

## Supplementary Table S1

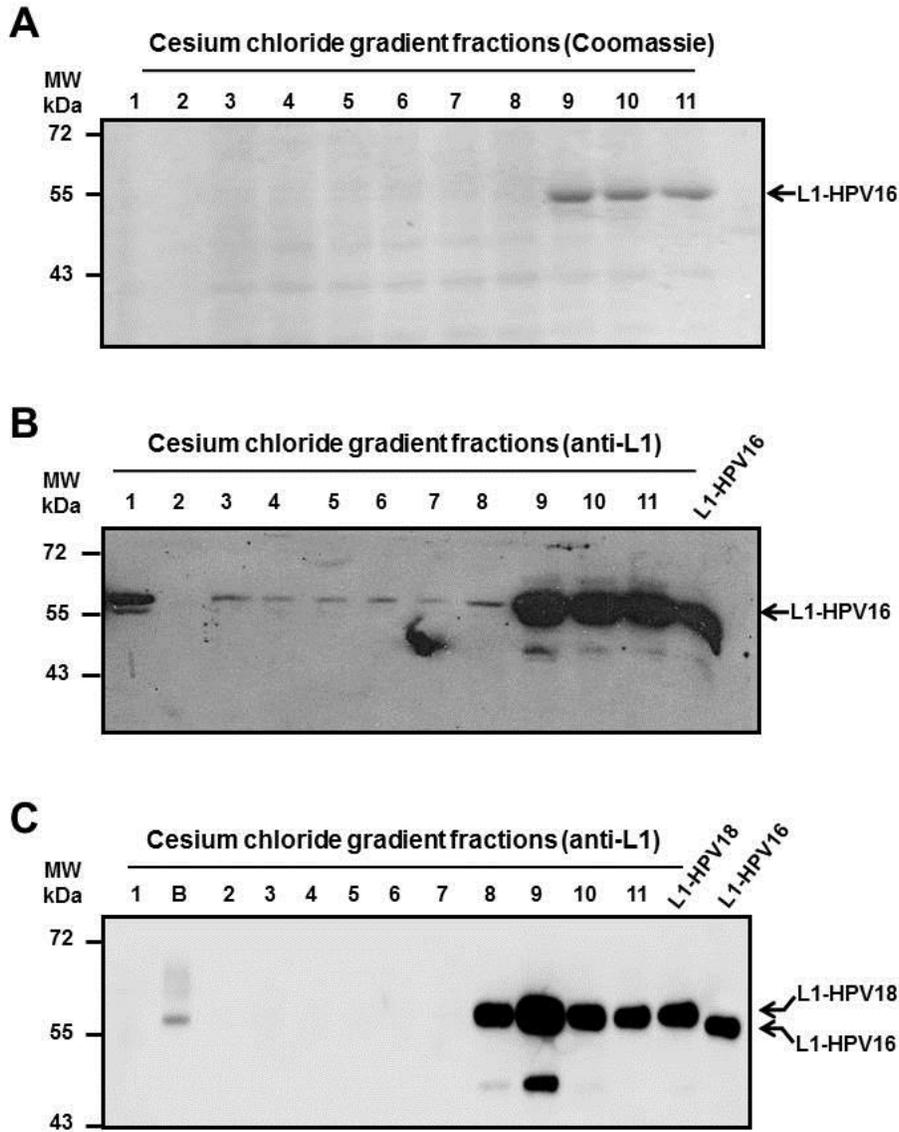
### Validation of ELISA system to detect anti-VLPs and anti-L1 antibodies from HPV16/18.

Population group	VLPs Seropositivity		L1 Seropositivity	
	HPV16	HPV18	HPV16	HPV18
<i>Vaccinated vs Control</i> <sup>1</sup> (n= 36/50)	n= 41	n= 22	n= 33	n= 28
<i>Sensitivity</i>	100.0	61.1	88.9	69.4
<i>Specificity</i>	100.0	98.0	100.0	100.0
<i>Accuracy</i>	100.0	82.6	95.4	87.2
<i>AUC</i> <sup>2</sup>	<b>1.00</b>	<b>0.79</b>	<b>0.94</b>	<b>0.84</b>

<sup>1</sup> Control, this is the group of young girls non-sexually active, and presumably HPV-naïve.

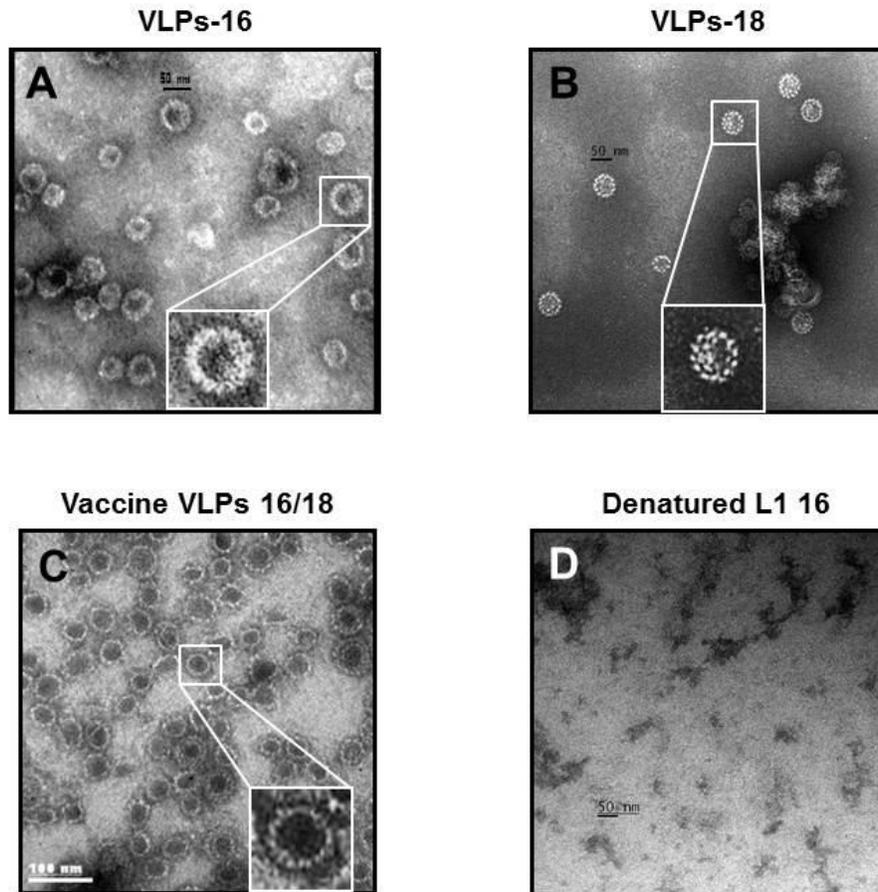
<sup>2</sup> AUC, Area under the ROC curve.

Supplementary Figure S1



**Figure S1. Identification of HPV16-L1 and HPV18-L1 proteins from CsCl gradient fractions.** (A) Fractions from CsCl gradient of High Five cells infected with BACU-L1-16 were separated on a 10 % acrylamide gel and Coomassie blue stained to visualize protein purity. The L1 proteins were visualized by Western blot in the different gradient fractions using an anti-L1 mouse monoclonal antibody (dil. 1:2000) (DAKO) that recognized the L1 proteins from HPV16 (B) and HPV18 (C). The specific antibody complex was developed by chemiluminescence and membranes exposed to X-OMAT film. Arrows show the specific L1 proteins.

Supplementary Figure S2



**Figure S2.** Electron micrographs of VLPs from HPV16/18 produced in baculovirus. Purified CsCl gradient VLPs 16/18 were prepared for TEM as described under Material and Methods and photographed at a magnification of  $\times 25,000$  and  $\times 14,000$ . (A) Purified VLPs from HPV16; (B) VLPs from HPV18; (C) VLPs-16/18 from HPV vaccine were used as a positive control. (D) Denatured L1 was obtained (monomers and aggregates) by treatment of VLPs with carbonate buffer (pH 9.6) for 16 h at 40 C. Scale bars: A, B, and D, 50 nm; C, 100 nm.