



Targeting Peripheral N-Methyl-D-Aspartate Receptor (NMDAR): A Novel Strategy for the Treatment of Migraine

Veberka Kalatharan 🔟 and Mohammad Al-Mahdi Al-Karagholi *🔟

Danish Headache Center, Department of Neurology, Rigshospitalet Glostrup, Faculty of Health and Medical Sciences, University of Copenhagen, Valdemar Hansens Vej 5, DK-2600 Glostrup, Denmark * Correspondence: mahdi.alkaragholi@gmail.com; Tel.: +45-31-19-16-47

Abstract: Backgrounds: Several acute and preventive medications were developed for the treatment of migraine. Yet, a significant proportion of patients reports an inadequate response and a lack of tolerability, emphasizing the need for new options. Glutamate is the most important excitatory neurotransmitter in the brain, and glutamate receptors including N-Methyl-D-Aspartate Receptor (NMDAR) are expressed at several levels of the trigeminovascular system, which is the anatomical and physiological substrate of migraine pain. Objective: To review preclinical and clinical studies investigating the role of the NMDAR in migraine pathophysiology. Methods: No protocol was registered for this study. References for the present review were identified from a narrative search of the PubMed database. Search terms such as glutamate, migraine, N-Methyl-D-Aspartate Receptor, and NMDAR were used. No restrictions were made in terms of the language and date of publication. Results: In animal models, administration of monosodium glutamate (MSG) activated and sensitized trigeminovascular neurons. In healthy human participants, consumption of MSG caused headaches, craniofacial sensitivity, and nausea. In in vivo models and through immunolabeling, NMDAR subunits NR1, NR2A, and NR2B were expressed in trigeminal ganglion neurons. In humans, NMDAR antagonists such as ketamine and memantine caused a significant reduction in pain intensity and monthly headache frequency. Conclusions: Accumulative evidence indicates that NMDAR is a promising new target for the treatment of migraine. Selective NMDAR antagonists without central effects are needed to investigate their therapeutic benefit in the treatment of migraine.

Keywords: glutamate; headache; pain; aura; CSD

1. Introduction

Migraine is a primary headache disorder, affecting more than 15% of the global adult population in their most productive years of life with a health and economic burden of billions of dollars globally [1]. The clinical manifestation of migraine is recurrent attacks with a severe, and usually unilateral and throbbing headache, lasting 4–72 h and associated with nausea and/or light and sound sensitivity [2]. In one-third of individuals with migraine, the headache phase is preceded by transient focal neurological disturbances, the so-called migraine aura-phase [3], whose underlying mechanism is considered to be cortical spreading depression (CSD) [4]. CSD is a self-sustaining, slowly propagating wave of transient neuronal and glial depolarization [3] that silences brain and electrical activity for several minutes [5–7]. In the past three decades, advances made in migraine research led to the development of several acute and preventive medications. Yet, a significant proportion of patients reports an inadequate response and a lack of tolerability [8].

Glutamate is the principal excitatory neurotransmitter in the central nervous system [9], and its role in migraine pathophysiology has been under the spotlight for more than three decades [10–13]. Glutamate receptors are pharmacologically classified as ionotropic and metabotropic receptors. N-Methyl-D-Aspartate Receptor (NMDAR) is an ionotropic glutamate receptor [9] that modulates excitatory neurotransmission by conducting sodium (Na⁺)



Citation: Kalatharan, V.; Al-Karagholi, M.A.-M. Targeting Peripheral N-Methyl-D-Aspartate Receptor (NMDAR): A Novel Strategy for the Treatment of Migraine. J. Clin. Med. 2023, 12, 2156. https://doi.org/10.3390/jcm12062156

Academic Editors: Filippo Baldacci and Ciro De Luca

Received: 23 December 2022 Revised: 23 February 2023 Accepted: 2 March 2023 Published: 10 March 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). and calcium (Ca^{2+}) ions into the cell and potassium (K^+) ions outside the cell. Key structures related to migraine pain, including the trigeminal ganglion (TG), trigeminal nucleus caudalis (TNC), and thalamus contain high densities of NMDAR-positive neurons, pinpointing a potential connection between the NMDAR and migraine pathophysiology [9,12,14–16].

Here, we provide a concise overview of the molecular structure and physiological function of NMDAR, highlight recent mechanistic insights into the role of this receptor in migraine pathophysiology, and discuss the possible complementarity and interdependence of signaling pathways involved in migraine attack initiation.

2. Methods

No protocol was registered for this study. References for the present review were identified through a narrative search of the PubMed database regarding glutamate and migraine on 27th of November 2022. The search terms "glutamate signaling AND migraine", "migraine AND glutamate AND target", "Migraine AND glutamate", "Migraine AND glutamate and "Migraine AND glutamate" and "Migraine AND NMDAR" were used. No restrictions were made in terms of the language and date of publication. Additionally, references from relevant articles were identified. The final reference list was generated based on relevance to the topic by reading the title and abstract.

3. Ion Channels in Migraine Pathophysiology

The origin of migraine pain involves activation and sensitization of trigeminal afferents innervating meningeal and large pial arteries, denoted as the trigeminovascular system (TVS) [2]. First-order neurons in the TG project nociceptive signals to second-order central neurons in the TNC [17] and axons from their relay to multiple nuclei in the brainstem, hypothalamus, and thalamus [18]. Using neuroimaging in spontaneous human migraine attacks, the hypothalamus was activated during the prodromal symptoms (changes in attention and concentration, mood swings, tiredness, frequent yawning, craving certain foods, and multiple others) which occur hours to days before headache onset in up to 90% of individuals with migraine [19]. Whether migraine pain is of peripheral provenance or whether hypothalamus and thalamus network to integrate peripheral influences with central mechanisms, facilitating the precipitation of migraine pain is yet to be elucidated.

The human genome encodes at least 400 ion channel family members, which represent the second largest class of membrane proteins for drug discovery after G-protein-coupled receptors (GPCRs) [20]. Yet, ion channels remain considerably under-exploited as therapeutic targets [21]. Ion channels are expressed in migraine-related structures including cranial arteries, where they principally regulate the vascular tone. Dysfunction or abnormal regulation of ion channels might result in disruption of the excitation–inhibition balance, neuronal excitability, and peripheral and central sensitization [20,22]. Balance control impairment in neuronal function causes a heterogeneous group of disorders called channelopathies and includes epilepsy, episodic ataxia, and familial hemiplegic migraine (FHM) [23]. These findings and the observation that migraine is more prevalent in individuals with channelopathies including epilepsy and episodic ataxia [24] indicate that migraine might be a channelopathy disease.

4. Glutamate and Migraine

The role of glutamate amino acid in migraine has been investigated in human models in the past three decades. Glutamate plays a role in neurogenic inflammation and excitation of the trigeminal pain pathway. All classes of glutamate receptors are located in superficial laminae I and II of the trigeminocervical complex (TCC) [12,18], which forms the base of the complex interaction and modulation of neuronal pain signals. Moreover, extracellular glutamate concentrations increase during CSD [25,26], and this increase further charges CSD and facilitates its propagation by activating NMDARs [27–29]. Ferrari and colleagues showed that individuals with migraine with aura had higher plasma glutamate levels compared to healthy participants, individuals with migraine without aura, and individuals with tension-type headache [10]. Plasma glutamate levels were further increased during attacks in individuals with migraine with and without aura [10]. D'Andrea and colleagues showed that, platelet glutamate levels (platelet dense granules carry a considerable amount of glutamate) were significantly higher in individuals with migraine with aura compared to healthy participants and other headache groups, such as individuals with migraine without aura, or with tension headaches and cluster headaches [30].

Additional studies supported that individuals with migraine with and without aura had higher plasma glutamate levels compared to healthy participants [31,32]. Interestingly, migraine preventive therapies were found to reduce plasma glutamate levels [32]. The involvement of glutamate signaling in headache and migraine was further reinforced by the finding that (1) the consumption of a single dose (150 mg/kg) of monosodium glutamate (MSG) caused headache, craniofacial sensitivity, and nausea in healthy participants [33], and (2) repeated MSG intake (150 mg/kg) for five daily sessions for one-week reduced pressure pain thresholds and caused headache in healthy participants compared to placebo [34].

5. N-Methyl-D-Aspartate Receptor (NMDAR)

Glutamate receptors are pharmacologically classified as ionotropic (ligand-gated ion channels) and metabotropic (GPCRs). NMDAR along with alpha-amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid receptor (AMPAR) and Kainate receptor (KAR) are categorized as ionotropic glutamate receptors [9]. NMDAR modulates excitatory neuro-transmission by conducting Na⁺ and Ca²⁺ ions into the cell and K⁺ ions outside the cell (Figure 1).

The NMDAR is a heterotetrameric complex assembled from three different NMDAR subunits (NR1, NR2, and NR3) [35] (Figure 2). The NR1 subunit is essential for a functional NMDAR complex [35,36] and is represented by eight splice variants, which influence the channel properties. Four NR2 subunits (NR2A, NR2B, NR2C and NR2D) and two NR3 subunits (NR3A and NR3B) have been mapped. The NR1-NR2A heterodimer suggests a mechanism for ligand-induced ion channel opening and is the functional unit in tetrameric NMDARs [37]. Table 1 presents the predominant localization of NMDAR subunits. All NM-DAR subunits share a common membrane structure characterized by a large N-terminus domain (NTD), an agonist/ligand binding domain (ABD/LBD), a transmembrane region (TMD), and a cytoplasmic region (CTD) (Figure 2) [35]. NMDAR activation requires the simultaneous binding of glycine and glutamate, together with the removal of the endogenous channel-pore blocker, magnesium ion (Mg^{2+}) , in a voltage-dependent manner [38,39]. The particular importance of the NMDAR is due to the high permeability to Ca²⁺ ions that gives NMDARs a significant role in both synaptic plasticity under physiological conditions and neuronal death under excitotoxic pathological conditions [35]. A typical NMDAR complex consists of two glycine-binding NR1 subunits and two glutamate-binding NR2 subunits [36]. Incorporating either NR3A or NR3B subunits showed decreased channel activity via reduced single-channel conductance, reduced Ca²⁺ permeability and increased Mg^{2+} blockade [35,40–42]. Notably, the NR3 subunit can bind to glycine rather than glutamate, which is similar to the NR1 subtype. Thus, a novel type of NMDAR complex composed of NR1 and NR3 subunits would only require glycine and not glutamate for activation [43]. The entry of Ca^{2+} into dendritic spines through NMDARs is essential for long-term potentiation (LTP) and long-term depression (LTD) [44]. LTP is induced by a high-frequency synaptic activity that causes postsynaptic membrane depolarization, a decrease in voltage-dependent Mg²⁺ blockage of the NMDAR pore, and a massive entry of Ca^{2+} ions into dendritic spines leading to calmodulin (CaM) and CaM-dependent kinase II activation (Figure 1).



Figure 1. Tripartite signaling of N-Methyl-D-Aspartate Receptor (NMDAR). This model illustrates three parallel signaling pathways of NMDAR transduction. Binding of glutamate and glycine results in ionotropic function and membrane depolarization through efflux of K⁺ ions and influx of Na⁺ and Ca²⁺ ions mediating downstream Ca²⁺-dependent pathways. NMDAR signaling can also happen non-ionotropically [45] when glycine or glutamate bind independently and induce conformational changes and downstream protein interactions. Glycine stimulation enhances NMDAR association with the AP2 and subsequent activation of a downstream endocytic pathway. Lastly, glutamate binding can result in LTD through p38 MAPK and excitotoxicity though an activation of Src. AP2 = endocytic adaptor protein 2, CaMKII = calcium/calmodulin kinase II, LTD = long-term depression, LTP = long-term potentiation, p38 MAPK = p38 mitogen-activated protein kinase, PP1 = protein phosphatase 1, Rap = ras-related protein, Src = thyrosin kinase family.



 Table 1. Predominant localization of the NMDAR subunits.

sites that either work as an NMDAR agonist or antagonist.

NMDAR Subtype	Predominant Localization	References
NR1	Present throughout CNS.	[9]
NR2A	Thalamus; prominently in the lateral thalamic nuclei.	[9]
NR2B	Thalamus.	[9]
NR2C	High expression in cerebellum, low elsewhere.	[9]
NR2D	High expression early during development, low expression in adults.	[9]
NR3A	Spinal cord, thalamus, hypothalamus, brainstem, hippocampus (CA1), amygdala, and certain parts of cortex cerebri.	[40,46]
NR3B	In adults: expressed in somatic motor neurons in the brainstem and spinal cord, cerebellum, and hippocampus. The distribution of NR3B seems to be as ubiquitous as NR1.	[40-42,47]

5.1. Evidence Implicating NMDAR in Migraine

NMDARs are expressed in trigeminovascular neurons (trigeminal neurons innervating dural blood vessels) [47]. Glutamate transporter (excitatory amino acid transporter-2 (*EAAT-2*)) was identified in dural blood vessels [48]. In rat models, the expression of NM-DAR subunits NR1, NR2A, and NR2B have been demonstrated in TG [49–52]. Specifically,

Receptor (NMDAR). The NMDAR is assembled as tetramers; two NR1 and two NR2 subunits forming a dimer–dimer structure or two NR1 subunits associating with one NR2 and NR3 subunit. The channel construction with different domains gives rise to ligand binding or allosteric modulatory

the NR2B subunit was expressed in nerve fibers that innervated dural blood vessels [53]. In a recent study, Guerrero-Toro and colleagues investigated the role of peripherally released glutamate in trigeminal nociception by using TG neurons from rodent models. The authors revealed (1) the presence of NR2A and NR2B subunits in TG neurons through immunolabeling, (2) the expression of other glutamate receptors in TG neurons, (3) the presence of calcitonin gene-related peptide (CGRP) enhanced the fraction of NMDARpositive neurons, and (4) the removal of Mg^{2+} increased the nociceptive firing of trigeminal afferents by NMDAR in in vivo conditions [54]. Moreover, NMDAR activation mediated nociceptive transmission in the TNC [55], and the stimulation of dural structures caused a co-release of glutamate and CGRP in the TCC [56]. In rats, intravenous administration of 50 mg/kg MSG activated and sensitized the trigeminovascular neurons associated with dural vasodilation. Co-administration of a peripherally restricted NMDAR antagonist, (2R)-amino-5-phosphonovaleric (APV) attenuated the MSG-induced effects [53]. Glutamate is a negatively charged, polar amino acid. The central uptake of glutamate and other anionic excitatory amino acids from the peripheral circulation is limited by the blood-brain barrier (BBB). At physiologic plasma concentrations, glutamate flux from plasma into the brain is mediated by a high-affinity transport system at the BBB. Efflux from the brain back into plasma appears to be driven in large part by a Na⁺-dependent active transport system at the capillary abluminal membrane. The glutamate concentration in brain interstitial fluid is only a fraction of that of plasma and is maintained fairly independently of small fluctuations in plasma concentration [57]. Thus, glutamate-induced headache is likely due to peripheral activation of NMDAR, indicating that targeting peripheral NMDAR might be a possible strategy for the treatment of individuals with migraine to avoid side effects due to central action.

5.2. NMDAR Antagonists in Preclinical Studies

Based on the knowledge about NMDAR subunits and their distinctive domains, several potential therapeutic approaches such as allosteric modulators, and competitive and non-competitive antagonists have been proposed for anti-migraine therapy (Figure 2). The negative allosteric modulators of NMDAR include the endogenous zinc ion (Zn^{2+}) and ifenprodil-like compounds [35,58], which both act on the NR2 NTD as non-competitive antagonists. Zn²⁺ has been shown to be a selective antagonist of the subunits NR2A and NR2B, whereas ifenprodil and its derivatives are selective antagonists of NR2B with more than a 100-fold higher affinity [59]. The pore domain of the NMDAR forms binding sites for non-competitive pore blockers such as the endogenous Mg²⁺ and exogenous compounds such as MK-801 (also known as dizocilpine), memantine, and ketamine. Memantine and Mg²⁺ are low-affinity NMDAR blockers with faster blocking and unblocking kinetics, which are linked to fewer and less severe side effects than substances with slower receptor kinetics such as ketamine and MK-801 [38]. The administration of dizocilpine maleate, an NDMAR antagonist, resulted in a substantial blockade of neuronal firing and the inhibition of trigeminovascular-evoked responses in the trigeminovascular complex in cats [47]. Apart from being involved in the trigeminal pain pathway, the NMDAR seems to be implicated in the initiation and propagation of CSD, which is believed to be the pathophysiological mechanism underlying migraine aura [5,55,60]. Mg²⁺ selectively suppressed glutamate-induced CSD [61], and MK-801 (non-specific NMDAR antagonist) and ifenprodil (NR2B selective antagonist) inhibited experimental-induced CSD [27,60,62,63] (Box 1). In 2019, Hoffmann and colleagues investigated the efficacy of Mg²⁺ and memantine in rat models by comparing microiontophoretic application and systemic administration to modulate trigeminovascular nociception. They observed that intravenous systemic administration of moderate concentrations of low-affinity NMDAR channel blockers such as Mg²⁺ and memantine does not inhibit stimulus-evoked firing significantly compared to microiontophoretic application [38].

Box 1. Preclinical evidence.

- 1. Glutamate N-Methyl-D-Aspartate Receptor (NMDAR) is expressed in nerve fibers innervating dural blood vessels. Glutamate transporter (excitatory amino acid transporter-2 (*EAAT-2*)) is expressed in dural blood vessels [48].
- In rats, intravenous administration of 50 mg/kg monosodium glutamate (MSG) activated and sensitized trigeminovascular neurons associated with dural vasodilation. Co-administration of a peripherally restricted NMDAR antagonist (2R)-amino-5-phosphonovaleric (APV) attenuated MSG-induced effects [53].
- 3. Intraperitoneal injections of 1000 mg/kg MSG caused headache-like and nausea symptoms in association with an increase in plasma calcitonin-gene related peptide (CGRP) concentration. MSG-induced headache-like symptoms were inhibited by APV and partly attenuated by olcegepant, a selective CGRP receptor antagonist [64].
- 4. Administration of dizocilpine maleate, a NDMAR antagonist, and GYKI 52466, a non-NMDAR antagonist, resulted in a substantial showed blockade of neuronal firing and inhibition of trigeminovascular-evoked responses in the trigeminovascular complex in cats [47].

5.3. NMDAR Antagonists in Clinical Studies

NMDAR antagonists such as Mg²⁺, ketamine, and memantine have been investigated for migraine treatment and prophylaxis, respectively [65]. Administration of Mg²⁺ has been examined in numerous clinical trials either perorally or intravenously [66]. In a prospective quasi-experimental study, Baratloo and colleagues found that intravenous magnesium sulfate might be superior to intravenous caffeine for the acute management of migraine headaches [67]. A double-blinded, cross-over multicenter pilot study reported a significant reduction in the incidence of migraine attacks when administering peroral triagnesium dietrate compared to placebo [68] (Table 2). Additionally, the administration of transdermal MgCl₂ solution is described as an ideal approach in acute migraine treatment due to the immediate absorption through the skin [66,69]. Collectively, Mg²⁺ seems to have beneficial effect for the acute and preventive treatment of migraine.

Table 2. Clinical studies investigating NMDAR antagonists.

NMDAR Antagonist	Author and Year	Study Design	Number of Participants	Dose (Administration Form)	Type of migraine (With or Without Aura)	Findings
Magnesium	Baratloo A, et. al. (2017) [67]	Prospective quasi- experimental study	Total $n = 70$ Completed study $n = 70$	2 g Magnesium Sulfate (intravenous)	Individuals with migraine (Aura not reported)	Intravenous magnesium sulfate (2g) might be superior to intravenous caffeine (60 mg) for the short-term management of migraine headaches.
Magnesium	K. Taubert, et. al. (1994) [68]	Double- blinded, cross-over multicenter pilot study	Total $n = 43$ Completed study $n = 43$	600 mg/day Trimagnesium Dieitrate (oral)	Individuals with migraine (Aura not reported)	Significant reduction in the incidence of migraine attacks was observed.
Ketamine	Nicolodi et. al. (1995) [70]	Randomized double- blinded, cross-over study	Total $n = 34$ Completed study $n = 34$	80 μg/kg Ketamine hydrochloride (subcutaneous)	Individuals with migraine (Aura not reported)	Compared to the placebo group, Individuals in the ketamine group experienced a marked relief of pain both as an acute and prophy- lactic treatment.

NMDAR Antagonist	Author and Year	Study Design	Number of Participants	Dose (Administration Form)	Type of migraine (With or Without Aura)	Findings
Ketamine	Kaube et. al. (2000) [71]	Open label study	Total $n = 11$ Completed study $n = 11$	25 mg (Intranasal)	Familial hemiplegic migraine	5 out of 11 participants experienced a reduction in severity and duration of migraine attacks after ketamine administration, whereas 6 out of 11 participants had no beneficial effect.
Ketamine	Afridi et. al. (2013) [72]	Randomized double-blinded placebo- controlled trial	Total $n = 30$ Completed study $n = 18$	25 mg (Intranasal)	Individuals with migraine (With aura)	Intranasal ketamine is effective at reducing aura severity in individuals with prolonged aura.
Ketamine	Lauritsen et. al. (2016) [73]	Retrospective study	Total $n = 6$ Completed study $n = 6$	0.1 mg/kg (Intravenous)	Refractory migraine	6 out of 6 participants obtained the target pain relief endpoint after infusion of ketamine. The mean ketamine infusion rate at the time of pain relief was 0,34 mg/kg/hour.
Ketamine	Etchinson et. al. (2018) [74]	Randomized double-blinded placebo- controlled trial	Total $n = 34$ Completed study $n = 34$	0.2 mg/kg (Intravenous)	Individuals with migraine (With and without aura)	The difference in NRS pain scores was neither statistically nor clinically significant between ketamine group (n = 16) and placebo group $(n = 18)$.
Memantine	Charles et. al. (2007) [65]	Retrospective study	Total $n = 60$ Completed study $n = 54$	5 mg to 20 mg/day (Peroral)	Individuals with migraine (With and without aura)	67% reported a greater than 50% reduction in estimated monthly headache frequency.
Memantine	Bigal et. al. (2008) [75]	Prospective, open label study	Total $n = 38$ Completed study $n = 23$	10 mg to 20 mg/day (Peroral)	Refractory migraine	A significant reduction in headache frequency, severity, and MIDAS score was observed after 3 months of treatment.
Memantine	Noruzzadeh et. al. (2016) [76]	Randomized double-blinded placebo- controlled trial	Total $n = 52$ Completed study $n = 52$	10 mg/day (Peroral)	Individuals with migraine (Without aura)	Individuals in the memantine group $(n = 25)$ showed significantly greater reduction in monthly headache attack frequency and headache severity than the placebo group $(n = 27)$.

Table 2. Cont.

Several studies have investigated the use of ketamine or AMPA (LY293558; BGG492) antagonists as abortive therapies in migraine with aura and familial hemiplegic migraine [71,73,77]. Although most of these studies concluded that these drugs possessed limited or at best moderate efficacy in the reduction in severity of headaches and analgesic consumption, the presence of side effects owing to their effects on central glutamate receptors limited future exploration of their use in migraine. Regarding migraine prophylaxis, the efficacy of memantine has been investigated in retrospective (in 2007, [65]), open-label (in 2008, [75]), and randomized double-blinded placebo-controlled studies (in 2016, [76]). The participants in the three mentioned studies were individuals with migraine with and without aura as well as individuals with refractory migraine who received peroral doses of memantine 5 to 20 mg/day (Table 2). Together, these studies reported a memantine-induced significant reduction in (1) the monthly headache frequency, (2) the mean number of days with severe pain, and (3) the mean disability score.

For the treatment of migraine with severe and prolonged aura, ketamine was studied in a small open-label study (n = 11) (in 2000, [71]) and a randomized double-blinded placebocontrolled trial (in 2013 [72] and 2018 [74]). The intranasal administration of ketamine (25 mg) reduced the severity, and to a lesser extent, the duration of aura [71,72]. However, intravenous administration of a low-dose (0.2 mg/kg) of ketamine for the treatment of acute migraine did not produce a significant reduction in pain or disability-related scores [74]. In 1995, Nicolodi and colleagues investigated the administration of subcutaneous ketamine hydrochloride in individuals with migraine and stated that there was noticeable pain relief in the ketamine group compared to the placebo group [70]. In 2016, Lauritsen and colleagues reported in a retrospective study that six out of six individuals with refractory migraine achieved the target pain relief endpoint with intravenous ketamine infusion [73] (Table 2 and Box 2). Collectively, it can be assumed that migraine attacks [16].

The most common side effects reported from the above-mentioned trials with ketamine and memantine are asthenia, dizziness, nausea, psychiatric symptoms such as hallucinations, rashes, cognitive dysfunctions, somnolence, and changes in cardiovascular and respiratory stability [65,70,73,75]. So far, the American Headache Society [78,79] and European Headache Federation [80] have not recommended ketamine or memantine for the treatment of migraine. Mg²⁺, on the contrary, takes place as a nutraceutical in the migraine prevention treatment [78,80,81].

Box 2. Clinical evidence.

- 1. Genome-wide association studies have shown significant polymorphisms in glutamate receptor and transporter genes that are associated with migraine [82].
- 2. Consumption of a single dose (150 mg/kg) of monosodium glutamate (MSG) caused headache, craniofacial sensitivity, and nausea in healthy participants [33].
- 3. Repeated MSG intake (150 mg/kg) in five daily sessions for one week reduced pressure pain thresholds and caused headaches in healthy participants compared to placebo [34].
- 4. Glutamate levels measured in plasma and cerebrospinal fluid were elevated both ictally and interictally in individuals with migraine compared to healthy controls [10], and migraine preventive therapies reduced plasma glutamate levels [32].
- 5. N-Methyl-D-Aspartate Receptor (NMDAR) antagonist drugs such as ketamine and memantine showed some efficacy in acute and prophylaxis migraine treatment [65,71,73,77].

6. Perspective and Conclusions

Glutamate and NMDAR are involved in the initiation of migraine aura and migraine pain. NMDAR subunits expressed in the trigeminal afferents are involved in trigeminal nociception. NMDAR antagonists such as memantine and ketamine might be applicable for individuals with migraine with aura [62] despite some studies showing low efficacy, which can be explained by their central mechanism of action. As migraine is increasingly being recognized as a heterogeneous disorder, available NMDAR antagonists might have beneficial effects in a subgroup of individuals with migraine, including in individuals with difficult-to-treat migraine. So far, ketamine or memantine have not been recommended in the treatment of migraine [78–80]. Mg^{2+} (NMDAR blocker), on the contrary, takes place as a nutraceutical in migraine prevention treatment [78,80,81]. More selective antagonists

targeting peripheral NMDAR are needed to explore their beneficial effects in the treatment of migraine.

Funding: This research was funded by Rigshospitalet, Grant number E-23327-04.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The study did not report any data.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

AMPAR	Alpha-amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid receptor
CSD	Cortical spreading depression
KAR	Kainate receptor
LTP	Long-term potentiation
LTD	Long-term depression
NMDAR	N-Methyl-D-Aspartate Receptor
NTD	N-terminus domain
TG	Trigeminal ganglion
TVS	Trigeminovascular system
TMD	Transmembrane domain
TNC	Trigeminal nucleus caudalis

References

- Ashina, M.; Katsarava, Z.; Do, T.P.; Buse, D.C.; Pozo-Rosich, P.; Özge, A.; Krymchantowski, A.V.; Lebedeva, E.R.; Ravishankar, K.; Yu, S.; et al. Migraine: Epidemiology and systems of care. *Lancet* 2021, 397, 1485–1495. [CrossRef]
- 2. Ashina, M. Migraine. N. Engl. J. Med. 2020, 383, 1866–1876. [CrossRef]
- Eikermann-Haerter, K.; Ayata, C. Cortical spreading depression and migraine. *Curr. Neurol. Neurosci. Rep.* 2010, 10, 167–173. [CrossRef]
- Holland, P.R.; Akerman, S.; Goadsby, P.J. Cortical spreading depression-associated cerebral blood flow changes induced by mechanical stimulation are modulated by AMPA and GABA receptors. *Cephalalgia* 2010, 30, 519–527. [CrossRef]
- 5. Pietrobon, D. Ion channels in Migraine disorders. Curr. Opin. Physiol. 2018, 2, 98–108. [CrossRef]
- 6. Olesen, J.; Friberg, L.; Olsen, T.S.; Iversen, H.K.; Lassen, N.A.; Andersen, A.R.; Karle, A. Timing and topography of cerebral blood flow, aura, and headache during migraine attacks. *Ann. Neurol.* **1990**, *28*, 791–798. [CrossRef]
- Olesen, J.; Larsen, B.; Lauritzen, M. Focal hyperemia followed by spreading oligemia and impaired activation of rCBF in classic migraine. *Ann. Neurol.* 1981, 9, 344–352. [CrossRef]
- Lee, M.J.; Al-Karagholi, M.A.; Reuter, U. New migraine prophylactic drugs: Current evidence and practical suggestions for non-responders to prior therapy. *Cephalalgia* 2023, 43, 3331024221146315. [CrossRef]
- 9. Vikelis, M.; Mitsikostas, D.D. The role of glutamate and its receptors in migraine. *CNS Neurol. Disord. Drug Targets* 2007, 6, 251–257. [CrossRef]
- Ferrari, M.D.; Odink, J.; Bos, K.D.; Malessy, M.; Bruyn, G.W. Neuroexcitatory plasma amino acids are elevated in migraine. *Neurology* 1990, 40, 1582–1586. [CrossRef]
- 11. Lauritzen, M. Spreading depression and migraine. Pathol. Biol. 1992, 40, 332–337. [PubMed]
- 12. Tallaksen-Greene, S.J.; Young, A.B.; Penney, J.B.; Beitz, A.J. Excitatory amino acid binding sites in the trigeminal principal sensory and spinal trigeminal nuclei of the rat. *Neurosci. Lett.* **1992**, *141*, 79–83. [CrossRef] [PubMed]
- 13. Alam, Z.; Coombes, N.; Waring, R.H.; Williams, A.C.; Steventon, G.B. Plasma levels of neuroexcitatory amino acids in patients with migraine or tension headache. *J. Neurol. Sci.* **1998**, *156*, 102–106. [CrossRef] [PubMed]
- Kalia, L.V.; Kalia, S.K.; Salter, M.W. NMDA receptors in clinical neurology: Excitatory times ahead. *Lancet Neurol.* 2008, 7, 742–755. [CrossRef]
- Waung, M.W.; Akerman, S.; Wakefield, M.; Keywood, C.; Goadsby, P.J. Metabotropic glutamate receptor 5: A target for migraine therapy. Ann. Clin. Transl. Neurol. 2016, 3, 560–571. [CrossRef]
- 16. Chan, K.; MaassenVanDenBrink, A. Glutamate receptor antagonists in the management of migraine. *Drugs* **2014**, *74*, 1165–1176. [CrossRef]
- 17. Mayberg, M.; Langer, R.S.; Zervas, N.T.; Moskowitz, M.A. Perivascular meningeal projections from cat trigeminal ganglia: Possible pathway for vascular headaches in man. *Science* **1981**, *213*, 228–230. [CrossRef]

- 18. Akerman, S.; Holland, P.R.; Goadsby, P.J. Diencephalic and brainstem mechanisms in migraine. *Nat. Rev. Neurosci.* 2011, 12, 570–584. [CrossRef]
- 19. Schulte, L.H.; Peng, K.P. Current understanding of premonitory networks in migraine: A window to attack generation. *Cephalalgia* **2019**, *39*, 1720–1727. [CrossRef]
- 20. Hutchings, C.J.; Colussi, P.; Clark, T.G. Ion channels as therapeutic antibody targets. MAbs 2019, 11, 265–296. [CrossRef]
- 21. Wickenden, A.; Priest, B.; Erdemli, G. Ion channel drug discovery: Challenges and future directions. *Future Med. Chem.* **2012**, *4*, 661–679. [CrossRef] [PubMed]
- 22. Yan, J.; Dussor, G. Ion channels and migraine. *Headache* 2014, 54, 619–639. [CrossRef] [PubMed]
- 23. Pietrobon, D. Calcium channels and migraine. Biochim. Biophys. Acta 2013, 1828, 1655–1665. [CrossRef] [PubMed]
- 24. Ferrari, M.D. Migraine. Lancet 1998, 351, 1043–1051. [CrossRef]
- 25. Fabricius, M.; Jensen, L.H.; Lauritzen, M. Microdialysis of interstitial amino acids during spreading depression and anoxic depolarization in rat neocortex. *Brain Res.* **1993**, *612*, 61–69. [CrossRef]
- Molchanova, S.; Kööbi, P.; Oja, S.S.; Saransaari, P. Interstitial concentrations of amino acids in the rat striatum during global forebrain ischemia and potassium-evoked spreading depression. *Neurochem. Res.* 2004, 29, 1519–1527. [CrossRef]
- 27. Lauritzen, M.; Hansen, A.J. The effect of glutamate receptor blockade on anoxic depolarization and cortical spreading depression. *J. Cereb. Blood Flow Metab.* **1992**, *12*, 223–229. [CrossRef]
- 28. Ayata, C.; Lauritzen, M. Spreading Depression, Spreading Depolarizations, and the Cerebral Vasculature. *Physiol. Rev.* 2015, 95, 953–993. [CrossRef]
- Marrannes, R.; Willems, R.; De Prins, E.; Wauquier, A. Evidence for a role of the N-methyl-D-aspartate (NMDA) receptor in cortical spreading depression in the rat. *Brain Res.* 1988, 457, 226–240. [CrossRef]
- 30. D'Andrea, G.; Cananzi, A.R.; Joseph, R.; Morra, M.; Zamberlan, F.; Milone, F.F.; Grunfeld, S.; Welch, K. Platelet glycine, glutamate and aspartate in primary headache. *Cephalalgia* **1991**, *11*, 197–200. [CrossRef]
- Cananzi, A.R.; D'Andrea, G.; Perini, F.; Zamberlan, F.; Welch, K. Platelet and plasma levels of glutamate and glutamine in migraine with and without aura. *Cephalalgia* 1995, 15, 132–135. [CrossRef] [PubMed]
- 32. Ferrari, A.; Spaccapelo, L.; Pinetti, D.; Tacchi, R.; Bertolini, A. Effective prophylactic treatments of migraine lower plasma glutamate levels. *Cephalalgia* **2009**, *29*, 423–429. [CrossRef]
- 33. Baad-Hansen, L.; Cairns, B.; Ernberg, M.; Svensson, P. Effect of systemic monosodium glutamate (MSG) on headache and pericranial muscle sensitivity. *Cephalalgia* 2010, *30*, 68–76. [CrossRef]
- Shimada, A.; Cairns, B.E.; Vad, N.; Ulriksen, K.; Pedersen, A.M.L.; Svensson, P.; Baad-Hansen, L. Headache and mechanical sensitization of human pericranial muscles after repeated intake of monosodium glutamate (MSG). J. Headache Pain 2013, 14, 2. [CrossRef]
- Paoletti, P.; Neyton, J. NMDA receptor subunits: Function and pharmacology. *Curr. Opin. Pharmacol.* 2007, 7, 39–47. [CrossRef]
 [PubMed]
- 36. Ye, R.; Kong, X.; Han, J.; Zhao, G. N-methyl-D-aspartate receptor antagonists for migraine: A potential therapeutic approach. *Med. Hypotheses* **2009**, *72*, 603–605. [CrossRef] [PubMed]
- 37. Furukawa, H.; Singh, S.K.; Mancusso, R.; Gouaux, E. Subunit arrangement and function in NMDA receptors. *Nature* 2005, 438, 185–192. [CrossRef]
- Hoffmann, J.; Storer, R.J.; Park, J.W.; Goadsby, P.J. N-Methyl-d-aspartate receptor open-channel blockers memantine and magnesium modulate nociceptive trigeminovascular neurotransmission in rats. *Eur. J. Neurosci.* 2019, 50, 2847–2859. [CrossRef]
- 39. Kleckner, N.W.; Dingledine, R. Requirement for glycine in activation of NMDA-receptors expressed in Xenopus oocytes. *Science* **1988**, 241, 835–837. [CrossRef]
- Ciabarra, A.M.; Sullivan, J.M.; Gahn, L.G.; Pecht, G.; Heinemann, S.; Sevarino, K. Cloning and characterization of chi-1: A developmentally regulated member of a novel class of the ionotropic glutamate receptor family. *J. Neurosci.* 1995, *15*, 6498–6508. [CrossRef]
- 41. Nishi, M.; Hinds, H.; Lu, H.P.; Kawata, M.; Hayashi, Y. Motoneuron-specific expression of NR3B, a novel NMDA-type glutamate receptor subunit that works in a dominant-negative manner. *J. Neurosci.* 2001, 21, Rc185. [CrossRef] [PubMed]
- 42. Matsuda, K.; Kamiya, Y.; Matsuda, S.; Yuzaki, M. Cloning and characterization of a novel NMDA receptor subunit NR3B: A dominant subunit that reduces calcium permeability. *Brain Res. Mol. Brain Res.* **2002**, 100, 43–52. [CrossRef] [PubMed]
- 43. Low, C.M.; Wee, K.S. New insights into the not-so-new NR3 subunits of N-methyl-D-aspartate receptor: Localization, structure, and function. *Mol. Pharmacol.* **2010**, *78*, 1–11. [CrossRef]
- 44. Bliss, T.V.; Collingridge, G.L. A synaptic model of memory: Long-term potentiation in the hippocampus. *Nature* **1993**, *361*, 31–39. [CrossRef] [PubMed]
- 45. Rajani, V.; Sengar, A.S.; Salter, M.W. Tripartite signalling by NMDA receptors. Mol. Brain 2020, 13, 23. [CrossRef]
- Sucher, N.J.; Akbarian, S.; Chi, C.L.; Leclerc, C.L.; Awobuluyi, M.; Deitcher, D.; Wu, M.K.; Yuan, J.P.; Jones, E.G.; A Lipton, S. Developmental and regional expression pattern of a novel NMDA receptor-like subunit (NMDAR-L) in the rodent brain. *J. Neurosci.* 1995, 15, 6509–6520. [CrossRef]
- 47. Storer, R.J.; Goadsby, P.J. Trigeminovascular nociceptive transmission involves N-methyl-D-aspartate and non-N-methyl-D-aspartate glutamate receptors. *Neuroscience* **1999**, *90*, 1371–1376. [CrossRef]

- Shin, H.E.; Han, S.J.; Lee, K.S.; Park, J.-W. Polymorphism of the Glutamate Transporter Protein EAAT2 and Migraine Transformation into Chronic Daily Headache. J. Clin. Neurol. 2011, 7, 143–147. [CrossRef]
- Dong, X.D.; Mann, M.K.; Kumar, U.; Svensson, P.; Arendt-Nielsen, L.; Hu, J.; Sessle, B.; Cairns, B. Sex-related differences in NMDA-evoked rat masseter muscle afferent discharge result from estrogen-mediated modulation of peripheral NMDA receptor activity. *Neuroscience* 2007, 146, 822–832. [CrossRef]
- 50. Lee, J.; Ro, J.Y. Differential regulation of glutamate receptors in trigeminal ganglia following masseter inflammation. *Neurosci. Lett.* **2007**, *421*, 91–95. [CrossRef]
- Ivanusic, J.J.; Beaini, D.; Hatch, R.J.; Staikopoulosl, V.; Sesslel, B.; Jenningsl, E.; Ivanusic, J.J.; Beaini, D.; Hatch, R.J.; Staikopoulos, V.; et al. Peripheral N-methyl-d-aspartate receptors contribute to mechanical hypersensitivity in a rat model of inflammatory temporomandibular joint pain. *Eur. J. Pain* 2011, *15*, 179–185. [CrossRef] [PubMed]
- Fernandez-Montoya, J.; Buendia, I.; Martin, Y.B.; Egea, J.; Negredo, P.; Avendaño, C. Sensory Input-Dependent Changes in Glutamatergic Neurotransmission- Related Genes and Proteins in the Adult Rat Trigeminal Ganglion. *Front. Mol. Neurosci.* 2016, 9, 132. [CrossRef] [PubMed]
- 53. O'Brien, M.; Cairns, B.E. Monosodium glutamate alters the response properties of rat trigeminovascular neurons through activation of peripheral NMDA receptors. *Neuroscience* **2016**, *334*, 236–244. [CrossRef]
- Guerrero-Toro, C.; Koroleva, K.; Ermakova, E.; Gafurov, O.; Abushik, P.; Tavi, P.; Sitdikova, G.; Giniatullin, R. Testing the Role of Glutamate NMDA Receptors in Peripheral Trigeminal Nociception Implicated in Migraine Pain. *Int. J. Mol. Sci.* 2022, 23, 1529. [CrossRef] [PubMed]
- 55. Wang, X.M.; Mokha, S.S. Opioids modulate N-methyl-D-aspartic acid (NMDA)-evoked responses of trigeminothalamic neurons. *J. Neurophysiol.* **1996**, *76*, 2093–2096. [CrossRef] [PubMed]
- 56. Xiao, Y.; Richter, J.A.; Hurley, J.H. Release of glutamate and CGRP from trigeminal ganglion neurons: Role of calcium channels and 5-HT1 receptor signaling. *Mol. Pain* **2008**, *4*, 12. [CrossRef]
- 57. Mészáros, M.; Phan, T.H.M.; Vigh, J.P.; Porkoláb, G.; Kocsis, A.; Páli, E.K.; Polgár, T.F.; Walter, F.R.; Bolognin, S.; Schwamborn, J.C.; et al. Targeting Human Endothelial Cells with Glutathione and Alanine Increases the Crossing of a Polypeptide Nanocarrier through a Blood-Brain Barrier Model and Entry to Human Brain Organoids. *Cells* 2023, 12, 503. [CrossRef]
- Paoletti, P.; Ascher, P.; Neyton, J. High-affinity zinc inhibition of NMDA NR1-NR2A receptors. J. Neurosci. 1997, 17, 5711–5725. [CrossRef]
- 59. Faerber, L.; Drechsler, S.; Ladenburger, S.; Gschaidmeier, H.; Fischer, W. The neuronal 5-HT3 receptor network after 20 years of research—Evolving concepts in management of pain and inflammation. *Eur. J. Pharmacol.* 2007, 560, 1–8. [CrossRef]
- 60. Shatillo, A.; Salo, R.A.; Giniatullin, R.; Gröhn, O.H. Involvement of NMDA receptor subtypes in cortical spreading depression in rats assessed by fMRI. *Neuropharmacology* **2015**, *93*, 164–170. [CrossRef]
- 61. Lauritzen, M. Pathophysiology of the migraine aura. The spreading depression theory. *Brain* **1994**, *117 Pt 1*, 199–210. [CrossRef] [PubMed]
- 62. Peeters, M.; Gunthorpe, M.J.; Strijbos, P.J.; Goldsmith, P.; Upton, N.; James, M.F. Effects of pan- and subtype-selective N-methyl-D-aspartate receptor antagonists on cortical spreading depression in the rat: Therapeutic potential for migraine. *J. Pharmacol. Exp. Ther.* **2007**, *321*, 564–572. [CrossRef] [PubMed]
- 63. Faria, L.C.; Mody, I. Protective effect of ifenprodil against spreading depression in the mouse entorhinal cortex. *J. Neurophysiol.* **2004**, 92, 2610–2614. [CrossRef] [PubMed]
- 64. Benbow, T.; Teja, F.; Sheikhi, A.; Exposto, F.G.; Svensson, P.; Cairns, B.E. Peripheral N-methyl-D-aspartate receptor activation contributes to monosodium glutamate-induced headache but not nausea behaviours in rats. *Sci. Rep.* **2022**, *12*, 13894. [CrossRef] [PubMed]
- 65. Charles, A.; Flippen, C.; Romero Reyes, M.; Brennan, K.C. Memantine for prevention of migraine: A retrospective study of 60 cases. *J. Headache Pain* 2007, *8*, 248–250. [CrossRef]
- Dolati, S.; Rikhtegar, R.; Mehdizadeh, A.; Yousefi, M. The Role of Magnesium in Pathophysiology and Migraine Treatment. *Biol. Trace Elem. Res.* 2020, 196, 375–383. [CrossRef]
- 67. Baratloo, A.; Mirbaha, S.; Delavar Kasmaei, H.; Payandemehr, P.; Elmaraezy, A.; Negida, A. Intravenous caffeine citrate vs. magnesium sulfate for reducing pain in patients with acute migraine headache; a prospective quasi-experimental study. *Korean J. Pain* **2017**, *30*, 176–182. [CrossRef]
- 68. Taubert, K. Magnesium in migraine. Results of a multicenter pilot study. Fortschr. Med. 1994, 112, 328–330.
- 69. Shin, H.J.; Na, H.S.; Do, S.H. Magnesium and Pain. Nutrients 2020, 12, 2184. [CrossRef]
- 70. Nicolodi, M.; Sicuteri, F. Exploration of NMDA receptors in migraine: Therapeutic and theoretic implications. *Int. J. Clin. Pharmacol. Res.* **1995**, *15*, 181–189.
- 71. Kaube, H.; Herzog, J.; Kaufer, T.; Dichgans, M.; Diener, H.C. Aura in some patients with familial hemiplegic migraine can be stopped by intranasal ketamine. *Neurology* **2000**, *55*, 139–141. [CrossRef] [PubMed]
- Afridi, S.K.; Giffin, N.J.; Kaube, H.; Goadsby, P. A randomized controlled trial of intranasal ketamine in migraine with prolonged aura. *Neurology* 2013, 80, 642–647. [CrossRef] [PubMed]
- Lauritsen, C.; Mazuera, S.; Lipton, R.B.; Ashina, S. Intravenous ketamine for subacute treatment of refractory chronic migraine: A case series. J. Headache Pain 2016, 17, 106. [CrossRef] [PubMed]

- 74. Etchison, A.R.; Bos, L.; Ray, M.; McAllister, K.B.; Mohammed, M.; Park, B.; Phan, A.V.; Heitz, C. Low-dose Ketamine Does Not Improve Migraine in the Emergency Department: A Randomized Placebo-controlled Trial. West J. Emerg. Med. 2018, 19, 952–960. [CrossRef]
- 75. Bigal, M.; Rapoport, A.; Sheftell, F.; Tepper, D.; Tepper, S. Memantine in the preventive treatment of refractory migraine. *Headache* **2008**, *48*, 1337–1342. [CrossRef] [PubMed]
- 76. Noruzzadeh, R.; Modabbernia, A.; Aghamollaii, V.; Ghaffarpour, M.; Harirchian, M.H.; Salahi, S.; Nikbakht, N.; Noruzi, N.; Tafakhori, A. Memantine for Prophylactic Treatment of Migraine Without Aura: A Randomized Double-Blind Placebo-Controlled Study. *Headache* 2016, 56, 95–103. [CrossRef]
- 77. Gomez-Mancilla, B.; Brand, R.; Jurgens, T.P.; Göbel, H.; Sommer, C.; Straube, A.; Evers, S.; Sommer, M.; Campos, V.; O Kalkman, H.; et al. Randomized, multicenter trial to assess the efficacy, safety and tolerability of a single dose of a novel AMPA receptor antagonist BGG492 for the treatment of acute migraine attacks. *Cephalalgia* 2014, 34, 103–113. [CrossRef]
- 78. Oskoui, M.; Pringsheim, T.; Holler-Managan, Y.; Potrebic, S.; Billinghurst, L.; Gloss, D.; Hershey, A.D.; Licking, N.; Sowell, M.; Victorio, M.C.; et al. Practice guideline update summary: Acute treatment of migraine in children and adolescents: Report of the Guideline Development, Dissemination, and Implementation Subcommittee of the American Academy of Neurology and the American Headache Society. *Neurology* 2019, *93*, 487–499. [CrossRef]
- 79. Ailani, J.; Burch, R.C.; Robbins, M.S. The American Headache Society Consensus Statement: Update on integrating new migraine treatments into clinical practice. *Headache J. Head Face Pain* **2021**, *61*, 1021–1039. [CrossRef]
- 80. Antonaci, F.; Dumitrache, C.; De Cillis, I.; Allena, M. A review of current European treatment guidelines for migraine. *J. Headache Pain* **2010**, *11*, 13–19. [CrossRef]
- 81. AHS-First-Contact-PreventativeTreatment; American Headache Society: Mount Royal, NJ, USA, 2021.
- Hautakangas, H.; Winsvold, B.S.; Ruotsalainen, S.E.; Bjornsdottir, G.; Harder, A.V.E.; Kogelman, L.J.A.; Thomas, L.F.; Noordam, R.; Benner, C.; Gormley, P.; et al. Genome-wide analysis of 102,084 migraine cases identifies 123 risk loci and subtype-specific risk alleles. *Nat. Genet.* 2022, 54, 152–160. [CrossRef] [PubMed]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.