further explored. The lipid composition was reported to be significantly different among flowers' organs and tissues [7]. Furthermore, the majority of studies on edible flowers lipids have focused on their essential oils including basil (*Ocimum basilicum* L.) [8], chrysanthemum (*Chrysanthemum indicum* L.) [9], marigold (*Tagetes minuta* L.) [10], yarrow (*Achillea millefolium* L.) [11], calendula (*Calendula officinalis* L.) [12], and rose (*Rosa × damascena* Herrm.) [13]. However, the literature assigns more importance to pollen compared to other flowers' parts (petals, sepals, and buds), because of their distinctive fatty acid profiles, characteristically dominant in one or more fatty acids [14]. However, pollen may detract flower's flavor and cause allergies in some people, and so it is usually removed when edible flowers are marketed.

In order to improve knowledge about the lipid composition of edible flowers, the purpose of this paper was to provide an overview of published data on the lipid content, fatty acids profile, tocopherols, and carotenoids in edible flowers, to increase their acceptability as potential food ingredients and therefore

their production for food purposes. The authors of the present review want to state that inflorescences, such as cauliflower, broccoli, and artichoke, were not included in this discussion.

## 2. Edible Flowers

The edible flowers market is gaining interest from consumers and chefs of restaurants because they add color, fragrance, and flavor to food, and due being a potential source of nutrients and bioactive compounds [15]. The use of flowers as a food ingredient has been traced back to ancient civilizations. For example, edible flowers were especially popular in Victorian era in England. Other cultures incorporated edible flowers as ingredients in a wide variety of recipes: ancient Romans used violets and roses in dishes and lavender in sauces, native Americans ate blossoms from pumpkin and squash, medieval French put calendula in salads, and Europeans prepared drinks and salads with dandelion flowers [16,17]. Nowadays, edible flowers are most often consumed fresh, but they can also be consumed dried, in ice cubes, canned in sugar, and preserved in distillates [16]. In general, edible flowers can be eaten whole, but depending on the flower species, in some cases, only some parts should be consumed. For example, only the petals of *Tulipa*, *Chrysanthemum*, and *Rosa* spp., or the flower buds of daisies (*Bellis perennis* L.) or garden nasturtium (*Tropaeolum majus* L.) are consumed. Furthermore, in some flowers, it is necessary to remove some parts due to their bitterness, such as the white base in petals of roses and chrysanthemums.

Many flowers are edible but proper identification is essential because some are poisonous. In Figure 1 some edible flowers are represented. Popular edible flowers include hibiscus (*Hibiscus rosa-sinensis* L.), calendula (*Calendula officinalis*), nasturtium (*Tropaeolum majus*), pansy (*Viola × wittrockiana* Gams), rose (*Rosa* spp.), borago (*Borago officinalis* L.), begonia (*Begonia × tuberhybrida* Voss), busy lizzie (*Impatiens walleriana* Hook.f.), and viola (*Viola cornuta* L., *hybrida* Wiesb., *tricolor* L., *odorata* L.) [18]. Some herb flowers are also edible: alliums (leeks, chives, garlic), thyme (*Thymus vulgaris* L.), summer savory (*Satureja hortensis* L.), marjoram (*Origanum majorana* L.), mint (*Mentha* spp.), and common sage (*Salvia officinalis* L.), as well as flowers of some fruit trees, such as elderberry blossoms (*Sambucus* spp.) and citrus blossoms (orange, lemon, lime, grapefruit, and kumquat). Moreover, some flowers are recognized by consumers as vegetables, such as artichoke (*Cynara scolymus* L.), broccoli, and cauliflower (*Brassica oleracea* L.). Even though these are inflorescences, they are not discussed in this review.



**Figure 1.** Examples of edible flowers. Sources: Flora-On: Flora de Portugal Interactiva. (2014). Sociedade Portuguesa de Botânica [19].

### 3. Lipid Content and Composition

Lipid contents reported in the literature for edible flowers are described in Table 1. Hibiscus flowers showed the highest fat content at 19 and 26 g/100 g dry weight (dw). In general, the fat content in other edible flowers ranges from 0.1 to 8.5 g/100 g dw for *Centaurea cyanus* and *Antirrhinum majus* L., respectively. However, since the main component is water, varying between 70% and 95%, the fat content in fresh edible flowers is low [20]. Furthermore, the fat content in edible flowers is not very distinct from other aerial parts of the plant; on a 100 g dw basis, moringa flowers had 2.91 g of fat, leaves 4.96 g, and immature pods 1.28 g [21]. Common mallow flowers had 2.84 g of fat, leaves 2.76 g, and leafy flowered stems 3.09 g [22]. When comparing the fat contents of edible flowers with other vegetables, such as asparagus (3.99 g/100 g dw) [23], lettuce (0.25 g/100 g dw), cabbage (0.2 g/100 g dw), and spinach (0.38 g/100 g dw) [24], the values are similar.

**Table 1.** Fat content (g/100 g dry weight) reported in the literature for some edible flowers.

Scientific Name	Flower	Common Name	Fat (g/100 g Dry Weight)	References
<i>Agave salmiana</i> Otto ex Salm-Dyck		Agave	2.8	[25]
<i>Allium schoenoprasum</i>		Chives	3.4	[26]
<i>Antirrhinum majus</i>		Snapdragon	4.2–8.5	[25,27]
<i>Arbutus xalapensis</i> Kunth		Texas madrone	3.9	[25]
<i>Azadirachta indica</i> L.		Neem	5.2	[28]
<i>Calendula officinalis</i>	Calendula/common marigold/pot marigold		3.6–5.6	[29–31]
<i>Centaurea cyanus</i>		Centaurea	0.1	[30]
<i>Cucurbita pepo</i> L.		Pumpkin	5.0	[25]
<i>Cynara scolymus</i>		Artichoke	2.8	[29]
<i>Dahlia mignon</i>		Dahlia	2.2	[30]
<i>Erythrina americana</i> Mill.		Coral tree	2.3	[25]
<i>Erythrina caribaea</i> Krukoff & Barneby		Erythrina	1.5	[25]
<i>Euphorbia radians</i> Benth.		Sun spurge	4.9	[25]
<i>Hibiscus esculentus</i> L.		Hibiscus	19.0	[32]
<i>Hibiscus sabdariffa</i>		Mahua	26.0	[32]
<i>Madhuca indica</i> J.F.Gmel.		Common mallow	6.1	[15]
<i>Malva sylvestris</i>		Moringa	2.8	[22]
<i>Moringa oleifera</i> Lam.		Rosa canina	2.9	[21]
<i>Rosa canina</i> L.		Rose	2.0	[30]
<i>Rosa micrantha</i>		Sechuan button	1.3	[33]
<i>Spilanthes oleracea</i> L.		Tagetes erecta	2.2	[34]
<i>Tagetes erecta</i> L.		Mexican marigold	1.9	[34]
<i>Tropaeolum majus</i>	Garden nasturtium		3.1–3.6	[29,34]
<i>Viola × wittrockiana</i>		Pansies	5.0–6.0	[27,29]
<i>Yucca filifera</i> Chabaud		Yucca	2.1	[25]

The major lipid classes—fatty acids and lipid-soluble components (vitamin E and carotenoids) are detailed in the next sections.

#### 3.1. Fatty Acids

The fatty acids profile of some edible flowers, including total saturated fatty acids (SFA), monounsaturated fatty acids (MUFA), and polyunsaturated fatty acids (PUFA) contents, are summarized in Tables 2 and 3. Thirty-six fatty acids were identified and quantified in edible flowers. The major fatty acids found in flowers were palmitic acid (C16:0), linoleic (C18:2), and  $\alpha$ -linolenic (C18:3) (Figure 2) with a high proportion of essential fatty acids. However, their ranges varied widely among species: palmitic acid ranged from 0.08% to 53.9% for *Hibiscus sabdariffa* and *Chrysanthemum morifolium* Ramat., respectively; linoleic acid varied between 0.05% and 57.02% for *Hibiscus sabdariffa* and *Punica granatum* L., respectively; and,  $\alpha$ -linolenic acid between 0.02% and 36.9% for *Hibiscus esculentus* and *Calendula officinalis* (petals), respectively.

Palmitic acid is one of the most common SFA found in plants. Although associated with increased risk of developing cardiovascular diseases [35], oxidative DNA damage, DNA strand breakage, necrosis, and apoptosis in human cells in vitro [36,37], when consumed with other fatty acids, like PUFAs, which were also detected in edible flowers, SFA are unlikely to have any

significant impact on human health [37,38]. Furthermore, a recent review reported that more rigorous investigations are needed to understand the advantages and disadvantages induced by palmitic acid consumption, because there are some controversial results [39]. Edible flowers also contain low amounts of other saturated fatty acids, such as stearic (C18:0) (0.01–16.8% for *Hibiscus sabdariffa* and *Rosa canina*, respectively), lauric (C12:0) (0.09–3.66% for *Moringa oleifera* and *Calendula officinalis* flowers, respectively), and myristic (C14:0) acids (0.1–24.9% for *Chrysanthemum morifolium* and *Calendula officinalis* flowers, respectively).

Among the PUFAs, two of the most important fatty acids, linoleic acid (omega ( $\omega$ )-6 group) and  $\alpha$ -linolenic acid (omega( $\omega$ )-3 group), are essential fatty acids. So, humans must obtain them through diet because the body lacks the desaturase enzymes required for their production. These PUFAs are present in high proportions in some flowers (>50%), such as *Calendula officinalis* (petals), *Taraxacum* sect. *Ruderalia*, *Punica granatum*, *Rosa micrantha*, and *Trifolium angustifolium* (Table 2). Both fatty acids have important roles in human growth and development, as well as in the prevention and treatment of coronary artery diseases, hypertension, diabetes, arthritis, other inflammatory and autoimmune disorders, and cancer [40–44]. So, the presence of  $\omega$ -3 and  $\omega$ -6 fatty acids in edible flowers could be a way to promote their consumption and inclusion in the human diet. Regarding MUFA, they are mainly represented by oleic acid (C18:1), ranging from 0.01% (*Hibiscus sabdariffa*) to 28.5% (*Gundelia tournefortii* L.), followed by eicosenoic (C20:1) and erucic (C22:1) acids in low quantities.

In Table 3, PUFA and SFA predominate over MUFA due to the significant contribution of  $\alpha$ -linolenic and linoleic, and palmitic acids, respectively. However, in all cases, unsaturated fatty acids predominate over saturated ones (generally higher than 53%), with one exception observed in calendula flowers (23.3%). Furthermore, high PUFA/SFA ratios reduce the risk of cardiovascular diseases [45]. In general, all edible flowers studied until now showed high ratios of PUFA/SFA (higher than 0.45) [46] and low  $\omega$ -6/ $\omega$ -3 ratios (lower than 4.0) [33], which are recommended for the human diet, except calendula (PUFA/SFA ratio equal to 0.27) and dahlia ( $\omega$ -6/ $\omega$ -3 ratio equal to 4.25). Additionally, a  $\omega$ -6/ $\omega$ -3 ratio equal or lower than 4 is beneficial for reducing serum “bad cholesterol”, and inhibiting a major receptor for oxidized low-density lipoprotein (ox-LDL) uptake [47], with potential to protect against obesity, insulin resistance, and inflammation [48]. However, Tsoupras et al. [49] presented data that supports inflammation induced by several factors, with platelet-activating factor (PAF), as being strongly implicated in cardiovascular diseases, rather than serum cholesterol alone. Therefore, food antioxidants might be lipid counterparts on the onset of cardiovascular diseases. So, edible flowers are a healthy lipid source (rich in oleic, linoleic, and linolenic fatty acids), offering potential health benefits.

**Table 2.** Fatty acids (%) profile of some edible flowers mentioned in the literature.

Fatty Acids	<i>Allium schoenoprasum</i>	<i>Anchusa azurea</i>	<i>Azadirachta indica</i>	<i>Calendula officinalis</i>	<i>Capparis spinosa L.</i>	<i>Cassia fistula L.</i>	<i>Centaurea cyanus</i>	<i>Chrysanthemum morifolium</i>	<i>Cichorium intybus</i>	<i>Taraxacum sect. Ruderalia</i>	<i>Dahlia mignon</i>	<i>Gundelia tournefortii</i>
	Flowers	Flowers	Flowers	Petals	Flowers	Flowers	Flowers	Petals	Flowers	Flowers	Petals	Bud
Caproic (C6:0)				0.3	0.5		0.2				0.9	nd
Caprylic (C8:0)			0.3	0.3	0.7		0.07				0.9	nd
Capric (C10:0)			0.1	0.2	0.3		0.1				1	nd
Undecylic (C11:0)			0.3	0.1			nd				nd	nd
Lauric (C12:0)			1	1.6	3.7		nd	nd	0.1–1.1		0.7	nd
Myristic (C14:0)	11.6	1.9	9.9	24.9	1.9	2.1	0.9	0.1–14.8	0		3.1	nd
Myristoleic (C14:1ω5)				nd	0.1		0.2		7		0.6	nd
Pentadecylic (C15:0)	7.9–16.9	0.6	0.8	0.2	0.5	1.6		0.4		1.8	0.7	nd
Palmitic (C16:0)	14.3	31.8	23.4	35.6	25.7	34.5	23.4	0.6–53.9	18.5	17	23.4	23.4
Palmitoleic (C16:1ω7)				0.2	0.2		0.3				0.9	nd
Margaric (C17:0)	nd	0.3	0.2	0.5	tr		0.8		0.4		0.9	nd
Stearic (C18:0)	5.5	2.9	3.9	5.9	3.3	15.2	9.7	1	1.7		7.6	2.5
Oleic (C18:1ω9)	5.8	9.7	1.6	2.5	0.4	nd	4.4		1		5.8	28.5
Linoleic (C18:2ω6)	7.6–13.4	6	18.6	20.3	9.3	0.7	41.2	6.7		1.9	33	36.5
α-Linolenic (C18:3ω3)	7.3	12.6	36.9	11.1	0.5		18.8		0.6	23.1	8.6	0.1
Arachidic (C20:0)	tr	1.3	0.63	0.8	2.6	2.8	5.3		0.8		1.6	0.3
Eicosenoic (C20:1ω9)		0.4	nd	0.1		nd	nd				nd	nd
Eicosadienoic (C20:2ω6)			nd				nd				0.4	nd
Eicosatrienoic (C20:3ω3)			0.26				0.5				0.6	nd
Dihomo-γ-linolenic (C20:3ω6)							26.9				nd	nd
Eicosapentaenoic (C20:5ω3)			nd								nd	nd
Behenic (C22:0)	1.7	2.1	0.56	0.3	5.9	0.9	2		1.5		nd	nd
Erucic (C22:1ω9)			nd			nd	6				nd	nd
Docosadienoic (C22:2ω6)		5.7										
Tricosylic (C23:0)	1.8		0.1		nd		0.2		0.3		0.2	nd

Table 2. Cont.

Fatty Acids	<i>Allium schoenoprasum</i>	<i>Anchusa azurea</i>	<i>Azadirachta indica</i>	<i>Calendula officinalis</i>	<i>Capparis spinosa L.</i>	<i>Cassia fistula L.</i>	<i>Centaurea cyanus</i>	<i>Chrysanthemum morifolium</i>	<i>Cichorium intybus</i>	<i>Taraxacum sect. Ruderalia</i>	<i>Dahlia mignon</i>	<i>Gundelia tournefortii</i>
	Flowers	Flowers	Flowers	Petals	Flowers	Flowers	Flowers	Petals	Flowers	Flowers	Petals	Bud
Lignoceric (C24:0)		nd	1.7	0.9	0.9	tr	nd	1.1		nd	2.3	nd
References	[26]	[50]	[28]	[29]	[31]	[50]	[51]	[30]	[52]	[50]	[53]	[30]
Fatty Acids	<i>Hedysarum coronarium</i>	<i>Hibiscus esculentus</i>	<i>Hibiscus sabdariffa</i>	<i>Malva sylvestris</i>	<i>Moringa oleifera</i>	<i>Punica granatum</i>	<i>Robinia pseudoacacia</i>	<i>Rosa canina</i>	<i>Rosa micrantha</i>	<i>Rosmarinus officinalis</i>	<i>Sambucus nigra</i>	<i>Trifolium angustifolium</i>
	Flowers	Flowers	Flowers	Flowers	Flowers	Flowers	Flowers	Petals	Petals	Flowers	Flowers	Flowers
Caproic (C6:0)				0.6	nd			0.2	0.1		0.2	
Caprylic (C8:0)				0.03	0.09			0.2	0.4		0.8	
Capric (C10:0)				0.02	nd			0.3	0.3		0.2	
Undecylic (C11:0)					nd			nd	nd		nd	
Lauric (C12:0)				0.12	0.09	1.2–6.6		1.22	0.9		2.3	
Tridecylic (C13:0)					nd			0.03	nd		0.02	
Myristic (C14:0)		tr	tr	0.9	0.59	0.6–3.7		2.6	1.5		4.8	
Myristoleic (C14:1)	6.4			0.2–0.8	1.96		2.4	0.3	nd	3.7	1.7	0.2
Pentadecylic (C15:0)	1			0.07–1.5	nd		nd	0.3	0.2	0.4	1.2	0.3
Palmitic (C16:0)	7.7	3.4	0.08	17.2–22.4	23.43	27.7–43.6	9.1	23.4	11.3	7.1	17.7	15.4
Palmitoleic (C16:1)		0.03	nd	0.62	nd	nd–0.95		0.22	nd			0.3
Margaric (C17:0)	nd			nd–0.3	nd		0.7	0.53	0.7	0.3	nd	0.4
Stearic (C18:0)	4.4	0.3	0.01	2.4–3.6	4.52	4.8–10.1	2.3	16.8	0.6	2.2	nd	3.2
Oleic (C18:1ω9)	0.4	1.8	0.01	1.9–6.1	21.55	6.3–20.5	0.7	1.95	1.8	1.1	1.5	6.1
Linoleic (C18:2ω6)	1.3	2.5	0.05	0.8–23.5	18.96	47.4–57.0	2.7	31.9	21.2	1.5	0.8	20.2
α-Linolenic (C18:3ω3)	0.7	0.02	0.03	3.1–33.5	23.01	14.8–25.4	0.6	19.5	32.3	1.9	1.3	34.7
Arachidic (C20:0)	1.2	0.02	tr	1.2–1.6	0.98	2.3–8.4	0.8	3.6	3.7	0.5	0.2	2.6
Eicosenoic (C20:1)					0.07	nd	0.9–4.9		nd	0.6		0.2
Eicosadienoic (C20:2ω6)					0.1	0.75	0.8–1.8		nd	nd	1	0.1
Eicosatrienoic (C20:3ω3)					nd	nd–0.5		0.33	nd			

**Table 2.** Cont.

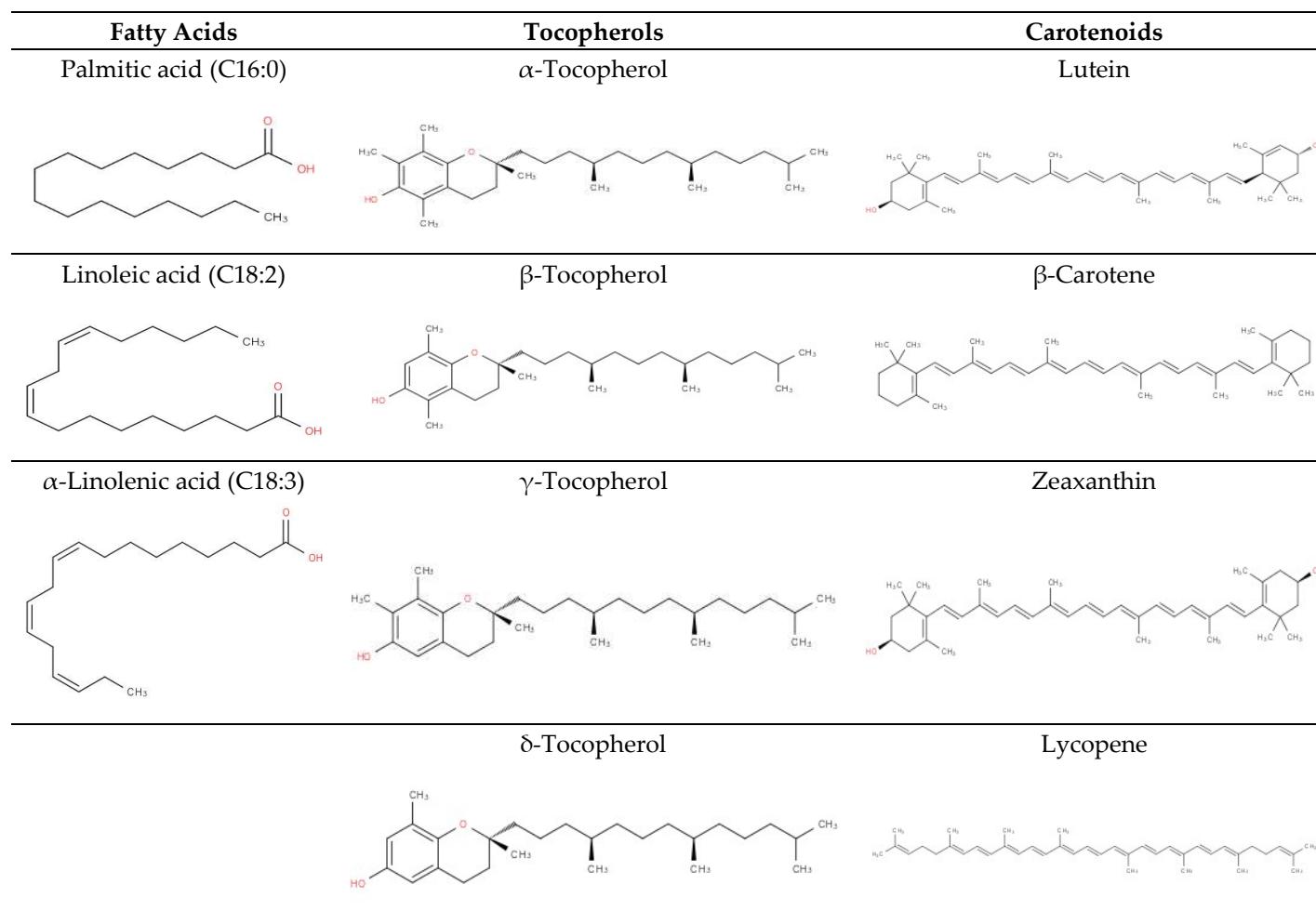
Fatty Acids	<i>Hedysarum coronarium</i>	<i>Hibiscus esculentus</i>	<i>Hibiscus sabdariffa</i>	<i>Malva sylvestris</i>	<i>Moringa oleifera</i>	<i>Punica granatum</i>	<i>Robinia pseudoacacia</i>	<i>Rosa canina</i>	<i>Rosa micrantha</i>	<i>Rosmarinus officinalis</i>	<i>Sambucus nigra</i>	<i>Trifolium angustifolium</i>
	Flowers	Flowers	Flowers	Flowers	Flowers	Flowers	Flowers	Petals	Petals	Flowers	Flowers	Flowers
Dihomo- $\gamma$ -linolenic (C20:3 $\omega$ 6)						0.6–0.9						
Eicosapentaenoic (C20:5 $\omega$ 3)					nd			nd	nd			
Behenic (C22:0)	2.1			1–1.5	2.06	0.3–5.5	0.5	1.8	4.4	nd	nd	2.4
Erucic (C22:1)					nd	0.7–1.8		nd				
Docosadienoic (C22:2 $\omega$ 6)						0.9–1.6						
Tricosylic (C23:0)	nd			10	nd		nd	0.08	9.3	0.3	nd	0.2
Lignoceric (C24:0)	0.2			1	nd	0.3–1.5	0.3	1	3.4	nd	nd	3.2
References	[50]	[32]	[32]	[22,50]	[21]	[55]	[50]	[30]	[33]	[50]	[50]	[56]

nd—not detected; tr—trace amounts.

**Table 3.** Saturated, monounsaturated, polyunsaturated and unsaturated fatty acids in edible flowers \*.

Flowers		SFA (%)	MUFA (%)	PUFA (%)	UFA (%)	PUFA/SFA	$\omega$ -6/ $\omega$ -3	References
Scientific Name	Common Name							
<i>Azadirachta indica</i>	Neem	45.1	10.2	44.7	54.9	1.0	1.5	[28]
<i>Bauhinia variegata</i> L.	Cow's foot	42.3	6.2	51.4	57.6	1.2	1.2	[57]
<i>Calendula officinalis</i>	Calendula (common marigold)	40.7–76.7	1.8–2.9	20.5–57.5	23.3–59.3	0.3–1.4	0.6–0.8	[30,31]
<i>Centaurea cyanus</i>	Centaurea	36.2	10.9	52.9	63.8	1.5	0.4	[30]
<i>Dahlia mignon</i>	Dahlia	46.6	7.2	46.2	53.4	1.0	4.2	[30]
<i>Moringa oleifera</i>	Moringa	31.8	26.3	42.2	68.4	1.3	0.8	[21]
<i>Punica granatum</i> cv. Chelfi	Punica	15.0	59.2	25.8	85.0	1.7	2.6	[55]
<i>Punica granatum</i> cv. Gabsi	Punica	33.5	14.4	52.0	66.5	1.6	2.8	[55]
<i>Punica granatum</i> cv. Tounsi	Punica	33.8	8.8	57.4	66.2	1.7	3.2	[55]
<i>Punica granatum</i> cv. Nabli	Punica	31.6	8.1	14.4	59.2	1.4	2.3	[55]
<i>Rosa canina</i>	Rose	45.8	2.5	51.7	54.2	1.1	1.6	[30]
<i>Rosa micrantha</i>	Rose	11.2	13.4	75.4	88.8	6.7	1.4	[33]
<i>Taraxacum officinale</i>	Dandelion	33.5	3.0	63.5	66.5	1.9	1.1	[53]

\* SFA—Saturated fatty acids; MUFA—Monounsaturated fatty acids, PUFA—Polyunsaturated fatty acids, UFA—Unsaturated fatty acids.



**Figure 2.** Chemical structures of the main lipophilic compounds detected in edible flowers. Source: Hastings, et al. [58].

### 3.2. Tocopherols

Vitamin E is a class of lipid-soluble antioxidants synthesized by plants and photosynthetic organisms [59]. There are four isoforms ( $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$ ) of tocopherols and tocotrienols, which differ in the number and positions of the methyl groups in the chromanol ring. In flowers, tocols are mostly located in petal leucoplasts [59]. Tocopherols are also essential components of the human diet because they perform numerous critical functions, including quenching and scavenging of various reactive oxygen species (ROS) and free radicals, and protecting PUFA from lipid peroxidation [60].

Tocopherols identified and quantified in edible flowers are listed in Table 4. In the majority of the flowers analysed, only the four isoforms of tocopherols ( $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$ ) were detected (Figure 2), with  $\alpha$ -tocopherol being the major compound. *Calendula officinalis* was the flower that had the highest content of  $\alpha$ -tocopherol (56.78 mg/100 g dw), followed by *Rosa micrantha* (26.72 mg/100 g dw) and *Taraxacum* sect. Ruderalia (21.60 mg/100 g dw). These flowers presented higher contents of  $\alpha$ -tocopherol when compared with some vegetables, such as wild asparagus (0.75–4.51 mg/100 g dw) or leafy vegetables (2.59–10.12 mg/100 g dw) [61]. The Academy of Sciences reports a Recommended Dietary Allowance (RDA) value for  $\alpha$ -tocopherol of 15 mg/day [62], whereas the daily recommended dose for tocopherols consumption in adults is 300 mg/day [63]. Despite the low amounts of tocopherols in edible flowers, their daily consumption may contribute to supplying this vitamin to the organism. In parallel,  $\gamma$ -tocopherol was also detected in almost all flowers studied, ranging from 0.16 to 7.68 mg/100 g dw in *Gundelia tournefortii* and *Rosa micrantha*, respectively.

### 3.3. Carotenoids

Carotenoids are lipophilic pigments widely distributed in nature, and they have different roles in the plant life cycle including photo-protective functions, and provision of substrates for plant growth, regulator of abscisic acid and other hormones [64–67], as well as, in human nutrition and health, providing provitamin A and having anti-cancer activities [68]. Carotenoids can be classified into two classes: carotenes ( $\alpha$ -carotene,  $\beta$ -carotene, and lycopene) and xanthophylls ( $\beta$ -cryptoxanthin, lutein, and zeaxanthin) [69]. In flowers, carotenoids are found in all anatomical parts: sepals, pollen, anthers, stamens, and petals [70]. Flowers offer distinct carotenoids profiles that depend on species and variety [71], as shown in Tables 5 and 6. The edible flowers studied have shown a very different range in values between species.

Table 5 also shows that different flowers of the same species may have different amounts of total carotenoids. This may be due to their different colors, cultivars, soil characteristics, conditions of production, and parts of the flower (petals or whole flowers), or other factors including analytical ones. Among published studies in edible flowers, the majority of carotenoids are xanthophylls (Table 6), such as lutein and zeaxanthin (Figure 2). Lutein was the main carotenoid in chrysanthemum (11.78–307.22  $\mu$ g/g dw), snapdragon (14.1  $\mu$ g/g dw), garden nasturtium (350–450  $\mu$ g/g fw), Mexican marigold (1062  $\mu$ g/g fw), crem (243.23  $\mu$ g/g dw), and pansies (51.1  $\mu$ g/g dw). Epoxy xanthophylls, such as flavoxanthin (calendula), violaxanthin (yellow bloom), auroxanthin (golden aster), antheraxanthin (rose), and neoxanthin (flame tree), are also common and can be found in high contents in some flowers (Table 6). Edible flowers also contain carotenes, such as lycopene and  $\beta$ -carotene (Figure 2). In some cases, edible flowers showed higher values of  $\beta$ -carotene than green leafy and root vegetables. For example, garden petunias cv. Summer Sun (358.1 mg/100 g fw), Mexican marigold (8.55 mg/100 g fw), and squash flowers (1.01–13.35 mg/100 g fw) showed higher values than spinach (4 mg/100 g fw), carrot (2.2 mg/100 g fw), coriander (6.1 mg/100 g fw), and mint (4.3 mg/100 g fw) [72].

As expected, the carotenoid amounts were different according to the distinct parts of the plants. For example, leaves of caper (mean 5.02 and 8.09 mg/100 g fw,  $\beta$ -carotene and lutein respectively) had higher concentrations of  $\beta$ -carotene and lutein than flower buds (mean 1.17 and 2.24 mg/100 g fw,  $\beta$ -carotene and lutein respectively) [73]; petals of calendula (7.71 mg/g dw) had higher values of total carotenoids than pollen (1.61 mg/g dw), stems (0.18 mg/g dw), and leaves (0.85 mg/g dw) [74].

Furthermore, different carotenoids are also detected in different parts of the flower: the stems of calendula contained carotenoids typical of photosynthetic tissue (e.g., lutein and  $\beta$ -carotene), whereas petals and stems showed more furanoid-oxides (e.g., flavoxanthin, auroxanthin, luteoxanthin, and 9Z-antheraxanthin) [74]. So, consumers of edible flowers can obtain different carotenoids according to the part of flower they eat.

Some studies found correlations between the color of the flowers and their carotenoids content [75,76], and that carotenoids in flowers are responsible for the yellow, orange, and red color classes of pigments [71]. Pintea et al. [75] found that calendula flowers with distinct colors (different varieties) contain the same pigments but in different amounts. For example, the variety that is dark orange (Double Esterel Orange) presented the highest total content of carotenoids (276 mg/100 g fw). Similar results were detected by Park et al. [77], who reported that between different cultivars and colors of chrysanthemums, the yellow-orange flowers were those that showed the highest content of carotenoids, namely Il Weol (345.56  $\mu$ g/g dw) and popcorn ball (189.57  $\mu$ g/g dw). So, carotenoids are important compounds in edible flowers, because the color of the flower is an essential attribute that influences the commercial acceptance of consumers [78]. Some colors of edible flowers may induce a reluctant attitude by consumers during the purchase, whereas others are more appealing. Since color may influence taste, reddish flowers may suggest to the consumer that they have a “sweet cherry or strawberry flavor”, whereas yellowish flowers may be associated with a sour or citrus flavour [79]. Furthermore, at the time of purchase of edible flowers, color influences the consumers because they may like one color or combination of colors more than others. According to Kelley et al. [79], consumers prefer dark colors, such as orange (associated to carotenoids) and crimson, because they are more appealing.

**Table 4.** Tocopherols determined in edible flowers mentioned in the literature.

Flowers		Tocopherols (mg/100 g dw)				Ref.
Scientific Name	Common Name	$\alpha$ -Tocopherol	$\beta$ -Tocopherol	$\gamma$ -Tocopherol	$\delta$ -Tocopherol	
<i>Musa × sapientum</i> L.	Banana flower	5.1				[80]
<i>Calendula officinalis</i>	Calendula (Common marigold)	19.4–56.8	1.1–1.5	2.4–2.9	nd	[30,31]
<i>Capparis spinosa</i>	Caper (different regions)	1.8–2.7		0.4–1.1		[81]
<i>Centaurea cyanus</i>	Centaurea	0.6	nd	0.3	nd	[30]
<i>Matricaria recutita</i>	Chamomile	3.5	0.2	2.6–4.0	1.2	[56]
<i>Trifolium angustifolium</i>	Clover	12.7	0.6	5.4	0.4	[56]
<i>Malva sylvestris</i>	Common mallow	14.0	0.6	2.5	0.2	[22]
<i>Tropaeolum pentaphyllum</i> Lam.	Crem	2.8		1.0		[82]
Dahlia mignon	Dahlia	4.4	1.8	0.7	0.4	[30]
<i>Bauhinia variegata</i>	Cow's foot	1.7				[57]
<i>Taraxacum</i> sect. Ruderalia	Dandelion	21.6	11.2	5.6	6.3	[53]
<i>Gomphrena globosa</i> L.	Globe amaranth	0.4		3.0	5.2	[83]
<i>Rosa canina</i>	Rose	8.2	0.2	0.8	0.1	[30]
<i>Rosa micrantha</i>	Rose	26.7	0.7	7.7	0.2	[33]
<i>Gundelia tournefortii</i>	Tumbleweed	7.9	0.2	0.2		[54]

nd: not detected.

**Table 5.** Total carotenoids contents in edible flowers reported in the literature.

Edible Flowers		Part Flower	Color	Total Carotenoids	Reference
Scientific Name	Common Name		Results Expressed in µg/g Fresh Weight		
<i>Calendula officinalis</i>	Calendula/Common marigold	Flowers	Dark orange to yellow	57–2760	[75,84–86]
		Petals	Yellow and orange	1073–1696	[87]
<i>Chrysanthemum morifolium</i>	Chrysanthemum	Petals	Yellow and orange	122–343	[87]
<i>Cucurbita pepo</i>	Squash flower	Flowers	ns	768	[88]
<i>Cucurbita maxima</i> Duchesne	Squash flower	Flowers	Yellow	12–188	[76]
<i>Dianthus caryophyllus</i> L.	Carnation	Petals	Green and red	2–12	[71]
<i>Helianthus annuus</i> L.	Sunflower	Petals	Yellow and orange	144–1600	[87]
<i>Helianthus tuberosus</i> L.	Jerusalem artichoke	Flower	ns	15.6	[85]
<i>Hibiscus rosa-sinensis</i>	Hibiscus	Flowers (different cultivars)	ns	2 × 10 <sup>3</sup> –40 × 10 <sup>3</sup>	[89]
<i>Matricaria recutita</i>	Chamomile	Flowers (different varieties)	ns	135–162	[90]
<i>Petunia × hybrida</i> Vilm.	Garden petunias	Flowers	Solid colour or bicolour (Red, rose, pink, blue, burgundy, white, yellow)	0.32–96.8	[91]
<i>Tagetes erecta</i>	Mexican marigold African Marigold	Petals	Yellow and orange	48–2130	[87]
		Flowers	Deep orange	6.3–1304	[85,92]
<i>Tagetes patula</i> L.	French marigold	Petals	Yellow and orange	270–2020	[86]
<i>Rosa</i> spp.	Rose	Flowers	Pink; Yellow; Red; Orange; White	0.1–61.7	[93]
Results expressed in µg/g dry weight					
<i>Antirrhinum majus</i>	Snapdragon	Flowers	ns	29 <sup>1</sup>	[27]
<i>Calendula officinalis</i>	Calendula/Common marigold	Petals	ns	7.71 × 10 <sup>3</sup>	[74]
		Flowers	ns	1405	[94]
<i>Dendranthema grandiflorum</i> (Ramat.) Kitam.	Chrysanthemum	Flowers (different cultivars)	Purple, White, Green, Red, Yellow	19–346	[77]
<i>Tropaeolum pentaphyllum</i>	Crem	Flowers	ns	396	[82]
<i>Viola × wittrockiana</i>	Pansies	Flowers	ns	146 <sup>1</sup>	[27]
<i>Viola tricolor</i>	Viola	Petals	ns	23 <sup>2</sup>	[95]

ns—not specified; <sup>1</sup> Lutein equivalent; <sup>2</sup> Composition of chromoplast globules (% of total dry weight).

**Table 6.** Individual carotenoids values in edible flowers.

Scientific Name	Common Name	Part Flower	Color	Carotenoids		Reference
				Results Expressed in µg/g fw		
<i>Calendula officinalis</i>	Calendula/ Common marigold	Petals (different var.)	Orange Yellow	Luteoxanthin	186.6–195.0 <sup>1</sup>	
				Lutein-5,6-epoxide	27.1–40.0 <sup>1</sup>	
				Flavoxanthin	483.4–532.5 <sup>1</sup>	
				Auroxanthin	120.4–133.7 <sup>1</sup>	
				(9'Z)-Lutein-5,6-epoxide	84.8–106.2 <sup>1</sup>	
				Lutein	33.9–62.5 <sup>1</sup>	
				Antheraxanthin	17.0–31.2 <sup>1</sup>	
				(9Z)-Lutein	10.2–18.7 <sup>1</sup>	
				(5'Z,9'Z)-Rubixanthin	67.8 <sup>1</sup>	
				α-Carotene	13.6 <sup>1</sup>	[87]
<i>Capparis spinosa</i>	Caper	Flower buds	—	β-Carotene	12.5–57.7 <sup>1</sup>	
				(5'Z)-Rubixanthin	50.9 <sup>1</sup>	
				δ-Carotene	23.7 <sup>1</sup>	
				(5Z,9Z,5'Z,9'Z)-Lycopene	69.5 <sup>1</sup>	
				γ-Carotene	33.9 <sup>1</sup>	
				(5'Z)-γ-Carotene	74.6 <sup>1</sup>	
				(5Z,9Z,5'Z)-Lycopene	59.4 <sup>1</sup>	
				(5Z,9Z)-Lycopene	68.5 <sup>1</sup>	
				Lycopene	147.6 <sup>1</sup>	
				β-carotene	4–23.3	
<i>Chrysopsis scabrella</i> Torr. & A.Gray	Golden aster	Flowers	—	Lutein	5.2–40.8	[73]
				Auroxanthin	29.1	
<i>Cucurbita maxima</i>	Squash flower	Petals	Yellow	Bixin	3.5	[69]
				β-Carotene	10.1–133.5	[76]
<i>Delonix regia</i> (Hook.) Raf.	Flame tree	Flower	—	Astaxanthin	2.9	
				Violaxanthin	38.7	
				Neoxanthin	38.7	[69]
				Zeaxanthin	36.7	
				Violaxanthin	12	
<i>Delonix regia</i> var. <i>flavida</i> Stehle	Yellow bloom	Flowers	—	Canthaxanthin	0.13	[69]
				Antheraxanthin	11.9	
<i>Gerbera jamesonii</i> Bolus ex Adlam	Gerbera	Flowers	—	Crocetin	3.7	[69]
				β-Carotene	0.14–35.8	
<i>Petunia hybrida</i>	Garden petunias	Flowers	Solid color/ bicolor (Red, rose, pink, blue, burgundy, white yellow)	Lutein	0.00–13.9	
				Zeaxanthin	0.00–3.3	[91]

Table 6. Cont.

Edible Flower		Part Flower	Color	Carotenoids		Reference
Scientific Name	Common Name			Results Expressed in µg/g fw	Results Expressed in µg/g dw	
<i>Rosa</i>	Rose	Petals	—	Antheraxanthin Crocetin Auroxanthin Bixin β-cryptoxanthin Lutein Neoxanthin Violaxanthin Zeaxanthin β-Carotene Neoxanthin	10.2 2.7 22.1 5.7 31.6 1062 nd 43.7 53.7 85.5 nd	[69]
<i>Solidaster lutens</i> M.L.Green	Solid aster	Petals	—	Bixin β-cryptoxanthin Lutein Neoxanthin Violaxanthin Zeaxanthin β-Carotene Neoxanthin	5.7 31.6 1062 nd 43.7 53.7 85.5 nd	[69]
<i>Tagetes erecta</i>	Mexican marigold	Flowers	Deep orange	Violaxanthin Lutein β-Carotene	tr 350–450 tr	[92]
<i>Tropaeolum majus</i>	Garden nasturtium	Flowers	Yellow Brownish orange	Violaxanthin Lutein β-Carotene	nd	[96]
Results expressed in µg/g dw						
<i>Antirrhinum majus</i>	Snapdragon	Flowers	—	Violaxanthin Antheraxanthin Lutein Zeaxanthin β-carotene Lutein Zeaxanthin	nd nd 14.1 7.4 7.7 11.8–307 0.14–2.9	[27]
<i>Dendranthema grandiflorum</i>	Chrysanthemum	Flowers (different cultivars)	Purple White Green Red Yellow	β-Cryptoxanthin 13-cis-β-Carotene α-carotene Trans-β-carotene 9-cis-β-carotene β-carotene Neoxanthin Violaxanthin Antheraxanthin Lutein Zeaxanthin α-cryptoxanthin β-carotene 5,6-epoxide β-carotene 9Z-β-carotene β-carotene Lutein Zeaxanthin α-cryptoxanthin β-carotene 5,6-epoxide β-carotene 9Z-β-carotene β-carotene Lutein Zeaxanthin α-carotene β-carotene	0.09–2.1 0.13–5.6 0.04–3.5 1.4–55.8 0.3–5.12 1277 tr 2.81 2.83 8.96 4.79 tr 0.48 5.46 0.76 342–388 243 14.2 2.6 3.6 132	[77]
<i>Matricaria recutita</i>	Chamomile	Flowering aerial parts	White	Violaxanthin Antheraxanthin Lutein Zeaxanthin α-cryptoxanthin β-carotene 5,6-epoxide β-carotene 9Z-β-carotene β-carotene Lutein Zeaxanthin α-carotene β-carotene	2.81 2.83 8.96 4.79 tr 0.48 5.46 0.76 342–388 243 14.2 2.6 3.6 132	[56]
<i>Viola declinata</i> Waldst. & Kit.	Viola	Aerial parts	—	Violaxanthin α-cryptoxanthin β-carotene 5,6-epoxide β-carotene 9Z-β-carotene β-carotene Lutein Zeaxanthin α-carotene β-carotene	4.79 tr 0.48 5.46 0.76 342–388 243 14.2 2.6 3.6 132	[97]
<i>Trifolium angustifolium</i>	Clover	Flowering aerial parts	—	Violaxanthin α-cryptoxanthin β-carotene 5,6-epoxide β-carotene 9Z-β-carotene β-carotene Lutein Zeaxanthin α-carotene β-carotene	2.81 2.83 8.96 4.79 tr 0.48 5.46 0.76 342–388 243 14.2 2.6 3.6 132	[56]
<i>Tropaeolum pentaphyllum</i>	Crem	Flowers	—	Violaxanthin α-cryptoxanthin β-carotene 5,6-epoxide β-carotene 9Z-β-carotene β-carotene Lutein Zeaxanthin α-carotene β-carotene	2.81 2.83 8.96 4.79 tr 0.48 5.46 0.76 342–388 243 14.2 2.6 3.6 132	[82]

**Table 6.** Cont.

				Results expressed in µg/g dw	
			—		
<i>Viola × wittrockiana</i>	Pansies	Flowers	—	Violaxanthin Antheraxanthin Lutein Zeaxanthin β-carotene	8.9 8.5 51.1 38.2 41.5
					[27]
				Results expressed in % of peak area of carotenoids in the HPLC chromatogram	
<i>Calendula officinalis</i>	Calendula/ Common marigold	Petals	—	Neoxanthin Z-Neoxanthin Violaxanthin Luteoxanthin Auroxanthin 9Z-Violaxanthin Flavoxanthin Mutatoxanthin 9Z-Antheraxanthin Lutein 9/9VZ-Lutein 13/13VZ-Lutein α-Cryptoxanthin β-Cryptoxanthin Lycopene α-Carotene β-Carotene (8'R)-Luteoxanthin Lutein-5,6-epoxide Flavoxanthin (8R,8'R)-Auroxanthin (9'Z)-Lutein-5,6-epoxide Lutein Antheraxanthin (9Z)-Lutein (5'Z,9'Z)-Rubixanthin α-Carotene β-Carotene (5'Z)-Rubixanthin δ-Carotene (5Z,9Z,5'Z,9'Z)-Lycopene γ-Carotene (5'Z)-Carotene (5Z,9Z,5'Z)-Lycopene (5Z,9Z)-Lycopene (all-E)-Lycopene	0.52 1.2 0.3 11.8 9.5 2.6 21.1 3 5.1 5.7 2.6 1.8 5.5 2.11 7.4 5.7 6.5 11 1.6 28.5 7.1 5 2 1 0.6 4 0.8 3.4 3 1.4 4.1 2 4.4 3.5 4.1 8.7
					[74]
			Orange		[98]

**Table 6.** Cont.

Results expressed in % of peak area of carotenoids in the HPLC chromatogram						
Flowers (four varieties)	Yellow-orange Lemon yellow Orange Dark orange	Neoxanthin Luteoxanthin + Auro Antheraxanthin Flavoxanthin Mutatoxanthin Lactucaxanthin Lutein Zeaxanthin Rubixanthin Lycopene $\gamma$ -carotene $\alpha$ -carotene $\beta$ -carotene	0.9–2.8 8.9–19.0 2.1–6.8 14.1–42.0 0.4–2.2 4.5–11.3 8.3–12.3 0.11–0.23 4.6–14.4 0.6–14.0 5.1–12.2 0.1–1.9 2.4–17.5	[75]		
Identification without quantification						
<i>Chrysanthemum morifolium</i>	Florist's daisy	Petals (different cultivars)	Yellow/White	Violaxanthin all-E-lutein $\beta$ -carotene 5,6-dihydro-5,6-dihydroxylutein 9Z-L, (9Z)-lutein 9'Z-L, (9'Z)-lutein 9Z-Le, (9Z)-lutein 5,6-epoxide 9'Z-Le, (9'Z)-lutein 5,6-epoxide Neoxanthin Zeaxanthin Antheraxanthin Zeaxanthin	—	[99]
<i>Cucurbita pepo</i>	Squash flower	Flowers	Yellow-orange	Flavoxanthin Cryptoxanthin	—	[100]
<i>Dianthus caryophyllus</i>	Carnation	Petals	Green/Red (during development)	Lutein and violaxanthin Zeaxanthin	Decreased Increased	[71]
<i>Taraxacum officinale</i>	Dandelion	Petals	Yellow	Isomers of lutein epoxide $\beta$ -Carotene Lycopene Xanthophyll	—	[101]
<i>Viola × wittrockiana</i>	Viola	Petals	—	—	[102]	

dw: dry weight, fw: fresh weight; tr: trace amounts. <sup>1</sup> Lutein equivalent.

#### 4. Oil Extraction

Until now, the studies performed on oil extraction of edible flowers have been performed for identification and characterization purposes only. To the best of our knowledge, there is no information on oil extraction from edible flowers for commercial purposes, only for essential oils (topic not discussed in the present review). Concerning the completed experimental studies, only solvent-based extractions are reported, and most of the studies involved hot-Soxhlet extraction or cold maceration. Edible flowers used in extraction can be prepared either from fresh or dried flowers. In case of dried samples, flowers are initially subjected to drying (e.g., air and freeze-drying), followed by grinding, milling, or homogenization to reduce sample particle size and enhance the extraction efficiency. Various solvents are commonly used in the Soxhlet extraction, such as petroleum ether [22,30,52,57] and hexane [51], whereas in maceration, a mixture of solvents is mainly used, such as chloroform-methanol [21,28,55]. However, in the future, new extraction technologies may be tested in edible flowers to improve extraction time, oil yield, and reduce the amount of solvent or even use green solvents. Based on its compositional data, edible flowers can be a potential source of fat and oil that are currently unexploited that could complement the existing sources. Furthermore, different edible flowers can offer a diversity of products that can be used for food flavoring and drink products.

#### 5. Conclusions

In general, fresh edible flowers show low nutrient content, including fat, because water is their main component. Nevertheless, their oils have an interesting composition from a health point of view, supporting an increased use for food purposes or as food supplements. The fatty acid profile of most flowers is rich in essential fatty acids and their vitamin E is mainly represented by  $\alpha$ -tocopherol—the vitamin E compound with the highest biological activity. Most carotenoids found in flowers are xanthophylls, such as lutein, although carotenes have also been reported (lycopene and  $\beta$ -carotene), all with interesting health attributes. Regarding different species and varieties of edible flowers, some variability in their lipid profiles and compositions has been observed.

In summary, and based on the available literature, it is evident that a wide gap still persists in the scientific knowledge regarding many edible flowers used for culinary and therapeutic purposes. The lipid composition of edible flowers merits further investigation to search for prospective food industry applications. This increased demand for edible flowers, as is or for oil extraction, will require a strong response from agriculture to increase productivity and quality.

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#### References

1. Lim, G.H.; Singhal, R.; Kachroo, A.; Kachroo, P. Fatty acid—And lipid-mediated signaling in plant defense. *Annu. Rev. Phytopathol.* **2017**, *55*, 505–536. [[CrossRef](#)] [[PubMed](#)]
2. Chapman, K.D.; Dyer, J.M.; Mullen, R.T. Biogenesis and functions of lipid droplets in plants—Thematic review series: Lipid droplet synthesis and metabolism: From yeast to man. *J. Lipid Res.* **2012**, *53*, 215–226. [[CrossRef](#)] [[PubMed](#)]
3. Velasco, L.; Goffman, F.D. Chemotaxonomic significance of fatty acids and tocopherols in Boraginaceae. *Phytochemistry* **1999**, *52*, 423–426. [[CrossRef](#)]

4. Szentmihályi, K.; Vinkler, P.; Lakatos, B.; Illés, V.; Then, M. Rose hip (*Rosa canina* L.) oil obtained from waste hip seeds by different extraction methods. *Bioresour. Technol.* **2002**, *82*, 195–201. [CrossRef]
5. Younis, Y.M.H.; Ghirmay, S.; Al-Shihry, S.S. African *Cucurbita pepo* L.: Properties of seed and variability in fatty acid composition of seed oil. *Phytochemistry* **2000**, *54*, 71–75. [CrossRef]
6. Rezig, L.; Chouaibi, M.; Msada, K.; Hamdi, S. Chemical composition and profile characterisation of pumpkin (*Cucurbita maxima*) seed oil. *Ind. Crops. Prod.* **2012**, *37*, 82–87. [CrossRef]
7. Yunus, I.S.; Cazenave-Gassiot, A.; Liu, Y.C.; Lin, Y.C.; Wenk, M.R.; Nakamura, Y. Phosphatidic acid is a major phospholipid class in reproductive organs of *Arabidopsis thaliana*. *Plant Signal. Behav.* **2015**, *10*, e1049790. [CrossRef] [PubMed]
8. Chalchat, J.C.; Ozcan, M.M. Comparative essential oil composition of flowers, leaves and stems of basil (*Ocimum basilicum* L.) used as herb. *Food Chem.* **2008**, *110*, 501–503. [CrossRef] [PubMed]
9. Shunying, Z.; Yang, Y.; Huaidong, Y.; Yue, Y.; Guolin, Z. Chemical composition and antimicrobial activity of the essential oils of *Chrysanthemum indicum*. *J. Ethnopharmacol.* **2005**, *96*, 151–158. [CrossRef] [PubMed]
10. Chamorro, E.R.; Ballerini, G.; Sequeira, A.F.; Velasco, G.A.; Zalazar, M.F. Chemical composition of essential oil from *Tagetes minuta* L. leaves and flowers. *J. Agric. Chem. Soc.* **2008**, *96*, 80–86.
11. Figueiredo, A.C.; Barroso, J.G.; Pais, M.S.S.; Scheffer, J.J.C. Composition of the essential oils from leaves and flowers of *Achillea millefolium* L. ssp. *Millefolium*. *Flavor Fragr. J.* **1992**, *7*, 219–222. [CrossRef]
12. Gazim, Z.C.; Rezende, C.M.; Fraga, S.R.; Svidzinski, T.I.E.; Cortez, D.A.G. Antifungal activity of the essential oil from *Calendula officinalis* L. (Asteraceae) growing in Brazil. *Braz. J. Microbiol.* **2008**, *39*, 61–63. [CrossRef] [PubMed]
13. Babu, K.G.D.; Singh, L.B.; Joshi, V.P.; Singh, V. Essential oil composition of Damask rose (*Rosa damascena* Mill.) distilled under different pressures and temperatures. *Flavour Fragr. J.* **2002**, *17*, 136–140.
14. Manning, R. Fatty acids in pollen: A review of their importance for honey bees. *Bee World* **2001**, *82*, 60–75. [CrossRef]
15. Patel, M.; Naik, S.N. Flowers of *Madhuca indica* J.F. Gmel, Present status and future perspectives. *Indian J. Nat. Prod. Resour.* **2010**, *1*, 438–443.
16. Mlcek, J.; Rop, O. Fresh edible flowers of ornamental plants—A new source of nutraceutical foods. *Trends Food Sci. Technol.* **2011**, *22*, 561–569. [CrossRef]
17. Rop, O.; Mlcek, J.; Jurikova, T.; Neugebauerova, J.; Vabkova, J. Edible flowers—A new promising source of mineral elements in human nutrition. *Molecules* **2012**, *17*, 6672–6683. [CrossRef] [PubMed]
18. Bastin, S. Edible Flowers. Available online: <http://www2.ca.uky.edu/hes/fcs/factshts/FN-SSB.025.pdf> (accessed on 2 August 2018).
19. Flora de Portugal Interactiva (2014), Sociedade Portuguesa de Botânica. Available online: <http://www.florapon.pt> (accessed on 10 September 2018).
20. Fernandes, L.; Casal, S.; Pereira, J.A.; Saraiva, J.A.; Ramalhosa, E. Edible flowers: A review of the nutritional, antioxidant, antimicrobial properties and effects on human health. *J. Food Compost. Anal.* **2017**, *60*, 38–50. [CrossRef]
21. Sánchez-Machado, D.I.; Núñez-Gastélum, J.A.; Reyes-Moreno, C.; Ramírez-Wong, B.; López-Cervantes, J. Nutritional quality of edible parts of *Moringa oleifera*. *Food Anal. Methods* **2010**, *3*, 175–180. [CrossRef]
22. Barros, L.; Carvalho, A.M.; Ferreira, I.C.F.R. Leaves, flowers, immature fruits and leafy flowered stems of *Malva sylvestris*: A comparative study of the nutraceutical potential and composition. *Food Chem. Toxicol.* **2010**, *48*, 1466–1472. [CrossRef] [PubMed]
23. Martins, D.; Barros, L.; Carvalho, A.M.; Ferreira, I.C.F.R. Nutritional and in vitro antioxidant properties of edible wild greens in Iberian Peninsula traditional diet. *Food Chem.* **2017**, *125*, 488–494. [CrossRef]
24. Hanif, R.; Iqbal, Z.; Iqbal, M.; Hanif, S.; Rasheed, M. Use of vegetables as nutritional food: Role in human health. *Res. J. Agric. Biol. Sci.* **2006**, *1*, 18–22.
25. Sotelo, A.; López-García, S.; Basurto-Peña, F. Content of nutrient and antinutrient in edible flowers of wild plants in Mexico. *Plant Foods Hum. Nutr.* **2007**, *62*, 133–138. [CrossRef] [PubMed]
26. Grzeszczuk, M.; Wesolowska, A.; Jadcak, D.; Jakubowska, B. Nutritional value of chive edible flowers. *Acta Sci. Pol. Hortorum Cultus* **2011**, *10*, 85–94.
27. González-Barrio, R.; Periago, M.J.; Luna-Recio, C.; Javier, G.-A.F.; Navarro-González, I. Chemical composition of the edible flowers, pansy (*Viola × wittrockiana*) and snapdragon (*Antirrhinum majus*) as new sources of bioactive compounds. *Food Chem.* **2018**, *252*, 373–380. [CrossRef] [PubMed]

28. Rao, G.N.; Rao, P.G.P.; Satyanarayana, A. Chemical, fatty acid, volatile oil composition and antioxidant activity of shade dried neem (*Azadirachta indica* L.) flower powder. *Int. Food Res. J.* **2014**, *21*, 807–813.
29. Vieira, P.M. Avaliação da Composição Química, dos Compostos Bioativos e da Atividade Antioxidante em seis Espécies de Flores Comestíveis. Master's Thesis, Universidade Estadual Paulista, São Paulo, Brazil, 2013.
30. Pires, T.C.S.P.; Dias, M.I.; Barros, L.; Ferreira, I.C.F.R. Nutritional and chemical characterization of edible petals and corresponding infusions: Valorization as new food ingredients. *Food Chem.* **2017**, *220*, 337–343. [CrossRef] [PubMed]
31. Miguel, M.; Barros, L.; Pereira, C.; Calhelha, R.C.; Garcia, P.A.; Castro, M.Á.; Santos-Buelga, C.; Ferreira, I.C.F.R. Chemical characterization and bioactive properties of two aromatic plants: *Calendula officinalis* L. (flowers) and *Mentha cervina* L. (leaves). *Food Funct.* **2016**, *7*, 2223–2232. [CrossRef] [PubMed]
32. Glew, R.H.; VanderJagt, D.J.; Lockett, C.; Grivetti, L.E.; Smith, G.C.; Pastuszyn, A.; Millson, M. Amino acid, fatty acid, and mineral composition of 24 indigenous plants of Burkina Faso. *J. Food Compos. Anal.* **1997**, *10*, 205–217. [CrossRef]
33. Guimarães, R.; Barros, L.; Carvalho, A.M.; Ferreira, I.C.F.R. Studies on chemical constituents and bioactivity of *Rosa micrantha*: An alternative antioxidants source for food, pharmaceutical, or cosmetic applications. *J. Agric. Food Chem.* **2010**, *58*, 6277–6284. [CrossRef] [PubMed]
34. Navarro-González, I.; González-Barrio, R.; García-Valverde, V.; Bautista-Ortíz, A.B.; Periago, M.J. Nutritional composition and antioxidant capacity in edible flowers: Characterization of phenolic compounds by HPLC-DAD-ESI/MS<sup>n</sup>. *Int. J. Mol. Sci.* **2015**, *16*, 805–822. [CrossRef] [PubMed]
35. WHO. *Diet, Nutrition and the Prevention of Chronic Diseases*; WHO Technical Report Series 916, Report of a Joint WHO/FAO Expert Consultation; World Health Organization: Geneva, Switzerland, 2003; p. 88.
36. Ricchi, M.; Odoardi, M.R.; Carulli, L.; Anzivino, C.; Ballestri, S.; Pinetti, A.; Fantoni, L.I.; Marra, F.; Bertolotti, M.; Barni, S.; et al. Differential effect of oleic and palmitic acid on lipid accumulation and apoptosis in cultured hepatocytes. *J. Gastroenterol. Hepatol.* **2009**, *24*, 830–840. [CrossRef] [PubMed]
37. Carvalho, I.S.; Teixeira, M.C.; Brodelius, M. Fatty acids profile of selected *Artemisia* spp. plants: Health promotion. *LWT-Food Sci. Technol.* **2011**, *44*, 293–298. [CrossRef]
38. French, M.A.; Sundram, K.; Clandinin, M.T. Cholesterolaemic effect of palmitic acid in relation to other dietary fatty acids. *Asia Pac. J. Clin. Nutr.* **2002**, *11*, S401–S407. [CrossRef] [PubMed]
39. Mancini, A.; Imperlini, E.; Nigro, E.; Montagnese, C.; Daniele, A.; Orrù, S.; Buono, P. Biological and nutritional properties of palm oil and palmitic acid: Effects on health. *Molecules* **2015**, *20*, 17339–17361. [CrossRef] [PubMed]
40. Boden, G.; Sargrad, K.; Homko, C.; Mozzoli, M.; Stein, T.P. Effect of a low-carbohydrate diet on appetite, blood glucose levels, and insulin resistance in obese patients with type 2 diabetes. *Ann. Intern. Med.* **2005**, *142*, 403–411. [CrossRef] [PubMed]
41. Peyron-Caso, E.; Taverna, M.; Guerre-Millo, M.; Veronese, A.; Pacher, N.; Slama, G.; Rizkalla, S.W. Dietary (n-3) polyunsaturated fatty acids up-regulate plasma leptin in insulin-resistant rats. *J. Nutr.* **2002**, *132*, 2235–2240. [CrossRef] [PubMed]
42. Simopoulos, A.P. Essential fatty acids in health and chronic disease. *Am. J. Clin. Nutr.* **1999**, *70*, 560S–569S. [CrossRef] [PubMed]
43. Tortosa-Caparrós, E.; Navas-Carrillo, D.; Marín, F.; Orenes-Piñero, E. Anti-inflammatory effects of omega 3 and omega 6 polyunsaturated fatty acids in cardiovascular disease and metabolic syndrome. *Crit. Rev. Food Sci. Nutr.* **2017**, *57*, 3421–3429. [CrossRef] [PubMed]
44. Mozaffarian, D.; Geelen, A.; Brouwer, I.A.; Geleijnse, J.M.; Zock, P.L.; Katan, M.B. Effect of fish oil on heart rate in humans: A meta-analysis of randomized controlled trials. *Circulation* **2005**, *112*, 1945–1952. [CrossRef] [PubMed]
45. Kang, M.J.; Shin, M.S.; Park, J.N.; Lee, S.S. The effects of polyunsaturated:saturated fatty acids ratios and peroxidisability index values of dietary fats on serum lipid profiles and hepatic enzyme activities in rats. *Br. J. Nutr.* **2005**, *94*, 526–532. [CrossRef] [PubMed]
46. HMSO, U.K. Department of Health. Nutritional aspects of cardiovascular disease. London. *Rep. Health Soc. Subj.* **1994**, *46*, 37–46.
47. Liu, L.; Hu, Q.; Wu, H.; Xue, Y.; Cai, L.; Fang, M.; Liu, Z.; Yao, P.; Wu, Y.; Gong, Z. Protective role of n6/n3 PUFA supplementation with varying DHA/EPA ratios against atherosclerosis in mice. *J. Nutr. Biochem.* **2016**, *32*, 171–180. [CrossRef] [PubMed]

48. Liu, H.Q.; Qiu, Y.; Mu, Y.; Zhang, X.J.; Liu, L.; Hou, X.H.; Zhang, L.; Xu, X.N.; Ji, A.L.; Cao, R.; et al. A high ratio of dietary n-3/n-6 polyunsaturated fatty acids improves obesity-linked inflammation and insulin resistance through suppressing activation of TLR4 in SD rats. *Nutr. Res.* **2013**, *33*, 849–858. [CrossRef] [PubMed]
49. Tsoupras, A.; Lordan, R.; Zabetakis, I. Inflammation, not cholesterol, is a cause of chronic disease. *Nutrients* **2018**, *10*, 604. [CrossRef] [PubMed]
50. Loizzo, M.R.; Pugliese, A.; Bonesi, M.; Tenuta, M.C.; Menichini, F.; Xiao, J.; Tundis, R. Edible flowers: A rich source of phytochemicals with antioxidant and hypoglycemic properties. *J. Agric. Food Chem.* **2016**, *64*, 2467–2474. [CrossRef] [PubMed]
51. Barnaby, A.G.; Reid, R.; Warren, D. Antioxidant activity, total phenolics and fatty acid profile of *Delonix regia*, *Cassia fistula*, *Spathodea campanulata*, *Senna siamea* and *Tibouchina granulosa*. *J. Anal. Pharm. Res.* **2016**, *3*, 2–7.
52. Ukiya, M.; Akihisa, T.; Yasukawa, K.; Kasahara, Y.; Kimura, Y.; Koike, K.; Nikaido, T.; Takido, M. Constituents of compositae plants. 2. Triterpene diols, triols, and their 3-o-fatty acid esters from edible Chrysanthemum flower extract and their anti-inflammatory effects. *J. Agric. Food Chem.* **2001**, *49*, 3187–3197. [CrossRef] [PubMed]
53. Dias, M.I.; Barros, L.; Alves, R.C.; Oliveira, M.B.P.P.; Santos-Buelga, C.; Ferreira, I.C.F.R. Nutritional composition, antioxidant activity and phenolic compounds of wild *Taraxacum* sect. Ruderalia. *Food Res. Int.* **2014**, *56*, 266–271. [CrossRef]
54. Matthäus, B.; Özcan, M.M. Chemical evaluation of flower bud and oils of tumbleweed (*Gundelia tournefortii* L.) as a new potential nutrition sources. *J. Food Biochem.* **2011**, *35*, 1257–1266. [CrossRef]
55. Meknia, M.; Flaminii, G.; Garra, M.; Hmida, R.B.; Cheraiefa, I.; Mastouri, M.; Hammamia, M. Aroma volatile components, fatty acids and antibacterial activity of four Tunisian *Punica granatum* L. flower cultivars. *Ind. Crops. Prod.* **2013**, *48*, 111–117. [CrossRef]
56. Barros, L.; Oliveira, S.; Carvalho, A.M.; Ferreira, I.C.F.R. In vitro antioxidant properties and characterization in nutrients and phytochemicals of six medicinal plants from the Portuguese folk medicine. *Ind. Crops Prod.* **2010**, *32*, 572–579. [CrossRef]
57. Villavicencio, A.L.C.H.; Heleno, S.A.; Calhelha, R.C.; Santos-Buelga, C.; Barros, L.; Ferreira, I.C.F.R. The influence of electron beam radiation in the nutritional value, chemical composition and bioactivities of edible flowers of *Bauhinia variegata* L. var. candida alba Buch.-Ham from Brazil. *Food Chem.* **2018**, *241*, 163–170. [PubMed]
58. Hastings, J.; Owen, G.; Dekker, A.; Ennis, M.; Kale, N.; Muthukrishnan, V.; Turner, S.; Swainston, N.; Mendes, P.; Steinbeck, C. ChEBI in 2016: Improved services and an expanding collection of metabolites. *Nucleic Acids Res.* **2015**, *44*, D1214–D1219. [CrossRef] [PubMed]
59. Mokrošnop, V.M. Functions of tocopherols in the cells of plants and other photosynthetic organisms. *Ukr. Biochem. J.* **2014**, *86*, 26–36. [CrossRef] [PubMed]
60. Murkovic, M.; Hillebrand, A.; Winkler, J.; Pfannhauser, W. Variability of vitamin E content in pumpkin seeds (*Cucurbita pepo* L.). *Z. Lebensm. Unters. Forsch.* **1996**, *202*, 275–278. [CrossRef] [PubMed]
61. Morales, P.; Carvalho, A.M.; Sánchez-Mata, M.C.; Cámera, M.; Molina, M.; Ferreira, I.C.F.R. Tocopherol composition and antioxidant activity of Spanish wild vegetables. *Genet. Resour. Crops. Evol.* **2012**, *59*, 851–863. [CrossRef]
62. Health and Medicine Division the National Academies (Former IOM). *Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids*; National Academy Press: Washington, DC, USA, 2000.
63. European Food Safety Authority (EFSA). Opinion on mixed tocopherols, tocotrienol tocopherol and tocotrienols as sources for vitamin E added as a nutritional substance in food. Scientific opinion of the panel on food additives, flavourings, processing aids and materials in contact with food. *EFSA J.* **2008**, *640*, 1–34.
64. Nambara, E.; Marion-Poll, A. Abscisic acid biosynthesis and catabolism. *Annu. Rev. Plant Biol.* **2005**, *56*, 165–185. [CrossRef] [PubMed]
65. Green, B.R.; Durnford, D.G. The chlorophyll-carotenoid proteins of oxygenic photosynthesis. *Annu. Rev. Plant Biol.* **1996**, *47*, 685–714. [CrossRef] [PubMed]
66. Niyogi, K. Safety valves for photosynthesis. *Curr. Opin. Plant Biol.* **2000**, *3*, 455–460. [CrossRef]
67. Auldrige, M.E.; McCarty, D.R.; Klee, H.J. Plant carotenoid cleavage oxygenase and their apocarotenoid products. *Curr. Opin. Plant Biol.* **2006**, *9*, 315–332. [CrossRef] [PubMed]

68. Mayne, S.T. β-Carotene, carotenoids and disease prevention in humans. *FASEB J.* **1996**, *10*, 690–701. [[CrossRef](#)] [[PubMed](#)]
69. Kamalambigeswari, R.; Rebecca, L.J. Extraction of major carotenoids from flower petals. *Int. J. Pharm. Sci. Rev. Res.* **2016**, *39*, 37–39.
70. Britton, G. Chapter 10: Functions of intact carotenoids. In *Carotenoids: Natural Functions*; Britton, G., Liaaen-Jensen, S., Pfander, H., Eds.; Birkhäuser: Berlin, Germany, 2008; Volume 4, pp. 189–221.
71. Ohmiya, A.; Tanase, K.; Hirashima, M.; Yamamizo, C.; Yagi, M. Analysis of carotenogenic gene expression in petals and leaves of carnation (*Dianthus caryophyllus* L.). *Plant Breed.* **2013**, *132*, 423–429. [[CrossRef](#)]
72. Singh, G.; Kawatra, A.; Sehgal, S. Nutritional composition of selected green leafy vegetables, herbs and carrots. *Plant Food. Hum. Nutr.* **2001**, *56*, 359–364. [[CrossRef](#)]
73. Tlili, N.; Nasri, N.; Saadaoui, E.; Khaldi, A.; Triki, S. Carotenoid and Tocopherol Composition of Leaves, Buds, and Flowers of *Capparis spinosa* grown wild in Tunisia. *J. Agric. Food Chem.* **2009**, *57*, 5381–5385. [[CrossRef](#)] [[PubMed](#)]
74. Bakó, E.; Deli, J.; Tóth, G. HPLC study on the carotenoid composition of Calendula products. *J. Biochem. Biophys. Methods* **2002**, *53*, 241–250. [[CrossRef](#)]
75. Pintea, A.; Bele, C.; Andrei, S.; Socaciu, C. HPLC analysis of carotenoids in four varieties of *Calendula officinalis* L. flowers. *Acta Biol. Szeged.* **2003**, *47*, 37–40.
76. Seroczyńska, A.; Korzeniewska, A.; Sztangret-Wiśniewska, J.; Niemirowicz-Szczytt, K.; Gajewski, M. Relationship between carotenoids content and flower or fruit flesh colour of winter squash (*Cucurbita maxima* Duch.). *Folia Hortic.* **2006**, *18*, 51–61.
77. Park, C.H.; Chae, S.C.; Park, S.-Y.; Kim, J.K.; Kim, Y.J.; Chung, S.O.; Arasu, M.V.; Al-Dhabi, N.A.; Park, S.U. Anthocyanin and carotenoid contents in different cultivars of chrysanthemum (*Dendranthema grandiflorum* Ramat.) flower. *Molecules* **2015**, *20*, 11090–11102. [[CrossRef](#)] [[PubMed](#)]
78. Kelley, K.M.; Behe, B.K.; Biernbaum, J.A.; Poff, K.L. Consumer preference for edible flower color, container size, and price. *HortScience* **2001**, *36*, 801–804.
79. Kelley, K.M.; Behe, B.K.; Biernbaum, J.A.; Poff, K.L. Combinations of colors and species of containerized edible flowers: Effect on consumer preferences. *HortScience* **2002**, *37*, 218–221.
80. Ching, L.S.; Mohamed, S. Alpha-tocopherol content in 62 edible tropical plants. *J. Agric. Food Chem.* **2001**, *49*, 3101–3105. [[CrossRef](#)] [[PubMed](#)]
81. Tlili, N.; Khaldi, A.; Triki, S.; Munné-Bosch, S. Phenolic compounds and vitamin antioxidants of caper (*Capparis spinosa*). *Plant Food Hum. Nutr.* **2010**, *65*, 260–265. [[CrossRef](#)] [[PubMed](#)]
82. Bona, G.S.; Boschetti, W.; Bortolin, R.C.; Vale, M.G.R.; Moreira, J.C.F.; Rios, O.A.; Flôres, S.H. Characterization of dietary constituents and antioxidant capacity of *Tropaeolum pentaphyllum* Lam. *J. Food Sci. Technol.* **2017**, *54*, 3587–3597. [[CrossRef](#)] [[PubMed](#)]
83. Roriz, C.L.; Barros, L.; Carvalho, A.M.; Ferreira, I.C.F.R. HPLC-Profiles of tocopherols, sugars, and organic acids in three medicinal plants consumed as infusions. *Int. J. Food Sci.* **2014**. [[CrossRef](#)] [[PubMed](#)]
84. Sausserde, R.; Kampuss, K. Composition of carotenoids in calendula (*Calendula officinalis* L.) flowers. In Proceedings of the 9th Baltic Conference on Food Science and Technology “Food for Consumer Well-Being”, Jelgava, Latvia, 9 May 2014; pp. 13–18.
85. Petrova, I.; Petkova, N.; Ivanov, I. Five Edible flowers—valuable source of antioxidants in human nutrition. *Int. J. Phytochem. Res.* **2016**, *8*, 604–610.
86. Toiu, A.; Benedec, D.; Duda, M.; Hangau, D.; Oniga, I. Determination of total carotenoid content in some *Calendula officinalis* and *Tagetes patula* varieties. *Hop Med. Plant* **2016**, *24*, 57–62.
87. Kishimoto, S.; Sumitomo, K.; Yagi, M.; Nakayama, M.; Ohmiya, A. Three routes to orange petal color via carotenoid components in 9 compositae species. *J. Jpn. Soc. Horitic. Sci.* **2007**, *76*, 250–257. [[CrossRef](#)]
88. Aquino-Bolaños, E.N.; Urrutia-Hernández, T.; Castillo-Lozano, M.L.; Chavéz-Servia, J.; Verdálet-Guzmán, I. Physicochemical parameters and antioxidant compounds in edible squash (*Cucurbita pepo*) flower stored under controlled atmospheres. *J. Food Qual.* **2013**, *36*, 302–308. [[CrossRef](#)]
89. Trivellini, A.; Vernieri, P.; Ferrante, A.; Serra, G. Physiological characterization of flower senescence in long life and ephemeral Hibiscus (*Hibiscus rosa-sinensis* L.). *Acta Hortic.* **2007**, *755*, 457–464. [[CrossRef](#)]
90. Telesiński, A.; Grzeszczuk, M.; Jadcak, D.; Zakrzewska, H. Fluoride content and biological value of flowers of some chamomile (*Matricaria recutita* L.) cultivars. *J. Elem.* **2012**, *703*–712.

91. Murakami, Y.; Fukui, Y.; Watanabe, H.; Kokubun, H.; Toya, Y.; Ando, T. Distribution of carotenoids in the flower of non-yellow commercial petunia. *J. Hortic. Sci. Biotechnol.* **2003**, *78*, 127–130. [[CrossRef](#)]
92. Tinoi, J.; Rakariyatham, N.; Deming, R.L. Determination of major carotenoid constituents in petal extracts of eight selected flowering plants in the north of Thailand. *Chiang Mai J. Sci.* **2006**, *33*, 327–334.
93. Prata, G.G.B.; Souza, K.O.; Lopes, M.M.A.; Oliveira, L.S.; Aragao, F.A.S.; Alves, R.E.; Silva, S.M. Nutritional characterization, bioactive compounds and antioxidant activity of Brazilian roses (*Rosa* spp.). *J. Agric. Sci. Technol.* **2017**, *19*, 929–941.
94. Komes, D.; Belščak-Cvitanović, A.; Horžić, D.; Marković, K.; Kovačević, G.K. Characterisation of pigments and antioxidant properties of three medicinal plants dried under different drying conditions. In Proceedings of the 11th International Congress on Engineering and Food, Atenas, Costa Rica, 22–26 May 2011.
95. Hansmann, P.; Sitte, P. Composition and molecular structure of chromoplast globules of *Viola tricolor*. *Plant Cell Rep.* **1982**, *1*, 111–114. [[CrossRef](#)] [[PubMed](#)]
96. Niizu, P.Y.; Rodriguez-Amaya, D.B. Flowers and leaves of *Tropaeolum majus* L. as rich sources of lutein. *J. Food Sci.* **2005**, *70*, S605–S609. [[CrossRef](#)]
97. Toiu, A.; Muntean, E.; Oniga, I.; Tămaş, M. Pharmacognostic research on *Viola declinata* Waldst. et Kit. (Violaceae). *Farmacia* **2009**, *57*, 218–222.
98. Kishimoto, S.; Maoka, T.; Sumitomo, K.; Ohmiya, A. Analysis of carotenoid composition in petals of calendula (*Calendula officinalis* L.). *Biosci. Biotechnol. Biochem.* **2005**, *69*, 2122–2128. [[CrossRef](#)] [[PubMed](#)]
99. Kishimoto, S.; Ohmiya, A. Regulation of carotenoid biosynthesis in petals and leaves of chrysanthemum (*Chrysanthemum morifolium*). *Physiol. Plant.* **2006**, *128*, 436–447. [[CrossRef](#)]
100. Azimova, S.S.; Glushenkova, A.I. *Lipids, Lipophilic Components and Essential Oils from Plant Sources*; Springer: New York, NY, USA, 2012; p. 307.
101. Meléndez-Martínez, A.J.; Britton, G.; Vicario, I.M.; Heredia, F.J. HPLC analysis of geometrical isomers of lutein epoxide isolated from dandelion (*Taraxacum officinale* F. Weber ex Wiggers). *Phytochemistry* **2006**, *67*, 771–777. [[CrossRef](#)] [[PubMed](#)]
102. Gamsjaeger, S.; Baranska, M.; Schulz, H.; Heiselmayer, P.; Musso, M. Discrimination of carotenoid and flavonoid content in petals of pansy cultivars (*Viola × wittrockiana*) by FT-Raman spectroscopy. *J. Raman Spectrosc.* **2011**, *42*, 1240–1247. [[CrossRef](#)]



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