



Supplementary Materials

Ezrin Regulates the Cell Surface Localization of PD-L1 in HEC-151 Cells

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Original source images for immunoblots.

PD-L1

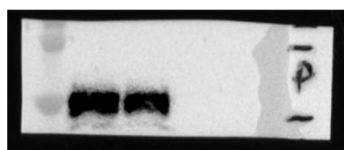


Figure 1c

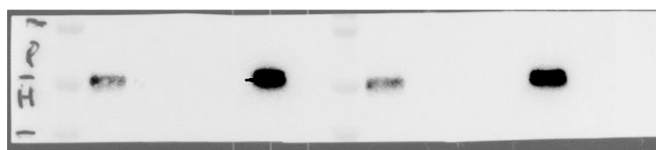


Figure 4

Ezrin

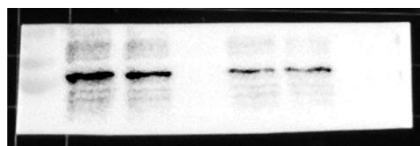


Figure 1c

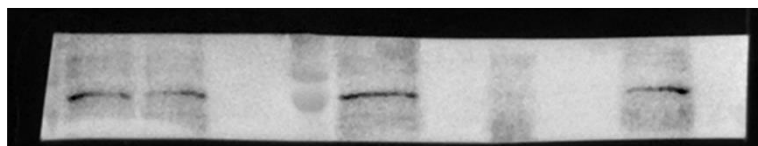


Figure 4

Radixin

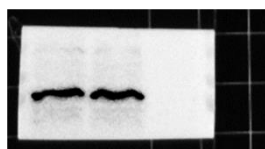


Figure 1c

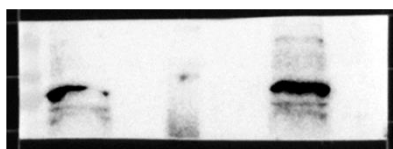


Figure 4

Moesin



Figure 1c

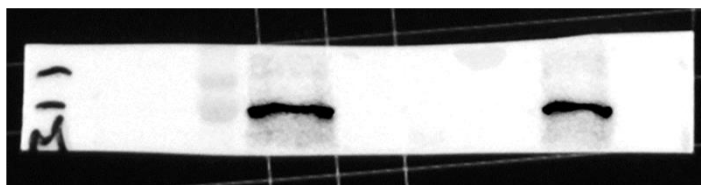


Figure 4

GAPDH

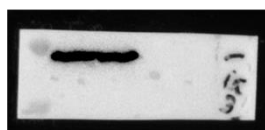


Figure 1c
(PD-L1)



Figure 1c
(Ezrin)

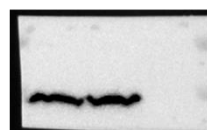


Figure 1c
(Radixin)

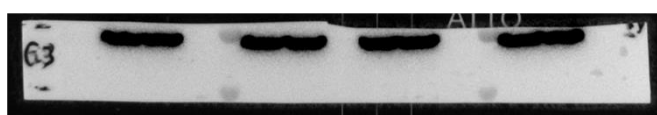


Figure 1c
(Moesin)

Figure S1. Original source images for immunoblots. The original western blotting membrane to detect the protein expression of programmed death ligand-1 (PD-L1), ezrin, radixin, and moesin as well as the corresponding glyceraldehyde-3-phosphate dehydrogenase (GAPDH) used as a loading control shown in Figure 1c or Figure 4. Blots are labeled according to the corresponding Figure panel within the main manuscript.

RNA interference-mediated knockdown of target genes in HEC-151 cells.

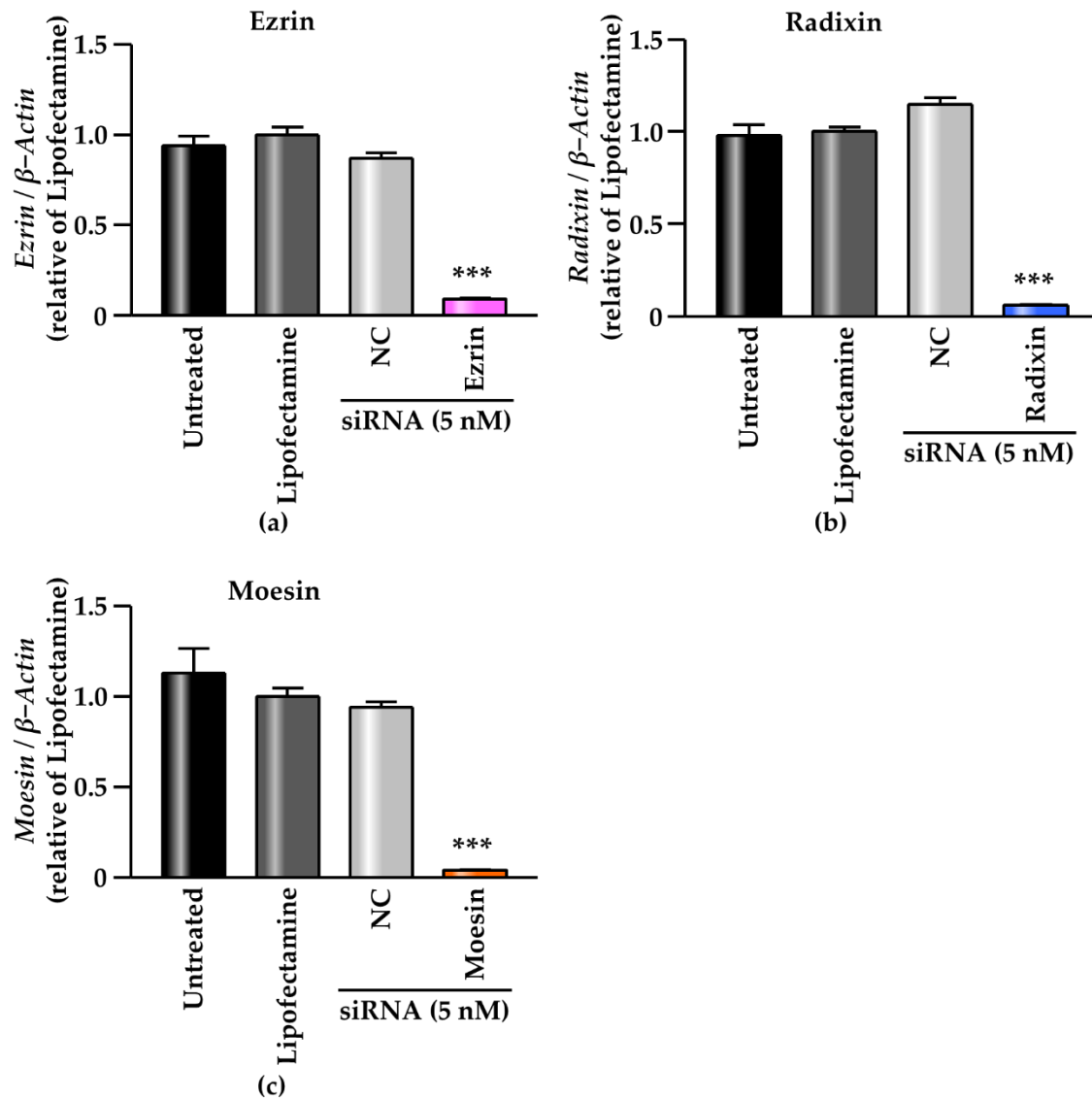


Figure S2. RNA interference-mediated knockdown of target genes in HEC-151 cells. Cells were treated with the transfection medium (Untreated), transfection reagent (Lipofectamine), nontargeting control (NC) siRNA, and specific siRNAs for ezrin, radixin, or moesin at the concentrations of 5 nM, and then cultured for 4 days. Each column shows the mRNA expression levels of (a) ezrin, (b) radixin, and (c) moesin normalized with β -Actin in cells from each treatment group relative to that in cells treated with Lipofectamine alone as determined by real-time reverse transcription-polymerase chain reaction. $n = 3$, *** $p < 0.001$ vs. Lipofectamine. All data were expressed as the mean \pm SEM and analyzed by one-way ANOVA followed by Dunnett's test.

Table S1. List of primers used in the present study.

Gene	Primer sequence (5'→3')
<i>h-β-Actin</i> (forward)	TGGCACCCAGCACAATGAA
<i>h-β-Actin</i> (reverse)	CTAAGTCATAGTCCGCCTAGAAGCA
<i>h-Ezrin</i> (forward)	ACCATGGATGCAGAGCTGGAG
<i>h-Ezrin</i> (reverse)	CATAGTGGAGGCCAAAGTACCACA
<i>h-Radixin</i> (forward)	GAATTTGCCATTTCAGCCCAATA
<i>h-Radixin</i> (reverse)	GCCATGTAGAATAACCTTTGCTGTC
<i>h-Moesin</i> (forward)	CCGAATCCAAGCCGTGTGTA
<i>h-Moesin</i> (reverse)	GGCAAACCTCCAGCTCTGCATC
<i>h-PD-L1</i> (forward)	CAATGTGACCAGCACACTGAGAA
<i>h-PD-L1</i> (reverse)	GGCATAATAAGATGGCTCCCAGAA

Table S2. List of antibodies used in the present study.

Antibodies	Source	Cat#	Dilution
rabbit anti-ezrin	Cell Signaling Technology	3145	1:2,000 (WB) 1:50 (IF)
rabbit anti-radixin	Gene Tex	GTX105408	1:2,000 (WB) 1:100 (IF)
rabbit anti-moesin	Cell Signaling Technology	3150	1:2,000 (WB) 1:50 (IF)
Alexa Fluor 488-conjugated rabbit anti-PD-L1	Cell Signaling Technology	25048	1:50 (IF)
Alexa Fluor 488-conjugated goat anti-rabbit IgG	Thermo Fisher Scientific	R37116	1:25 (IF)
Alexa Fluor 594-conjugated goat anti-rabbit IgG	Thermo Fisher Scientific	R37117	1:25 (IF)
HRP-conjugated rabbit anti-PD-L1	Cell Signaling Technology	51296s	1:1,000 (WB)
mouse anti-GAPDH	Merck	MAB374	1:20,000 (WB)
HRP-conjugated anti-rabbit IgG (heavy + light)	SeraCare Life Sciences	5220-0336	1:10,000 (WB)
HRP-conjugated anti-mouse IgG (heavy + light)	SeraCare Life Sciences	5220-0341	1:10,000 (WB)
rabbit anti-PD-L1	Cell Signaling Technology	13684	1:30 (IP)
rabbit IgG isotype control	Cell Signaling Technology	3900	1:30 (IP)
APC-conjugated mouse anti-human PD-L1	BioLegend	329708	4.0 µg/test (FC)

western blotting; WB, immunofluorescence; IF, immunoprecipitation; IP, flow cytometry; FC, horseradish peroxidase; HRP, allophycocyanin; APC